

ECM extracellular matrix secreted by senescent fibroblasts induced by UVB promotes cell proliferation in HaCaT cells through PI3K/AKT and ERK signaling pathways

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Abstract. Chronic exposure to solar ultraviolet radiation (UV) induces photoaging, and ultimately photocarcinogenesis. Senescent human skin fibroblasts (HSFs) in UVB stress-induced premature senescence (UVB-SIPS) share a similar extracellular matrix (ECM) phenotype with other types of senescent fibroblast. ECM from senescent fibroblasts induced by a variety of stresses has been shown to promote preneoplastic and neoplastic epithelial cell growth, a potential mechanism in carcinogenesis. We undertook this study to explore whether the extracellular matrices from UVB-induced senescent fibroblasts have any effect on the proliferation of HaCaT cells. The results showed that ECM secreted from HSFs in UVB-SIPS has 13.15 and 29.27% more stimulatory effect on proliferation than ECM secreted from presenescent HSFs and non-ECM, respectively. ECM from fibroblasts in UVB-SIPS activates FAK, ERK, and AKT in HaCaT cells. ERK and PI3K/AKT inhibitors inhibit ECM-induced ERK, AKT activation and cell proliferation. Cytochalasin D, a destructive agent of the cytoskeleton, inhibits ECM-induced FAK activation and cell proliferation

in HaCaT cells. Collectively, we conclude that ECM secreted from HSFs in UVB-SIPS promotes cell proliferation via ERK and PI3K/AKT pathways and modulation of FAK and cytoskeletal proteins in HaCaT cells. Pharmacological manipulation of those signaling components may lead to the prevention and treatment of skin cancer induced by chronic solar exposure.

Introduction

Chronic exposure of solar ultraviolet radiation (UV) induces photoaging, and ultimately photocarcinogenesis. UVB has been implicated as the main cause for skin cancer (1,2). Pathologically, photoaging is characterized by the abnormal structure of the dermal connective tissue and alteration of dermal extracellular matrix (ECM) resulting from chronic UV irradiation (2). UVB crosses the epidermis, reaching the upper dermis composed mainly of fibroblast and ECM. Higher levels of senescent cells are found in fibroblasts from skin biopsies from older donors compared with younger counterparts (3). Moreover, *ex vivo* experiments have shown that the repeated exposure of human skin fibroblasts to UVB at subcytotoxic level triggers UVB stress-induced premature senescence (UVB-SIPS) (4).

Fibroblasts are well known for their functions in maintaining the homeostasis of ECM in tissues, which can be disturbed by repeated UV exposure. Over the past few decades, studies have focused on the mechanism through which UVB-induced premature senescence fibroblasts influence dermal ECM. Little has been known about how the alteration of ECM in photoaging skin affects the cellular activities of epithelial cells, such as survival, proliferation, cell cycle progression and relevant signal transduction pathways, despite the fact that all these processes have been implicated as essential mechanisms in the development of skin cancer, involving the clonal expansion of initiated cells giving rise to pre-malignant and then to malignant lesions (5,6).

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Several reports have cast light on the relationship of senescence and tumorigenesis, suggesting that senescent fibroblasts induced by a variety of stresses promote preneoplastic and neoplastic epithelial cell growth, but not normal epithelial cell growth, by the secretion of dissoluble factors, the deposition of ECM and direct cell-cell interaction. Extracellular matrix deposited by senescent fibroblasts accounts for at least 40% of growth stimulation (7). Senescent human skin fibroblasts (HSFs) induced by successive UVB exposure also share a similar phenotype with other types of senescent fibroblasts, in which the increase of certain extra-cellular matrix components, such as collagens, fibronectin and osteonectin, have been found (4,8,9). However, whether ECM deposited by HSFs in UVB-SIPS exhibits a similar effect on growth stimulation or cell proliferation remains unknown.

HaCaT cells are human preneoplastic epidermal keratinocytes, having acquired only some mutations that predispose to malignancy. Thus, it is a commonly used cell model for the understanding of the cellular and molecular mechanisms of tumorigenesis (7,10). Understanding the effects of ECM on HaCaT cell proliferation may offer a better understanding of cell progression. Thus far, two well-established models have been available to investigate how the ECM secreted by presenescent HSFs and HSFs in UVB-SIPS influence the functions of HaCaT cells. One is the induction of premature senescence fibroblasts by repeated UVB exposure (11), and another is the preparation of the extracellular matrix of fibroblasts (7).

Previous observations have suggested that ECM secreted by senescent fibroblasts can preferentially promote HaCaT cell proliferation and progression (7). Although, the mechanisms responsible for these effects have yet to be understood, ERK1/2 (12) and PI3K/AKT (13) signaling pathways have been implicated in the regulation of survival and proliferation in response to cell-ECM interaction in many types of cells (14). Focal adhesion kinase (FAK), a 125-kD tyrosine kinase, appears to also play a central role in integrin-mediated signal transduction, which normally localizes with surface integrin receptors, and is activated by cell binding to ECM (15), and PI3K/AKT and RAS-RAF-ERK1/2 are well-known downstream effectors of FAK (16-18). Activation of these pathways has been found to regulate cell survival and growth (18,19).

