

GSK-3 β -mediated fatty acid synthesis enhances epithelial to mesenchymal transition of TLR4-activated colorectal cancer cells through regulation of TAp63

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Abstract. Glycogen synthase kinase-3 β (GSK-3 β) in cancer cells is a critical regulatory component of both cellular metabolism and epithelial-mesenchymal transition (EMT) processes via regulation of the β -catenin/E-cadherin and phosphoinositide 3-kinase (PI3K)/AKT signaling pathway. Lipogenesis of cancer cells also plays a critical role in survival and metastasis. We investigated the role of GSK-3 β -mediated intracellular fatty acid synthesis to control EMT in TLR4-activated colorectal cancer cells and the underlying regulatory mechanism. Engagement of TLR4 with lipopolysaccharide (LPS) in colon cancer cells promoted the induction of phosphorylated GSK-3 β and related lipogenic enzymes as well as the expression of CD74, CD44 and macrophage inhibitory factor (MIF), but decreased expression of transcriptionally active p63 (TAp63). In addition, targeted inhibition of GSK-3 β using SB216763 was accompanied by decreased intracellular fatty acid synthesis and blockage of CD74 and CD44 expression, whereas it reversed the level of TAp63. Although TAp63 overexpression had no effect on the expression of CD74 and CD44 in LPS-treated colon cancer cells, GSK-3 β -dependent fatty acid synthesis and invasive activity were significantly suppressed. Notably, inhibition of CD44 or CD74 by siRNA not only attenuated *de novo* lipogenesis and migratory activity but also restored the expression of TAp63 in LPS-activated colon cancer cells. These results suggest that TAp63-mediated GSK-3 β activation induced by TLR4 stimulation triggers

migration and invasion of colon cancer cells through the regulation of lipid synthesis and GSK-3 β -mediated CD74/CD44 expression could be a target to control fatty acid-related EMT process through the modulation of TAp63 expression.

Introduction

GSK-3 β is believed to play an important regulatory role in cell metabolism, carcinogenesis and targeted therapy in cancer cells (1). GSK-3 β is expressed ubiquitously, regulated by diverse stimuli and involved in various signaling pathways, including antitumor protein p53 (TP53) activity (2,3). TP53 regulates Snail expression, which is also regulated by GSK-3 β . Epithelial-mesenchymal transition (EMT) is promoted through the activation of Snail protein expression after loss of TP53 activity (4). TP53 is normally a suppressor of Wnt/ β -catenin signaling, which is associated with activation of intracellular β -catenin regulated by GSK-3, resulting in loss of E-cadherin and mesenchymal phenotype (5-7). Expression of p63, a member of the p53 family, is regulated by the *TP63* gene (8), which is expressed as multiple isoforms with distinct properties. The transcriptionally active p63 (TAp63) isoform contains a transcription domain and can induce cell cycle arrest and apoptosis (9); however, the N-terminal-deleted p63 isoform (Δ Np63) has distinct functions, including induction of stem cell characteristics, progression of the cell cycle and enhancement of cell mobility (10). More invasive metastatic tumors exhibit decreased levels of p63 expression, suggesting that p63 loss upregulates the mechanism of tumorigenesis and metastatic spread (11,12). However, the relationship between GSK-3 β and TAp63, p53 family proteins in the migration and invasion of colon cancer cells remains unclear.

The expression of CD74 (major histocompatibility complex class II invariant chain) and CD44 are positively correlated with macrophage migration inhibitory factor (MIF) expression and lymph node metastasis in gastric cancer (13,14). The expression of CD74 significantly increases in gastrointestinal tumors compared to the level of normal tissue (15). Furthermore, stimulation of CD74 in colon cancer cells by MIF enhances cell survival and upregulates Bcl-2 expression (14).

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Engagement of MIF to CD74/CD44 induces upregulation of TAp63 expression, leading to tumor survival and progression (16). However, the role of TAp63 and its association with CD74/CD44 expression in the regulation of cancer metastasis in colon cancer cells remains unclear.

TAp63 also plays an important role in the regulation of energy metabolism: it accumulates in response to metabolic stress and transcriptionally activates NAD-dependent deacetylase sirtuin-1 (SirT1), adenosine monophosphate (AMP)-activated protein kinase (AMPK), and liver kinase B1 (LKB1), resulting in increased fatty acid synthesis (17). Fatty acid synthase (FASN) is selectively expressed in certain human cancers, including breast, stomach and colon cancer. The serum FASN levels of cancer patients in various clinical stages are significantly higher than those of healthy subjects (18). FASN expression is also controlled by p53 family proteins (19). The induction of FASN not only increases tumor growth and invasion but also confers resistance to chemotherapy (20,21). In addition, activation of GSK-3 β in murine embryonic fibroblasts induces the phosphorylation of acetyl-CoA carboxylase (ACC), one of the key intracellular lipogenic enzymes (22). The role of GSK-3 β , and its relationship to TAp63, in fatty acid synthesis for inducing EMT in colon cancer cells also remains unknown.

High levels of TLR4 expression and stimulation are associated with an increased incidence of metastasis and poor prognosis in colorectal cancer (23). Stimulation of TLR4 by lipopolysaccharide (LPS) activates the phosphatidylinositol 3-kinase (PI3K)/AKT (also known as protein kinase B) pathway, leading to an increase in metastasis of colon cancer cells (24,25). PI3K/AKT activation inhibits GSK-3 β function by phosphorylation at Ser-9 (26). The LPS-activated GSK-3 β signaling pathway is critical for carcinogenesis and metastasis of human cancers (27,28). However, the role of TAp63 in EMT processes and its association with GSK-3 β in TLR4-stimulated colorectal cancers (CRCs) remains unclear. Based on previous reports, we investigated the relationship between GSK-3 β and TAp63 and the role of these proteins in *de novo* fatty acid synthesis to regulate invasive activity in LPS-stimulated colon cancer cells. We also examined whether CD74/CD44 expression influences the activity of TAp63 and lipogenesis-mediated EMT of LPS-stimulated CRCs.

