

Association of genetic variants with chronic kidney disease in Japanese individuals with type 2 diabetes mellitus

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Abstract. Although diabetes mellitus has been recognized as a risk factor for chronic kidney disease (CKD), genetic factors for predisposition to CKD in individuals with diabetes mellitus remain elucidated. The purpose of the present study was to identify genetic variants that confer susceptibility to CKD among individuals with type 2 diabetes mellitus. The study population comprised 1742 Japanese individuals, including 636 subjects with CKD [estimated glomerular filtration rate (eGFR) <60 ml/min/1.73 m²] and 1106 controls (eGFR ≥60 ml/min/1.73 m²). The genotypes for 24 polymorphisms of 22 candidate genes were determined. An initial screen of allele frequencies by the Chi-square test revealed that four polymorphisms were significantly (false discovery rate <0.05) associated with the prevalence of CKD in individuals with type 2 diabetes mellitus. Subsequent multi-variable logistic regression analysis with adjustment for covariates as well as a stepwise forward selection procedure revealed that the 8733T→C polymorphism of *ALOX5AP* (rs3803278), the C→T (Ser532Leu) polymorphism of *IRAK1* (rs1059703), and the 2445G→A (Ala54Thr) polymorphism of *FABP2* (rs1799883) were significantly (P<0.05) associated with the prevalence of CKD. Our results suggest that *ALOX5AP*, *IRAK1*, and *FABP2* are susceptibility loci for CKD among Japanese individuals with type 2 diabetes mellitus.

Introduction

Chronic kidney disease (CKD) is a major risk factor for end stage renal disease (ESRD) and also an independent risk factor for all-cause death or cardiovascular disease in the general population (1-3), as well as in high-risk populations (4,5). Identification of genetic markers for CKD risk is thus important both for risk prediction and for intervention to avert future ESRD and cardiovascular events.

Diabetes mellitus has been recognized as an important risk factor for developing proteinuria (6,7) and progression to renal dysfunction (8,9) as well as a major source of morbidity and mortality in patients with established CKD (10). In addition, diabetic nephropathy is one of the most serious chronic complications among patients with diabetes mellitus (11), and is associated with increased cardiovascular mortality (12). Although diabetes mellitus is an important risk factor for CKD, genetic factors for predisposition to CKD in individuals with type 2 diabetes mellitus remain largely unknown. Furthermore, given the ethnic differences in lifestyle and environmental factors as well as in renal function and genetic background, it is important to examine genetic variants related to CKD in individuals with type 2 diabetes mellitus in each ethnic group.

We have now performed an association study for 24 polymorphisms of 22 candidate genes and CKD in 1742 Japanese individuals with type 2 diabetes mellitus. The purpose of the present study was to identify genetic variants that confer susceptibility to CKD among individuals with type 2 diabetes mellitus and thereby to provide a basis for the personalized prevention of this condition.

Materials and methods

Study population. The study population comprised 1742 unrelated Japanese individuals (1158 men, 584 women) who either visited outpatient clinics of or were admitted to one of

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Key words: genetics, polymorphism, chronic kidney disease, type 2 diabetes mellitus, end stage renal disease

Table I. The 24 polymorphisms of 22 genes examined in the study.

