

# Rab27: Molecular switch of tumor exosome secretion (Review)

MENGTIAN TANG<sup>1\*</sup>, ZHONGJIAN ZHU<sup>1\*</sup>, HUAXIN DUAN<sup>1</sup>, SHIMING TAN<sup>2</sup>, MENGZHOU SHEN<sup>1</sup>,  
XIANJIE JIANG<sup>2</sup>, QIU PENG<sup>2</sup>, LINDA OYANG<sup>2</sup>, ZONGYAO REN<sup>2</sup>, YUJUAN ZHOU<sup>2,3</sup> and QIANJIN LIAO<sup>1,3</sup>

<sup>1</sup>Department of Oncology, Hunan Provincial People's Hospital and The First Affiliated Hospital of Hunan Normal University, Hunan Normal University Health Science Center, Changsha, Hunan 410005, P.R. China; <sup>2</sup>The Affiliated Cancer Hospital of Xiangya School of Medicine, Central South University/Hunan Cancer Hospital, Hunan Key Laboratory of Cancer Metabolism, Changsha, Hunan 410013, P.R. China; <sup>3</sup>Hunan Engineering Research Center of Tumor Organoid Technology and Applications, Public Service Platform of Tumor Organoids Technology, Changsha, Hunan 410013, P.R. China

Received February 10, 2026; Accepted April 24, 2026

DOI: 10.3892/ijmm.2026.5853

**Abstract.** Rab proteins are core regulators of vesicle trafficking during membrane transport. As key members of this family, Rab27 comprises two functionally distinct isoforms, Rab27a and Rab27b, and participates in the regulation of exosome secretion by mediating the selective anchoring of multivesicular bodies to the plasma membrane. In tumor cells, Rab27 promotes malignant progression by modulating exosome secretion. Aberrantly expressed Rab27 is associated with the prognosis of patients with cancer, suggesting its potential as a prognostic biomarker for various malignant tumors. Furthermore, the development of novel small-molecule drugs targeting Rab27 is promising because these drugs can exert antitumor effects through interference with the exosome secretory pathway. The present review systematically summarizes the molecular mechanisms underlying the Rab27-mediated regulation of exosome secretion and its role in tumorigenesis and cancer progression and discusses the application prospects and challenges of Rab27-targeted antitumor therapeutic strategies, aiming to provide a reference for basic research and clinical translation of Rab27.

## Contents

1. Introduction
2. Structural characteristics and localization of Rab27
3. Exosome secretion and biogenesis mechanisms
4. Role of Rab27-mediated exosome secretion in tumors
5. Diagnostic and therapeutic potential of Rab27 in tumors
6. Conclusions

## 1. Introduction

The Rab family consists of small G proteins within the Ras superfamily (1). They possess structural characteristics, including a GTPase-folded structure and a C-terminal anchored structure (1,2). The GTPase-folded structure generally consists of six  $\beta$ -folded sheets and five  $\alpha$ -helices, which form the essential structure for its biological function (3). The C-terminal anchoring structure refers to the process of attaching to the membrane through a lipid moiety (such as an isoprenoid group) that is covalently linked to two cysteines at the C-terminus. This anchoring mechanism creates a close connection between Rab family proteins and the structural composition of lipid membranes (4,5). Typically, Rab family proteins are bound to GDP and are in an inactivated state. The guanine nucleotide exchange factor (GEF) promotes the conversion of Rab proteins to the active form by replacing GDP with GTP in a dynamically reversible process (6-8). To date, ~60 Rab proteins have been identified in humans, and one of their main functions is to regulate vesicular transport (9,10).

Due to their biological stability and diverse cargo, exosomes facilitate intercellular communication, thereby remodeling the tumor microenvironment (TME) and driving disease progression (11-17). Furthermore, exosomes offer dual potential as diagnostic biomarkers for cancer and as drug delivery vehicles (18-20). While various Rab proteins isoforms perform specific regulatory functions at different stages of exosome biogenesis (21). Among these, Rab27 is recognized as a key molecular switch that directly governs exosome secretion and is widely implicated in the progression of diverse pathological conditions, including cancer, immune-related disorders

---

*Correspondence to:* Professor Qianjin Liao, Department of Oncology, Hunan Provincial People's Hospital and The First Affiliated Hospital of Hunan Normal University, Hunan Normal University Health Science Center, 61 Jiefang West Road, Changsha, Hunan 410005, P.R. China  
E-mail: march-on@126.com

Professor Yujuan Zhou, The Affiliated Cancer Hospital of Xiangya School of Medicine, Central South University/Hunan Cancer Hospital, Hunan Key Laboratory of Cancer Metabolism, 283 Tongzipo Road, Changsha, Hunan 410013, P.R. China  
E-mail: yujany\_zhou@163.com

\*Contributed equally

**Key words:** Rab27, exosomes, transport, secretion, biomarker, antitumor treatment

(for example, rheumatoid arthritis and inflammatory bowel disease), neurological conditions (for example, Parkinson's disease), metabolic and endocrine diseases (for example, diabetes) and cardiovascular diseases (22-26). Although the role of Rab27 in regulating exosome secretion has been reported, a systematic overview of how Rab27 contributes to tumorigenesis and cancer progression via exosome secretion, as well as its translational potential as a diagnostic biomarker and therapeutic target, is lacking. The present review systematically summarizes the molecular mechanisms underlying Rab27-mediated exosome secretion in tumors and highlights critical issues that must be addressed to achieve therapeutic benefits by targeting Rab27.

## 2. Structural characteristics and localization of Rab27

Rab27 belongs to the Rab protein family and exists in two isoforms in humans: Rab27a and Rab27b (27,28). Rab27a (221 aa) shares 61% identity with Rab27b (218 aa) at the nucleotide level within the open reading frame and 71% identity at the amino acid level (29). The differences are located mainly at the carboxy terminus (30). Structurally, both Rab27a and Rab27b contain four GTP binding sites (16-24 aa, 74-78 aa, 133-136 aa and 163-165 aa) (31). Moreover, they have a shared effector region (38-46 aa). The first structural distinction between Rab27a and Rab27b lies in the disordered regions (31). Compared with Rab27a, Rab27b has a longer disordered region, mainly at the carboxy terminus. Second, slight disparities in the post-translational modification (PTM) sites of Rab27a and Rab27b exist. Compared with Rab27b, Rab27a has more ubiquitination sites (31,32). Moreover, the expression patterns of Rab27a and Rab27b differ markedly across various cancer types. For example, in certain malignancies, the expression of Rab27a is upregulated, whereas that of Rab27b is downregulated, and in other malignancies, the opposite pattern is observed. This differential expression highlights the heterogeneity of the regulation of Rab27 expression in cancer (33) (Fig. 1 and Table I).

As previously reported, Rab family proteins can be anchored to membrane-structured suborganelles, and Rab27a and Rab27b are no exception (30,34-36). However, some subtle differences exist. Rab27a is distributed in a relatively large number of subcellular organelles (30). Specifically, Rab27a is localized to melanosomes, secretory granules, late endosomes and lysosomes (37,38), whereas Rab27b is considered to be localized to the membrane of Golgi stacks and vesicles located in the trans-Golgi network (TGN) area, secretory granules and late endosomes (30,39). The distinct subcellular localization of Rab27 contributes critically to exosome biogenesis and secretion by mediating specific interactions with a range of effector proteins (23) (Fig. 2).

## 3. Exosome secretion and biogenesis mechanisms

Small extracellular vesicles (EVs), including exosomes, microvesicles and small ectosomes, are generated through the outward growth of the plasma membrane (PM) (40-42). Currently, there is no clear demarcation between EVs and exosomes (43,44). In the present study, the term 'exosomes' was used to denote EVs with biological activity. Exosomes are

small vesicles with a diameter of 40-160 nanometers and are characterized by a bi-layered lipid membrane (45). They play a critical role as mediators of intercellular communication by transferring bioactive cargo. The proteins, nucleic acids, lipids, metabolites, and even organelles such as mitochondria within them are transported to recipient cells, thereby influencing their biological functions (46-52). The production of exosomes is linked to the formation of multivesicular bodies (MVB) within cells (53-55). Initially, the PM undergoes endocytosis, or 'inward budding', to form early-sorting endosomes (ESEs); the TGN and the endoplasmic reticulum also contribute to their biogenesis. ESEs subsequently mature within the cell into late-sorting endosomes and MVBs. These MVBs then fuse with the cell membrane, and the intraluminal vesicles (ILVs) are expelled from the cell through exocytosis; subsequently, the expelled ILVs become exosomes (56-58).

Exosome biogenesis primarily involves endosomal sorting complex required for transport (ESCRT)-dependent and ESCRT-independent mechanisms (57,59-61). ESCRTs consist of five complexes, including ESCRT-0, ESCRT-I, ESCRT-II, ESCRT-III and VPS4 (62,63). First, ubiquitinated cargo is recognized and bound by the ESCRT-0 complex. It is then transferred to the ESCRT-I and ESCRT-II complexes for sorting and processing, ensuring the selective incorporation of specific cargo into the ILV. The ESCRT-III complex subsequently mediates membrane curvature and scission of the endosomal membrane, leading to ILV formation (60,64-66). Additionally, VPS4 supplies energy for this process by hydrolyzing ATP (67).

Rab protein-mediated exosome secretion is among the important steps in the ESCRT-independent mechanism (21,68,69). For instance, Rab5 is involved in the formation and transport of early endosomes, whereas Rab7 is involved in the maturation of MVBs and serves as a critical regulator of the fusion of MVBs with lysosomes, playing an essential role in lysosome biogenesis (70-73). Rab proteins can interact with the cytoskeleton and mediate the transport of exosomes along the cytoskeleton (74,75). Rab proteins collaborate with other proteins involved in membrane fusion, such as soluble N-ethylmaleimide-sensitive factor attachment protein receptors (SNAREs) proteins (an essential family of proteins that mediate the fusion of MVBs with the PM), to promote the fusion of exosomal membranes with cell membranes (76,77). Moreover, Rab family proteins are associated with the cell membrane regions involved in exosome secretion (78,79). Studies have shown that Rab11 and Rab35 function in early or recycling endosomes (80-82). Rab11 can regulate calcium-dependent MVB-PM fusion, thereby promoting the secretion of exosomes (83). Interestingly, a recent study has shown that accessory ESCRT-III can regulate the formation of Rab11 and exosomes containing Rab11a through an ESCRT-dependent mechanism and does not rely on the ubiquitination/deubiquitination cycle for cargo loading (84). Rab35 and its GTPase-activating protein (GAP) can mediate the binding of endocytic vesicles to the PM, thereby regulating the secretion of exosomes (85). Research has indicated that Rab37 is involved in apical exosome secretion, whereas Rab39 is a specific regulator of basolateral exosome secretion (86).