Materials and methods

Cell lines and reagents. The human colorectal cancer cell lines HCT-116, HCT-8 and HT-29 were purchased from the American Type Culture Collection (ATCC; Manassas, VA, USA). These cells were maintained in RPMI-1640 medium (Corning Incorporated, Corning, NY, USA), supplemented with 10% fetal bovine serum (FBS; RMBIO, Missoula, MT, USA), antibiotics and glutamine at 37°C in 5% CO₂. LPS and baicalein were obtained from Sigma-Aldrich (St. Louis, MO, USA). SB216763 was purchased from Selleckchem (Houston, TX, USA).

Western blotting. Cells were washed in phosphate-buffered saline (PBS) and lysed in NP-40 buffer (Elpis Biotech, Inc., Daejeon, Korea) supplemented with a protease inhibitor cocktail (Sigma-Aldrich). Protein phosphorylation states

were preserved through the addition of phosphatase inhibitors (Cocktail II; Sigma-Aldrich) to the NP-40 buffer. Protein concentrations were quantified using a BCA assay kit (Pierce, Rockford, IL, USA). Proteins (10 μ g/sample) were separated by 10-15% SDS-PAGE and transferred to nitrocellulose membranes (Millipore Corp., Billerica, MA, USA). The membranes were blocked with 5% skim milk prior to western blot analysis. Chemiluminescence was detected using an ECL kit (Advansta, Inc., Menlo Park, CA, USA) and the multiple Gel DOC system (Fujifilm, Tokyo, Japan). Primary antibodies against the following proteins were used: E-cadherin, N-cadherin, Snail, vimentin, TCF8/Zeb1, β -actin, MyD88, LKB1, SirT1, AceCS1, ACSL1, FASN, phospho-ACLY (Ser⁴⁵⁵), ACLY, phospho-ACC (Ser⁷⁹), ACC, phospho-AMPK (Thr¹⁷²), AMPK, phospho-GSK3 β (Ser⁹), GSK3 β , phospho-Lyn (Tyr⁵⁰⁷), Lyn, phospho-PI3K (Tyr⁴²⁸), PI3K, phospho-Akt (Ser⁴⁷³), Akt, phospho-Src (Tyr⁴¹⁶) and Src (Cell Signaling Technology, Beverly, MA, USA); TAp63 (BioLegend, Inc., San Diego, CA); α -SMA (Bioss, Inc., Woburn, MA, USA); FLAG (Sigma-Aldrich); CD74, CD44, phospho-PTEN (Ser³⁸⁰/Thr^{382/383}), and PTEN (Santa Cruz Biotechnology, Santa Cruz, CA, USA).

Small interfering RNA (siRNA) transfection. Experimentally verified human CD44-small interfering RNA (siRNA) duplex, CD74-siRNA duplex, and negative control-siRNA were obtained from Bioneer (Daejeon, Korea). Cells were seeded at a concentration of 1x10⁵/well in a 6-well plate and grown overnight prior to transfection with 200 nM siRNA using Lipofectamine RNAiMAX reagent (Invitrogen, Carlsbad, CA, USA) according to the manufacturer's instructions. Cells were used for further experiments 48 h after transfection.

TAp63 overexpression using transient transfection. Transient transfection of cultured cells was performed using Lipofectamine 2000 according to the manufacturer's instructions. Cells were plated at a density of 2x10⁵/well in a 6-well culture plate and transfected the next day. Typically, 10 ng of construct DNA was transfected with 9 μ l Lipofectamine. Cultured cells were transiently transfected with either a TAp63 expression vector of TAp63 cDNA cloned into pcDNA3.1 (Addgene, Cambridge, MA, USA) or empty vector pcDNA3.1 (Invitrogen). Cells were transfected for 48 h and analyzed by western blotting.

Quantification of human cytokines by ELISA. PGE2 and MIF concentrations in the culture supernatants were quantified using a single cytokine ELISA kit from Abcam (Cambridge, UK). VEGF and IL-8 were quantified by a single cytokine ELISA kit from R&D Systems (Minneapolis, MN, USA) according to the manufacturer's protocol. The data are expressed as the average of the number of biological replicates \pm standard deviation (SD).

Invasion assay. Cell invasion was determined using the CultreCoat 96-well Medium BME Cell Invasion Assay kit (R&D Systems) according to the manufacturer's protocol. Cells (2.5x10⁴) in serum-free RPMI-1640 containing 0.1% FBS were seeded into the upper chamber and the lower compartment was filled with RPMI-1640 containing 10% FBS as a chemoattractant. After incubation for 24 h, non-invading

