

Genetic variants at the miR-124 binding site on the cytoskeleton-organizing *IQGAP1* gene confer differential predisposition to breast cancer

HONG ZHENG^{1,2*}, FENGJU SONG^{1,2*}, LINA ZHANG^{1,2}, DA YANG³, PING JI³, YINGMEI WANG³, MARIA ALMEIDA⁴, GEORGE A. CALIN⁴, XISHAN HAO^{1,2}, QINGYI WEI⁵, WEI ZHANG³ and KEXIN CHEN^{1,2}

¹Department of Epidemiology and Biostatistics, Tianjin Medical University Cancer Hospital and Institute;

²Key Laboratory of Breast Cancer Prevention and Therapy, Tianjin Medical University, Ministry of Education, Tianjin, P.R. China; Departments of ³Pathology, ⁴Experimental Therapeutics, and ⁵Epidemiology, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

Received October 20, 2010; Accepted December 17, 2010

DOI: 10.3892/ijo.2011.940

Abstract. *IQGAP1* knockout mice develop gastric cancer, but the *IQGAP1* protein is associated with some advanced-stage human cancers. *IQGAP1* expression is regulated by a microRNA, miR-124, through a binding site at the 3'-untranslated region, where a single nucleotide polymorphism (SNP) exists in the core binding region. We asked whether *IQGAP1* expression is associated with breast cancer development and whether genetic variants at the miR-124 binding site are important. We genotyped the *IQGAP1* SNP rs1042538 A/T in 1,541 breast cancer cases and 1,598 controls and analyzed the frequency of the variant and interactions with major risk factors in these populations. We also measured the expression of *IQGAP1* at both mRNA and protein levels in different *IQGAP1* genotypes. The *IQGAP1* TT genotype, compared with the AA genotype, was associated with a significantly lower risk of developing breast cancer [P=0.049, odds ratio (OR), 0.78; 95% confidence interval (CI), 0.61-0.99]. In case-only analyses, the TT, compared with the AA, genotype was associated with progesterone receptor-positive subjects (OR, 1.35; 95% CI, 1.00-1.83). The expression levels of *IQGAP1* protein were significantly higher in the TT genotype compared to the AA genotype. The presence of SNPs at the miR-124 binding site may be a marker for predicting breast cancer risk and prognosis.

Introduction

During embryonic development and throughout adult life in humans, gene expression is tightly regulated by a complex biological network in a tissue-specific manner and in close interaction with the environment to ensure accurate spatial and temporal differentiation of multiple organs with distinct functions. Significant deviation in the key regulators of this network via mutations often results in pathogenesis, including cancer. Subtle differences in the key regulatory genes via single nucleotide polymorphisms (SNPs) can result either in non-pathologic differences, such as hair color, or in increased propensity for a diseased state, such as cancer. Revealing the roles of these SNPs is a major area of research in the realm of molecular epidemiology in the era of postgenomic medicine (1).

In recent years, studies have revealed the importance of a class of small non-coding RNAs, microRNAs (miRNAs), that are critically involved in regulating gene expression (2). miRNAs directly regulate about 30% of the genes in the human genome via degradation or translational inhibition of their target messenger RNAs (mRNAs) and are thus important regulators of cellular processes such as differentiation, proliferation, mobility, and apoptosis (3,4). The first miRNA-target mRNA pair to be verified *in vivo* was *let-7* miRNA and its target, *lin-41*; and the natural interaction between this miRNA and its target is one of the best understood (5). miRNAs suppress gene expression mainly by binding to the complementary sequences in the 3' untranslated regions (UTRs) of mRNA of their target genes (6), although some miRNAs have been shown to act as an enhancer element to increase gene expression (7). It is generally believed that nucleotides 2-8 from the 5' end of miRNA, called the 'seed region', are the most critical for miRNA binding (8). Perfect Watson-Crick complementarity is observed in these 7 consecutive base pairs in most cases. Therefore, a single nucleotide change in this region may cause sufficient disruption in binding to deregulate the target genes (9). If the target gene is an oncogene, a tumor suppressor gene, or other regulatory genes that are critical for homeostasis, the single nucleotide change in the miRNA binding site may alter

Correspondence to: Dr Kexin Chen, Department of Epidemiology and Biostatistics, Tianjin Medical University Cancer Hospital and Institute, Tianjin 300060, P.R. China
E-mail: chenkexin1963@yahoo.com

Dr Wei Zhang, Department of Pathology, Unit 85, The University of Texas M.D. Anderson Cancer Center, 1515 Holcombe Blvd., Houston, TX 77030, USA
E-mail: wzhang@mdanderson.org

*Contributed equally

Key words: breast cancer, microRNA, SNP, *IQGAP1*, case-control study

the expression of these genes and shift the normal cellular program to a cancer-prone state, thus increasing cancer risk and/or conferring a specific cancer phenotype (10,11).