Given all aforementioned information, we hypothesized that ECM secreted by HSFs promotes cell proliferation via the activation of ERK and PI3K/AKT pathways in HaCaT cells. Our results provide evidence to support this hypothesis. The modulation of these signaling pathways may lead to better prevention or clinical management of UV-induced skin aging and skin cancer.

Material and methods

Cell culture. Human skin fibroblasts (HSFs) at early cumulative population doublings (CDPs 25) were used for this experiment. HSFs were routinely cultivated in 175-cm² culture flasks containing 35 ml of DMEM/F12 (Invitrogen, USA) supplemented with 10% fetal calf serum (FCS) (Invitrogen, AU). HaCaT cells were cultured in serum-free KBM medium, as previously described (7,20,21). Cultures

were grown at 37°C in a humidified atmosphere containing 5% CO₂. When 80% confluent, the cells were subcultured.

UVB irradiation and identification of HSFs in UVB-SIPS. UVB irradiation of fibroblast cultures was accomplished using a Philips SS-04P/UVB source (wavelength 290-320 nm). The intensity of the radiation from the UVB source was measured prior to each experiment using UVR radiometer with a UVB sensor (Bioblock Scientific, Belgium). As described previously (11), presenescent HSFs (5x10⁶ cells) were subcultured 10-cm culture dishes at 50% confluence in DMEM/F12 + 1% (v/v) FCS, and 2 mM L-glutamine and 100 µg/ml streptomycin. At 72 h after plating, they were washed once with phosphate-buffered saline (PBS) pH 7.4, and exposed to UVB radiation in a thin layer of PBS with a dose of 150 J/m². After irradiation, PBS was replaced by DMEM/F12 + 1% FCS. The radiation stress was performed once a day for 5 days. At 3 days after the last radiation, a senescent phenotype of HSFs was observed under a microscope. Presenescent HSFs were kept in the same culture conditions without UVB exposure. HSFs in UVB-SIPS were identified at 3 days after the last UVB stress. The fibroblasts treated or untreated were transferred to 35-mm culture dishes at a density of 700 cells/cm² in DMEM/F12 + 1% FCS.

Senescence-associated β-galactosidase (SA β-gal) activity assay. Activity was determined 24 h later as described previously (3). The population of SA β-gal-positive cells was determined by counting 400 cells per dish. The proportions of cells positive for SA β-gal activity were given as a percentage of the total number of cells counted in each dish. The results are expressed as the mean of triplicates ± SD. Presenescent and senescent HSFs in UVB-SIPS were identified as containing <10% and >60% senescence-associated SA β-gal-positive, respectively (4).

The preparation of extracellular matrix (ECM) deposited by HSFs and the treatment of HaCaT cells. After identification, presenescent (4x10⁴/cm²) and senescent (5x10⁴/cm²) fibroblasts were allowed to attach to 3.5- or 6-cm culture dishes overnight and were incubated in serum-free medium for 3-4 days to generate similar cell numbers. Cells were treated with 2 mM EDTA for 3 h at 37°C, followed by three rapid washes with PBS to remove cells, leaving the ECM behind. Then, microscopic examination was performed to ensure the complete removal of fibroblasts and the presence of intact ECM in the wells or dishes (10). Fifty percent confluent HaCaT cells were incubated in growth factor-free KGM (Clonetics Co., USA) medium for 2 days, and detached with 0.05% trypsin and 0.05% EDTA. Trypsin activity was neutralized by 0.5% soybean trypsin inhibitor. HaCaT cells then were re-suspended in growth factor-free KGM medium which contained KGM, 1.8 mM CaCl₂, 5 mg/ml insulin and 0.5 mg/ml hydrocortisone, incubated at 37°C for 60 min in a rotator to allow intracellular signaling to basal level, and then plated on non-ECM dishes or dishes prepared above. In some studies, pharmacological agents were added to the HaCaT cells 1 h before seeding (22), while the cells were in suspension. The dose (1 µM of U0126, 400 nM of wortmannin and 2 µM of cytochalasin D, respectively) used in this experiment did

 SPANDIDOS PUBLICATIONS to have a conspicuous cytotoxic effect on HaCaT routinely (MTT assay, data not shown). At the directed times, HaCaT cells were harvested according to the needs of the study.

Pharmacological agents. ERK1/2 inhibitor, U0126, PI3K inhibitor, wortmannin and cytochalasin D used in this study were obtained from Sigma (St. Louis, MO, USA).