Notably, the role of Rab27 in exosome secretion cannot be ignored. Rab27 can mediate the binding of MVBs to the

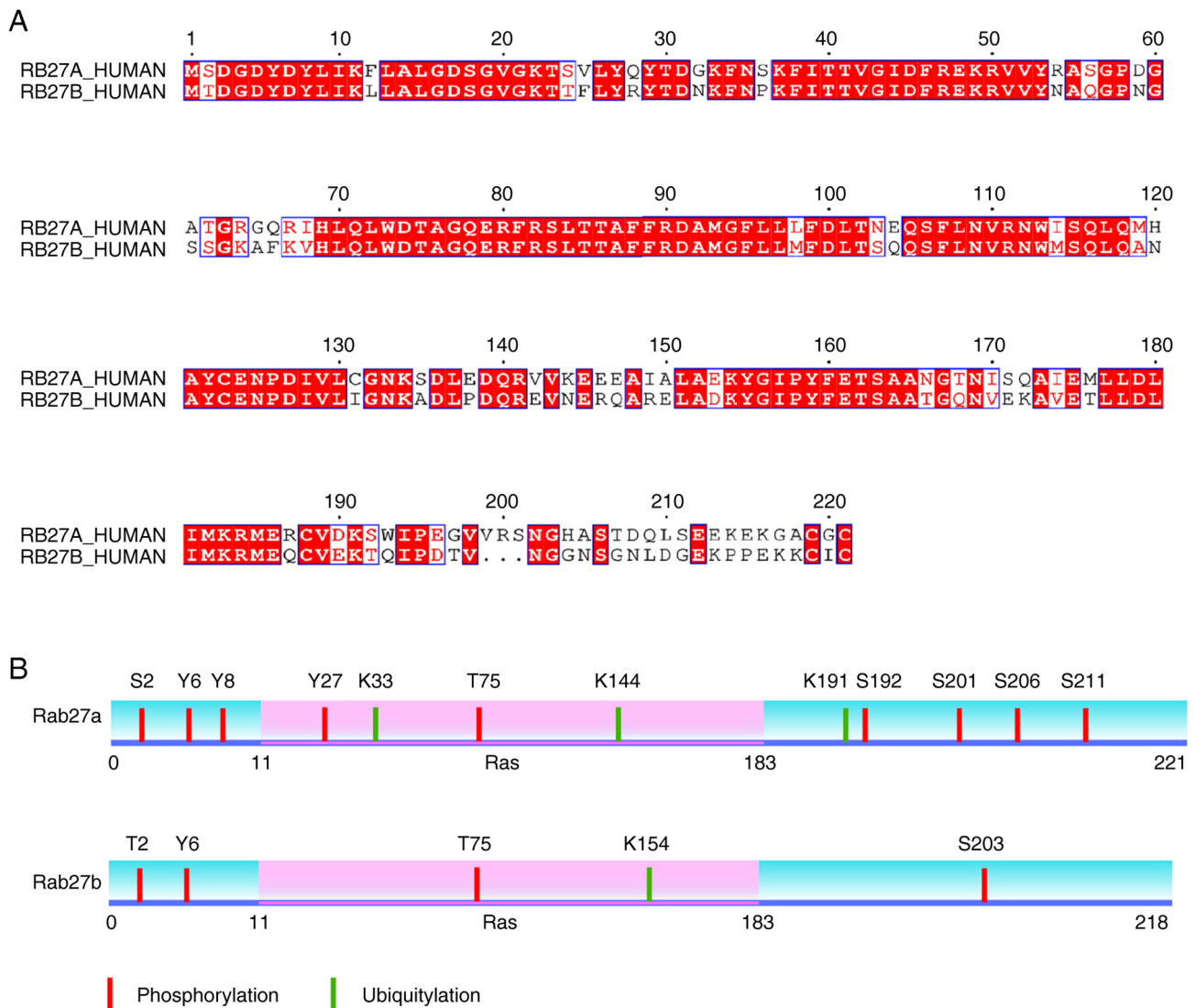


Figure 1. Structural characteristics of Rab27a and Rab27b. (A) Sequence alignment of the region between the Rab27a and Rab27b domains. Identical amino acids are highlighted with a red background and white font; functionally similar amino acids are indicated with a white background and red font; and divergent amino acids are shown in black font. (B) Post-translational modification sites of Rab27a and Rab27b.

PM and the secretion of exosomes (87,88). Research has demonstrated that Rab27a mediates the transport of MVBs to the PM (89). Moreover, Rab27a is localized to peripheral CD63-positive MVBs, preventing MVBs from fusing with one another or with other types of vesicles. This, in turn, promotes the fusion of MVBs with the PM (30). Rab27a can regulate the dynamics of cortical actin at the docking sites of MVBs by inhibiting the localization of coronin 1b (90). Rab27b is localized in the perinuclear region and mediates the transfer of MVBs from the TGN to the membrane. Additionally, Rab27b can promote the maintenance of MVBs at the cell periphery by regulating their transfer from microtubules to the actin-rich PM (57). The regulation of exosome secretion mediated by Rab is also intertwined with autophagy, resulting in the formation of a complex and coordinated regulatory network. Rab-dependent intracellular trafficking controls the sorting and transport of MVBs, which not only directs the transport and release of exosomes at the cell surface but also promotes the fusion of autophagosomes with MVBs through the recruitment of autophagy-related proteins (ATGs). This process

mediates the selective loading of exosomal cargoes, including inflammatory factors, microRNAs and ATGs (91-94).

Rab27 typically requires cooperation with other Rab proteins to perform its functions. For instance, in tumor cells, Rab5 mediates the sorting of PD-L1 into endocytic vesicles, whereas Rab27 facilitates the subsequent secretion of exosomal PD-L1. Their synergistic action reduces the surface expression of PD-L1, thereby enhancing the cytotoxic activity of T cells against tumors (87). Evidence has suggested that the cooperative interaction between Rab27a and Rab3a may represent a critical step in vesicle trafficking. Rab27a can bind to Rabphilin3A to form a complex, which mediates the transport of vesicles toward the PM while recruiting a GEF to catalyze the activation of Rab3a. Activated Rab3a then associates with Rabphilin3A, further recruiting the SNARE complex, which ultimately triggers the fusion of vesicles with the PM (95,96). Furthermore, Rab27 can cooperate with Rab35 to promote the formation of MVBs (97). Conversely, Rab27 also has antagonistic interactions with certain Rab proteins. In mast cells, Rab37, Munc13-4 and Rab27 can

Table I. Differential expression of Rab27a and Rab27b across various cancer types in the GEPIA database.

Cancer type	Number of tumor samples	Number of normal samples	Rab27a expression	Rab27b expression
ACC	77	128	Down	-
BLCA	404	28	-	-
BRCA	1085	291	-	Up
CESC	306	13	-	Up
CHOL	36	9	-	-
COAD	275	349	-	-
DLBC	47	337	-	-
ESCA	182	286	-	-
GBM	163	207	Up	Down
HNSC	519	44	-	-
KICH	66	53	-	Up
KIRC	523	100	-	-
KIRP	286	60	-	-
LAML	173	70	-	-
LGG	518	207	-	-
LIHC	369	160	-	-
LUAD	483	347	-	-
LUSC	486	338	Down	-
MESO	87	-	-	-
OV	426	88	-	-
PAAD	179	171	Up	Up
PCPG	182	3	-	-
PRAD	492	152	-	-
READ	92	318	-	-
SARC	262	2	-	-
SKCM	460/461	558	Up	Down
STAD	408	211	-	-
TGCT	137	165	-	Down
THCA	512	337	Up	-
THYM	118	339	Down	-
UCEC	174	91	-	-
UCS	57	78	Down	-
UVM	79	-	-	-

ACC, adrenocortical carcinoma; BLCA, bladder cancer; BRCA, breast cancer; CESC, cervical squamous cell carcinoma and endocervical adenocarcinoma; CHOL, cholangiocarcinoma; COAD, colon adenocarcinoma; DLBC, diffuse large B-cell lymphoma; ESCA, esophageal adenocarcinoma; GBM, glioblastoma; HNSC, head and neck squamous cell carcinoma; KICH, kidney chromophobe; KIRC, kidney renal clear cell carcinoma; KIRP, kidney renal papillary cell carcinoma; LAML, acute myeloid leukemia; LGG, low-grade gliomas; LIHC, liver hepatocellular carcinoma; LUAD, lung adenocarcinoma; LUSC, lung squamous cell carcinoma; MESO, mesothelioma; OV, ovarian carcinoma; PAAD, pancreatic adenocarcinoma; PCPG, pheochromocytoma and paraganglioma; PRAD, prostate adenocarcinoma; READ, rectum adenocarcinoma; SARC, sarcoma; SKCM, skin cutaneous melanoma; STAD, stomach adenocarcinoma; TGCT, testicular germ cell tumors; THCA, thyroid carcinoma; THYM, thymoma; UCEC, uterine corpus endometrial carcinoma; UCS, uterine carcinosarcoma; UVM, uveal melanoma.

form a complex, counteracting the vesicle-priming activity of Rab27-Munc13-4 (98). In the context of Sjögren's syndrome, an antagonistic relationship between Rab3D and Rab27 has been identified, in which the relative activities of these two proteins in secretory vesicles correlate with the enzymatic activity of secretory vesicle cathepsin S in the tears of patients (99). In addition, the abundance of ceramides, a class of lipid signaling molecules, is positively correlated with exosome biogenesis. Proteins in the tetraspanin family, which form a complex with

syntenin and participate in the sorting of exosomal cargo and modulate membrane curvature, are both critically involved in the exosome secretion process (100-104) (Fig. 3).

In the TME, exosomes serve as critical mediators of inter-cellular communication between tumor cells and non-tumor cells (105,106). They not only reshape the extracellular matrix, tumor vasculature and immune microenvironment but also modulate the physical and chemical properties of the TME and induce the malignant differentiation of nontumor cells (such as

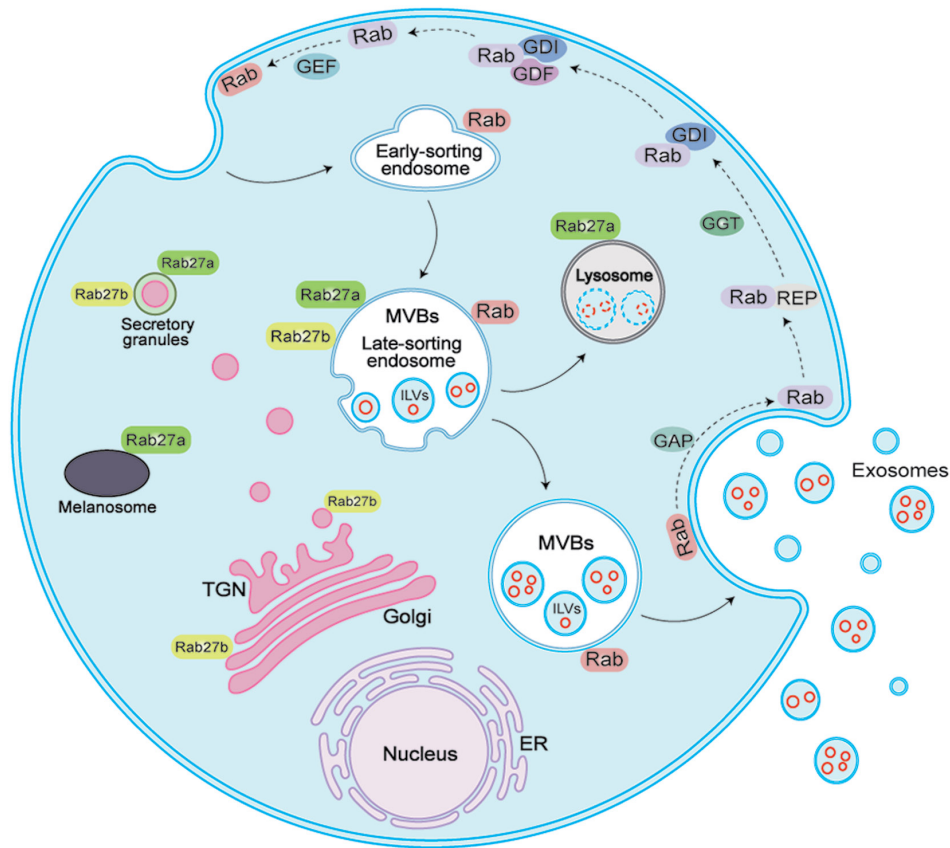


Figure 2. Rab switch form and subcellular localization of Rab27. Rab proteins exist in two reversible states: The inactive GDP-bound form and the active GTP-bound form. In the inactive state, Rab is bound to GDP. GEFs catalyze the exchange of GDP with GTP, thereby activating Rab. The GTP-bound active form of Rab plays a crucial role in vesicular transport. Subsequently, GAPs promote the hydrolysis of GTP to GDP, returning Rab to its inactive state. Inactive GDP-bound Rab is then recognized by the REP, and in the presence of geranyl-geranyl-transferase, it associates with GDIs and GDFs, which regulate the membrane cycling of Rab proteins. Rab27a is localized to melanosomes, secretory granules, late endosomes and lysosomes. Rab27b is predominantly found in the membrane of Golgi stacks and in vesicles located in the TGN area, secretory granules, and late endosomes. GEFs, guanine nucleotide exchange factors; GAPs, GTPase-activating proteins; GDIs, guanine nucleotide dissociation inhibitors; REP, Rab escort protein; GDFs, GDP dissociation stimulator factors; TGN, trans-Golgi network; ER, endoplasmic reticulum; MVBs, multivesicular bodies.

stellate cells and adipocytes) (107-113). Given the pivotal role of exosomes in intercellular communication within the TME, the role of Rab27 as a key regulator of exosome secretion indicates that not only is it a molecular switch for exosome secretion from tumor cells but also it may be a key molecule in regulating the overall communication of the TME.