Breast cancer is the most common cancer among women in developed countries, and an alarming increase in incidence has been seen in developing countries (12). Germline mutations in *BRCA1* and *BRCA2* genes account for only 5% of all breast cancer cases in the general population (13,14). Other low-penetrance genetic variants, especially in as-yet unknown combinations, are expected to explain most breast cancer incidence (15). Investigators, including those in our group, have hypothesized that the 3' UTRs of miRNA target genes may harbor such important variants (10,16). Saunders *et al* (17) conducted a bioinformatic survey of the human genome for SNPs in putative miRNA target sites and found an appreciable level of variations within predicted as well as experimentally verified miRNA targets. One of the SNPs highlighted in the study of Saunders *et al* was SNP rs1042538 in the *IQGAP1* gene (encoding IQ motif-containing GTPase-activating protein 1), and this SNP has been experimentally verified to disrupt a miRNA target site sequence for miRNA-124 (miR-124) (18).

The scaffold protein IQGAP1 integrates signaling pathways and participates in diverse cellular activities that are important for both normal development and diseased states (19-24). IQGAP1 has attracted attention from cancer and developmental biologists because IQGAP1 expression appears to play an opposite role in normal development and in cancer progression. Studies with human tumor tissues have suggested that *IQGAP1* is an oncogene, which is overexpressed in a number of human solid neoplasms, including cancers of the colon, ovaries, stomach, and breast as well as glioblastoma (25-29). Functional studies have established the fact that IQGAP1 interacts with and regulates the actin-Cdc42/Rac1-mitogen-activated protein kinase pathway, thus contributing to its role in cell migration and invasion (19). Furthermore, another report suggested that IQGAP1 is involved in the expansion of cancer stem cells in glioblastoma and that together with IGFBP2 was associated with shorter survival in glioblastoma patients (27). Similarly, IQGAP1 was shown to be a marker of poor prognosis in ovarian cancer (26). In gastric cancer, it was shown that IQGAP1 was up-regulated by gene amplification (30).

However, the putative oncogenic properties of IQGAP1 have been contradicted by the finding that the deletion of *IQGAP1* in mice resulted in hyperplasia in gastric epithelial cells (31), suggesting that the *IQGAP1* gene plays an important role in normal development and actually has an antiproliferative function in normal epithelial cells. In other words, the *IQGAP1* gene may have a tumor suppression function in normal cells but may turn into an oncogene in tumor cells through an unknown mechanism. These conflicting properties of IQGAP1 suggest that IQGAP1 function may be dependent on developmental stage and cell type. However, determining the exact role and regulation of IQGAP1 in normal and cancer development will require more extensive studies, including population-based and functional studies.

A large number of SNPs have been found within the sequence of the *IQGAP1* gene, but SNP rs1042538 is recognized as the only one targeted by a miRNA with a high frequency of variation in Chinese population. This SNP (A/T variant) is one of the seven consecutive nucleotides

corresponding to the seed region of miR-124 (Fig. 1). The A allele, together with the other six, form a perfect pairing with the seed, which is responsible for the down-regulation of IQGAP1 by miR-124 (17). Therefore, we conducted a case-control study to investigate whether this SNP at the miRNA-binding site on the IQGAP1 gene plays a role in breast cancer development and prognosis in a Chinese population.

Patients and methods

Patients and controls. Study patients were recruited from the Breast Cancer Research Center in Tianjin Medical University Cancer Hospital, and clinical information was acquired from the Tianjin Cancer Registry upon the approval of the Institutional Review Board (32). This study included 1,541 patients with newly diagnosed and histologically confirmed breast cancer, who were consecutively recruited between January 1, 2007 and February 28, 2008. The response rate of the eligible patients we recruited was ~95%.

We also recruited 1,598 cancer-free women (controls) during this study period who were genetically unrelated to one another and living in the nearby community. The response rate of the eligible controls who were approached for recruitment was ~90%. The controls were frequency-matched to the cases by age (± 5 years).

After the study participants signed an informed consent form, they were interviewed for demographic data and information about major risk factors, including family history. For the cases, we also collected information about tumor features and disease severity, including morphologic characteristics, mean age at diagnosis, infiltrating/invasive ductal carcinoma (IDC) status, tumor size, presence of lymph node and/or organ metastasis, clinical stage, and estrogen receptor (ER) and progesterone receptor (PR) status. Each eligible subject donated 20 ml of blood, which was collected into heparinized tubes and used for biomarker assays, including DNA extraction and genotyping.

Genotyping. From each blood sample, a leukocyte cell pellet, obtained from the buffy coat by centrifugation of 1 ml of whole blood, was used for DNA extraction. Genomic DNA was isolated with the Qiagen DNA Blood Mini Kit (Qiagen Inc., Valencia, CA) according to the manufacturer's instructions. For genotyping the rs1042538 A/T SNP, both the amplifying primers and the TaqMan MGB probes were designed for the TaqMan SNP Genotyping assays (Applied Biosystems, Foster City, CA). More than 10% of the samples were randomly selected for repeated assays, and the results were 100% concordant.