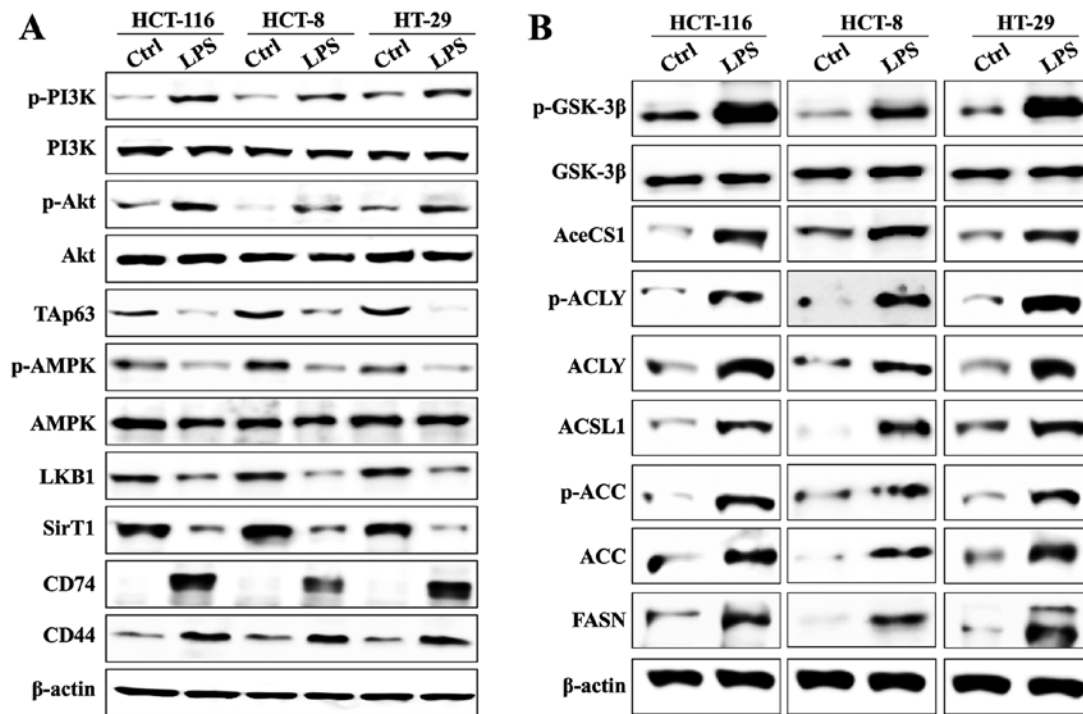


Figure 1. TLR4 stimulation increases the expression of CD74, CD44 and lipogenic enzymes in colon cancer cells. HCT-116, HCT-8 and HT-29 cells were treated with 500 ng/ml LPS for 24 h. Sterile distilled water was used as a vehicle control. Total cell lysates were subjected to western blot analysis with antibodies against (A) p-PI3K, PI3K, p-Akt, Akt, p-AMPK, AMPK, LKB1, SirT1, Tap63, CD74, CD44 and (B) p-GSK-3β, GSK-3β, AceCS1, p-ACLY, ACLY, ACSL1, p-ACC, ACC or FASN. β-actin served as an internal control. Data are representative of 3 independent experiments.

cells on the upper membrane surface were removed by wiping with a cotton swab. Invaded cells were stained with calcein-AM and quantified using a microplate reader.

Measurement of free fatty acid level. For assessment of free fatty acids, colorectal cancer cells were harvested, homogenized with chloroform/Triton X-100 solution (1% Triton X-100 in pure chloroform), and centrifuged at top speed to collect the organic phase (lower phase). The supernatant was vacuum dried to remove the chloroform and the dried lipid samples were dissolved in fatty acid assay buffer. Concentrations of free fatty acids were determined using the Free Fatty Acid Quantification kit (Abcam) according to the manufacturer's instructions.

Statistical analysis. Data were expressed as the mean ± standard deviation (SD). Statistical analysis was conducted using one-way analysis of the variance. A P-value <0.05 was considered statistically significant.

Results

TLR4 stimulation increases the expression of GSK-3β, CD74, CD44 and lipogenic enzymes in colon cancer cells. First, we investigated whether TLR4 stimulation affects the expression of Tap63-related signal molecules and the expression of lipogenic enzymes in CRC cells. LPS-treated colon cancer cells (HCT-116, HCT-8 and HT-29) showed activation of the PI3K/AKT signaling pathway, whereas phosphorylation of AMPK, LKB1 and SirT1, key metabolic regulators, was significantly reduced (Fig. 1A). Although the expression

of Tap63 was downregulated, upregulation of CD74 and CD44 was observed in LPS-activated HCT-116, HCT-8 and HT-29 cells (Fig. 1A). Enzymes associated with lipogenesis in CRC were activated after engagement of LPS with TLR4. Cytoplasmic acetyl-CoA synthetase (AceCS1), ATP-citrate lyase (ACLY), phospho-ACLY (p-ACLY), long-chain acyl-CoA synthetase (ACSL1), ACC, phospho-ACC (p-ACC), and fatty acid synthase (FASN) were all upregulated. In addition, LPS activation induced phosphorylation of GSK-3β in colon cancer cells (Fig. 1B). Our results suggest that the TLR4-mediated signaling pathway is one of the key regulators of metastasis-related mechanisms in colon cancer cells.

Inhibition of GSK-3β restores the expression of Tap63 and attenuates fatty acid synthesis, secretion of MIF and generation of PGE2 in LPS-activated colon cancer cells. To confirm the correlation of GSK-3β with lipogenic enzyme activity and invasion of LPS-activated colon cancer cells, the cells were pretreated with the GSK-3β inhibitor SB216763. Treatment with SB216763 blocked phosphorylation of GSK-3β and attenuated activation of lipogenic enzymes in LPS-treated colon cancer cells (Fig. 2A). The levels of intracellular long chain fatty acid increased >3-fold compared to non-treated control cells after stimulation with LPS. In addition, pretreatment with SB216763 significantly blocked fatty acid synthesis in LPS-activated colon cancer cells (Fig. 2B). Prostaglandin E₂ (PGE₂) increases the phosphorylation of GSK-3β and consequently stimulates β-catenin/T cell factor (TCF)-mediated transcription for growth and invasion in colon cancer (29). We observed that stimulation with LPS increased the secretion of PGE₂ in colon cancer cells; however, inhibition of GSK-3β