#### 4. Role of Rab27-mediated exosome secretion in tumors

Although the molecular mechanism of exosome biogenesis remains incompletely understood, the expression of the Rab27 protein as a key molecular switch that governs exosome biogenesis has emerged as an important approach for constructing a cell model of differential exosome secretion. Research has indicated that in hepatocellular carcinoma (HCC) cells, overexpression of Rab27a increases exosome secretion (114). Conversely, the inhibition of SDC4 and Rab27a by mutant  $\beta$ -catenin reduces exosome secretion, thereby facilitating immune escape and cancer cell communication in HCC (115). Moreover, in primary breast cancer (BC) cells, the suppression of Rab27a or Rab27b expression can reduce the secretion of exosomes by primary BC cells (116). Similarly, Hannafon *et al* (117) reported that in BC, silencing Rab27a expression significantly decreased the secretion of exosomes,

inhibited the antiangiogenic effect of docosahexaenoic acid on endothelial cells, and promoted tumor angiogenesis. In melanoma, silencing Rab27a was also shown to reduce exosome secretion (118). In addition to its role in tumor diseases, Rab27 also plays a role in regulating exosome secretion in other disorders. For instance, enterovirus A71 interacts with Rab27a through non-structural protein 3A, which can decrease Rab27a ubiquitination and promote exosome secretion (119). In cardiomyocytes, activation of the AKT/Rab27a signaling pathway can promote the secretion of inflammatory exosomes (120). These findings indicate that Rab27 plays a crucial role in the regulation of exosome secretion. Because exosome secretion is a complex physiological process, altering the expression or interaction of Rab27 with other exosome marker proteins (for example, tetraspanin superfamily members and the Rho family of small GTPases) could be more effective for regulating exosome secretion. For instance, Linc01703 can facilitate the secretion of CD81<sup>+</sup> exosomes via the formation of the Rab27a/SYTL1/CD81 complex (121).

Rab27a and Rab27b are oncogenes. They can regulate diverse malignant biological behaviors of cancer cells via different molecular mechanisms (122,123). Research on melanoma revealed that silencing Rab27 expression significantly decreased the levels of various intracellular proteins associated

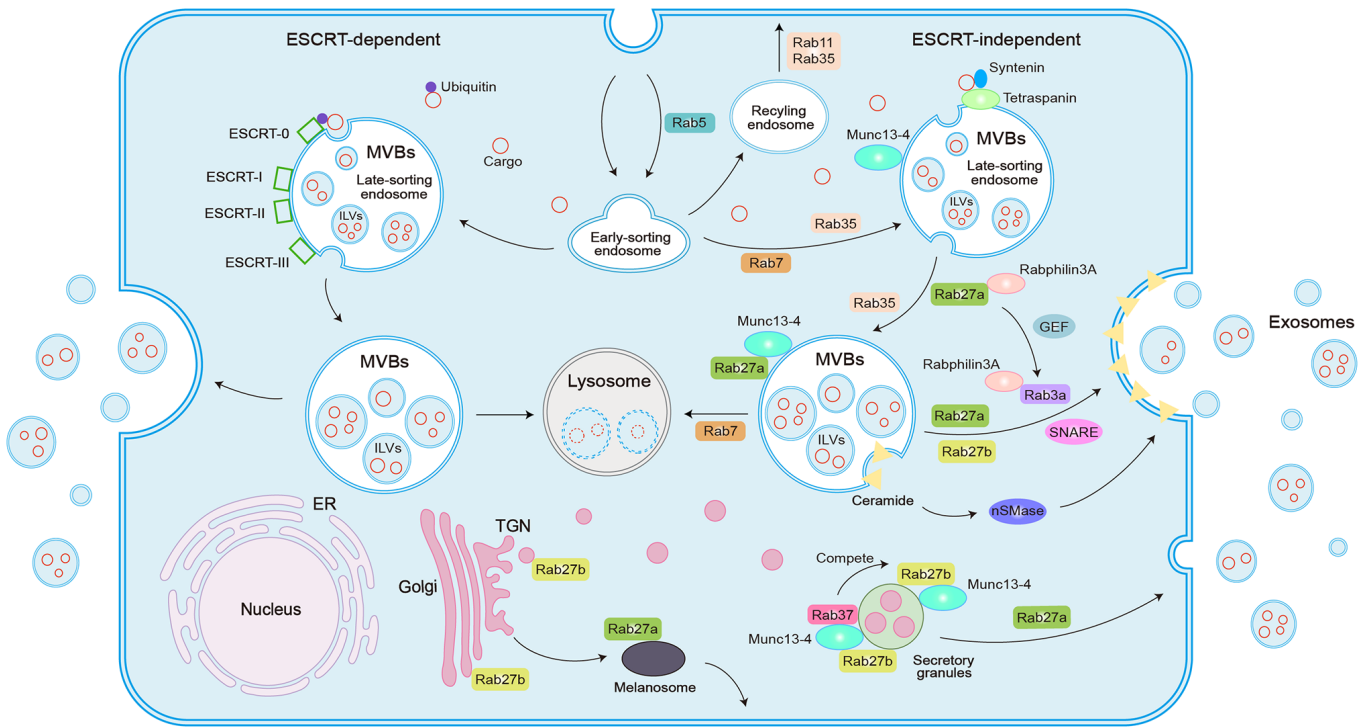


Figure 3. Mechanisms of exosome biogenesis and the functional role of Rab27 in exosome formation. Exosome biogenesis primarily involves two distinct pathways: The ESCRT-dependent mechanism and the ESCRT-independent mechanism. The ESCRT machinery comprises five essential complexes, namely, ESCRT-0, ESCRT-I, ESCRT-II and ESCRT-III, which orchestrate the formation of ILVs and facilitate cargo sorting. VPS4, an ATPase, provides the requisite energy for this intricate process through ATP hydrolysis. The ESCRT-independent pathway predominantly involves a ceramide-dependent mechanism and tetraspanin and Rab proteins. Specifically, Rab27 regulates the binding of MVBs and subsequent exosome secretion and acts in concert with other Rab proteins to orchestrate the entire process of exosome secretion. Rab5 is instrumental in the formation and trafficking of early endosomes, whereas Rab7 plays a crucial role in MVB maturation and lysosome biogenesis. Additionally, Rab37 is involved in the secretion of apical exosomes. During exosome secretion, Rab5, Rab3a and Rab35 act synergistically with Rab27, whereas Rab37 and Rab3D have antagonistic effects on Rab27 function. ESCRT, endosomal sorting complex required for transport; ILVs, intraluminal vesicles; TGN, trans-Golgi network; ER, endoplasmic reticulum; MVBs, multivesicular bodies; GEFs, guanine nucleotide exchange factors.

with cancer cell proliferation, invasion, angiogenesis, adhesion and epithelial-mesenchymal transition (EMT). These findings indicate that Rab27 is closely linked to a variety of malignant biological behaviors of cancer cells (124). The aforementioned study demonstrated that silencing Rab27 in mice bearing brain tumors induced vascular malformations and increased tumor vascular permeability, thereby facilitating immune cell infiltration across the blood-brain barrier and enhancing the efficacy of anti-brain tumor immunotherapy (125). Most studies have demonstrated that Rab27 is closely associated with the proliferation, migration and invasion of cancer cells. Li *et al* reported that Rab27a promoted the proliferation, migration and invasion of colorectal cancer (CRC) cells (126). *In vitro* experiments have shown that Rab27a can increase the proliferation, migration and invasion of oral squamous cell carcinoma cells. The underlying molecular mechanism might be related to the regulation of ZDHHC13-mediated epidermal growth factor receptor palmitoylation and membrane retention by Rab27a (127). Moreover, Rab27a can increase the proliferation and migration of bladder cancer cells by activating the NF- $\kappa$ B pathway. It can also inhibit apoptosis, thereby inducing chemotherapy resistance (128). Rab27b can interact with ZDHHC9 to regulate NRAS palmitoylation and facilitate the progression of myeloid leukemia (129). Interestingly, in a xenograft mouse model, Nambara *et al* (130) demonstrated that Rab27b promoted the peritoneal metastasis of gastric cancer (GC) cells. However, it did not affect the proliferation or invasion of cancer

cells *in vitro* (130). These findings suggest that in a mouse model, Rab27 may promote peritoneal metastasis through exosome-mediated remodeling of the TME rather than through its direct effect on the cancer cells themselves.

Rab27 can affect tumor progression in an exosome-dependent manner (122). In triple-negative BC (TNBC) cells, silencing Rab27a significantly reduced exosome secretion, whereas silencing Rab27b did not significantly affect exosome secretion. The inhibition of Rab27a inhibited cell proliferation, invasion and adhesion (131). Alt *et al* (132) demonstrated that in TNBC, silencing Rab27a reduces exosome secretion and suppresses the interaction between tumor cells and mesenchymal stem cells, thereby inhibiting tumor growth and metastasis. A study regarding non-small cell lung cancer (NSCLC) showed that Rab27b mediates exosome secretion by NSCLC cancer stem cells (CSCs) and proposed that Rab27b is required for the maintenance of a highly tumorigenic, cancer-inducing and aggressive stem cell population (133). A study on HCC has shown that CSCs secrete exosomes in a Rab27a-dependent manner and promote the development of HCC resistance to regorafenib (134). Furthermore, Rab27 knockout has been shown to decrease the secretion of macrophage-derived exosomes, which increases the sensitivity of pancreatic ductal adenocarcinoma (PDAC) to gemcitabine (135). Although these studies suggest that Rab27 can induce tumor progression by promoting the secretion of exosomes, the underlying molecular mechanisms remain

unclear. The genetic material contained in exosomes may be important for the ability of Rab27 to promote the occurrence and development of tumors. Song *et al* (136) reported that Rab27a overexpression could increase the migration and adhesion of renal cell carcinoma (RCC) cells, promote exosomal miR-127-3p secretion, and promote the metastasis of RCC. Moreover, silencing Rab27 in bladder cancer cells significantly decreases the secretion of exosomal miR-23b and miR-921, leading to the suppression of tumor cell invasion (137). In BC cells, the suppression of Rab27 expression can reduce the secretion of exosomal mitochondrial DNA, leading to inhibition of receptor cell invasion and delaying the progression of this disease (138). Upregulated Rab27a promotes the secretion of exosomes and induces the production of IFN $\alpha$  in the culture medium, which then activates the TYK2/STAT/HSPA5 signaling pathway to promote NSCLC cell proliferation and metastasis (139). Rab27 is regulated by upstream regulatory genes, which modulate exosome secretion through Rab27 signaling, thereby influencing tumorigenesis and tumor progression. In lung adenocarcinoma, Linc01703 has been demonstrated to suppress lung cancer metastasis through the formation of the Rab27a/SYTL1/CD81 complex, which facilitates the secretion of CD81<sup>+</sup> exosomes (121). Furthermore, Zhang *et al* (140) revealed that PRR34-AS1 upregulates Rab27a expression by recruiting DDX3X, thereby promoting the secretion of vascular endothelial-derived growth factor and TGF- $\beta$ -containing exosomes in HCC cells and enhancing their malignant phenotype.

Nevertheless, the function of Rab27 is not absolute. Its tumor-suppressive role in PDAC contrasts sharply with its aforementioned tumor-promoting functions, indicating its remarkable functional plasticity and dependence on the TME. The expression level of Rab27a in PDAC gradually decreases with disease progression. Ablation of Rab27a in genetically engineered mouse models promotes liver metastasis of cancer cells, supporting a tumor-suppressive role for Rab27a in PDAC (141). This may be closely linked to PDAC-specific epigenetic regulation. Hypermethylation of the SMC3 gene in PDAC tissues represses its expression, thereby weakening the binding of SMC3 to the promoter and enhancer regions of Rab27a and reducing the transcriptional activity of Rab27a, ultimately resulting in downregulated Rab27a expression in PDAC (141). In addition, disparities in the genetic background of different tumor cell types may also contribute to the divergent functions of Rab27. Collectively, these findings suggest that Rab27 acts neither as a pure oncoprotein nor as a strict tumor suppressor and that its function is coordinately modulated by the intrinsic properties of tumor cells, the TME, and disease stage. These findings provide a theoretical basis for targeting Rab27 and exosome-mediated intercellular communication for precision cancer therapy. Clinical intervention should consider tumor type-specific microenvironmental characteristics and molecular regulatory mechanisms to develop individualized therapeutic regimens, thereby overcoming the limitations of conventional single-target therapies (Table II).