Quantitative measurement of *IQGAP1* mRNA and protein expression. Total RNA was isolated from 37 frozen breast cancer tissues from patients with known genotypes of *IQGAP1* (AA or TT), as determined from their blood samples. The extraction and purification of total RNA were performed with TRIzol reagent (Invitrogen) according to the manufacturer's instructions. RNA quality and concentration were determined with the Agilent 2100 Bioanalyzer (Agilent Technologies). Real-time quantitative polymerase chain reaction (PCR) was performed in a 96-well reaction plate (MicroAmp® Optical

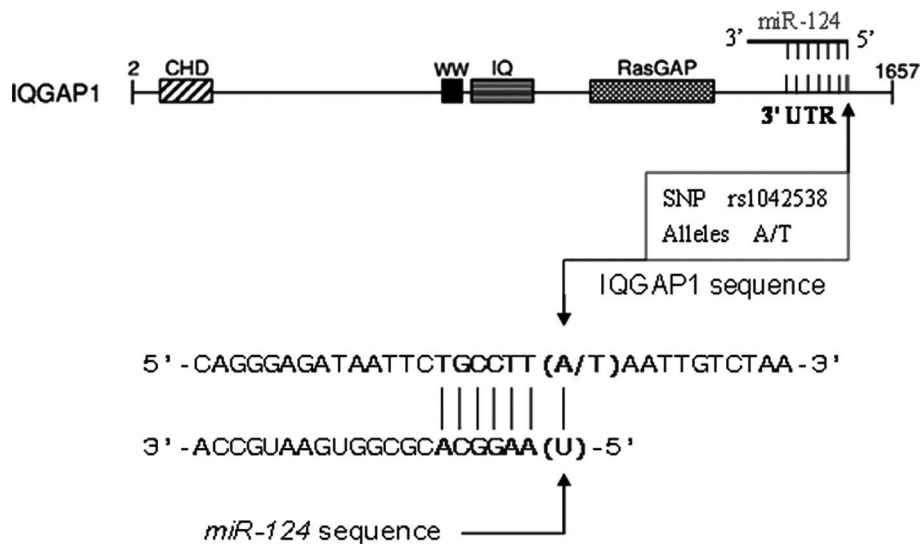


Figure 1. The base-pairing of the miR-124 seed region and the 3'-UTR of the *IQGAP1* gene, including the rs1042538 polymorphism. SNP rs1042538 has an A/T variation, which is one of the seven consecutive nucleotides corresponding to the seed region of miR-124. The A allele, together with the other six, forms a perfect pairing with the seed, which is responsible for the down-regulation of IQGAP1 by miR-124; the T allele forms a non-perfect pairing with the seed and may escape the regulation of miR-124.

96-Well Reaction Plate, Applied Biosystems) on an ABI PRISM® 7500 Sequence Detector System (Applied Biosystems), according to the manufacturer's instructions.

RT-PCR for IQGAP1 expression was done using power SYBR Green one-step RT-PCR master mix reagent kit (P/N 4391178). All primers were synthesized by Sangon Corp. (Shanghai, China). In order to normalize the differences in the amount of total RNA used in each reaction, glyceraldehyde-3-phosphate dehydrogenase (GAPDH) RNA was measured as endogenous control. Each sample was analyzed in duplicate and the coefficient of variation of all reactions was <5%. The relative expression level of IQGAP1 to GAPDH was described using the equation: expression = $2^{-\Delta Ct}$. Western blotting was done to evaluate the effect of miR-124 on the expression of IQGAP1 in cell lines, MM231 and LN299, using actin as a control. IQGAP1 protein expression was measured through Western blotting using actin as a loading control in 48 breast cancer tissues with IQGAP1 AA or TT genotype.

Statistical analyses. We used the χ^2 test to compare differences in frequency distributions of demographic variables, risk factors, and alleles of the *IQGAP1* polymorphism between the cases and controls. We also tested the Hardy-Weinberg equilibrium of genotype distributions in the controls. In addition, we used unconditional univariate and multivariate logistic regression analyses to examine the association between the SNP and breast cancer risk by estimating odds ratios (ORs) and 95% confidence intervals (CIs) with and without adjustment for age and other risk factors. Finally, we stratified the genotype data according to age, family history, and clinical variables (including morphologic characteristics, tumor size, presence of lymph node and/or organ metastases, tumor stage, and ER and PR status) of breast cancer patients by using the χ^2 test and logistic regression. The expression levels of IQGAP1 between AA and TT genotypes were compared with use of the rank-sum test. All statistical analyses were two-sided and

performed with use of SAS software (version 9.0; SAS Institute, Cary, NC), and a P=0.05 was considered statistically significant.

Results

Case-control analysis of the *IQGAP1* SNP. This case-control study included 1,541 breast cancer cases and 1,598 controls, and the distributions of known risk factors between cases and controls are shown in Table I. Age was adequately matched between cases and controls (P=0.242).

Genomic DNA for *IQGAP1* SNP genotyping was isolated from the peripheral blood of all participants with use of the TaqMan assay. The distribution of the *IQGAP1* genotypes (rs1042538) is shown in Table II. The SNP was in Hardy-Weinberg equilibrium in controls (P=0.469). The AA genotype was found in 33.42% of cases and 32.17% of controls, whereas the TT genotype was found in 17.26% of cases and 19.52% of controls. The TT genotype, compared with the AA genotype, was associated with a significantly lower risk of developing breast cancer (P=0.049, OR, 0.78; 95% CI, 0.61-0.99). In other words, women with the AA genotype were more likely than those with the TT genotype to develop breast cancer.

We also separated the cases into those with IDC (about 2/3 of all cases) and others (mainly including adenocarcinoma, carcinoma simplex, mucinous carcinoma, medullary carcinoma, lobular carcinoma). The two groups of cases were used in case-control analyses, and results revealed that in non-IDC cases only, those with the TT genotype had a significantly lower risk of developing cancer (OR=0.69; 95% CI, 0.50-0.94) than did those with the AA genotype (Table II). Thus, it appears that the effect of the *IQGAP1* SNP is less apparent in more aggressive and invasive stages of breast cancer, when many genetic and epigenetic factors become involved.