Locus	Gene	Symbol	Polymorphism	dbSNP ^a
1q23-q25	Selectin E	<i>SELE</i>	561A→C (Ser128Arg)	rs5361
1q31-q32	Interleukin 10	<i>IL10</i>	-819T→C	rs1800871
1q31-q32	Interleukin 10	<i>IL10</i>	-592A→C	rs1800872
2q31	Collagen, type III, alpha-1	<i>COL3A1</i>	3730A→G (Ile1205Val)	rs2271683
3p21.31	SREBF cleavage-activating protein	<i>SCAP</i>	2386A→G (Ile796Val)	rs12487736
3q27	Adipocyte, C1Q, and collagen domain containing	<i>ADIPOQ</i>	62G→T	rs1501299
4q28-q31	Fatty acid-binding protein 2	<i>FABP2</i>	2445G→A (Ala54Thr)	rs1799883
5q34	Potassium channel, calcium-activated, large conductance, subfamily M, β member 1	<i>KCNMB1</i>	G→A (Glu65Lys)	rs11739136
6p21.3	Tumor necrosis factor	<i>TNF</i>	-850C→T	rs1799724
6p12	Vascular endothelial growth factor	<i>VEGF</i>	936C→T	rs3025039
7q21.3-q22	Plasminogen activator inhibitor 1	<i>PAI1</i>	A→G (Tyr243Cys)	rs13306846
7q22.1	Cytochrome P450, subfamily IIIA, polypeptide 4	<i>CYP3A4</i>	13989A→G (Ile118Val)	(NC_000007.12)
8p12	Plasminogen activator, tissue	<i>PLAT</i>	-7351C→T	rs2020918
9q32-q33.3	Prostaglandin-endoperoxide synthase 1	<i>PTGS1</i>	C→T	rs883484
13q12	Arachidonate 5-lipoxygenase-activating protein	<i>ALOX5AP</i>	8733T→C	rs3803278
13q34	Factor VII	<i>F7</i>	11496G→A (Arg353Gln)	rs6046
16p11.2	Vitamin K epoxide reductase complex, subunit 1	<i>VKORC1</i>	2255T→C	rs2359612
17q23	Platelet-endothelial cell adhesion molecule 1	<i>PECAMI</i>	1454C→G (Leu125Val)	rs668
19q13.2	Apolipoprotein E	<i>APOE</i>	-219G→T	rs405509
19q13.2	Apolipoprotein E	<i>APOE</i>	3932T→C (Cys112Arg)	rs429358
22q11.2	Catechol-O-methyltransferase	<i>COMT</i>	G→A (Val158Met)	rs4680
Xq28	Interleukin 1 receptor-associated kinase 1	<i>IRAK1</i>	C→T	rs7061789
Xq28	Interleukin 1 receptor-associated kinase 1	<i>IRAK1</i>	C→T (Ser532Leu)	rs1059703
Xq28	Interleukin 1 receptor-associated kinase 1	<i>IRAK1</i>	A→C	rs3027898

^aIn instances in which rs numbers in dbSNP were not detected, NCBI GenBank accession numbers are shown in parentheses.

the participating hospitals (Gifu Prefectural General Medical Center and Gifu Prefectural Tajimi Hospital in Gifu Prefecture, Japan; and Hirosaki University Hospital, Reimeiko Rehabilitation Hospital, and Hirosaki Stroke Center in Aomori Prefecture, Japan) between October 2002 and March 2008 because of various symptoms or for an annual health checkup, or who were recruited to a population-based prospective cohort study of aging and age-related diseases in Gunma Prefecture and Tokyo, Japan. Glomerular filtration rate was estimated with the use of the simplified prediction equation derived from the modified version of that described in the Modification of Diet in Renal Disease (MDRD) Study as proposed by the Japanese Society of Nephrology (13): $\text{eGFR (ml min}^{-1} \text{ 1.73 m}^{-2}) = 194 \times [\text{age (years)}]^{-0.287} \times [\text{serum creatinine (mg/dl)}]^{-1.094} \times [0.739 \text{ if female}]$. The National Kidney Foundation-Kidney Disease Outcomes Quality Initiative guidelines recommend a diagnosis of CKD if $\text{eGFR is } <60 \text{ ml min}^{-1} \text{ 1.73 m}^{-2}$ (14). Nonlinear relations between GFR and the risk of adverse events, such as death, cardiovascular events, and hospitalization, have been demonstrated, with an increased risk being associated with an $\text{eGFR of } <60 \text{ ml min}^{-1} \text{ 1.73 m}^{-2}$ (1). We thus adopted the criterion of an $\text{eGFR of } <60 \text{ ml min}^{-1} \text{ 1.73 m}^{-2}$ for diagnosis of CKD in the present study. On the basis of this criterion, 636 subjects (421 men, 215 women) were diagnosed

with CKD. The control subjects comprised 1106 controls (737 men, 369 women) whose $\text{eGFR was } \geq 60 \text{ ml min}^{-1} \text{ 1.73 m}^{-2}$. The diagnosis of diabetes mellitus was defined as fasting blood glucose of $\geq 6.93 \text{ mmol/l}$ or hemoglobin A_{1c} of $\geq 6.5\%$, or both, or taking antidiabetes medication.