## 5. Diagnostic and therapeutic potential of Rab27 in tumors

The expression of Rab27 in tumor tissues is a valuable biomarker for predicting poor prognosis across various malignant

neoplasms. In gastric carcinoma research, Rab27b expression has been found to be negatively correlated with both overall survival and recurrence-free survival, indicating its potential as a prognostic biomarker for adverse outcomes (130). With respect to lung squamous cell carcinoma, increased Rab27b expression has been identified as a potential negative prognostic indicator for patients with NSCLC (142). In pancreatic cancer studies, Rab27b expression has been shown to be associated with poor survival outcomes in a subgroup of patients who may exhibit favorable responses to adjuvant chemotherapy (143). Notably, the roles of Rab27a and Rab27b are distinct in cancer tissues with different pathological characteristics. A study by An *et al* (144) further indicated that Rab27a and Rab27b may serve as potential biomarkers for predicting lymph node metastasis and prognosis in patients with GC. In GC tissues, Rab27a is highly expressed and predominantly localized in the nucleus, and its expression level is significantly positively correlated with lymph node metastasis. Rab27b is also globally overexpressed, is distributed mainly in the cytoplasm and PM, and has pathological subtype-specific prognostic value (144). In well-differentiated GC, high Rab27b expression is associated with reduced patient survival, whereas in poorly differentiated adenocarcinoma, low Rab27b expression predicts a worse prognosis (144). However, the aforementioned study lacked comparative analyses of Rab27a and Rab27b expression and localization in paired adjacent non-tumor tissues, which may explain the divergent prognostic performance of these two proteins in GCs with distinct differentiation statuses. In a study by Kottorou *et al* (145), the expression of both Rab27a and Rab27b was shown to be downregulated in CRC tissues and correlated with poor patient prognosis. Notably, low Rab27b expression is closely associated with advanced tumor stage, lymph node metastasis, and distant metastasis, whereas high Rab27b expression is linked to poor tumor differentiation and high malignancy (145). These findings suggest that Rab27 may act as a double-edged sword in relation to cancer progression. This phenomenon may be attributed to tumor heterogeneity and the bidirectional nature of signaling networks downstream of Rab27a and could also involve differential responses of tumor cells to diverse external stimuli, including chemotherapy and targeted therapy (Table III).

In the field of cancer therapy, as a key regulator of exosome secretion, Rab27 holds great promise for targeted therapeutic applications. By mediating exosome secretion and remodeling the TME, targeting Rab27 may not only directly suppress the malignant phenotypes of tumor cells but also improve vascular function and reverse the immunosuppressive state (125). Currently, therapeutic strategies targeting Rab27 are under active development and are expected to provide new breakthroughs for precision cancer therapy.

Given the critical role of Rab27 in tumor progression and its complex downstream regulatory networks, downregulating Rab27 expression using gene silencing technologies is theoretically associated with favorable antitumor effects. Researchers have strategically utilized the unique tropism of Epstein-Barr virus (EBV) for human B cells by incorporating Rab27a small interfering RNA into inactivated EBV particles, which effectively reduces the release of CD19<sup>+</sup> exosomes from B cells, consequently enhancing the antitumor efficacy of chemotherapeutic interventions (146). Furthermore, targeting Rab27 in

Table II. Rab27/Rab27 mediate the role of exosome in tumors.

Authors, year	Cancer type	Rab27 subtype	Exosome secretion	Rab27 expression	Downstream molecules	Phenotype	(Refs.)
Horodecka <i>et al.</i> , 2025	Melanoma	Rab27a/b	-	Up	HER/AKT, HER/ERK	Proliferation, metastasis, angiogenesis, EMT	(124)
Adnani <i>et al.</i> , 2022	Brain tumor	Rab27a/b	-	Up	TJ/Claudin5	Angiogenesis, vascular permeability, immunotherapy	(125)
Li <i>et al.</i> , 2022	CRC	Rab27a	-	Up		Proliferation, metastasis	(126)
Huang <i>et al.</i> , 2023	OSCC	Rab27a	-	Up	ZDHHC13/EGFR	Proliferation, metastasis	(127)
Liu <i>et al.</i> , 2017	Bladder cancer	Rab27a	-	Up	p-κB/p-p65/NF-κB	Proliferation, metastasis, chemoresistance	(128)
Ren <i>et al.</i> , 2023	ML	Rab27b	No	Up	ZDHHC9/NRAS, c-RAF/MEK/ERK	Proliferation	(129)
Nambara <i>et al.</i> , 2023	GC	Rab27b	-	Up		Metastasis	(130)
Wang <i>et al.</i> , 2023	TNBC	Rab27a	Yes	Up		Proliferation, metastasis	(131)
Alt <i>et al.</i> , 2020	TNBC	Rab27a	Yes	Up		Proliferation, metastasis	(132)
Meneses <i>et al.</i> , 2023	NSCLC	Rab27b	Yes	Up		Proliferation, metastasis, angiogenesis	(133)
Huang <i>et al.</i> , 2021	HCC	Rab27a	Yes	Up		Regorafenib resistance	(134)
Binenbaum <i>et al.</i> , 2021	PDAC	Rab27a/b	Yes	Up	miR-365	Gemcitabine resistance	(135)
Song <i>et al.</i> , 2024	RCC	Rab27a	Yes	Up	miR-127-3p	Metastasis	(136)
Ostenfeld <i>et al.</i> , 2014	Bladder cancer	Rab27a/b	Yes	Up	miR-23b, miR-921	Metastasis	(137)
Rabas <i>et al.</i> , 2021	Breast cancer	Rab27a/b	Yes	-	TLR 9	Metastasis	(138)
Zeng <i>et al.</i> , 2024	NSCLC	Rab27a	Yes	Up	IFNα, TYK2/STAT/HSPA5	Proliferation, metastasis	(139)
Huang <i>et al.</i> , 2023	LUAD	Rab27a	Yes	Up	Rab27a/SYTL1/CD81	Metastasis	(121)
Zhang <i>et al.</i> , 2022	HCC	Rab27a	Yes	Up	VEGF, TGF-β	Proliferation, metastasis, EMT	(140)
Bastos <i>et al.</i> , 2023	PDAC	Rab27a		Down		Metastasis	(141)

CRC, colorectal cancer; OSCC, oral squamous cell carcinoma; GC, gastric cancer; TNBC, triple-negative breast cancer; ML, myeloid leukemia; NSCLC, non-small cell lung cancer; HCC, hepatocellular carcinoma; PDAC, pancreatic ductal adenocarcinoma; RCC, renal cell carcinoma; LUAD, lung adenocarcinoma.

Table III. The potential of Rab27 as a tumor biomarker.

Authors, year	Cancer	Rab27 subtype	Rab27 expression	Type of biomarker	Correlation	(Refs.)
Nambara <i>et al</i> , 2023	GC	Rab27b	Up	Survival rate	Negative correlation	(130)
Koh and Song, 2019	SQCC	Rab27b	Up	Survival rate	Negative correlation	(142)
Pecqueux <i>et al</i> , 2023	Pancreatic cancer	Rab27b	Up Up	Survival rate Chemotherapy sensitivity	Negative correlation Positive correlation	(143)
An <i>et al</i> , 2022	GC (Well-differentiated/ moderately differentiated adenocarcinoma) (Poorly differentiated adenocarcinoma)	Rab27a Rab27b Rab27b	Up Up Down	Lymphatic metastasis Survival rate Survival rate	Positive correlation Negative correlation Positive correlation	(144)
Kottorou <i>et al</i> , 2023	Colorectal cancer	Rab27a/b	Down	Survival rate	Positive correlation	(145)

GC, gastric cancer.

combination with other agents has synergistic effects. Depletion of Rab27a diminishes tumor-derived exosome secretion, reverses hepatic metabolic reprogramming, alleviates chemoresistance, and reduces adverse effects of chemotherapy (147). In BC cells, silencing Rab27a decreases 6J1-induced PD-L1 secretion in exosomes, potentiates antitumor immune responses, and improves therapeutic efficacy (87). Nevertheless, RNA interference-based strategies generally suffer from insufficient target specificity and poor *in vivo* stability, restricting their clinical application to local administration or reliance on highly efficient delivery carriers (148). In addition, inhibition of a single target may trigger compensatory signaling pathways, resulting in suboptimal therapeutic outcomes (28,149).

Given the limitations associated with single-targeted inhibition of Rab27, the development of multitarget, multi-pathway small-molecule strategies has distinct advantages. Tipifarnib, an inhibitor of exosome biogenesis, simultaneously targets Rab27a, nSMase2 and ALIX. By suppressing the PTM of Rab27 and acting cooperatively on multiple core proteins involved in exosome biogenesis, it overcomes acquired resistance resulting from single-target interventions (150). However, the pharmacokinetic properties and toxicological profiles of such synthetic small molecules still require comprehensive evaluation. Natural small-molecule products have shown promising translational potential in targeting Rab27 because of their multitarget characteristics and favorable cost effectiveness (151). *Acorus calamus* effectively suppresses exosome secretion in TNBC cells through dual targeting of Rab27a and nSMase2 (152). Resveratrol reduces exosome secretion by suppressing Rab27a expression, thereby inhibiting proliferation, migration and EMT in Huh7 cells and blocking HCC progression. Furthermore, resveratrol-induced exosomes further suppress the malignant phenotypes of tumor cells by inhibiting  $\beta$ -catenin nuclear translocation and autophagy activation (153). In the future, identifying and structurally optimizing the core pharmacophores that target Rab27 from natural products is expected to emerge as an important direction for the development of drugs targeting Rab27.

In addition to directly targeting Rab27, disrupting its interaction with downstream effector molecules or blocking downstream signaling pathways represents an effective intervention strategy. The novel compound BHMPS [(E)-N-benzyl-6-(2-(3,4-dihydroxy-benzylidene)hydrazinyl)-N-methylpyridine-3-sulfonamide] has been identified as an effective inhibitor of the Rab27a-Slp4 interaction. The inhibition of Rab27a-mediated exosome secretion represents a promising therapeutic strategy for suppressing BC metastasis and invasion (154). Molecular studies have revealed that MUC1-C interacts with the Rab27a protein and that targeted inhibition of MUC1-C expression significantly suppresses the secretion of MICA/B-containing exosomes (155). Notably, both aforementioned strategies remain at the basic research stage. Camptothecin, a widely used chemotherapeutic agent, targets Topo I to block DNA replication and repair and induce apoptosis in cancer cells (156). Moreover, it has been shown that camptothecin can also inhibit proliferation and migration in head and neck squamous cell carcinoma by blocking the Rab27a-mediated activity of the PI3K/AKT pathway (157). These findings support the feasibility of drug repurposing for cancer therapy.

As inhibitors of exosome biogenesis and secretion, Nexinhib20 and GW4869 exhibit considerable potential in antitumor therapy. Nexinhib20 suppresses exosome secretion by specifically blocking the interaction between Rab27a and JFC1, whereas GW4869 effectively inhibits the expression of nSMase2 and thereby reduces exosome biogenesis (158-160). It has been demonstrated that combining Nexinhib20 or GW4869 with cisplatin/etoposide enhances the inhibitory effects of first-line chemotherapy against small cell lung cancer cells (161). Moreover, Nexinhib20 selectively inhibits granule and exosome release without compromising critical neutrophil functions such as phagocytosis and neutrophil extracellular traps formation, theoretically conferring a lower risk of immunosuppression (162). However, both agents remain at the preclinical stage of cellular and animal models, and their safety profiles and pharmacokinetic properties require further

Table IV. Antitumor approach targeting Rab27.

Authors, year	Cancer	Molecular target	Medication/Method	Function	Current stage	(Refs.)
Zhang <i>et al.</i> , 2023	Colon cancer, GC, lymphoma	Rab27a	iEBV-Rab27a siRNA	Chemotherapy sensitivity	Preclinical stage	(146)
Wang <i>et al.</i> , 2023	Melanoma, osteosarcoma	Rab27a	lentiCRISPR	Metabolic reprogramming of the liver and chemotherapy sensitivity	Preclinical stage	(147)
Gupta <i>et al.</i> , 2025	Triple-negative BC	Rab27a	<i>Acorus calamus</i>	Exosome secretion	Clinical stage	(152)
Tong <i>et al.</i> , 2024	Hepatocellular carcinoma	Rab27a	Resveratrol	Cell proliferation, migration, EMT	Preclinical stage	(153)
Ye <i>et al.</i> , 2023	BC	Rab27a	Triazine compound 6J1	Antitumor immune response	Preclinical stage	(87)
Park <i>et al.</i> , 2022	BC	Rab27a-Slp4 complex	BHMP5	Migration and invasion	Preclinical stage	(154)
Morimoto <i>et al.</i> , 2023	Colorectal cancer	MUC1-C-Rab27a complex	GO-203 inhibitor	Exosome secretion	Preclinical stage	(155)
Zhao <i>et al.</i> , 2024	Head and neck squamous cell carcinoma	Rab27a	Camptothecin	Proliferation and migration	Clinical stage	(157)
Irep <i>et al.</i> , 2024	Small cell lung cancer	Rab27a	Nexinhib20	Exosome secretion	Preclinical stage	(161)

BC, breast cancer.