Case-only analysis of *IQGAP1* genotypes. The patient data we collected included information on mean age at diagnosis,

Table I. Frequency distributions of selected variables in breast cancer cases and cancer-free controls in Chinese women.

Variables	No. (%) of subjects		OR (95% CI)	P-value ^a
	Cases (n=1,541)	Controls (n=1,598)		
Age (years)				
≤50	739 (47.96)	733 (45.87)		0.242
>50	802 (52.04)	865 (54.13)		
Frequency of pregnancy				
≤2	710 (46.07)	728 (45.56)	1.00	0.771
>2	831 (53.93)	870 (54.44)	0.94 (0.80-1.11)	
Duration of breastfeeding (months)				
≤12	733 (47.57)	604 (37.80)	1.00	<0.001
>12	808 (52.43)	994 (62.20)	0.66 (0.56-0.77)	
Menopause ^c				
No	731 (47.65)	706 (44.54)	1.00	0.081
Yes	803 (52.35)	879 (55.46)	0.81 (0.64-1.03)	
Oral contraception ^c				
Never	1,197 (81.87)	1,321 (84.46)	1.00	0.057
Ever	265 (18.13)	243 (15.54)	1.14 (0.93-1.41)	
Smoking status ^c				
Never	1,309 (87.85)	1,478 (93.25)	1.00	<0.001
Ever	181 (12.15)	107 (6.75)	2.23 (1.70-2.93)	
Benign breast disease ^c				
Never	1,129 (73.79)	1,465 (92.78)	1.00	<0.001
Ever	401 (26.21)	114 (7.22)	4.41 (3.49-5.58)	
Family history of cancer ^{b,c}				
No	1,061 (68.90)	1,409 (88.45)	1.00	<0.001
Yes	479 (31.10)	184 (11.55)	3.24 (2.65-3.95)	

CI, confidence interval; OR, odds ratio. ^aTwo-sided χ^2 test. ^bFirst- and second-degree relatives. ^cDue to missing values, the number of cases and controls are less than 1,541 and 1,598, respectively.

IDC status, tumor size, presence of lymph node and/or organ metastasis, clinical stage, and ER/PR status. Our analysis did not reveal association between any genotypes and mean age at diagnosis ($P=0.387$), IDC status (IDC vs. non-IDC, $P=0.194$), tumor size ($P=0.821$), lymph node metastases ($P=0.652$), clinical stage ($P=0.773$), or ER status ($P=0.386$) (Table IV). Our analysis revealed significant association between genotypes and PR status of cases. Specifically, cases with the TT genotype were more likely to be PR-positive than were those with the AA genotype (OR, 1.35; 95% CI, 1.00-1.83) (Table III). Furthermore, in ER-negative cases, the IQGAP1 genotype was significantly associated with PR status. Cases with AT or TT genotypes, compared with the AA genotype, were more likely to be

PR-positive ($P=0.022$) (OR for AT genotype, 1.50; 95% CI, 1.01-2.23; OR for TT genotype, 1.91; 95% CI, 1.18-3.09) (Table IV).

Analysis of IQGAP1 expression. Our hypothesis was that IQGAP1 TT genotype, evading the regulation of miR-124, would have a higher level of IQGAP1 expression. We thus measured IQGAP1 expression at mRNA and protein levels using breast cancer tissue. IQGAP1 mRNA was higher in AA genotype than in TT genotype (0.040 vs. 0.025), the difference between the AA genotype and the TT genotype was not significant (rank-sum test, $P=0.109$) (Fig. 2A). The relative IQGAP1 protein levels were higher in TT genotype than in

Table II. Multivariate-adjusted ORs for risk of breast cancer associated with the *IQGAP1* polymorphism.

Genotype	No. (%) of all subjects			No. (%) of patients with IDC			No. (%) of patients with other morphology		
	Cases (n=1,541)	Controls (n=1,598)	OR ^a (95% CI)	Cases (n=1,065)	Controls (n=1,598)	OR ^a (95% CI)	Cases (n=476)	Controls (n=1,598)	OR ^a (95% CI)
<i>IQGAP1</i> (A/T)									
AA	515 (33.42)	514 (32.17)	1.00	345 (32.39)	514 (32.17)	1.00	170 (35.71)	514 (32.17)	1.00
AT	760 (49.32)	772 (48.31)	0.96 (0.79- 1.15)	525 (49.10)	772 (48.31)	1.01 (0.85-1.21)	235 (49.37)	772 (48.31)	0.92 (0.73-1.16)
TT	266 (17.26)	312 (19.52)	0.78 (0.61-0.99)^b	195 (18.31)	312 (19.52)	0.93 (0.74-1.17)	71 (14.92)	312 (19.52)	0.69 (0.50-0.94)^b
Trend test	P=0.160			P=0.604			P=0.026		

CI, confidence interval; OR, odds ratio. ^aORs are adjusted for age, duration of breastfeeding, menopause, oral contraception, smoking status, benign breast disease, and family history of cancer. ^bStatistically significant.

AA genotype (125.46 vs. 51.35), the difference between the TT genotype and the AA genotype was statistically significant (rank sum test, P=0.039) (Fig. 2B).