Type 2 diabetes mellitus was defined according to the criteria accepted by the World Health Organization and described previously (15,16). Individuals with type 1 diabetes mellitus, with maturity-onset diabetes of the young, with other metabolic or endocrinologic diseases, or with severe liver dysfunction were excluded from the study. Individuals taking drugs that cause secondary diabetes mellitus were also excluded. Subjects with CKD and controls either had or did not have other conventional risk factors for CKD, including hypertension (systolic blood pressure of $\geq 140 \text{ mmHg}$ or diastolic blood pressure of $\geq 90 \text{ mmHg}$, or both, or taking antihypertensive medication), or hypercholesterolemia (serum total cholesterol of $\geq 5.72 \text{ mmol/l}$ or taking lipid-lowering medication). The study protocol complied with the Declaration of Helsinki and was approved by the Committees on the Ethics of Human Research of Mie University Graduate School of Medicine, Hirosaki University Graduate School of Medicine, Gifu International Institute of Biotechnology, Tokyo Metropolitan Institute of Gerontology, and participating

Table II. Primers, probes, and other PCR conditions for genotyping of polymorphisms related (FDR <0.05) to chronic kidney disease in individuals with type 2 diabetes mellitus.

Gene	Polymorphism	Sense primer	Antisense primer	Probe 1	Probe 2	Annealing temp. (°C)	Cycles
<i>ALOX5AP</i>	8733T→C	GAGTGATGACCAACCCTTAATACCA	GTCCACAAAAGCCTCTCTGGTGAA	AGCCAAGTTTCTGAGC	AGCCAAGCTTCTGAGGCGC	60	50
<i>IRAK1</i>	C→T (Ser532Leu)	CCTCTTTTCCACTTGCAGGTGTAC	GTGCTGGACACGTAGGAGTTCTC	GGCGGCCTCCAAATGCCCGG	GGCGGCCTCCGAATGCC	60	50
<i>FABP2</i>	2445G→A (Ala54Thr)	AGCTGACAAATTACACAAAGAAGGAA	GTTGTAAATTAAAGGTGACACCAAG	AATGTTTCGAAAAGCGCTTGATT	TCAAAGAATCAAGCACTTTTCGA	60	50
<i>SERPINE1</i>	A→G (Tyr243Cys)	TCTTGTCGTCCTTCACACAGCTGAGT	AGGGGCAGCAATGAACATGCTG	CAGGATGTCGTAGTAATGGC	CAGGATGTCGTAGCAATGGC	60	50

hospitals. Written informed consent was obtained from each participant.

Selection of polymorphisms. Our aim was to identify genes associated with CKD in the Japanese population with type 2 diabetes mellitus in a case-control association study by examining the relations of one to three polymorphisms of each candidate gene to this condition. With the use of public databases, including PubMed (NCBI) and Online Mendelian Inheritance in Man (OMIM), we selected 22 candidate genes that have been characterized and suggested to be associated with CKD. On the basis of published studies or by searching PubMed and single nucleotide polymorphism (SNP) databases [dbSNP (NCBI) and Japanese SNP database (JSNP)], we further selected 24 polymorphisms of these genes, most located in the promoter region or exons, that might be expected to result in changes in the function or expression of the encoded protein (Table I). Wild-type and variant alleles of the polymorphisms were determined from the original sources.