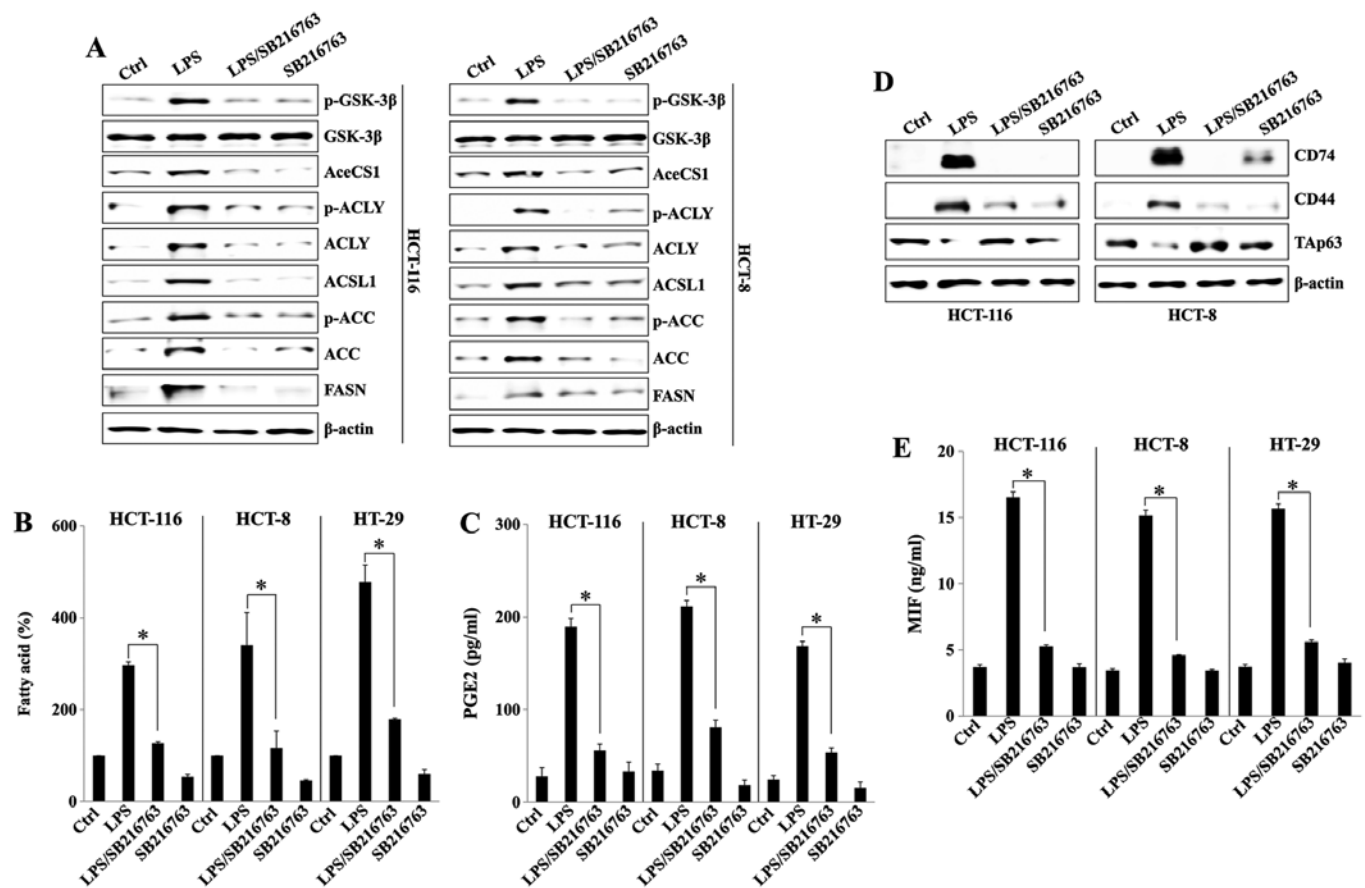


Figure 2. Inhibition of GSK-3 β attenuates fatty acid synthesis and secretion of MIF. The effect of GSK-3 β inhibitor on intracellular lipogenesis enzymes, fatty acid synthesis and PGE2 production was investigated. (A) Total protein was subjected to western blotting with antibodies against (A) p-GSK-3 β , GSK-3 β , AceCS1, p-ACLY, ACLY, ACSL1, p-ACC, ACC and FASN. (B) Production of intracellular long chain fatty acid levels in LPS-stimulated colon cancer cells compared to non-treated control cells was determined using a free fatty acid quantification kit. Data from at least three independent experiments performed in triplicate are represented as percentages of individual values in nmol/ μ l compared to control values (set at 100%) and are shown as means \pm SD. * P <0.05. (C) Concentrations of PGE2 in culture supernatants of HCT-116, HCT-8 and HT-29 cells were quantified by ELISA. Cells were seeded into 6-well plates (2×10^5 /well) and treated with 500 ng/ml LPS for 24 h. * P <0.01. Data are presented as the mean of three independent experiments and error bars represent SDs of the means. (D) Total protein was subjected to western blot analysis with antibodies against TAP63, CD44 and CD74. (E) Concentrations of MIF in the culture supernatants of HCT-116, HCT-8 and HT-29 cells were quantified by ELISA. Cells were seeded into 6-well plates (2×10^5 /well) and treated with 500 ng/ml LPS for 24 h. * P <0.01. Data are presented as the mean of three independent experiments and error bars represent the SD of the mean.

reduced the LPS-activated PGE2 production (Fig. 2C). Treatment with SB216763 effectively blocked the expression of CD74 and CD44 in LPS-treated colon cancer cells (Fig. 2D). SB216763 also reduced secretion of MIF in LPS-treated colon cancer cells (Fig. 2E). In addition, suppression of GSK-3 β recovered the expression of TAP63 in LPS-stimulated colon cancer cells (Fig. 2D). Our results suggest that GSK-3 β not only plays a critical role in lipogenesis and cancer metabolism, but also regulates the level of TAP63 in LPS-stimulated colon cancer cells.