Discussion

We began this case-control study to find an answer to the question of whether the SNP at the miR-124 binding site on the 3'-UTR of the cytoskeleton-organizing gene *IQGAP1* had any effect on breast cancer risk. We did not have an a priori prediction because the published results on the function of *IQGAP1* appeared to be contradictory. On the one hand, removal of the *IQGAP1* gene predisposed mice to the development of gastric cancer. On the other hand, *IQGAP1* expression was elevated in many cancers and in some tumors correlated with poor prognosis. Our case-control study showed that the AA genotype of the miR-124 binding site SNP on *IQGAP1*, which renders its down-regulation by miR-124, was associated with increased risk of breast cancer. Although this does not prove that *IQGAP1* plays a role in tumor suppression, the result is consistent with the knockout mouse studies that suggest that *IQGAP1* is a negative regulator for cancer development. We also examined *IQGAP1* expression in breast cancer tissues and their adjacent pathologically normal tissues and found that indeed, *IQGAP1* was up-regulated in breast cancer tissue, similar to findings in other previously reported studies (33). Moreover, *IQGAP1* knockdown experiments showed that reduction of *IQGAP1* resulted in increased cell growth in non-cancer breast epithelial cells but resulted in decreased cell growth in breast cancer cells, supporting that *IQGAP1* plays an opposite role in normal development and cancer. Or in other words, *IQGAP1* may indeed act as a tumor suppressor in breast cancer initiation but act as an oncogene after cancer has developed. Therefore, our findings may help explain the apparently contradictory results regarding *IQGAP1* in the same human populations, at least partially removing the argument that *IQGAP1* may work differently in humans and mice.

The A allele, which binds perfectly with miR-124 and leads to the down-regulation of *IQGAP1* expression, is associated with increased risk of breast cancer. This finding suggests that *IQGAP1*, and by extension cytoskeleton organization, is critical for normal breast cell development. A lack of *IQGAP1* and disrupted cytoskeleton organization contributed to breast cancer development or hyperplasia in gastric epithelial cells in the mouse studies (31), although it is not known whether there were any abnormalities in breast epithelial cells in the *IQGAP1* gene knockout mice or whether the A allele of our studied SNP had any effect on gastric cancer development. Nevertheless, because cytoskeleton structure is critical for maintaining highly organized and polarized epithelia cell sheets, the requirement for sufficient *IQGAP1* expression is quite logical. Further case-control studies are clearly warranted in other types of cancer.

Our population-based case-control study suggests that the T allele of the miR-124 binding site SNP on the *IQGAP1* gene is a 'good' variant that has a breast cancer protection function. Of interest, the T allele was found in frequencies of 40, 10, and 10%, respectively, for the HapMap panels of Asian, European, and African women. It is well established that breast cancer

Table III. Association of the *IQGAP1* polymorphism with clinical characteristics of breast cancer cases.

Variables	No. of patients	IQGAP1 rs1042538 (A/T) genotype frequency			P-value ^a
		AA (n=515)	AT (n=760)	TT (n=266)	
Age at diagnosis (years)	1,540 ^b	51.61±10.50	52.13±11.11	51.14±10.18	0.387
Morphology	1,541				
IDC		345 (66.99)	525 (69.08)	195 (73.31)	0.194
Others		170 (33.01)	235 (30.92)	71 (26.69)	
OR (95% CI)		1.00	0.91 (0.72-1.15)	0.74 (0.53-1.03)	
Tumor size	1,395 ^b				
≤2 cm		171 (36.93)	242 (35.12)	87 (35.80)	0.821
>2 cm		292 (63.07)	447 (64.88)	156 (64.20)	
OR (95% CI)		1.00	1.08 (0.85-1.38)	1.05 (0.76-1.45)	
Lymph node metastases	1,534 ^b				
No		309 (60.47)	473 (62.48)	159 (59.77)	0.652
Yes		202 (39.53)	284 (37.52)	107 (40.23)	
OR (95% CI)		1.00	0.92 (0.73-1.16)	1.03 (0.76-1.39)	
Clinical stage	1,394 ^b				
0+I		122 (26.41)	193 (27.97)	63 (26.03)	0.773
II+III+IV		340 (73.59)	497 (72.03)	179 (73.97)	
OR (95% CI)		1.00	0.92 (0.71-1.21)	1.02 (0.72-1.45)	
ER	1,515 ^b				
-		224 (44.09)	314 (42.32)	125 (47.17)	0.386
+		284 (55.91)	428 (57.68)	140 (52.83)	
OR (95% CI)		1.00	1.08 (0.86-1.35)	0.88 (0.66-1.19)	
PR	1,515 ^b				
-		241 (47.44)	314 (42.32)	106 (40.00)	0.085
+		267 (52.56)	428 (57.68)	159 (60.00)	
OR (95% CI)		1.00	1.23 (0.98-1.54)	1.35 (1.00-1.83)^c	

CI, confidence interval; ER, estrogen receptor; IDC, infiltrating ductal carcinoma; OR, odds ratio; PR, progesterone receptor. ^aTwo-sided χ^2 test for difference in frequency distribution of variables between *IQGAP1* genotypes. ^bDue to missing values, n is <1,541. ^cStatistically significant.

incidence is much lower among Asian women than among women of European or African descent. Such differences have often been attributed to different lifestyles in the absence of known genetic factors. However, if the T allele is indeed protective, this allele and other similar alleles may, at least in part, constitute the genetic basis for the varying incidence in breast cancer among Asian, European, and African women. Future population-based studies in European and African women cohorts will be needed to test this hypothesis.