Genotyping of polymorphisms. Venous blood (7 ml) was collected into tubes containing 50 mmol/l ethylenediamine-tetraacetic acid (disodium salt), and genomic DNA was isolated with a kit (Genomix; Talent, Trieste, Italy). Genotypes of the 24 polymorphisms were determined at G&G Science (Fukushima, Japan) by a method that combines the polymerase chain reaction (PCR) and sequence-specific oligonucleotide probes with suspension array technology (Luminex, Austin, TX). Primers, probes, and other PCR conditions for genotyping polymorphisms found to be related [false discovery rate (FDR) <0.05] to CKD by the Chi-square test are shown in Table II. Detailed genotyping methodology was described previously (17).

Statistical analysis. Quantitative data were compared between subjects with CKD and controls by the unpaired Student's t-test. Categorical data were compared by the Chi-square test. Allele frequencies were estimated by the gene counting method, and the Chi-square test was used to identify departures from Hardy-Weinberg equilibrium. In the initial screen, allele frequencies (2 x 2) of each polymorphism were compared between subjects with CKD and controls by the Chi-square test. Given the multiple comparisons of genotypes with CKD, the FDR was calculated from the distribution of P-values for allele frequency of the 24 polymorphisms (18).

Polymorphisms with an FDR of <0.05 were further examined by multivariable logistic regression analysis with adjustment for covariates. Multivariable logistic regression analysis was thus performed with CKD as a dependent variable and independent variables including age, gender (0 = woman, 1 = man), smoking status (0 = nonsmoker, 1 = smoker), history of hypertension (0 = no history, 1 = positive history), and genotype of each polymorphism; P-values, odds ratios, and 95% confidence intervals were calculated. Each genotype was assessed according to dominant (0 = wild-type homozygote, 1 = heterozygote = variant homozygote), recessive (0 = wild-type homozygote = heterozygote, 1 = variant homozygote), and additive [(0, 0) = wild-type homozygote, (1, 0) = heterozygote, (0, 1) = variant homozygote] genetic models. Additive models included the additive 1 model (heterozygotes

Table III. Characteristics of subjects with chronic kidney disease (CKD) and controls among individuals with type 2 diabetes mellitus.

Characteristic	CKD	Controls	P
No. of subjects	636	1106	
Age (years)	70.3±9.1	65.7±9.8	<0.0001
Gender (male/female, %)	66.2/33.8	66.6/33.4	0.8711
BMI (kg/m ²)	23.7±3.5	23.9±3.5	0.2782
Current or former smoker (%)	19.6	27.3	0.0003
Hypertension (%)	81.3	72.1	<0.0001
Systolic blood pressure (mmHg)	152±29	144±25	<0.0001
Diastolic blood pressure (mmHg)	80±15	79±15	0.6703
Hypercholesterolemia (%)	31.5	31.6	0.9412
Serum total cholesterol (mmol/l)	5.24±1.09	5.18±1.03	0.3655
Serum triglycerides (mmol/l)	1.82±1.27	1.66±1.09	0.0154
Serum HDL-cholesterol (mmol/l)	1.28±0.43	1.30±0.36	0.3991
Fasting plasma glucose (mmol/l)	9.42±3.99	9.41±3.89	0.9718
Glycosylated hemoglobin (%)	7.15±1.94	7.40 ±2.05	0.1119
Serum creatinine (mmol/l)	119.5±114.4	62.4±13.0	<0.0001
eGFR (ml/min/1.73 m ²)	46.5±11.8	81.1±16.8	<0.0001
End stage renal failure (%)	3.3	0	<0.0001

Quantitative data are means ± SD. Smoker, smoking of ≥10 cigarettes daily; hypertension, systolic blood pressure of ≥140 mmHg or diastolic blood pressure of ≥90 mmHg (or both), or taking of antihypertensive medication; hypercholesterolemia, serum total cholesterol of ≥5.72 mmol/l or taking lipid-lowering medication.

versus wild-type homozygotes) and the additive 2 model (variant homozygotes versus wild-type homozygotes), which were analyzed simultaneously with a single statistical model.