Western blot analysis. To determine the time course of the tyrosine phosphorylation of proteins, HaCaT cells were plated at a density of 80,000 cells/cm² onto the ECM as prepared above or non-ECM as a control. At 0.5, 1, 2 and 4 h, the medium was gently aspirated, and the cells were scraped into cell lysis buffer [20 mM Tris (pH 8.0), 150 mM NaCl, 1% NP-40, 1% deoxycholate, 1 mM EDTA, 1 mM phenylmethylsulfonyl fluoride, 40 mM glycerophosphate, 125 μM Na₃VO₄, 50 mM NaF, 2 μg of leupeptin/ml, 2 μg of aprotinin/ml, 2 μg of pepstatin/ml, and 1 mM dithiothreitol]. One hundred microliters per 6-cm dish of cell lysates were incubated in a cold room (4°C) for 30 min. After centrifugation at 14,000 x g for 10 min at 4°C, BCA (Pierce, USA) assays were performed to determine total protein concentrations, which were normalized to 2 μg/μl for all samples. The samples (30 μg) were then prepared in 6X SDS-PAGE sample loading buffer and denatured at 95°C for 5 min, the protein was separated by 10% SDS-PAGE and electrotransferred to Immobilon-P membranes (Millipore, Bedford, MA, USA). After blocking with 3% bovine serum albumin (BSA) in Tris-buffered saline containing 0.1% Tween-20 (TBST), the membranes were incubated with primary antibody directed against total FAK, phospho-FAK (Tyr 397) (sc-11765-R, Santa Cruz Biotechnologies, USA), phospho-ERK1/2 (E-4, Santa Cruz), phospho-Akt (Ser473) (Cell Signaling Technology, USA) and anti-GAPDH (Santa Cruz), in 2% BSA in TBST overnight at 4°C or 1 h at 37°C. After being washed three times with TBST, the membranes were then incubated with HRP-anti-mouse or rabbit (Santa Cruz) secondary antibody for 45 min at room temperature and subsequently washed. A chemiluminescent detection kit (Pierce) was then used to detect expression of these proteins and phospho-proteins, and the density of the bands was quantified using Quality One software (Bio-Rad, USA), as previously reported (23).

Cell proliferation assay. Cell proliferation was assessed by manual cell counting or MTT assay as previously reported (20,21). Briefly, HaCaT cells (1x10⁵ cells/well) were seeded to 3.5-cm dishes pre-coated with ECM prepared earlier or not, and treated as a control in EGF-free KBM medium. The medium was changed once every day. At 24, 48 and 96 h following seeding, attached HaCaT cells were harvested with trypsin and counted using a cell counter (Coulter, USA). For MTT assay (20,21,24,25) at 96 h following seeding, the medium was changed to the same medium supplied with 0.8 mg/ml of MTT, and the HaCaT cells then were incubated for 4 h. Washing with PBS was followed by the addition of 1 ml of dimethylsulfoxide (DMSO, Sigma), shaking for 10 min so that complete dissolution was achieved. Aliquots (150 μl) of the resulting solutions were transferred to 96-well plates

and absorbance was recorded at 560 nm using the Microplate Spectrophotometer System (Labsystems, Helsinki, Finland). In some experiments, pharmacological agents U0126, wortmannin and cytochalasin D were used as described above. Each experiment was repeated three times. No effects of either non-ECM or ECM deposited by both types of fibroblasts on stimulated apoptosis and proliferation was observed compared to DMSO as control.

Apoptosis assay. Apoptosis and cell cycle progression was evaluated 24 h after HaCaT cells being seeded with or without administration of the agents and determined with flow cytometry as described (26). Briefly, floating and adherent cells that were trypsinized were pooled and subsequently fixed in 80% ethanol (-20°C) overnight. After being washed twice with PBS, cells were stained in PBS/EDTA containing 7.5 mM propidium iodide and 10 mg/ml RNase A. After 30 min incubation at room temperature, the cell cycle was analyzed by flow cytometry (FACScalibur, Becton Dickinson), and the percentages of cells in apoptosis were determined using the ModFit LT 3.0 program. Apoptosis assays were performed three times and recorded photographically every time.

Statistical analysis. All experiments were performed at least three times in triplicate and data was statistically analysed using SPSS 11.0 Software. The values in the figures are expressed as mean ± SD. Student's t-test was used to determine if there was a significant difference between two means (p<0.05), and statistical differences are indicated. When multiple means were compared, significance was determined by a one-way analysis of variance (p<0.05) in conjunction with a Newman-Keuls *post hoc* test. For analysis of variance, letter designations are used to indicate significant differences. Means with a common letter designation are not different; those with a different letter designation are significantly different from all other means with different letter designations.

Results

Effect of different ECMs on the proliferation of HaCaT cells. To determine whether changes in ECM in response to repeated UVB exposure promote HaCaT cell proliferation, we treated HaCaT cells with ECM deposited by fibroblasts from different treatment. We preincubated HaCaT cells in growth factor-deficient KBM medium before seeding cells onto dishes coated by ECM secreted from HSFs in UVB-SIPS and presenescent HSFs or non-treated as a control. Proliferation experiments were performed as described in Materials and methods. Our results showed that ECM secreted from HSFs in UVB-SIPS had 13.15 and 29.27% (p<0.05) more stimulatory effect than ECM secreted from presenescent HSFs and non-ECM, respectively (Fig. 1). Interestingly, ERK inhibitor U0126, PI3K inhibitor wortmannin, and FAK inhibitor cytochalasin D, inhibited ECM-induced HaCaT cell proliferation.