Tap63 expression regulates GSK-3 β -dependent intracellular lipogenesis and invasion activity of LPS-treated colon cancer cells. Next, we investigated the role and connection of TAP63 in GSK-3 β -mediated lipogenesis and EMT processes in CRCs after LPS stimulation. After transfection with the TAP63-expressing plasmid, the levels of TAP63-related proteins did not significantly change compared to control cells or cells transfected with a control vector (Fig. 3A). TAP63 overexpression in LPS-activated HCT-116 cells not only blocked the attenuation of phosphorylated AMPK, LKB1 and SirT1, but

also prevented the upregulation of phosphorylated GSK-3 β and activation of lipogenic enzymes (Fig. 3B and C). Although, the upregulated CD74 and CD44 expression following LPS stimulation was maintained (Fig. 3B), the downregulation of lipogenic enzymes was accompanied by decreased intracellular fatty acid synthesis (Fig. 3D). Inhibition of GSK-3 β by TAP63 transfection also reduced the production of PGE2 (Fig. 3D). TAP63-expressing HCT-116 cells activated by LPS showed concomitant reduction of the mesenchymal markers N-cadherin, snail, vimentin, α -SMA and TCF8 and decreased migratory activity (Fig. 3E and F). Our results suggest that TAP63 expression regulates GSK-3 β -related intracellular lipogenesis and influences mesenchymal characteristics and invasion activity in LPS-activated CRCs.

TLR4-triggered CD74/CD44 induction contributes to lipogenesis-dependent EMT processes of colon cancer cells through the regulation of TAP63 expression. We found that treatment with SB216763 recovered the expression of TAP63 in LPS-activated colon cancer cells, whereas overexpression of TAP63 also reduced the levels of phosphorylated GSK-3 β .