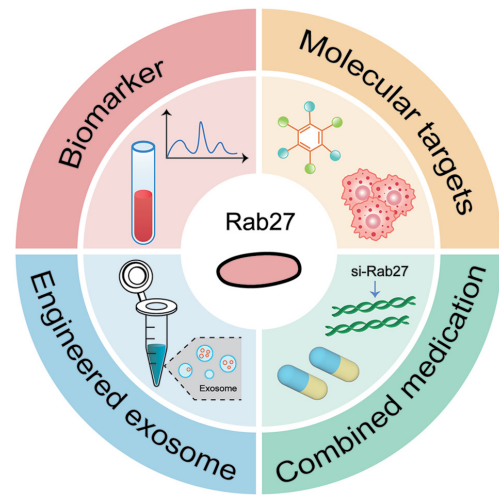


Figure 4. Potential applications of Rab27. Therapeutic prospects in clinical practice via molecular targets, biomarkers, combined medications and engineered exosome preparation.

validation. Additionally, strategies to achieve selective inhibition of tumor-derived exosomes rather than those from normal cells remain to be addressed.

While several Rab27-targeted strategies have been developed, their safety profiles require critical evaluation. Rab27 is widely involved in physiological vesicle secretion in normal cells and regulates key processes, including immune cytokine release, insulin secretion and neurotransmission. Non-specific targeting may readily lead to multisystem side effects such as immune dysregulation and metabolic disorders (122,163). Therefore, a balance must be maintained between therapeutic efficacy and target selectivity in the development of Rab27-targeted antitumor agents. Although existing studies have indicated that resveratrol at effective doses does not significantly affect Rab27 expression in normal cells, its selectivity and potential toxic side effects in complex human physiological environments still require careful evaluation (153,164). Furthermore, the regulation of Rab27 by camptothecin, resveratrol, and *Acorus calamus* is concentration-dependent. How to maintain potent antitumor activity while avoiding interference with insulin secretion and neurotransmission represents an urgent clinical translation challenge (152,153,157) (Fig. 4 and Table IV).

## 6. Conclusions

Typically, Rab27a and Rab27b can regulate the malignant progression of tumors by promoting the secretion of exosomes (149,163). Interestingly, the two play different roles in the process of exosome secretion. Rab27a is localized to peripheral CD63-positive MVBs and is essential for mediating the fusion of MVBs with the PM (30). By contrast, Rab27b is predominantly located in the perinuclear region, where it regulates the transfer of MVBs from microtubules to the actin-rich cortex, facilitating membrane turnover from the TGN to MVBs (30). While the majority of studies have identified Rab27 as an oncogene and a molecular switch for exosome release, alternative perspectives have emerged. For instance, recent research has indicated that the impact

of Rab27a on the migration and invasion of melanoma cell lines may be contingent upon the unique characteristics of the cell line and appears to be independent of exosome secretion (165). These findings not only reveal the complex roles of Rab27 in tumors and exosomes but also provided a direction for further exploration of its underlying mechanisms and application value.

The Rab27 protein plays a pivotal role in tumor biology, primarily through its regulation of exosome secretion, which significantly influences tumor cell proliferation, invasion and metastatic potential. Its expression levels are strongly correlated with tumor initiation, progression and clinical outcomes. The scientific community has reached a consensus that the modulation of Rab27 expression directly impacts exosome secretion dynamics. However, the precise molecular mechanisms underlying Rab27-mediated exosome biogenesis and secretion warrant further comprehensive investigation. Considering the critical role of exosomal genetic cargo in tumor regulation, exploring whether Rab27 facilitates the selective enrichment of specific genetic materials within exosomes, thereby influencing tumor progression, is imperative. Beyond its established potential as a diagnostic biomarker and therapeutic target in oncology, the upregulation of Rab27 expression may offer a novel approach for the efficient production of engineered exosomes, presenting innovative strategies for the large-scale preparation of artificial exosomes for therapeutic applications.

#### Acknowledgements

Not applicable.

#### Funding

The present study was supported in part by the National Natural Science Foundation of China (grant nos. 82472882, 82302987 and 82303534), the Natural Science Foundation of Hunan (grant nos. 2025JJ30047, 2024JJ4025, 2023ZJ1122, 2023JJ60469, 2023JJ40413, 2023JJ30372 and 2023JJ30375), the Science and Technology Innovation Program of Hunan (grant nos. 2023RC3199, 2023SK4034 and 2023RC1073), the National Key Clinical Specialty Scientific Research Project (grant nos. Z2023086 and Z2023017), the Hunan Provincial Health High-Level Talent Scientific Research Project (R2023040 and R2023093), and the Research Project of the Health Commission of Hunan (grant nos. 20255845 and 20255433).

#### Availability of data and materials

Not applicable.

#### Authors' contributions

MT and ZZ drafted the manuscript and prepared the figures. HD, ST, MS, XJ, QP, LO and ZR helped collect the related studies and participated in discussions. YZ and QL designed the review and revised the manuscript. All authors read and approved the final version of the manuscript. Data authentication is not applicable.

#### Ethics approval and consent to participate

Not applicable.

#### Patient consent for publication

Not applicable.

#### Competing interests

The authors declare that they have no competing interests.

#### References

- Morstein J, Bowcut V, Fernando M, Yang Y, Zhu L, Jenkins ML, Evans JT, Guiley KZ, Peacock DM, Krahnke S, *et al*: Targeting Ras-, Rho-, and Rab-family GTPases via a conserved cryptic pocket. *Cell* 187: 6379-6392.e17, 2024.
- Stenmark H: Rab GTPases as coordinators of vesicle traffic. *Nat Rev Mol Cell Biol* 10: 513-525, 2009.
- Hutagalung AH and Novick PJ: Role of Rab GTPases in membrane traffic and cell physiology. *Physiol Rev* 91: 119-149, 2011.
- Lu Q, Wang PS and Yang L: Golgi-associated Rab GTPases implicated in autophagy. *Cell Biosci* 11: 35, 2021.
- Parray ZA: A review on evolution, structural characteristics, interactions, and regulation of the membrane transport protein: The family of Rab proteins. *Int J Biol Macromol* 296: 139828, 2025.
- Barr F and Lambright DG: Rab GEFs and GAPs. *Curr Opin Cell Biol* 22: 461, 2010.
- Goody RS, Rak A and Alexandrov K: The structural and mechanistic basis for recycling of Rab proteins between membrane compartments. *Cell Mol Life Sci* 62: 1657-1670, 2005.
- Füllbrunn N, Nicastro R, Mari M, Griffith J, Herrmann E, Rasche R, Borchers AC, Auffarth K, Kümmel D, Reggiori F, *et al*: The GTPase activating protein Gyp7 regulates Rab7/Ypt7 activity on late endosomes. *J Cell Biol* 223: e202305038, 2024.
- Koike S and Jahn R: Rab GTPases and phosphoinositides fine-tune SNAREs dependent targeting specificity of intracellular vesicle traffic. *Nat Commun* 15: 2508, 2024.
- Borchers AC, Langemeyer L and Ungermann C: Who's in control? Principles of Rab GTPase activation in endolysosomal membrane trafficking and beyond. *J Cell Biol* 220: e202105120, 2021.
- Wu X, Li Y, Ren X and An X: Exosome-camouflaged inhalable (PDSA-HSA) nanocarrier for targeted disulfiram delivery in lung cancer therapy. *Biomater Adv* 180: 214597, 2026.
- Li S, Han H, Yang K, Li X, Ma L, Yang Z and Zhao YX: Exosome-mediated metabolic reprogramming: Effects on thyroid cancer progression and tumor microenvironment remodeling. *Mol Cancer* 24: 247, 2025.
- Yin Y, Ming L, Qin Y, Tang J, Liu B, Liu Y, Jin G, Jiang L, Yao S, Qi X and Huang Z: RAB22A triggers intercellular chemoresistance transmission in colorectal cancer by promoting exosome release via the PKM2-pSNAP23 axis. *Oncogene* 44: 4205-4217, 2025.
- Yang K, Zhou Q, Qiao B, Shao B, Hu S, Wang G, Yuan W and Sun Z: Exosome-derived noncoding RNAs: Function, mechanism, and application in tumor angiogenesis. *Mol Ther Nucleic Acids* 27: 983-997, 2022.
- Qi R, Bai Y, Li K, Liu N, Xu Y, Dal E, Wang Y, Lin R, Wang H, Liu Z, *et al*: Cancer-associated fibroblasts suppress ferroptosis and induce gemcitabine resistance in pancreatic cancer cells by secreting exosome-derived ACSL4-targeting miRNAs. *Drug Resist Updat* 68: 100960, 2023.
- Sun H, Zhang T, Zhang X, Liu Y, Wang X, Wang X, Tan C, Ni S, Weng W, Zhang M, *et al*: Exosomal CCT6A secreted by cancer-associated fibroblasts interacts with  $\beta$ -catenin to enhance chemoresistance and tumorigenesis in gastric cancer. *Adv Sci (Weinh)* 12: e06674, 2025.
- Zhang J, Chen W, Zhang C, He Q, Wang X, Han J, Gao P, Wang K, Xie H, Gao F, *et al*: Prostate cancer cells secrete PD-1 in exosomes to enhance myeloid-derived suppressor cell activity and promote tumor immune evasion. *Cancer Res* 85: 3435-3453, 2025.