Effects of different ECMs on cell-cycle progression and apoptosis of HaCaT cells. To further study whether the

proliferation effect is due to lower apoptosis or higher propagation, flow cytometry analysis was performed to evaluate the effects of different ECM on cell-cycle progression and apoptosis. The results showed that compared with non-ECM groups, the groups cultured in ECM secreted from HSFs in UVB-SIPS and presenescent HSFs had a 23.68 and 8.55% increase of cell proportion in S phase ($p < 0.05$) and a 6.23 and 6.20% decrease of cells in G₀/G₁ phase, respectively, indicating that both ECM from HSFs in UVB-SIPS and presenescent HSFs accelerated cell-cycle progression by promoting the G₁-S phase transition, but, there was a greater effect of ECM from HSFs in UVB-SIPS (Fig. 2C). Unexpectedly, apoptosis analysis showed an opposite result: that HaCaT cultured in non-ECM, ECM from presenescent HSFs and HSFs in UVB-SIPS had averages of 3.88, 5.53 and 7.04% apoptosis, respectively (Fig. 2A and B). As expected, ERK inhibitor U0126, PI3K inhibitor wortmannin, and FAK inhibitor cytochalasin D exacerbated ECM-induced apoptosis (Fig. 2A and B).

Effect of different ECMs on the level of FAK, ERK1/2, and AKT phosphorylation. To determine the activation of FAK, ERK1/2, and AKT in HaCaT cells seeded on ECM deposited by fibroblasts from different treatments, we measured the phosphorylation of FAK, ERK, and AKT in time course experiments using phospho-specific antibodies. The results showed that attachment to prepared ECM (non-ECM controls, ECM deposited by HSFs in UVB-SIPS and ECM by presenescent HSFs) induced both FAK and ERK1/2 phosphorylation in a time-dependent manner, both FAK and ERK1/2 phosphorylation began at 30 min, reached a peak at 120 min, and remained elevated for 4 h (Fig. 3). Densitometric analysis data showed that peak phosphorylation of ERK1/2 in UVB-SIPS groups and presenescent groups was 177 and 131% of non-ECM controls ($p < 0.05$), whereas FAK activity was 154 and 118%, respectively. The most significant difference of FAK phosphorylation occurred at 240 min; UVB-SIPS groups and presenescent groups exhibited 203% (Fig. 3F) and 145% of ERK phosphorylation ($p < 0.01$) (Fig. 3D) compared to controls, respectively. Collectively, in three groups, ECM deposited by HSFs in UVB-SIPS induced the most rapid and intense phosphorylation of ERK1/2 and FAK. For AKT, attachment to non-ECM and ECM deposited by HSFs in UVB-SIPS or by presenescent HSFs induced a slight but not significant phosphorylation of AKT (Fig. 3E).

The effects of ERK1/2 inhibitor U0126 and PI3K inhibitor wortmannin. The above data suggest that PI3K/AKT and ERK may be involved in ECM-mediated cell proliferation. PI3K/AKT functions as a protective signaling against apoptosis, including anoikis, in different cell types, while, it remains controversial whether ERK functions as an anti-apoptotic or a pro-apoptotic molecule (27). To further determine whether ERK1/2, and PI3K/AKT signaling pathway mediates this ECM-induced proliferation of HaCaT cells, we used selective ERK1/2 inhibitor U0126 and PI3K inhibitor wortmannin. The results showed that U0126 and wortmannin completely blocked phosphorylation of ERK1/2 and AKT at a concentration of 1 μ M and 400 nM

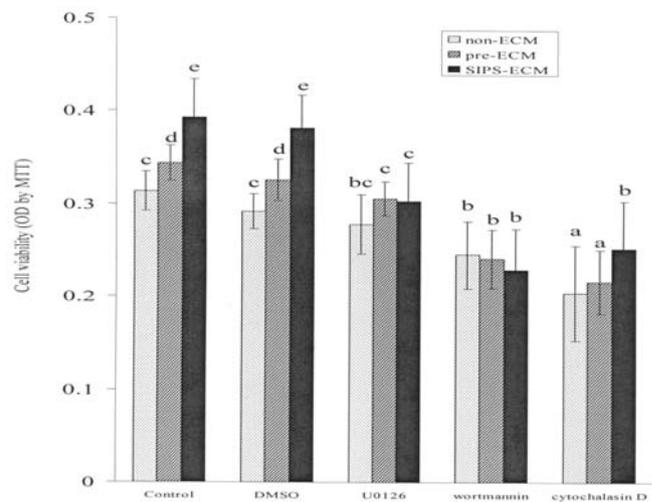


Figure 1. Effects of ECMs from different fibroblasts on the proliferation of HaCaT cells. Fifty percent confluent HaCaT cells were incubated in growth factor-free KBG medium for 2 days before suspension and being seeded onto the non-modified dishes and the dishes coated by ECM from HDFs in UVB-SIPS or presenescent HDFs, as described in Materials and methods. This was performed in control EGF-free KBM medium (blank), medium supplemented with a DMSO vehicle control, or the medium supplemented with ERK inhibitor U0126, PI3K inhibitor wortmannin or cytochalasin D, respectively. After 4 days incubation, MTT assay was performed to evaluate the effect of proliferation. The data are means \pm SD from one experiment representative of three. Letter designations above the bar denote statistical significance ($p < 0.05$) when the bars are labeled with different letters. Where labeled with two or more letters, the data are not significantly different from other data labeled with any of the same letters.

respectively (Fig. 4). The treatment at such concentration did not induce vacuolization or otherwise affect the cells morphologically, at least at the light microscopic level (data not shown).