In our study, we also evaluated the potential association of this SNP with breast cancer prognosis. This was because

IQGAP1 has been shown to be overly expressed in cancer, and expression levels have been associated with poor prognosis in several other cancer types, including glioblastoma (34). This association is not surprising because enhanced cell migration, invasion, and metastasis of cancer cells require heightened cytoskeleton reorganizing activities. However, we were initially surprised with our analysis results that showed a lack of correlation of this SNP with major clinical parameters such as metastasis and tumor invasiveness in our breast cancer cases. We did observe some association with PR status, but the significance of this association is currently unclear. A few

Table IV. Association of the *IQGAP1* polymorphism with clinical characteristics of breast cancer in estrogen receptor-negative cases.

Variables	No. of patients	IQGAP1 rs1042538 (A/T) genotype frequency			P-value ^a
		AA (n=224)	AT (n=314)	TT (n=125)	
Tumor size	603 ^b				
≤2 cm		64 (32.00)	97 (33.68)	36 (31.30)	0.873
>2 cm		136 (68.00)	191 (66.32)	79 (68.70)	
OR (95% CI)		1.00	0.93 (0.63-1.36)	1.03 (0.63-1.69)	
Lymph node metastases	663				
No		129 (57.59)	199 (63.38)	78 (62.40)	0.381
Yes		95 (42.41)	115 (36.62)	47 (37.60)	
OR (95% CI)		1.00	0.79 (0.55-1.11)	0.82 (0.52-1.28)	
Clinical stage	603 ^b				
0+I		41 (20.60)	74 (25.61)	27 (23.48)	0.441
II+III+IV		158 (79.40)	215 (74.39)	88 (76.52)	
OR (95% CI)		1.00	0.75 (0.49-1.16)	0.85 (0.48-1.47)	
Morphology	663				
IDC		158 (70.54)	223 (71.02)	93 (74.40)	0.721
Others		66 (29.46)	91 (28.98)	32 (25.60)	
OR (95% CI)		1.00	0.98 (0.67-1.42)	0.82 (0.50-1.35)	
PR	662 ^b				
-		173 (74.76)	217 (70.21)	80 (63.30)	0.022
+		51 (25.24)	96 (29.79)	45 (36.70)	
OR (95% CI)		1.00	1.50 (1.01-2.23)^c	1.91 (1.18, 3.09)^c	

CI, confidence interval; IDC, infiltrating ductal carcinoma; OR, odds ratio; PR, progesterone receptor. ^aTwo-sided χ^2 test for difference in frequency distribution of variables between *IQGAP1* genotypes. ^bDue to missing values, n is <663, the total number of patients with AA, AT, or TT genotypes. ^cStatistically significant.

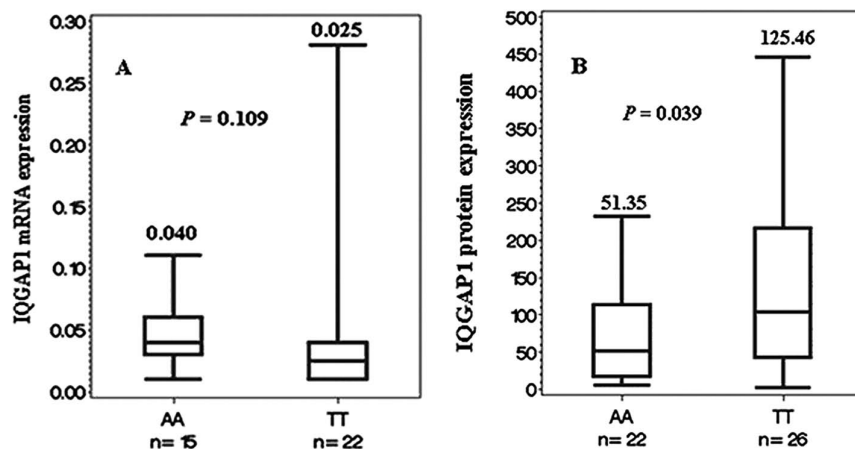


Figure 2. IQGAP1 expression in tumor tissue with IQGAP1 AA or TT genotypes. (A) Comparison of IQGAP1 expression at mRNA level between IQGAP1 genotypes (AA and TT) in breast cancer tissue. The difference of IQGAP1 mRNA between the TT genotype and the AA genotype was not significant (rank-sum test, $P=0.109$). (B) Comparison of IQGAP1 expression at protein level between IQGAP1 genotypes (AA and TT) in breast cancer tissue. IQGAP1 protein was higher in TT genotype than in AA genotype, the difference between the TT genotype and the AA genotype was statistically significant (rank-sum test, $P=0.039$).

other studies have also shown an association between genetic variations and PR status. In a case-control study with more than 9,000 subjects, Pooley *et al* (35) found that a coding SNP (rs3218536) in one of the DNA double-strand break repair genes, *XRCC2*, was strongly associated with risk of developing PR-positive breast cancer. In another study, Synowiec *et al* (36) reported an association between the *RAD51*-135G/C polymorphism and PR expression (OR=6.33; 95% CI=1.15-35.01).