18. Zhang H, Feng K, Han M, Shi Y, Zhang Y, Wu J, Yang W, Wang X, Di L and Wang R: Homologous magnetic targeted immune vesicles for amplifying immunotherapy via ferroptosis activation augmented photodynamic therapy against glioblastoma. *J Control Release* 383: 113816, 2025.
19. Greening DW, Xu R, Rai A, Suwakulsiri W, Chen M and Simpson RJ: Clinical relevance of extracellular vesicles in cancer-therapeutic and diagnostic potential. *Nat Rev Clin Oncol* 22: 924-952, 2025.
20. Yuan Y, Jiang H, Xue R, Feng XJ, Liu BF, Li L, Peng B, Ren CS, Li SM, Li N, *et al*: Identification of a biomarker panel in extracellular vesicles derived from non-small cell lung cancer (NSCLC) through proteomic analysis and machine learning. *J Extracell Vesicles* 14: e70078, 2025.
21. Jin H, Tang Y, Yang L, Peng X, Li B, Fan Q, Wei S, Yang S, Li X, Wu B, *et al*: Rab GTPases: Central coordinators of membrane trafficking in cancer. *Front Cell Dev Biol* 9: 648384, 2021.
22. Liu J, Shi J, Niu S, Liu Z, Cui X, Song Y, Tang X, Fan J, Xu H, Yu W, *et al*: Genistein alleviates rheumatoid arthritis by inhibiting fibroblast-like synovial exosome secretion regulated by the Rab27/nMase2/Mfge8 pathway. *Food Funct* 16: 1407-1422, 2025.
23. Bauer KM, Nelson MC, Tang WW, Chiaro TR, Brown DG, Ghazaryan A, Lee SH, Weis AM, Hill JH, Klag KA, *et al*: CD11c+ myeloid cell exosomes reduce intestinal inflammation during colitis. *JCI Insight* 7: e159469, 2022.
24. Scholz K, Pattanayak R, Ekkatine R, Pair FS, Nobles A, Stone WJ and Yacoubian TA: Rab27b promotes lysosomal function and alpha-synuclein clearance in neurons. *J Neurosci* 45: e1579242025, 2025.
25. Jiang S, Shen D, Jia WJ, Han X, Shen N, Tao W, Gao X, Xue B and Li CJ: GGPPS-mediated Rab27A geranylgeranylation regulates  $\beta$  cell dysfunction during type 2 diabetes development by affecting insulin granule docked pool formation. *J Pathol* 238: 109-119, 2016.
26. Soucy A, Potts C, Kaija A, Harrington A, McGilvrey M, Sutphin GL, Korstanje R, Tero B, Seeker J, Pinz I, *et al*: Effects of a global Rab27a null mutation on murine PVAT and cardiovascular function. *Arterioscler Thromb Vasc Biol* 44: 1601-1616, 2024.
27. Kukimoto-Niino M, Sakamoto A, Kanno E, Hanawa-Suetsugu K, Terada T, Shirouzu M, Fukuda M and Yokoyama S: Structural basis for the exclusive specificity of Slac2-a/melanophilin for the Rab27 GTPases. *Structure* 16: 1478-1490, 2008.
28. Menaceur C, Dusaïly O, Gosselet F, Fenart L and Saint-Pol J: Vesicular trafficking, a mechanism controlled by cascade activation of rab proteins: Focus on Rab27. *Biology (Basel)* 12: 1530, 2023.
29. Ramalho JS, Tolmachova T, Hume AN, McGuigan A, Gregory-Evans CY, Huxley C and Seabra MC: Chromosomal mapping, gene structure and characterization of the human and murine RAB27B gene. *BMC Genet* 2: 2, 2001.
30. Ostrowski M, Carmo N, Krumeich S, Fanget I, Raposo G, Savina A, Moita CF, Schauer K, Hume AN, Freitas RP, *et al*: Rab27a and Rab27b control different steps of the exosome secretion pathway. *Nat Cell Biol* 12: 19-30, 1-13, 2010.
31. Lussi YC, Magrane M, Martin MJ and Orchard S; UniProt Consortium: Searching and navigating UniProt databases. *Curr Protoc* 3: e700, 2023.
32. Oates ME, Romero P, Ishida T, Ghalwash M, Mizianty MJ, Xue B, Dosztányi Z, Uversky VN, Obradovic Z, Kurgan L, *et al*: D<sup>2</sup>P<sup>2</sup>: Database of disordered protein predictions. *Nucleic Acids Res* 41 (Database Issue): D508-D516, 2013.
33. Tang Z, Kang B, Li C, Chen T and Zhang Z: GEPIA2: An enhanced web server for large-scale expression profiling and interactive analysis. *Nucleic Acids Res* 47 (W1): W556-W560, 2019.
34. Zerial M and McBride H: Rab proteins as membrane organizers. *Nat Rev Mol Cell Biol* 2: 107-117, 2001.
35. Yin S, Tao Y, Li T, Li C, Cui Y, Zhang Y, Yin S, Zhao L, Hu P, Cui L, *et al*: Itaconate facilitates viral infection via alkylating GD12 and retaining Rab GTPase on the membrane. *Signal Transduct Target Ther* 9: 371, 2024.
36. Ebnet K and Gerke V: Rho and Rab family small GTPases in the regulation of membrane polarity in epithelial cells. *Front Cell Dev Biol* 10: 948013, 2022.
37. Hume AN, Collinson LM, Rapak A, Gomes AQ, Hopkins CR and Seabra MC: Rab27a regulates the peripheral distribution of melanosomes in melanocytes. *J Cell Biol* 152: 795-808, 2001.
38. Ménager MM, Ménasché G, Romao M, Knapnougel P, Ho CH, Garfa M, Raposo G, Feldmann J, Fischer A and de Saint Basile G: Secretory cytotoxic granule maturation and exocytosis require the effector protein hMunc13-4. *Nat Immunol* 8: 257-267, 2007.
39. Zhang J, Zhang K, Qi L, Hu Q, Shen Z, Liu B, Deng J, Zhang C and Zhang Y: DENN domain-containing protein FAM45A regulates the homeostasis of late/multivesicular endosomes. *Biochim Biophys Acta Mol Cell Res* 1866: 916-929, 2019.
40. Bizingre C, Arellano-Anaya Z, Picard F, Pietri M, Baudry A, Roussel F, Bianchi C, Alleaume-Butaux A, Ardila-Osorio H, Romao M, *et al*: ADAM sheddase activity promotes the detachment of small extracellular vesicles from the plasma membrane. *J Extracell Vesicles* 14: e70114, 2025.
41. Zhao T, Pelegrina-Hidalgo N, Edwards DC, Bąk KM, Karmakar U, Fernando AJ, Vendrell M, Rossi AG, Cockroft SL, Kunath T, *et al*: Fluorescence characterization of extracellular vesicles using single-molecule confocal microscopy. *Small Methods* 9: e00907, 2025.
42. Horbay R, Syrvatka V, Bedzay A, van der Merwe M, Burger D and Beug ST: From mitochondria to immunity: The emerging roles of mitochondria-derived vesicles and small extracellular vesicles in cellular communication and disease. *J Extracell Vesicles* 14: e70192, 2025.
43. Mondal J, Pillarisetti S, Junnuthula V, Surwase SS, Hwang SR, Park IK and Lee YK: Extracellular vesicles and exosome-like nanovesicles as pioneering oral drug delivery systems. *Front Bioeng Biotechnol* 11: 1307878, 2024.
44. Waqas MY, Javid MA, Nazir MM, Niaz N, Nisar MF, Manzoor Z, Bhatti SA, Hameed S and Khaliq MH: Extracellular vesicles and exosome: Insight from physiological regulatory perspectives. *J Physiol Biochem* 78: 573-580, 2022.
45. Kalluri R and LeBleu VS: The biology, function, and biomedical applications of exosomes. *Science* 367: eaau6977, 2020.
46. van Niel G, D'Angelo G and Raposo G: Shedding light on the cell biology of extracellular vesicles. *Nat Rev Mol Cell Biol* 19: 213-228, 2018.
47. Mathieu M, Martin-Jaular L, Lavieu G and Théry C: Specificities of secretion and uptake of exosomes and other extracellular vesicles for cell-to-cell communication. *Nat Cell Biol* 21: 9-17, 2019.
48. Bertolio M, Li Q, Mowry FE, Reynolds KE, Alananzeh R, Wei H, Keum K, Jarvis R, Wu J and Yang Y: Glutamatergic regulation of miRNA-containing intraluminal vesicle trafficking and extracellular vesicle secretion from cortical neurons. *J Extracell Vesicles* 14: e70100, 2025.
49. Myette RL, Holterman CE, Trentin-Sonoda M, Cooper TT, Lajoie GA, Cairns G, Burelle Y, El Khatib N, Raman-Nair J, Burger D and Kennedy CRJ: Extracellular vesicle mitochondrial DNA reflects podocyte mitochondrial stress and is associated with relapse in nephrotic syndrome. *Int J Mol Sci* 26: 7245, 2025.
50. Yang J, Wu S and He M: Extracellular vesicle-mediated delivery of mitochondrial circRNA MTCO2 protects against cerebral ischemia by modulating mPTP-dependent ferroptosis. *Redox Biol* 86: 103806, 2025.
51. Wang W, Yang Q, Li J, Li N, Xu J, Yu A, Zheng M, Wu M and Hua C: Exosomes and metabolic reprogramming crosstalk in breast cancer ecosystem: New insights and therapeutic prospects. *Sci Bull (Beijing)* 71: 23-27, 2026.
52. Lai JJ, Chau ZL, Chen SY, Hill JJ, Korpany KV, Liang NW, Lin LH, Lin YH, Liu JK, Liu YC, *et al*: Exosome processing and characterization approaches for research and technology development. *Adv Sci (Weinh)* 9: e2103222, 2022.
53. Wang W, Qiao S, Kong X, Zhang G and Cai Z: The role of exosomes in immunopathology and potential therapeutic implications. *Cell Mol Immununol* 22: 975-995, 2025.
54. Yanagawa K and Yoshimori T: Rubicon regulates exosome secretion via the non-autophagic pathway. *Autophagy* 21: 1160-1162, 2025.
55. Picchio V, Pontecorvi V, Dhori X, Bordin A, Floris E, Cozzolino C, Frati G, Pagano F, Chimenti I and De Falco E: The emerging role of artificial intelligence applied to exosome analysis: From cancer biology to other biomedical fields. *Life Sci* 375: 123752, 2025.
56. Théry C, Zitvogel L and Amigorena S: Exosomes: Composition, biogenesis and function. *Nat Rev Immunol* 2: 569-579, 2002.
57. Arya SB, Collie SP and Parent CA: The ins-and-outs of exosome biogenesis, secretion, and internalization. *Trends Cell Biol* 34: 90-108, 2024.
58. He C, Zheng S, Luo Y and Wang B: Exosome theranostics: Biology and translational medicine. *Theranostics* 8: 237-255, 2018.