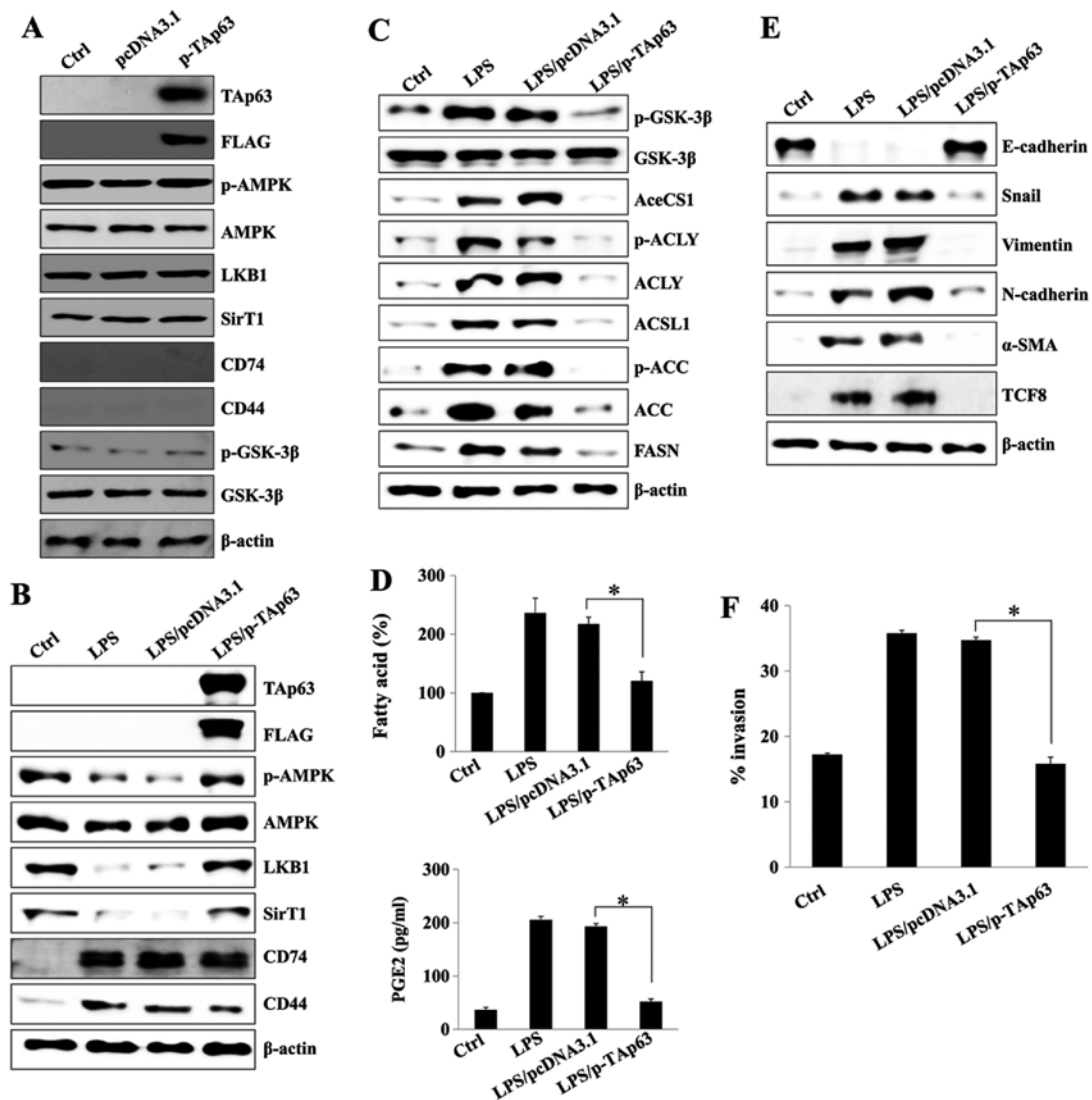


Figure 3. TAp63 expression regulates GSK-3 β -dependent intracellular lipogenesis and invasion activity of LPS-treated colon cancer cells. (A) The HCT-116 cells were transfected with either empty vector pcDNA3.1 or TAp63 expression vector. The cells were analyzed by western blotting with anti-TAp63 and anti-FLAG antibodies and then total protein was subjected to immunoblotting with antibodies against CD74, CD44, p-AMPK, AMPK, LKB1, SirT1, GSK-3 β and p-GSK-3 β . HCT-116 cells were treated with 500 ng/ml LPS for 24 h and then transfected with either empty vector pcDNA3.1 or TAp63 expression vector. (B and C) Total proteins in cell lysates were subjected to western blot analysis of TAp63, FLAG, CD74, CD44, p-AMPK, AMPK, LKB1, or SirT1 (B), and p-GSK-3 β , GSK-3 β , AceCS1, p-ACLY, ACLY, ACSL1, p-ACC, ACC or FASN (C). β -actin served as an internal control. (D) Production of intracellular long chain fatty acid levels in LPS-stimulated colon cancer cells was determined using a free fatty acid quantification kit and compared to that of non-treated control cells. Data from at least three independent experiments performed in triplicate are represented as percentage of individual values in nmol/ μ l compared to control values (set at 100%) and are shown as means \pm SD. * P <0.01. Concentration of PGE2 in the culture supernatants of HCT-116 cells was quantified by ELISA. Cells were seeded into 6-well plates (2×10^5 /well) and treated with 500 ng/ml LPS for 24 h. * P <0.01. Data are presented as the mean of three independent experiments and error bars represent SDs of the means. (E) Western blot analysis of EMT markers E-cadherin, N-cadherin, vimentin, Snail, α -SMA and TCF8/Zeb1. β -actin served as an internal control. (F) TAp63 overexpression decreased cell invasiveness detected by a BME cell invasion assay kit. Each value is the mean \pm standard deviation of 3 determinations. * P <0.01.

We estimated that another molecule plays an important role in regulating TAp63 expression and GSK-3 β -mediated lipogenesis. From previous results, targeted inhibition of GSK-3 β using SB216763 suppressed the expression of CD74 and CD44, but reversed the level of TAp63. Moreover, TAp63 overexpression and reduction of lipogenesis had no effect on the expression of CD74 and CD44 in LPS-treated colon cancer cells. Based on these results, we next evaluated whether LPS-induced expression of CD74 and CD44 affects the expression of TAp63 and phosphorylated GSK-3 β , the activation of lipogenic enzymes, fatty acid synthesis and MIF secretion. Although TLR4-stimulation of CD44 or CD74-knockdown HCT-116

cells induced PI3K/AKT activation, gene silencing of CD44 or CD74 with siRNA blocked the downregulation of TAp63, phosphorylated AMPK, LKB1 and SirT1 in LPS-treated HCT-116 cells (Fig. 4A and B). Expression of FASN and the production of long chain free fatty acids in LPS-activated HCT-116 cells were decreased after knockdown of CD44 or CD74 with siRNA (Fig. 4A-C). Secretion of PGE2 and MIF was also attenuated in CD44 or CD74 knockdown HCT-116 cells after stimulation with LPS (Fig. 4D and E). Expression of mesenchymal markers and invasion activity were significantly suppressed when induction of CD44 or CD74 expression was inhibited in LPS-stimulated HCT-116 cells (Fig. 5A