Our results suggest a number of possibilities. First, IQGAP1 expression in breast cancer may not be strongly regulated by miR-124, and thus the SNP site for miR-124 binding may be irrelevant. Although we do not have direct evidence to support this possibility, one study found that miR-124 promoter is methylated in acute lymphoblastic leukemia (ALL) and that the expression of miR-124 in ALL is very low (37). Our analysis of miR-124 expression supported this observation that miR-124 is decreased in cancer tissue compared with adjacent normal tissue. A second scenario is that IQGAP1 expression levels may be less important to breast cancer because other genes in the same pathway are more activated. If this is true, then the expression of IQGAP1 should not be correlated with prognosis of breast cancer. We tested this hypothesis by examining the gene expression profile data in the public domain and showed that indeed, IQGAP1 expression levels in two breast cancer datasets did not correlate with grade or metastasis. We also showed that IQGAP1 expression did not correlate with grade in gastric cancer. In contrast, IQGAP1 expression levels were associated with grade and poor survival in gliomas, consistent with the literature.

In this study, we could not evaluate the association between IQGAP1 SNP genotypes and breast cancer survival, mainly because the breast cancer cases recruited for our study were relatively recent, which meant that there was not enough follow-up time for analysis of survival or distant metastasis. However, in the van't Veer *et al* (38) study, clinical follow-up of 78 breast cancer patients for more than 5 years after lumpectomy showed that IQGAP1 expression levels were not correlated with distance metastasis. Therefore, the role of IQGAP1 in cancer prognosis is apparently dependent on cancer type.

In conclusion, we have conducted the first epidemiologic study of the association between the IQGAP1 SNP within the miR124 binding site and breast cancer risk. Our study showed that the regulation of IQGAP1 could be complex and that the role of IQGAP1 in cancer cannot be generalized. However, the role of this SNP in cancer risk may be broader because of the different frequency distributions of demographic variables, risk factors, and alleles of the IQGAP1 polymorphism between Chinese women and women of other ethnic groups. Because of its potential significance, this study should be replicated with populations of various ethnic backgrounds. Once validated, this SNP may be important for genetic testing and screening of individuals at high risk of breast cancer.

Acknowledgments

We thank Hongwei Han, Lei Lei, and Yanrui Zhao for their technical assistance. We also thank Ms. Tamara Locke of the Department of Scientific Publications at M.D. Anderson

Cancer Center for editing this manuscript. This work was supported partially by grants from the National Natural Science Foundation of China (No. 30872172; No. 30771844), the Tianjin Science and Technology Committee Foundation (No.08ZCZGHZ02000;09ZCZDSF04700and08JCZDJC23600), Important National Science and Technology Specific Projects (2011ZX09307-001-04) and the Major State Basic Research Development Program of China (973 Program) (No. 2009CB918903). The tissue bank is jointly supported by the Tianjin Cancer Institute and Hospital and National Foundation for Cancer Research (US).

References

1. Zhang C: MicroRNomics: a newly emerging approach for disease biology. *Physiol Genomics* 33: 139-147, 2008.
2. Kim VN: Small RNAs: classification, biogenesis, and function. *Mol Cells* 19: 1-15, 2005.
3. Tsuchiya S, Okuno Y and Tsujimoto G: MicroRNA: biogenetic and functional mechanisms and involvements in cell differentiation and cancer. *J Pharmacol Sci* 101: 267-270, 2006.
4. Wiemer EA: The role of microRNAs in cancer: no small matter. *Eur J Cancer* 43: 1529-1544, 2007.
5. Vella MC, Reinert K and Slack FJ: Architecture of a validated microRNA:target interaction. *Chem Biol* 11: 1619-1623, 2004.
6. Evans SC, Kourtidis A, Markham TS, Miller J, Conklin DS and Torres AS: MicroRNA target detection and analysis for genes related to breast cancer using MDLcompress. *EURASIP J Bioinform Syst Biol*: 43670, 2007.
7. Vasudevan S, Tong Y and Steitz JA: Switching from repression to activation: microRNAs can up-regulate translation. *Science* 318: 1931-1934, 2007.
8. Shivdasani RA: MicroRNAs: regulators of gene expression and cell differentiation. *Blood* 108: 3646-3653, 2006.
9. Yu Z, Li Z, Jolicoeur N, *et al*: Aberrant allele frequencies of the SNPs located in microRNA target sites are potentially associated with human cancers. *Nucleic Acids Res* 35: 4535-4541, 2007.
10. Landi D, Gemignani F, Barale R and Landi S: A catalog of polymorphisms falling in microRNA-binding regions of cancer genes. *DNA Cell Biol* 27: 35-43, 2008.
11. Song F, Zheng H, Liu B, *et al*: An miR-502-binding site single-nucleotide polymorphism in the 3'-untranslated region of the SET8 gene is associated with early age of breast cancer onset. *Clin Cancer Res* 15: 6292-6300, 2009.
12. Jemal A, Siegel R, Ward E, Hao Y, Xu J and Thun MJ: Cancer statistics, 2009. *CA Cancer J Clin* 59: 225-249, 2009.
13. Chen YC and Hunter DJ: Molecular epidemiology of cancer. *CA Cancer J Clin* 55: 45-57, 2005.
14. McPherson K, Steel CM and Dixon JM: ABC of breast diseases. Breast cancer-epidemiology, risk factors, and genetics. *BMJ* 321: 624-628, 2000.
15. Pharoah PD, Tyrer J, Dunning AM, Easton DF and Ponder BA: Association between common variation in 120 candidate genes and breast cancer risk. *PLoS Genet* 3: e42, 2007.
16. Chen K, Song F, Calin G A, Wei Q, Hao X and Zhang W: Polymorphisms in microRNA targets: a gold mine for molecular epidemiology. *Carcinogenesis* 29: 1306-1311, 2008.
17. Saunders MA, Liang H and Li WH: Human polymorphism at microRNAs and microRNA target sites. *Proc Natl Acad Sci USA* 104: 3300-3305, 2007.
18. Lim LP, Lau NC, Garrett EP, *et al*: Microarray analysis shows that some microRNAs downregulate large numbers of target mRNAs. *Nature* 433: 769-773, 2005.
19. Noritake J, Watanabe T, Sato K, Wang S and Kaibuchi K: IQGAP1: a key regulator of adhesion and migration. *J Cell Sci* 118: 2085-2092, 2005.
20. Meyer RD, Sacks DB and Rahimi N: IQGAP1-dependent signaling pathway regulates endothelial cell proliferation and angiogenesis. *PLoS One* 3: e3848, 2008.
21. Wang Y, Wang A, Wang F, *et al*: IQGAP1 activates Tcf signal independent of Rac1 and Cdc42 in injury and repair of bronchial epithelial cells. *Exp Mol Pathol* 85: 122-128, 2008.
22. Ren JG, Li Z and Sacks DB: IQGAP1 integrates Ca²⁺/calmodulin and B-Raf signaling. *J Biol Chem* 283: 22972-22982, 2008.