59. Jia S, Yang Y, Liu J, Wang Z and Bai L: PPAR $\gamma$  controls ESCRT-dependent fibroblast-like synoviocyte exosome biogenesis and alleviates chondrocyte osteoarthritis mediated by exosomal ANXA1. *J Orthop Translat* 53: 187-205, 2025.
60. Han QF, Li WJ, Hu KS, Gao J, Zhai WL, Yang JH and Zhang SJ: Exosome biogenesis: Machinery, regulation, and therapeutic implications in cancer. *Mol Cancer* 21: 207, 2022.
61. Kenific CM, Zhang H and Lyden D: An exosome pathway without an ESCRT. *Cell Res* 31: 105-106, 2021.
62. Huang LJ, Zhan ST, Pan YQ, Bao W and Yang Y: The role of Vps4 in cancer development. *Front Oncol* 13: 1203359, 2023.
63. Vietri M, Radulovic M and Stenmark H: The many functions of ESCRTs. *Nat Rev Mol Cell Biol* 21: 25-42, 2020.
64. Larios J, Mercier V, Roux A and Gruenberg J: ALIX- and ESCRT-III-dependent sorting of tetraspanins to exosomes. *J Cell Biol* 219: e201904113, 2020.
65. Wang X, Han S, Liang J, Xu C, Cao R, Liu S, Luan Y, Gu Y and Han P: Essential role of Alix in regulating cardiomyocyte exosome biogenesis under physiological and stress conditions. *J Mol Cell Cardiol* 190: 35-47, 2024.
66. Lee YJ, Shin KJ, Jang HJ, Ryu JS, Lee CY, Yoon JH, Seo JK, Park S, Lee S, Je AR, *et al*: GPR143 controls ESCRT-dependent exosome biogenesis and promotes cancer metastasis. *Dev Cell* 58: 320-334.e8, 2023.
67. Raiborg C and Stenmark H: The ESCRT machinery in endosomal sorting of ubiquitylated membrane proteins. *Nature* 458: 445-452, 2009.
68. Wang F, Oudaert I, Tu C, Maes A, Van der Vreken A, Vlummens P, De Bruyne E, De Veirman K, Wang Y, Fan R, *et al*: System Xc<sup>-</sup> inhibition blocks bone marrow-multiple myeloma exosomal crosstalk, thereby countering bortezomib resistance. *Cancer Lett* 535: 215649, 2022.
69. Fan B, Wang L and Wang J: RAB22A as a predictor of exosome secretion in the progression and relapse of multiple myeloma. *Aging (Albany NY)* 16: 4169-4190, 2024.
70. Fei X, Li Z, Yang D, Kong X, Lu X, Shen Y, Li X, Xie S, Wang J, Zhao Y, *et al*: Neddylolation of Corola determines the fate of multivesicular bodies and biogenesis of extracellular vesicles. *J Extracell Vesicles* 10: e12153, 2021.
71. Schuhmacher JS, Tom Dieck S, Christoforidis S, Landerer C, Davila Galleo J, Hersemann L, Seifert S, Schäfer R, Giner A, Toth-Petroczy A, *et al*: The Rab5 effector FERRY links early endosomes with mRNA localization. *Mol Cell* 83: 1839-1855.e13, 2023.
72. Peng W, Chen S, Ma J, Wei W, Lin N, Xing J, Guo W, Li H, Zhang L, Chan K, *et al*: Endosomal trafficking participates in lipid droplet catabolism to maintain lipid homeostasis. *Nat Commun* 16: 1917, 2025.
73. Jiang C, He X, Chen X, Huang J, Liu Y, Zhang J, Chen H, Sui X, Lv X, Zhao X, *et al*: Lactate accumulation drives hepatocellular carcinoma metastasis through facilitating tumor-derived exosome biogenesis by Rab7A lactylation. *Cancer Lett* 627: 217636, 2025.
74. Malankhanova T, Liu Z, Xu E, Bryant N, Sung KW, Li H, Strader S and West AB: LRRK2 interactions with microtubules are independent of LRRK2-mediated Rab phosphorylation. *EMBO Rep* 26: 3445-3466, 2025.
75. Serres MP, Shaughnessy R, Escot S, Hammich H, Cuvelier F, Salles A, Rocancourt M, Verdon Q, Gaffuri AL, Sourigues Y, *et al*: MiniBAR/GARRE1 is a dual Rac and Rab effector required for ciliogenesis. *Dev Cell* 58: 2477-2494.e8, 2023.
76. Xu M, Ji J, Jin D, Wu Y, Wu T, Lin R, Zhu S, Jiang F, Ji Y, Bao B, *et al*: The biogenesis and secretion of exosomes and multivesicular bodies (MVBs): Intercellular shuttles and implications in human diseases. *Genes Dis* 10: 1894-1907, 2022.
77. Liu C, Liu D, Wang S, Gan L, Yang X and Ma C: Identification of the SNARE complex that mediates the fusion of multivesicular bodies with the plasma membrane in exosome secretion. *J Extracell Vesicles* 12: e12356, 2023.
78. Xu S, Cao B, Xuan G, Xu S, An Z, Zhu C, Li L and Tang C: Function and regulation of Rab GTPases in cancers. *Cell Biol Toxicol* 40: 28, 2024.
79. Chen XQ, Zuo X, Becker A and Mobley WC: Hyperactivation of RAB5 disrupts the endosomal Rab cascade leading to endolysosomal dysregulation in Down syndrome: A necessary role for increased APP gene dose. *Alzheimers Dement* 21: e70046, 2025.
80. He CW and Diaz E: Loss of SynDIG4/PRRT1 alters distribution of AMPA receptors in Rab4- and Rab11-positive endosomes and impairs basal AMPA receptor recycling. *Front Pharmacol* 16: 1568908, 2025.
81. Lempicki C, Milosavljevic J, Laggner C, Tealdi S, Meyer C, Walz G, Lang K, Campa CC and Hermle T: Discovery of a small molecule with an inhibitory role for RAB11. *Int J Mol Sci* 25: 13224, 2024.
82. Hall BA, Senior KE, Ocampo NT and Samanta D: Coxiella burnetii-containing vacuoles interact with host recycling endosomal proteins Rab11a and Rab35 for vacuolar expansion and bacterial growth. *Front Cell Infect Microbiol* 14: 1394019, 2024.
83. Savina A, Fader CM, Damiani MT and Colombo MI: Rab11 promotes docking and fusion of multivesicular bodies in a calcium-dependent manner. *Traffic* 6: 131-143, 2005.
84. Marie PP, Fan SJ, Mason J, Wells A, Mendes CC, Wainwright SM, Scott S, Fischer R, Harris AL, Wilson C and Goberdhan DCI: Accessory ESCRT-III proteins are conserved and selective regulators of Rab11a-exosome formation. *J Extracell Vesicles* 12: e12311, 2023.
85. Hsu C, Morohashi Y, Yoshimura SI, Manrique-Hoyos N, Jung S, Lauterbach MA, Bakhti M, Grönberg M, Möbius W, Rhee J, *et al*: Regulation of exosome secretion by Rab35 and its GTPase-activating proteins TBC1D10A-C. *J Cell Biol* 189: 223-232, 2010.
86. Matsui T, Sakamaki Y, Nakashima S and Fukuda M: Rab39 and its effector UACA regulate basolateral exosome release from polarized epithelial cells. *Cell Rep* 39: 110875, 2022.
87. Ye Z, Xiong Y, Peng W, Wei W, Huang L, Yue J, Zhang C, Lin G, Huang F, Zhang L, *et al*: Manipulation of PD-L1 endosomal trafficking promotes anticancer immunity. *Adv Sci (Weinh)* 10: e2206411, 2023.
88. Verweij FJ, Bebelman MP, George AE, Couty M, Bécot A, Palmulli R, Heiligenstein X, Sirés-Campos J, Raposo G, Pegtel DM and van Niel G: ER membrane contact sites support endosomal small GTPase conversion for exosome secretion. *J Cell Biol* 221: e202112032, 2022.
89. Sun Q, Weng RX, Li JH, Li YC, Xu JT, Li R, Lu X and Xu GY: Rab27a-mediated exosome secretion in anterior cingulate cortex contributes to colorectal visceral pain in adult mice with neonatal maternal deprivation. *Am J Physiol Gastrointest Liver Physiol* 325: G356-G367, 2023.
90. Sinha S, Hoshino D, Hong NH, Kirkbride KC, Grega-Larson NE, Seiki M, Tyska MJ and Weaver AM: Cortactin promotes exosome secretion by controlling branched actin dynamics. *J Cell Biol* 214: 197-213, 2016.
91. Zubkova E, Kalinin A, Bolotskaya A, Beloglazova I and Menshikov M: Autophagy-dependent secretion: Crosstalk between Autophagy and exosome biogenesis. *Curr Issues Mol Biol* 46: 2209-2235, 2024.
92. Pizzimenti C, Fiorentino V, Ruggeri C, Franchina M, Ercoli A, Tuccari G and Ieni A: Autophagy involvement in non-neoplastic and neoplastic endometrial pathology: The state of the art with a focus on carcinoma. *Int J Mol Sci* 25: 12118, 2024.
93. Ieni A, Pizzimenti C, Fiorentino V, Franchina M, Germanò A, Raffa G, Martini M, Fadda G and Tuccari G: Immunohistochemical profile of p62/SQSTM1/sequestosome-1 in human low- and high-grade intracranial meningiomas. *Anal Cell Pathol (Amst)* 2024: 5573892, 2024.
94. Pizzimenti C, Fiorentino V, Franchina M, Martini M, Giuffrè G, Lentini M, Silvestris N, Di Pietro M, Fadda G, Tuccari G and Ieni A: Autophagic-related proteins in brain gliomas: Role, mechanisms, and targeting agents. *Cancers (Basel)* 15: 2622, 2023.
95. Martinez-Arroyo O, Flores-Chova A, Sanchez-Garcia B, Redon J, Cortes R and Ortega A: Rab3A/Rab27A system silencing ameliorates High glucose-induced injury in podocytes. *Biology (Basel)* 12: 690, 2023.
96. Cazares VA, Subramani A, Saldade JJ, Hoerauf W and Stuenkel EL: Distinct actions of Rab3 and Rab27 GTPases on late stages of exocytosis of insulin. *Traffic* 15: 997-1015, 2014.
97. Camelo C, Körte A, Jacobs T and Luschnig S: Tracheal tube fusion in *Drosophila* involves release of extracellular vesicles from multivesicular bodies. *J Cell Sci* 135: jcs259590, 2022.
98. Higashio H, Satoh YI and Saino T: Mast cell degranulation is negatively regulated by the Munc13-4-binding small-guanosine triphosphatase Rab37. *Sci Rep* 6: 22539, 2016.
99. Meng Z, Edman MC, Hsueh PY, Chen CY, Klinngam W, Tolmachova T, Okamoto CT and Hamm-Alvarez SF: Imbalanced Rab3D versus Rab27 increases cathepsin S secretion from lacrimal acini in a mouse model of Sjögren's syndrome. *Am J Physiol Cell Physiol* 310: C942-C954, 2016.
100. Arya SB, Chen S, Jordan-Javed F and Parent CA: Ceramide-rich microdomains facilitate nuclear envelope budding for non-conventional exosome formation. *Nat Cell Biol* 24: 1019-1028, 2022.

101. Hendley AM, Ashe S, Urano A, Ng M, Phu TA, Peng XL, Luan C, Finger AM, Jang GH, Kerper NR, *et al*: nSMase2-mediated exosome secretion shapes the tumor microenvironment to immunologically support pancreatic cancer. *bioRxiv* [Preprint]: 2024.09.23.614610, 2024.
102. Qiao Z, Wang X, Zhao H, Deng Y, Zeng W, Wu J and Chen Y: Research on the TSPAN6 regulating the secretion of ADSCs-Exos through syntenin-1 and promoting wound healing. *Stem Cell Res Ther* 15: 430, 2024.
103. Palmulli R, Couty M, Piontek MC, Ponnaiah M, Dingli F, Verweij FJ, Charrin S, Tantucci M, Sasidharan S, Rubinstein E, *et al*: CD63 sorts cholesterol into endosomes for storage and distribution via exosomes. *Nat Cell Biol* 26: 1093-1109, 2024.
104. Ai Y, Guo C, Garcia-Contreras M, Sánchez B LS, Saftics A, Shodubi O, Raghunandan S, Xu J, Tsai SJ, Dong Y, *et al*: Endocytosis blocks the vesicular secretion of exosome marker proteins. *Sci Adv* 10: eadi9156, 2024.
105. Ge YY, Xia XC, Wu AQ, Ma CY, Yu LH and Zhou JY: Identifying adipocyte-derived exosomal miRNAs as potential novel prognostic markers for radiotherapy of esophageal squamous cell carcinoma. *World J Gastrointest Oncol* 17: 98808, 2025.
106. Niu X, Tian W, Li Z, Zhang G and Zhang P: Rab27a<sup>+</sup>CAF exosomal miR-9-5p promotes osteosarcoma progression via CREBRF/MAPK signaling pathway. *Cell Signal* 134: 111964, 2025.
107. Liu C, Luo Y, Zhou H, Lin M, Zang D and Chen J: Immune cell-derived exosomal non-coding RNAs in tumor microenvironment: Biological functions and potential clinical applications. *Chin J Cancer Res* 37: 250-267, 2025.
108. Wang D, Wang S, Jin M, Zuo Y, Wang J, Niu Y, Zhou Q, Chen J, Tang X, Tang W, *et al*: Hypoxic exosomal circPLEKHM1-mediated crosstalk between tumor cells and macrophages drives lung cancer metastasis. *Adv Sci (Weinh)* 11: e2309857, 2024.
109. Wu B, Liu DA, Guan L, Myint PK, Chin L, Dang H, Xu Y, Ren J, Li T, Yu Z, *et al*: Stiff matrix induces exosome secretion to promote tumour growth. *Nat Cell Biol* 25: 415-424, 2023.
110. Tan S, Yang Y, Yang W, Han Y, Huang L, Yang R, Hu Z, Tao Y, Liu L, Li Y, *et al*: Exosomal cargos-mediated metabolic reprogramming in tumor microenvironment. *J Exp Clin Cancer Res* 42: 59, 2023.
111. Yang Y, Liu R, Lv H, Zhao Y, Wang X, Wang Z, Li J and Shi D: Targeting tumor stroma via ultrasound-activated nanodroplets: Disrupting exosome-driven microenvironment crosstalk for enhanced antitumor efficacy. *J Control Release* 386: 114113, 2025.
112. Xun X, Hu H, Liu Q, Su R and Ai J: CAFs exosomal circFOXO1 promotes TNBC autophagy and radioresistance via miR-27a-3p/BNIP3 axis. *Sci Rep* 15: 29273, 2025.
113. Zhou X, Xie F and Zhang L: Tumor-immune cell interactions in cancer progression. *Cancer Res* 85: 2963-2966, 2025.
114. Li J, Lin W, Huang T, Chen M and Lin Q: IL-12 improves the anti-HCC efficacy of dendritic cells loaded with exosomes from overexpressing Rab27a tumor cells. *Exp Cell Res* 439: 114073, 2024.
115. Dantzer C, Vaché J, Brunel A, Mahouche I, Raymond AA, Dupuy JW, Petrel M, Bioulac-Sage P, Perrais D, Dugot-Senan N, *et al*: Emerging role of oncogenic  $\beta$ -catenin in exosome biogenesis as a driver of immune escape in hepatocellular carcinoma. *Elife* 13: RP95191, 2024.
116. Gounis M, Campos AV, Shokry E, Mitchell L, Deshmukh R, Dornier E, Rooney N, Dhayade S, Pardo L, Moore M, *et al*: Metabolic adaptations of micrometastases alter EV production to generate invasive microenvironments. *J Cell Biol* 224: e202405061, 2025.
117. Hannafon BN, Carpenter KJ, Berry WL, Janknecht R, Dooley WC and Ding WQ: Exosome-mediated microRNA signaling from breast cancer cells is altered by the anti-angiogenesis agent docosahexaenoic acid (DHA). *Mol Cancer* 14: 133, 2015.
118. Somuncu ÖS, Taşlı PN and Şahin F: Protocol for testing human melanoma exosomes that shift the healthy phenotype of human dermal cells. *Methods Mol Biol* 2849: 149-160, 2024.
119. Wu J, Zhao Y, Chen Q, Chen Y, Gu J and Mao L: Enterovirus A71 promotes exosome secretion by the nonstructural protein 3A interacting with Rab27a. *Microbiol Spectr* 11: e0344622, 2023.
120. Liu D, Chen H, Fu Y, Yao Y, He S, Wang Y, Cao Z, Wang X, Yang M and Zhao Q: KCa3.1 promotes proinflammatory exosome secretion by activating AKT/Rab27a in atrial myocytes during rapid pacing. *Cardiovasc Ther* 2023: 3939360, 2023.
121. Huang Y, Guo S, Lin Y, Huo L, Yan H, Lin Z, Chen Z, Cai J, Wu J, Yuan J, *et al*: LincRNA01703 facilitates CD81<sup>+</sup> exosome secretion to inhibit lung adenocarcinoma metastasis via the Rab27a/SYTL1/CD81 complex. *Cancers (Basel)* 15: 5781, 2023.
122. Wang TT, Li ZZ, Cai Y, Ren JG and Zhao JH: Emerging roles of Rab27 proteins: From normal to cancer stem cells. *Biochem Biophys Res Commun* 775: 152109, 2025.
123. Dornier E, Rabas N, Mitchell L, Novo D, Dhayade S, Marco S, Mackay G, Sumpton D, Pallares M, Nixon C, *et al*: Glutaminolysis drives membrane trafficking to promote invasiveness of breast cancer cells. *Nat Commun* 8: 2255, 2017.
124. Horodecka K, Czernek L, Pęczek L and Klink M: Revealing the role of RAB27 in HER receptor family expression and signaling in melanoma cells. *Cell Commun Signal* 23: 118, 2025.
125. Adnani L, Meehan B, Kim M, Choi D, Rudd CE, Riazalhosseini Y and Rak J: Immune cell infiltration into brain tumor microenvironment is mediated by Rab27-regulated vascular wall integrity. *Sci Adv* 11: eadr6940, 2025.
126. Li Q, Zhao H, Dong W, Guan N, Hu Y, Zeng Z, Zhang H, Zhang F, Li Q, Yang J and Xiao W: RAB27A promotes the proliferation and invasion of colorectal cancer cells. *Sci Rep* 12: 19359, 2022.
127. Huang J, Yang JG, Ren JG, Xia HF, Chen GH, Fu QY, Zhang LZ, Liu HM, Wang KM, Xie QH and Chen G: Overexpression of RAB27A in oral squamous cell carcinoma promotes tumor migration and invasion via modulation of EGFR membrane stability. *Int J Mol Sci* 24: 13103, 2023.
128. Liu J, Gong X, Zhu X, Xue D, Liu Y and Wang P: Rab27A overexpression promotes bladder cancer proliferation and chemoresistance through regulation of NF- $\kappa$ B signaling. *Oncotarget* 8: 75272-75283, 2017.
129. Ren JG, Xing B, Lv K, O'Keefe RA, Wu M, Wang R, Bauer KM, Ghazaryan A, Burslem GM, Zhang J, *et al*: RAB27B controls palmitoylation-dependent NRAS trafficking and signaling in myeloid leukemia. *J Clin Invest* 133: e165510, 2023.
130. Nambara S, Masuda T, Hirose K, Hu Q, Tobo T, Ozato Y, Kurashige J, Hiraki Y, Hisamatsu Y, Iguchi T, *et al*: Rab27b, a regulator of exosome secretion, is associated with peritoneal metastases in gastric cancer. *Cancer Genomics Proteomics* 20: 30-39, 2023.
131. Wang L, Yan Z and Xia Y: Silencing RAB27a inhibits proliferation, invasion and adhesion of triple-negative breast cancer cells. *Nan Fang Yi Ke Da Xue Xue Bao* 43: 560-567, 2023 (In Chinese).
132. Alt EU, Wörner PM, Pfnür A, Ochoa JE, Schächtele DJ, Barabadi Z, Lang LM, Srivastav S, Burow ME, Chandrasekar B and Izadpanah R: Targeting TRAF3IP2, compared to Rab27, is more effective in suppressing the development and metastasis of breast cancer. *Sci Rep* 10: 8834, 2020.
133. Meneses KM, Pandya P, Lindemann JA, Al-Qasrawi DS, Argo RA, Weems CM, Beetler DJ, Vijay GV, Yan IK, Wolfram J, *et al*: RAB27B drives a cancer stem cell phenotype in NSCLC cells through enhanced extracellular vesicle secretion. *Cancer Res Commun* 3: 607-620, 2023.
134. Huang H, Hou J, Liu K, Liu Q, Shen L, Liu B, Lu Q, Zhang N, Che L, Li J, *et al*: RAB27A-dependent release of exosomes by liver cancer stem cells induces Nanog expression in their differentiated progenies and confers regorafenib resistance. *J Gastroenterol Hepatol* 36: 3429-3437, 2021.
135. Binenbaum Y, Fridman E, Yaari Z, Milman N, Schroeder A, Ben David G, Shlomi T and Gil Z: Transfer of miRNA in macrophage-derived exosomes induces drug resistance in pancreatic adenocarcinoma. *Cancer Res* 78: 5287-5299, 2018.
136. Song DH, Lee JS, Lee JH, Kim DC, Yang JW, Kim MH, Na JM, Cho HK, Yoo J and An HJ: Exosome-mediated secretion of miR-127-3p regulated by RAB27A accelerates metastasis in renal cell carcinoma. *Cancer Cell Int* 24: 153, 2024.
137. Ostenfeld MS, Jeppesen DK, Laurberg JR, Boysen AT, Bramsen JB, Primdal-Bengtson B, Hendrix A, Lamy P, Dagnaes-Hansen F, Rasmussen MH, *et al*: Cellular disposal of miR23b by RAB27-dependent exosome release is linked to acquisition of metastatic properties. *Cancer Res* 74: 5758-5771, 2014.
138. Rabas N, Palmer S, Mitchell L, Ismail S, Gohlke A, Riley JS, Tait SWG, Gammage P, Soares LL, Macpherson IR and Norman JC: PINK1 drives production of mtDNA-containing extracellular vesicles to promote invasiveness. *J Cell Biol* 220: e202006049, 2021.
139. Zeng Y, Zhao J, Wu Z, Huang Y, Wang A, Zhu J, Xu M, Zhang W, Zhang X, Li J, *et al*: Targeting TYK2 alleviates Rab27A-induced malignant progression of non-small cell lung cancer via disrupting IFN $\alpha$ -TYK2-STAT-HSPA5 axis. *NPJ Precis Oncol* 8: 74, 2024.