We also performed a stepwise forward selection procedure to examine the effects of genotypes as well as of other covariates on CKD; each genotype was examined according to a dominant or recessive model on the basis of statistical significance in the multivariable logistic regression analysis. The P-levels for inclusion in and exclusion from the model were 0.25 and 0.1, respectively. With the exception of the initial screen by the Chi-square test (FDR<0.05), a P-value of <0.05 was considered statistically significant. Statistical significance was examined by two-sided tests performed with JMP version 6.0 and JMP Genomics version 3.2 software (SAS Institute, Cary, NC).

Results

The characteristics of the 1742 study subjects are shown in Table III. Age, the prevalence of hypertension, systolic blood pressure, and the serum concentration of triglycerides were greater, whereas the percentage of smoker was lower, in subjects with CKD than in controls.

Comparisons of allele frequencies with the Chi-square test revealed that the 17 polymorphisms were related (allele frequency P<0.05) to the prevalence of CKD (Table IV). Among these polymorphisms, the 8733T→C polymorphism of *ALOX5AP* (rs3803278), the C→T (Ser532Leu) polymorphism of *IRAK1* (rs1059703), the 2445G→A (Ala54Thr) polymorphism of *FABP2* (rs1799883), and the A→G

(Tyr243Cys) polymorphism of *SERPINE1* (rs13306846) were significantly associated with the prevalence of CKD on the basis of an FDR for allele frequency of <0.05 (Table IV). The genotype distributions of the 17 polymorphisms in subjects with CKD and in controls are also shown in Table IV. The genotype distributions of four polymorphisms significantly (FDR <0.05) associated with CKD were in Hardy-Weinberg equilibrium both in CKD subjects and in controls except that for *SERPINE1* in controls because of lack of variants in this group (Table V).

Multivariable logistic regression analysis with adjustment for age, sex, and the prevalence of smoking and hypertension revealed that the 8733T→C polymorphism of *ALOX5AP* (dominant and additive 1 and 2 models), the C→T (Ser532Leu) polymorphism of *IRAK1* (dominant and additive 1 models), and the 2445G→A (Ala54Thr) polymorphism of *FABP2* (dominant and additive 2 models) were significantly (P<0.05) associated with the prevalence of CKD in individuals with type 2 diabetes mellitus (Table VI). The variant A allele of *FABP2* was a risk factor for CKD, whereas the variant C allele of *ALOX5AP* and T allele of *IRAK1* were protective against this condition.

Finally, we performed a stepwise forward selection procedure to examine the effects of genotypes for the four polymorphisms associated with CKD by the Chi-square test as well as of age, sex, and the prevalence of smoking and hypertension on CKD (Table VII). Age, hypertension, *SERPINE1* genotype (dominant model), *ALOX5AP* genotype (dominant model), *IRAK1* genotype (dominant model), and *FABP2* genotype (dominant model), in descending order of

Table IV. Genotype distributions of polymorphisms related (allele frequency $P < 0.05$) to chronic kidney disease (CKD) among individuals with type 2 diabetes mellitus as determined by the Chi-square test.