The effects of cytochalasin D. Previous studies have demonstrated that tyrosine phosphorylation of FAK in response to multiple stimuli requires an intact actin cytoskeleton. Specifically, treatment of the cells with cytochalasin D, which disrupts filamentous actin and promotes their depolymerization, completely inhibits FAK tyrosine phosphorylation (28). While the treatment with cytochalasin D (2 μ M) did not induce cell detachment and slightly affected HaCaT cell morphology, addition of cytochalasin D to the medium for 4 days completely inhibited the effect of ECM-induced proliferation as shown in Fig. 1. To further study the effect of cytochalasin D, we used Western blot analysis. Treatment with 2 μ M cytochalasin D inhibited AKT phosphorylation and dramatically decreased FAK phosphorylation induced by cell-ECM attachment, but unexpectedly activated ERK1/2 (Fig. 5).

Discussion

Although UVB is thought to contribute to both skin aging and cancer, the interplay between photoaging and photocarcinogenesis has not been thoroughly studied. Repeated sublethal doses of UVB can induce premature senescence of

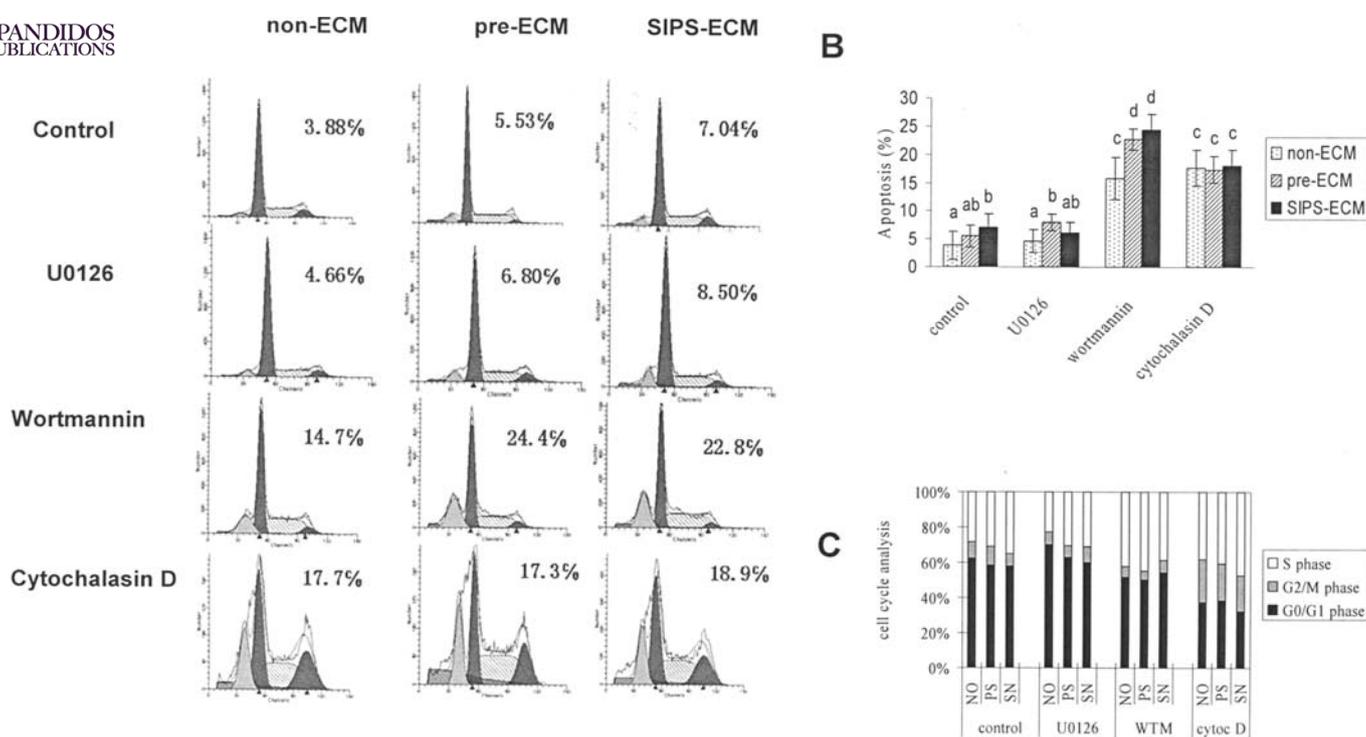


Figure 2. Effects of different ECMs on cell-cycle progression and apoptosis of HaCaT cells. (A) FACS analysis. HaCaT cells were prepared as above, with the presence of ERK inhibitor U0126, PI3K inhibitor wortmannin and cytochalasin D or DMSO control, were plated onto non-ECM dishes and the dishes were coated by ECM from HSFs in UVB-SIPS (SIPS-ECM) or presenescent HSFs (pre-ECM) in