23. Wang Y, Wang M, Wang F, *et al*: IQGAP1 promotes cell proliferation and is involved in a phosphorylation-dependent manner in wound closure of bronchial epithelial cells. *Int J Mol Med* 22: 79-87, 2008.
24. Mataraza JM, Briggs MW, Li Z, Entwistle A, Ridley AJ and Sacks DB: IQGAP1 promotes cell motility and invasion. *J Biol Chem* 278: 41237-41245, 2003.
25. Nabeshima K, Shimao Y, Inoue T and Koono M: Immunohistochemical analysis of IQGAP1 expression in human colorectal carcinomas: its overexpression in carcinomas and association with invasion fronts. *Cancer Lett* 176: 101-109, 2002.
26. Dong P, Nabeshima K, Nishimura N, *et al*: Overexpression and diffuse expression pattern of IQGAP1 at invasion fronts are independent prognostic parameters in ovarian carcinomas. *Cancer Lett* 243: 120-127, 2006.
27. Balenci L, Clarke ID, Dirks PB, *et al*: IQGAP1 protein specifies amplifying cancer cells in glioblastoma multiforme. *Cancer Res* 66: 9074-9082, 2006.
28. Johnson M, Sharma M and Henderson BR: IQGAP1 regulation and roles in cancer. *Cell Signal* (In press).
29. Nakamura H, Fujita K, Nakagawa H, *et al*: Expression pattern of the scaffold protein IQGAP1 in lung cancer. *Oncol Rep* 13: 427-431, 2005.
30. Walch A, Seidl S, Hermannstadter C, *et al*: Combined analysis of Rac1, IQGAP1, Tiam1 and E-cadherin expression in gastric cancer. *Mod Pathol* 21: 544-552, 2008.
31. Li S, Wang Q, Chakladar A, Bronson RT and Bernards A: Gastric hyperplasia in mice lacking the putative Cdc42 effector IQGAP1. *Mol Cell Biol* 20: 697-701, 2000.
32. Song F, He M, Li H, *et al*: A cancer incidence survey in Tianjin: the third largest city in China - between 1981 and 2000. *Cancer Causes Control* 19: 443-450, 2008.
33. Jadeski L, Mataraza JM, Jeong HW, Li Z and Sacks DB: IQGAP1 stimulates proliferation and enhances tumorigenesis of human breast epithelial cells. *J Biol Chem* 283: 1008-1017, 2008.
34. McDonald KL, O'Sullivan MG, Parkinson JF, *et al*: IQGAP1 and IGFBP2: valuable biomarkers for determining prognosis in glioma patients. *J Neuropathol Exp Neurol* 66: 405-417, 2007.
35. Pooley KA, Baynes C, Driver KE, *et al*: Common single-nucleotide polymorphisms in DNA double-strand break repair genes and breast cancer risk. *Cancer Epidemiol Biomarkers Prev* 17: 3482-3489, 2008.
36. Synowiec E, Stefanska J, Morawiec Z, Blasiak J and Wozniak K: Association between DNA damage, DNA repair genes variability and clinical characteristics in breast cancer patients. *Mutat Res* 648: 65-72, 2008.
37. Agirre X, Vilas ZA, Jimenez VA, *et al*: Epigenetic silencing of the tumor suppressor microRNA Hsa-miR-124a regulates CDK6 expression and confers a poor prognosis in acute lymphoblastic leukemia. *Cancer Res* 69: 4443-4453, 2009.
38. van't Veer LJ, Dai H, *et al*: Gene expression profiling predicts clinical outcome of breast cancer. *Nature* 415: 530-536, 2002.