Gene symbol	Polymorphism	dbSNP ^a	CKD	Controls	P	FDR
<i>ALOX5AP</i>	8733T→C	rs3803278			0.0051	0.0499
	TT		225 (35.4)	320 (28.9)		
	TC		304 (47.8)	563 (50.9)		
	CC		107 (16.8)	223 (20.2)		
<i>IRAK1</i>	C→T (Ser532Leu)	rs1059703			0.0056	0.0499
	CC		459 (72.2)	735 (66.5)		
	CT		71 (11.2)	155 (14.0)		
	TT		106 (16.7)	215 (19.5)		
<i>FABP2</i>	2445G→A (Ala54Thr)	rs1799883			0.0069	0.0499
	GG		248 (39.0)	497 (44.9)		
	GA		291 (45.8)	475 (43.0)		
	AA		97 (15.3)	134 (12.1)		
<i>SERPINE1</i>	A→G (Tyr243Cys)	rs13306846			0.0083	0.0499
	AA		632 (99.4)	1106 (100)		
	AG		4 (0.6)	0 (0)		
<i>IRAK1</i>	A→C	rs3027898			0.0137	0.0577
	AA		104 (16.4)	207 (18.7)		
	AC		70 (11.0)	152 (13.8)		
	CC		461 (72.6)	746 (67.5)		
<i>APOE</i>	-219G→T	rs405509			0.0144	0.0577
	GG		42 (6.6)	107 (9.7)		
	GT		250 (39.3)	453 (41.0)		
	TT		344 (54.1)	546 (49.4)		
<i>IRAK1</i>	C→T	rs7061789			0.0196	0.0605
	CC		464 (73.0)	751 (68.0)		
	CT		68 (10.7)	151 (13.7)		
	TT		104 (16.4)	203 (18.4)		
<i>SELE</i>	561A→C (Ser128Arg)	rs5361			0.0223	0.0605
	AA		582 (91.5)	1045 (94.5)		
	AC		54 (8.5)	60 (5.4)		
	CC		0 (0)	1 (0.1)		
<i>CYP3A4</i>	13989A→G (Ile118Val)	NC_000007.12			0.0227	0.0605
	AA		636 (100)	1096 (99.2)		
	AG		0 (0)	9 (0.8)		
<i>SCAP</i>	2386A→G (Ile796Val)	rs12487736			0.0270	0.0648
	AA		166 (26.1)	343 (31.0)		
	AG		311 (48.9)	517 (46.8)		
	GG		159 (25.0)	245 (22.2)		
<i>APOE</i>	3932T→C (Cys112Arg)	rs429358			0.0325	0.0696
	TT		503 (79.1)	916 (82.8)		
	TC		125 (19.7)	184 (16.6)		
	CC		8 (1.3)	6 (0.5)		
<i>VKORC1</i>	2255T→C	rs2359612			0.0356	0.0696
	TT		518 (81.5)	934 (84.8)		
	TC		109 (17.1)	160 (14.5)		
	CC		9 (1.4)	7 (0.6)		
<i>IL10</i>	-592A→C	rs1800872			0.0414	0.0696
	AA		304 (47.8)	470 (42.5)		
	AC		266 (41.8)	505 (45.7)		
	CC		66 (10.4)	131 (11.8)		

Table IV. Continued.

Gene symbol	Polymorphism	dbSNP ^a	CKD	Controls	P	FDR
<i>F7</i>	11496G→A (Arg353Gln)	rs6046			0.0434	0.0696
	GG		577 (90.7)	970 (87.7)		
	GA		58 (9.1)	131 (11.8)		
	AA		1 (0.2)	5 (0.5)		
<i>IL10</i>	-819T→C	rs1800871			0.0442	0.0696
	TT		304 (47.8)	471 (42.6)		
	TC		266 (41.8)	504 (45.6)		
	CC		66 (10.4)	131 (11.8)		
<i>COL3A1</i>	A→G (Ile1205Val)	rs2271683			0.0469	0.0696
	AA		525 (82.6)	872 (78.8)		
	AG		105 (16.5)	217 (19.6)		
	GG		6 (0.9)	17 (1.5)		
<i>COMT</i>	G→A (Val158Met)	rs4680			0.0498	0.0696
	GG		262 (41.2)	517 (46.8)		
	GA		306 (48.1)	481 (43.5)		
	AA		68 (10.7)	108 (9.8)		

Numbers in parentheses are percentages. ^aIn instances where rs numbers in dbSNP were not detected, NCBI GenBank accession numbers are shown.

Table V. Hardy-Weinberg P-values in subjects with chronic kidney disease (CKD) and controls.