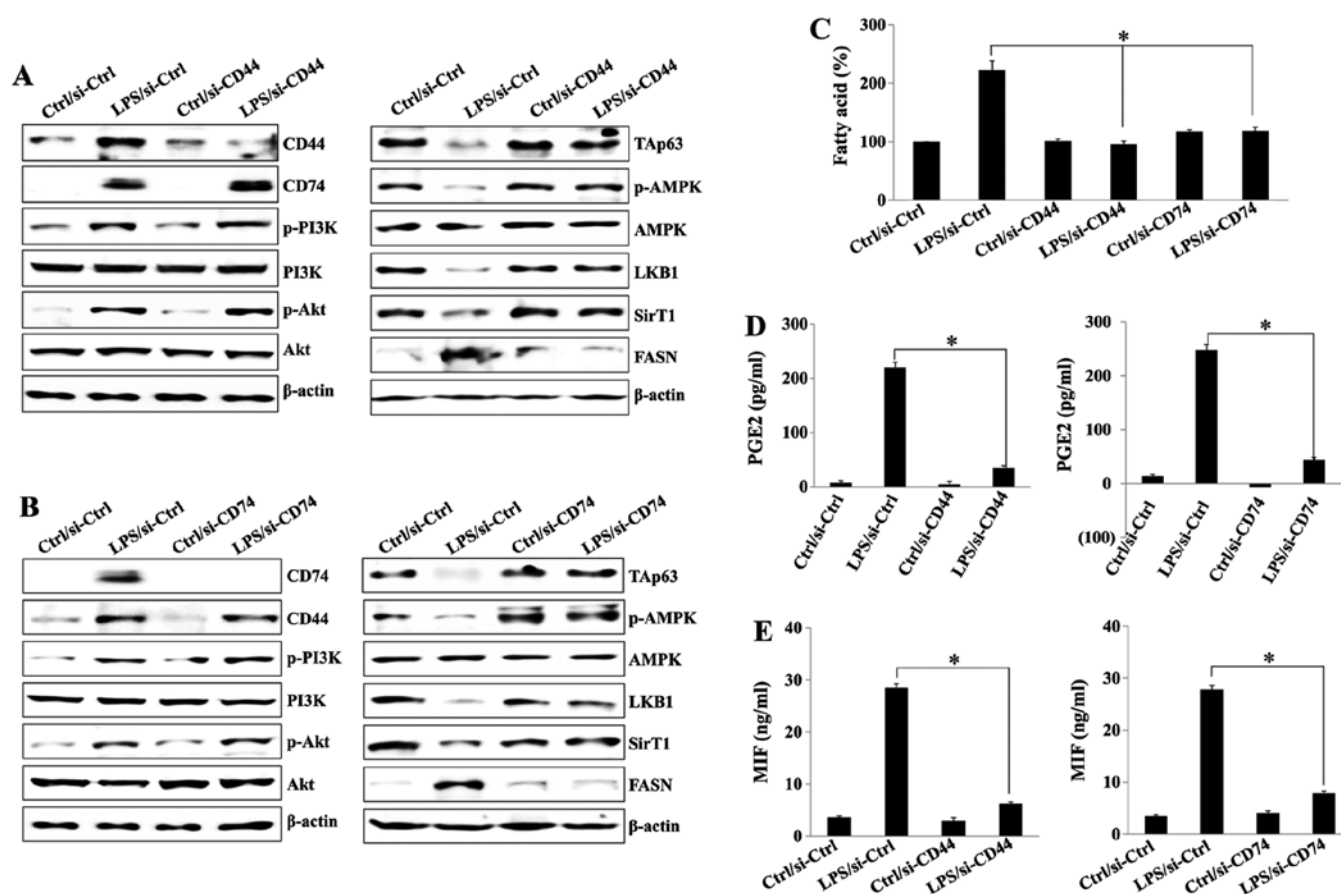


Figure 4. Induction of CD74 and CD44 by TLR4 stimulation contributes to fatty acid synthesis through PI3K/Akt activation. HCT-116 cells were treated with 500 ng/ml LPS for 24 h and then transfected with CD44-siRNA (200 nM), CD74-siRNA (200 nM), or control-siRNA for 48 h prior to experiments. (A and B) Total protein was subjected to western blot analysis with antibodies against CD44, CD74, p-PI3K, PI3K, p-Akt, Akt, Tap63, p-AMPK, AMPK, LKB1, SirT1 or FASN. β -actin served as an internal control. (C) Production of intracellular long chain fatty acids in LPS-stimulated colon cancer cells was determined using the free fatty acid quantification kit and compared to that of non-treated control cells. Data from at least three independent experiments performed in triplicate are represented as percentage of individual values in nmol/ μ l compared to control values (set at 100%) and are shown as means \pm SD. * P <0.01. (D and E) Concentrations of PGE2 (D) and MIF (E) in culture supernatants of HCT-116 cells were quantified by ELISA. Cells were seeded onto 6-well plates (2×10^5 /well) and treated with 500 ng/ml LPS for 24 h. * P <0.01. Data are presented as the mean of three independent experiments and error bars represent the SD of the mean.

and B). The production of EMT-related cytokines, including VEGF and IL-8, after LPS stimulation was also suppressed in HCT-116 cells transfected with CD44 or CD74 siRNA (Fig. 5C). Our results suggest that the CD74/CD44 expression of TLR4-stimulated colon cancer cells plays a critical role in the suppression of Tap63 as well as lipogenesis-related EMT processes.

Baicalein prevents migratory activity of LPS-treated colon cancer cells via modulation of Tap63/GSK-3 β -mediated cancer metabolism. Baicalein, a natural plant extract, suppresses EMT of breast epithelial cells and the tumorigenic activity of breast cancer cells (30). We previously reported that treatment with baicalein upregulates Tap63 expression, resulting in apoptosis of B-cell lymphoma cell lines (31). Treatment with baicalein also decreased the lipid level in the livers of mice fed a high-fat diet through PI3K/AKT-mediated AMPK activation (32). Based on these results, we investigated whether baicalein prevents the lipogenesis-related EMT processes through the regulation of Tap63-dependent GSK-3 β expression. Pre-exposure of LPS-activated HCT-116 cells to baicalein showed reduced phosphorylation of Syk/Src

tyrosine kinase, resulting in blockage of PI3K/AKT activation (Fig. 6A). Treatment with baicalein not only prevented the downregulation of Tap63 and phosphorylated AMPK, LKB1, and SirT1, but also suppressed the expression of CD44, CD74, and phosphorylated GSK-3 β in LPS-treated HCT-116 cells (Fig. 6B). The production of long chain fatty acid and PGE2 in TLR4-activated HCT-116 cells was efficiently blocked after treatment with baicalein (Fig. 6C and D). Expression of mesenchymal markers (Fig. 6E), production of MIF (Fig. 6F), and migratory activity (Fig. 6G) of LPS-stimulated HCT-116 cells were also inhibited by baicalein. Our results suggest that baicalein controls the EMT processes of LPS-stimulated CRCs through the regulation of Tap63/GSK-3 β -mediated lipogenesis.

Discussion

Epithelial to mesenchymal transition (EMT), a process in which epithelial cell layers lose their polarity through dramatic remodeling of the cytoskeleton, plays an important role in tumor metastasis (33). TLR4 expressed in several types of tumor cells plays a vital role in carcinogenesis, metastasis and