EGF-free KGM, respectively. At 24 h after being seeded, the floating and adherent cells were collected, fixed and PI stained, and FACS analysis was performed as described in Materials and methods. The rate of apoptosis was indicated as percentages of sub-G0/G1 phase cells. One representative experiment is illustrated. The numbers in the upper-right of each graph indicate the percentages of cells in apoptosis. (B) The average percentages of apoptosis. (C) Cell cycle analysis. The percentages of S-phase cells (white bars), G0/G1-phase cells (gray bars) and G2/M-phase cells (black bars) are shown. NO, non-ECM groups; PS, pre-ECM groups; SN, UVB-SIPS groups; WTM, wortmannin; cytoc D, cytochalasin D.

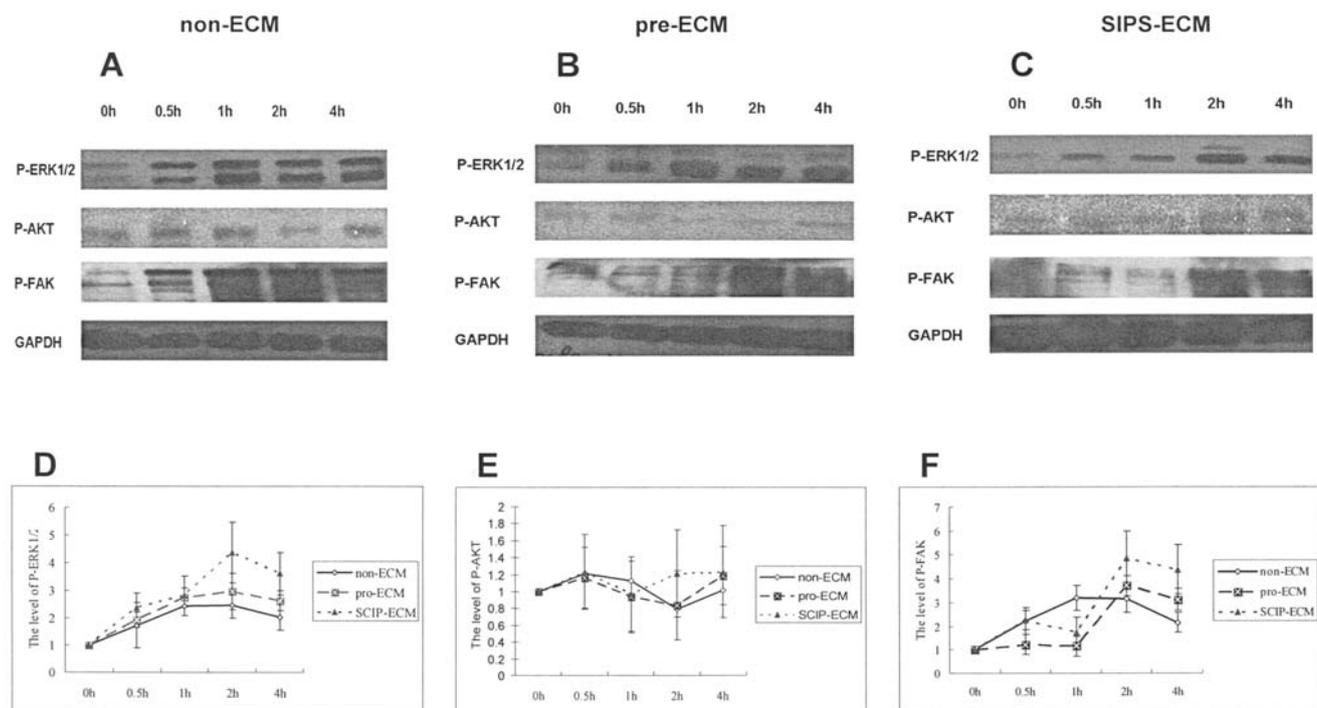


Figure 3. Effects of different ECMs on the levels of FAK, ERK1/2, and AKT phosphorylation. HaCaT cells were plated onto dishes coated by ECM from presenescent HSFs (pre-ECM) and from HSFs in SIPS-ECM (SIPS-ECM) prepared as described, or non-modified dishes (non-ECM). At 0.5, 1, 2 and 4 h, the cells were lysed, the proteins were separated by SDS-PAGE, probed with antibodies to phospho-ERK1/2, phospho-AKT, phospho-FAK and GAPDH and visualized using ECL [(A) non-ECM groups, (B) pre-ECM groups and (C) SIPS-ECM groups]. The densities of these bands were analyzed, and the values were quantified and normalized to the total amount of GAPDH. The time-course manner of ERK1/2, AKT and FAK phosphorylation was shown in D, E and F, respectively. The data were from three independent experiments.