140. Zhang Z, Zhou Y, Jia Y, Wang C, Zhang M and Xu Z: PRR34-AS1 promotes exosome secretion of VEGF and TGF- $\beta$  via recruiting DDX3X to stabilize Rab27a mRNA in hepatocellular carcinoma. *J Transl Med* 20: 491, 2022.
141. Bastos N, Castaldo SA, Adem B, Machado JC, Melo CA and Melo SA: SMC3 epigenetic silencing regulates Rab27a expression and drives pancreatic cancer progression. *J Transl Med* 21: 578, 2023.
142. Koh HM and Song DH: Prognostic role of Rab27A and Rab27B expression in patients with non-small cell lung carcinoma. *Thorac Cancer* 10: 143-149, 2019.
143. Pecqueux M, Wende B, Sommer U, Baenke F, Oehme F, Hempel S, Aust D, Distler M, Weitz J and Kahlert C: RAB27B expression in pancreatic cancer is predictive of poor survival but good response to chemotherapy. *Cancer Biomark* 37: 207-215, 2023.
144. An HJ, Lee JS, Yang JW, Kim MH, Na JM and Song DH: RAB27A and RAB27B expression may predict lymph node metastasis and survival in patients with gastric cancer. *Cancer Genomics Proteomics* 19: 606-613, 2022.
145. Kottorou A, Dimitrakopoulos FI, Diamantopoulou G, Kalofonou F, Stavropoulos M, Thomopoulos K, Makatsoris T, Koutras A and Kalofonos H: Small extracellular vesicles (sEVs) biogenesis molecular players are associated with clinical outcome of colorectal cancer patients. *Cancers (Basel)* 15: 1685, 2023.
146. Zhang F, Li R, Yang Y, Shi C, Shen Y, Lu C, Chen Y, Zhou W, Lin A, Yu L, *et al*: Specific decrease in B-cell-derived extracellular vesicles enhances post-chemotherapeutic CD8<sup>+</sup> T cell responses. *Immunity* 50: 738-750.e7, 2019.
147. Wang G, Li J, Bojmar L, Chen H, Li Z, Tobias GC, Hu M, Homan EA, Lucotti S, Zhao F, *et al*: Tumour extracellular vesicles and particles induce liver metabolic dysfunction. *Nature* 618: 374-382, 2023.
148. Tang Q and Khvorova A: RNAi-based drug design: Considerations and future directions. *Nat Rev Drug Discov* 23: 341-364, 2024.
149. Izumi T: In vivo roles of Rab27 and its effectors in exocytosis. *Cell Struct Funct* 46: 79-94, 2021.
150. Mallareddy V, Roy R, Cheng Z, Thej C, Benedict C, Truongcao M, Joladarashi D, Magadum A, Ibetti J, Cimini M, *et al*: Tipifarnib reduces extracellular vesicles and protects from heart failure. *Circ Res* 135: 280-297, 2024.
151. Meng L, Zhang C and Yu P: Treating cancer through modulating exosomal protein loading and function: The prospects of natural products and traditional Chinese medicine. *Pharmacol Res* 203: 107179, 2024.
152. Gupta S, Gupta S, Singh M and Patel AK: Role of *Acorus calamus* extract in reducing exosome secretion by targeting Rab27a and nSMase2: A therapeutic approach for breast cancer. *Mol Biol Rep* 52: 124, 2025.
153. Tong K, Wang P, Li Y, Tong Y, Li X, Yan S and Hu P: Resveratrol inhibits hepatocellular carcinoma progression through regulating exosome secretion. *Curr Med Chem* 31: 2107-2118, 2024.
154. Park JI, Song KH, Kang SM, Lee J, Cho SJ, Choi HK, Ahn J, Park JK, Kim J, Hwang SG, *et al*: BHMPS inhibits breast cancer migration and invasion by disrupting Rab27a-mediated EGFR and fibronectin secretion. *Cancers (Basel)* 14: 373, 2022.
155. Morimoto Y, Yamashita N, Daimon T, Hirose H, Yamano S, Haratake N, Ishikawa S, Bhattacharya A, Fushimi A, Ahmad R, *et al*: MUC1-C is a master regulator of MICA/B NKG2D ligand and exosome secretion in human cancer cells. *J Immunother Cancer* 11: e006238, 2023.
156. Gong J, Zhang W and Balthasar JP: Camptothecin-based anti-cancer therapies and strategies to improve their therapeutic index. *Cancers (Basel)* 17: 1032, 2025.
157. Zhao Y, Wang Y, Zhao L, Qu L and Zheng JH: Camptothecin suppresses cell proliferation and migration in head and neck squamous cell carcinoma by blocking RAB27A-mediated phosphatidylinositol 3 kinase (PI3K)/protein kinase B (AKT) pathway. *J Physiol Pharmacol* 75: 205-213, 2024.
158. Johnson JL, Ramadass M, He J, Brown SJ, Zhang J, Abgaryan L, Biris N, Gavathiotis E, Rosen H and Catz SD: Identification of neutrophil exocytosis inhibitors (nexinhibs), small molecule inhibitors of neutrophil exocytosis and inflammation: Druggability of the small GTPase Rab27a. *J Biol Chem* 291: 25965-25982, 2016.
159. Guenther AA, Ahn S, Min J, Zhang C, Lee HJ, Yang HK, Sung BH, Weaver AM and Choi E: Cortactin facilitates malignant transformation of dysplastic cells in gastric cancer development. *Cell Mol Gastroenterol Hepatol* 19: 101490, 2025.
160. Choezom D and Gross JC: Neutral sphingomyelinase 2 controls exosome secretion by counteracting V-ATPase-mediated endosome acidification. *J Cell Sci* 135: jcs259324, 2022.
161. Irep N, Inci K, Tokgun PE and Tokgun O: Exosome inhibition improves response to first-line therapy in small cell lung cancer. *J Cell Mol Med* 28: e18138, 2024.
162. Askari K, Johnson JL, Shukla A, Meneses-Salas E, Kiosses WB, Yu J and Catz SD: Nexinhib20 inhibits JFC1-mediated mobilization of a subset of CD11b/CD18<sup>+</sup> vesicles decreasing integrin avidity, but does not inhibit Racl. *J Leukoc Biol* 117: qiaf012, 2025.
163. Tanaka M and Nakamura T: Role of the RAB27/SYTL axis in tumor microenvironment construction. *Cancer Sci* 116: 1815-1822, 2025.
164. Ren ZQ, Zheng SY, Sun Z, Luo Y, Wang YT, Yi P, Li YS, Huang C and Xiao WF: Resveratrol: Molecular mechanisms, health benefits, and potential adverse effects. *MedComm (2020)* 6: e70252, 2025.
165. Horodecka K, Czernek L, Pęczek Ł, Gadzinowski M and Klink M: Impact of Rab27 on melanoma cell invasion and sEV secretion. *Int J Mol Sci* 25: 12433, 2024.



Copyright © 2026 Tang et al. This work is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International (CC BY-NC-ND 4.0) License.