Gene	Polymorphism	CKD	Controls
<i>ALOX5AP</i>	8733T→C	0.8020	0.3877
<i>IRAK1</i>	C→T (Ser532Leu)	0.2723 ^a	0.1738 ^a
<i>FABP2</i>	2445G→A (Ala54Thr)	0.4457	0.2142
<i>SERPINE1</i>	A→G (Tyr243Cys)	0.9366	

^aGenotype distributions in women.

statistical significance, were significant ($P < 0.05$) and independent determinants of CKD in individuals with type 2 diabetes mellitus.

Discussion

We examined the possible relations of 24 polymorphisms in 22 candidate genes to the prevalence of CKD in 1742 Japanese individuals with type 2 diabetes mellitus. Our present study showed that the 8733T→C polymorphism of *ALOX5AP* (rs3803278), the C→T (Ser532Leu) polymorphism of *IRAK1* (rs1059703), and the 2445G→A (Ala54Thr) polymorphism of *FABP2* (rs1799883) were significantly associated with CKD in such individuals.

Arachidonate 5-lipoxygenase-activating protein (*ALOX5AP*) is a regulator of a crucial pathway in the genesis of leukotriene inflammatory mediators (19), which are

implicated in atherosclerosis both in a mouse model (20) and in human studies (21,22). In a genome-wide study, the variants of *ALOX5AP* were associated with the prevalence of both myocardial infarction and stroke in Icelandic population by increasing leukotriene B₄ production and inflammation in the arterial wall (23). In a case-control study, sequence variants in *ALOX5AP* were significantly associated with stroke in a central European population (24). Furthermore, the haplotypes constructed from the 162A→C and 8733T→C polymorphisms of *ALOX5AP* were significantly associated with a reduced risk for myocardial infarction in Japanese population (25). We have now shown that the 8733T→C polymorphism of *ALOX5AP* was significantly associated with the prevalence of CKD in individuals with type 2 diabetes mellitus, with the C allele representing a protective factor for this condition. Effects of this polymorphism on the inflammatory processes of atherosclerosis may account for its association with CKD.

Interleukin 1 receptor-associated kinase 1 (*IRAK1*) is a key intracellular signaling protein that is activated by ligands of toll-like receptors. *IRAK1* activation by interleukin 6 results in phosphorylation and activation of the transcription factor STAT3 and consequent transcriptional activation of the gene for C-reactive protein in Hep3B cells (26). cDNA microarray analysis revealed that *IRAK1* is expressed at high levels in human coronary arteries (27), and *IRAK1* has been shown to be constitutively activated in blood mononuclear cells isolated from individuals with atherosclerosis (28). In addition, *IRAK1* is implicated in regulation of expression of the gene for the anti-inflammatory cytokine interleukin 10 (28). These observations suggest that activation of the innate immune system in general, and of *IRAK1* in particular, may contribute to the increased levels of inflammatory proteins associated

Table VI. Multivariable logistic regression analysis of polymorphisms related (FDR<0.05) to chronic kidney disease by the Chi-square test for individuals with type 2 diabetes mellitus.

Gene symbol	Polymorphism	Dominant		Recessive		Additive 1		Additive 2	
		P	OR (95% CI)	P	OR (95% CI)	P	OR (95% CI)	P	OR (95% CI)
<i>ALOX5AP</i>	8733T→C	0.0036	0.73 (0.59-0.90)	0.1257		0.0110	0.74 (0.59-0.93)	0.0109	0.68 (0.51-0.91)
<i>IRAK1</i>	C→T (Ser532Leu)	0.0066	0.73 (0.58-0.92)	0.0840		0.0283	0.67 (0.46-0.96)	0.0594	
<i>FABP2</i>	2445G→A (Ala54Thr)	0.0482	1.23 (1.00-1.51)	0.1126		0.1242		0.0422	1.38 (1.01-1.89)
<i>SERPINE1</i>	A→G (Tyr243Cys)	0.6559				0.6559			

OR, odds ratio; CI, confidence interval. Multivariable logistic regression analysis was performed with adjustment for age, sex, and the prevalence of smoking and hypertension.

Table VII. Effects of genotypes and other characteristics on chronic kidney disease among individuals with type 2 diabetes mellitus determined by a stepwise forward selection procedure (P<0.05).