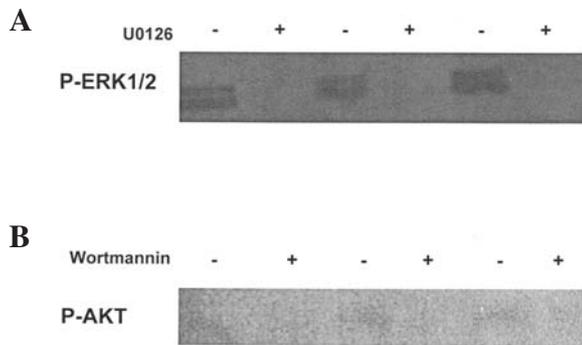


Figure 4. The effects of ERK1/2 inhibitor U0126 and PI3-K inhibitor wortmannin. HaCaT cells were kept in suspension and pretreated either in the absence or presence of ERK inhibitor U0126 or PI3K inhibitor wortmannin for 60 min, then were seeded in the non-modified dishes (lanes 1 and 2), the dishes coated by ECM from presenescent HDFs (lanes 3 and 4) and the dishes coated by ECM from HDFs in UVB-SIPS (lane 5 and 6) in EGF-free KGM, respectively. At 2 h (for analysis of ERK) and 4 h (for AKT) after treatment, the cells were collected and lysed, and phosphorylation of ERK (A) and AKT (B) was analyzed by Western blotting with specific antibodies against phospho-ERK1/2, phospho-AKT.

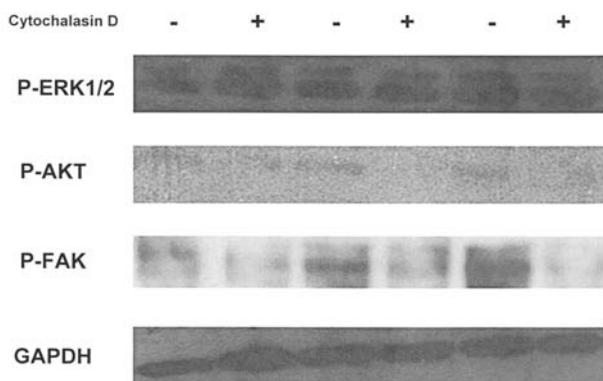


Figure 5. The effects of cytochalasin D. Pretreated HaCaT cells were plated onto dishes coated with prepared ECM or non-modified dishes either in the absence or presence of 2 M cytochalasin D, as described in Materials and methods. At 2 h after treatment, HaCaT cells were harvested and equal amounts of proteins were immunoblotted with antibodies to phospho-ERK1/2, phospho-AKT, phospho-FAK and GAPDH.

HSFs, which undergo a change of ECM-secreted phenotype similar to that of other types of senescent fibroblasts (4,8,9). ECM secreted by senescent human fibroblasts stimulated hyperproliferation and progression of preneoplastic epithelial cells, a potential mechanism for tumorigenesis (7). Whether a similar scenario occurs with ECM from HSFs in UVB stress-induced premature senescence (UVB-SIPS) and what the possible intracellular mechanisms involved are largely unknown. Our data demonstrate that ECM from HSFs in UVB-SIPS stimulates the proliferation of HaCaT cells compared to ECM from presenescent HSFs and controls, which is partially mediated by FAK and ERK, and in a PI3K/AKT-dependent manner.

The proliferation of HaCaT cells relies on cell survival and multiplication. Two mechanisms may be involved in the proliferation: the mechanism that rescues cells from apoptosis, and the mechanism that facilitates the growth of

cells. Fibroblasts are known to produce collagens, fibronectins, and laminins which have been proved to be essential for cell survival and proliferation in various cell types including keratinocytes. HSFs in UVB-SIPS increase the expression of fibronectin and certain forms of collagens. Among other ECM components are fibronectin and collagen (29,30) which function as rescuers from apoptosis and growth promoter. Although the hypothesis that ECM deposited by HSFs in UVB-SIPS can decrease apoptosis can be easily established, the result of the apoptosis assay described above showing that ECM from presenescent HSFs and HSFs in UVB-SIPS did not support more survival than non-ECM, seemingly does not support this. Indeed, one caveat is that ECM prepared as in our system is not composed of one single component. Among the components, some may act as rescuers, and others as inducers of apoptosis in specific cell types or experimental models. The rate of apoptosis in the presence of ECM derived differently may be the result of the coaction of these ECM components. Regardless of paradoxical results, our study showed that ECM secreted from HSFs in UVB-SIPS had 13 and 29% more proliferation effects than ECM secreted from presenescent HSFs and non-ECM, respectively (Fig. 1). Such proliferation effects were further supported by the results of the cell-cycle analysis, which indicated that ECM from HSFs in UVB-SIPS could more intensely promote the G1- to S-phase transition (Fig. 2).