Variable	P	R ²
Age	<0.0001	0.0419
Hypertension	0.0002	0.0060
<i>SERPINE1</i> (AG + GG versus AA)	0.0017	0.0043
<i>ALOX5AP</i> (TC + CC versus TT)	0.0021	0.0042
<i>IRAK1</i> (CT + TT versus CC)	0.0027	0.0039
<i>FABP2</i> (GA + AA versus GG)	0.0368	0.0019

R², contribution rate.

with atherosclerosis. Genetic variants of *IRAK1* were found to be associated with the plasma concentration of C-reactive protein in white women (29). Furthermore, we previously demonstrated that the A→C (rs3027898) and C→T (Ser532Leu, rs1059703) polymorphisms of *IRAK1* were associated with atherothrombotic cerebral infarction in men (30). We have now shown that the C→T (Ser532Leu) polymorphism of *IRAK1* was significantly associated with the prevalence of CKD in individuals with type 2 diabetes mellitus, with the T allele protecting against this condition. The T allele of this polymorphism was previously shown to be protective against atherothrombotic cerebral infarction (30). This association may be attributable to effects of this polymorphism on vascular inflammation, although the underlying mechanism remains elucidated.

Fatty acid-binding protein 2 (FABP2) is an intracellular protein that is expressed only in the columnar absorptive epithelial cells of the small intestine. It contains a single ligand site that has a high affinity for saturated and unsaturated fatty acids, and it contributes to the absorption and intracellular transport of long-chain fatty acids (31). The product of A allele of the 2445G→A (Ala54Thr) polymorphism of *FABP2* possesses a greater affinity for long-chain fatty acids *in vitro* than does that of the G allele (32). In addition, individuals with the A allele of this polymorphism were found to be more insulin resistant than were those with the G allele (32, 33). The A allele was also shown to be associated with higher plasma levels of low density lipoprotein-cholesterol (34) or with dyslipidemia (high plasma concentration of triglycerides and low concentration of high density lipoprotein-cholesterol) (35). In addition, the A allele of the 2445G→A (Ala54Thr) polymorphism was previously associated with atherothrombotic cerebral infarction in individuals with metabolic syndrome (36), and a parental history of stroke in the Swedish population (37). Moreover, it was associated with a 2- to 3.5-fold increase in cardiovascular risk in dyslipidemic men with diabetes compared with their dyslipidemic nondiabetic counterparts; for nonfatal MI, stroke, or death from coronary heart disease, the corresponding hazard ratio was 3.0, whereas for stroke alone it was 3.5 (38). We have now shown that the

2445G→A (Ala54Thr) polymorphism was significantly associated with the prevalence of CKD in individuals with type 2 diabetes mellitus, with the A allele representing a risk factor for this condition. The effects of this polymorphism on both insulin resistance and dyslipidemia may account for its association with the prevalence of CKD.

Our study has several limitations: i) We used an eGFR instead of a directly measured GFR to define CKD. ii) We were not able to obtain information about the underlying renal disease in each subject with CKD. Such information could be obtained by detailed clinical examination, including renal biopsy, but such diagnostic procedures are not considered feasible for a study whose subjects are recruited from the general population. iii) It is possible that one or more of the polymorphisms associated with CKD in the present study are in linkage disequilibrium with other polymorphisms in the same gene or in other nearby genes that are actually responsible for the development of this condition. iv) The functional relevance of the identified polymorphisms to gene transcription or to protein function was not determined in the present study.

In conclusion, our present results suggest that the 8733T→C polymorphism of *ALOX5AP*, the C→T (Ser532Leu) polymorphism of *IRAK1*, and the 2445G→A (Ala54Thr) polymorphism of *FABP2* are significantly associated with the prevalence of CKD in Japanese individuals with type 2 diabetes mellitus. Determination of genotypes for these polymorphisms may prove informative for assessment of the genetic risk for CKD in such individuals. Validation of our findings will require their replication with independent subject panels.

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