Our results are in agreement with the findings that ECM produced by senescent fibroblasts was more stimulatory than matrix produced by presenescent fibroblasts in HaCaT cells, and that this effect of growth stimulation is independent of the senescence inducer (7). It is worth noting, however, that such a proliferation effect could be underestimated, both because the methods employed for preparing ECM may decrease and inactivate the native matrix components, and because the relative amount of initial cells decreases due to higher apoptosis in UVB-SIPS groups. Overall, all those data results suggest that ECM from HSFs in UVB-SIPS has more stimulatory effects on the proliferation of HaCaT cells, depending much more on higher multiplication than higher survival. These findings also support the notion that cellular senescence is an example of evolutionary antagonistic pleiotropy, protecting complicated organisms from cancer early in life, but promoting cancer later in life (31).

In this study, we also measured ERK1/2, AKT and FAK phosphorylation in response to different ECMs in HaCaT cells. The focus on ERK1/2, AKT and FAK is based not only on their significance in regulating multiple cell functions such as apoptosis and proliferation, but also on their unresolved roles in cell-ECM interaction in previous studies (32). Among MAP kinase family members, ERK is most commonly linked to the regulation of cell proliferation, while JNK and p38 are more strongly tied to stresses (33). The ERK pathway lies at the heart of many signal transduction processes and constitutes a major pathway through which many cell surface receptors including growth factor receptors and integrin receptors transduce their signals into the nucleus. Our study showed that blockade of ERK1/2 fully inhibits the effect of proliferation induced by ECM, which is consistent with many previous observations that ECM-cell attachment leads to substantial activation of ERK and then

 SPANDIDOS PUBLICATIONS proliferation effect. However, both positive and roles of ERK in cell survival regulation have been

observed in several previous studies. In our model, the activation of ERK induced by cell-ECM interaction seemingly exhibited no effect on the apoptosis of HaCaT cells. Collectively, our results suggest that, activated ERK acts neither as an executioner of anoikis, nor as a survival factor, but likely acts as a promoter for the multiplication of HaCaT. Thus, ERK may play different roles in apoptosis in different cell types and experiment models. What determines whether ERK acts in a pro-apoptotic or anti-apoptotic fashion remains an important but unanswered question.

The PI3K/AKT pathway is believed to transduce cell survival signals through the phosphorylation-dependent suppression of intracellular apoptotic factors such as BAD, GSK3 and IKK α . Several studies have reported that environmental stresses capable of inducing apoptosis, such as UVC, ionizing radiation, and hyperosmotic stress lead to the downregulation of the PI3K/AKT pathway, and such downregulation is important in the apoptotic process. Treatment of cells with wortmannin blocks activation of AKT and increases cell death induced by various stresses (34). Our studies also showed that slight but detectable phosphorylated AKT was found in HaCaT cells under a condition of either attachment or suspension, and presented slight difference in the phosphorylated level among non-ECM groups, presenescent ECM groups and UVB-SIPS ECM groups (Fig. 3). Even though there were slight differences among the three groups, the phosphorylated form of AKT still worked as a critical protector of HaCaT cells from apoptosis in our system, the blockade of which could dramatically increase the apoptosis of cells and eventually inhibit the proliferation of HaCaT cells.

Our studies also showed that attachment to ECM rapidly and time-dependently stimulates FAK phosphorylation in HaCaT cells. FAK, a non-receptor PTK, has emerged as a critical link in integrin signal transduction as well as in the transduction of other signals, and has been implicated in the activation of AKT and ERK1/2 in many types of cells (17,35). FAK may be involved in early signaling in response to cell-ECM interaction. To further explore the role of FAK activation in ERK and PI3K/AKT signaling in HaCaT cells subjected to attachment, we pretreated HaCaT cells with cytochalasin D which has been proved to induce filamentous actin depolymerization and completely inhibits FAK tyrosine phosphorylation in other types of cell.

Our results showed that HaCaT cells pretreated by 2 μ M cytochalasin D displayed a dramatic decrease of FAK and AKT activation in response to attachment and notably, an increase in ERK activation which was observed in previous reports (32). Parallel to the blockade of AKT activation, treatment with cytochalasin D dramatically decreased the rate of survival in the three groups, resembling the results of wortmannin-treated groups. A proliferation assay indicated that treatment with cytochalasin D completely inhibited the stimulatory effect of ECM even though it appears to accelerate the cell-cycle progression (Figs. 1 and 2). It is obvious that the higher apoptosis induced by cytochalasin D overwhelmed such progression, further suggesting that AKT and ERK are both involved in the stimulatory effects of

ECM, the former acting as a rescuer, while the latter likely acting as a promoter of proliferation.

In conclusion, our data demonstrate that ECM from HSFs in UVB-SISP has more stimulatory effects on the proliferation of HaCaT cells, which appear to rely much more on higher multiplication than higher survival. In our system, low but detectable PI3K/AKT signaling can partially rescue HaCaT cells from apoptosis induced by EGF-withdrawing plus suspension, while attachment to ECM from HSFs of UVB-SIPS activates FAK and ERK1/2 signaling pathways which play an important role in promoting the multiplication of HaCaT cells, but not in enhancing cell survival. Our data provide insights into the understanding of the molecular mechanism involved in the stimulatory effects of UVB-SIPS ECM on HaCaT cell proliferation and further suggest that the modulation of AKT, ERK and FAK pathways could offer better clinical management or prevention of photocarcinogenesis.

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