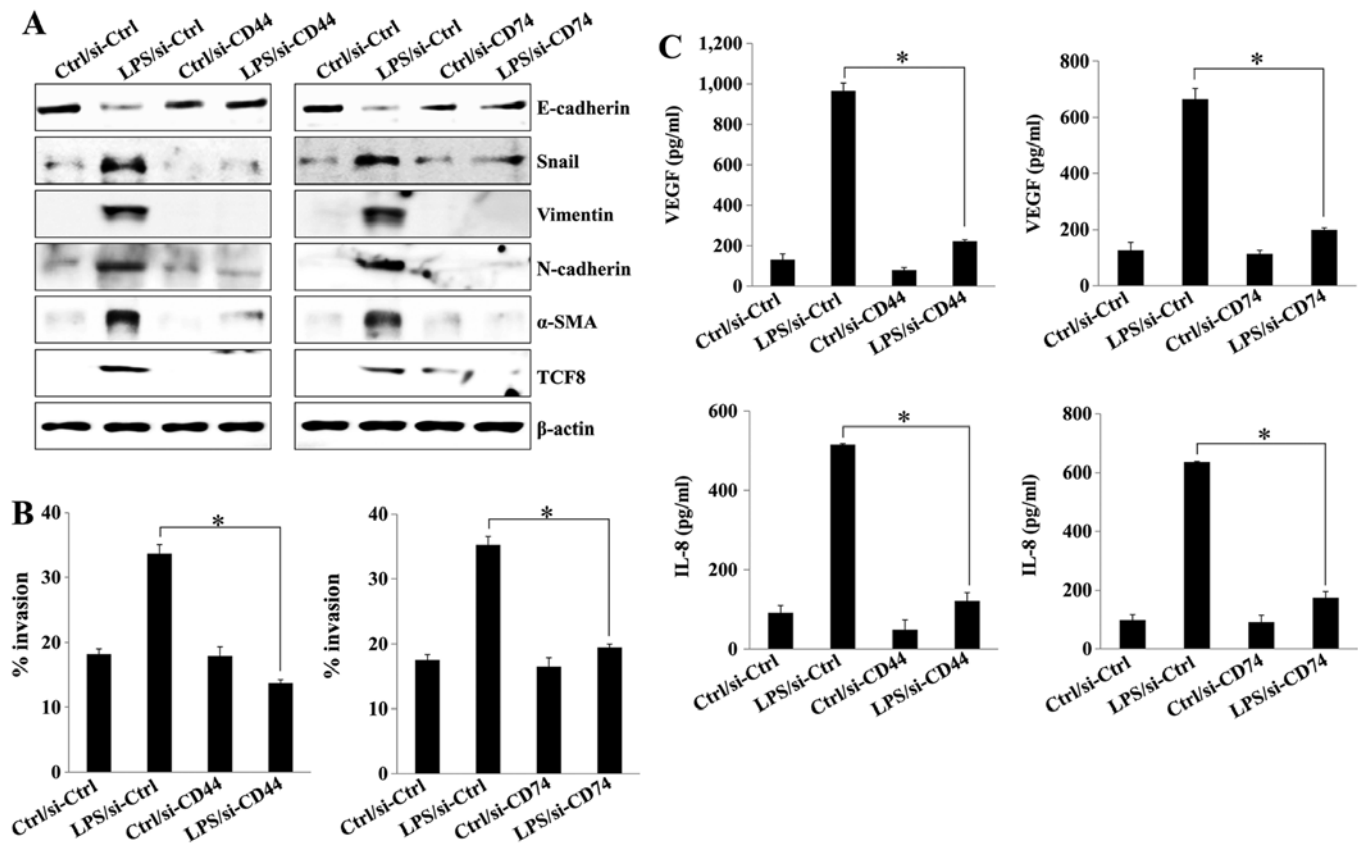


Figure 5. Induction of CD74 and CD44 by TLR4 stimulation contributes to EMT and invasiveness. HCT-116 cells were seeded into 6-well plates (2×10^5 /well), treated with 500 ng/ml LPS for 24 h, and then transfected with CD44-siRNA (200 nM), CD74-siRNA (200 nM), or control-siRNA for 48 h prior to experiments. (A) Western blot analysis of EMT markers (E-cadherin, N-cadherin, vimentin, Snail, α -SMA and TCF8/Zeb1). β -actin served as an internal control. (B) Gene silencing of CD44 or CD74 with siRNA transfection decreased the invasiveness of TLR4-stimulated HCT-116 cells. Migratory activity was detected using a BME cell invasion assay kit as described in Materials and methods. Each value is the mean \pm standard deviation of three determinations. * $P < 0.05$. (C) Concentrations of VEGF and IL-8 in culture supernatants of CD44- or CD74-knockdown HCT-116 cells were quantified by ELISA. * $P < 0.05$. Data are presented as the mean of three independent experiments and error bars represent the SD of the mean.

cancer progression (34). LPS enhances the invasive potential and induces EMT in HCC cells in a TLR4-dependent manner through activation of nuclear factor kappa B (NF- κ B) (28). Expression of p63 has been detected in several cancers, including bladder (35) and lung (36). Disruption of p63 in squamous cell lines results in upregulation of genes associated with increased invasiveness and metastasis in tumors (35). These results are consistent with the role of p63 as a marker of non-invasive epithelial tumors (37). Levels of p63 have wide-ranging influences on biological processes, including metabolism. TAp63 overexpressing cells are more glycolytically active than Δ Np63 cells, indicating that the two isoforms may regulate key steps of glycolysis in an opposite manner (38). However, the detailed regulatory mechanisms of TAp63 in colon cancer metastasis and its association with GSK-3 β -mediated intracellular lipogenesis remain obscure. We observed that TLR4-stimulated CRCs showed suppressed expression of TAp63, but upregulated levels of phosphorylated GSK-3 β and lipogenic enzymes. The migratory capacity and expression of EMT-related markers were also increased in a GSK-3 β -dependent manner after TLR4 activation. Overexpression of TAp63 significantly blocked GSK-3 β phosphorylation and fatty acid synthesis. Pharmacological inhibition of GSK-3 β also prevented lipogenesis. Our results

suggest that TLR4 activation in colon cancer cells enhances EMT-related intracellular lipid metabolism and migratory activity through modulation of TAp63-dependent GSK-3 β expression (Fig. 7).

FASN is highly expressed in many types of human cancer, including colorectal cancer (39). FASN levels in the serum of colorectal cancer patients are significantly higher than those of healthy individuals. Serum FASN levels in colorectal cancer patients are associated with tumor extent, liver metastasis and tumor clinical stage (39-41). Nuclear accumulation of GSK-3 β is also significantly correlated with poor prognosis, distant metastasis, and loss of membranous β -catenin colon cancer (42). However, the relationship between lipogenesis and GSK-3 β in TLR4-stimulated colorectal cancer remains unclear. The LKB1/AMPK signaling pathway regulates energy homeostasis through its ability to balance catabolic and anabolic activity to regulate lipid and glucose metabolism (43). Phosphorylation of the lipogenic enzyme ACC by AMPK is critical for the production of malonyl-CoA as a substrate for fatty acid biosynthesis. Expression of TAp63 is required for transcriptional activation of AMPK, LKB1 and Sirt1 (17). TLR4 stimulation of CRCs reduced TAp63 expression as well as activation of AMPK, LKB1 and Sirt1. In addition, we showed that upregulation of phosphorylated GSK-3 β by

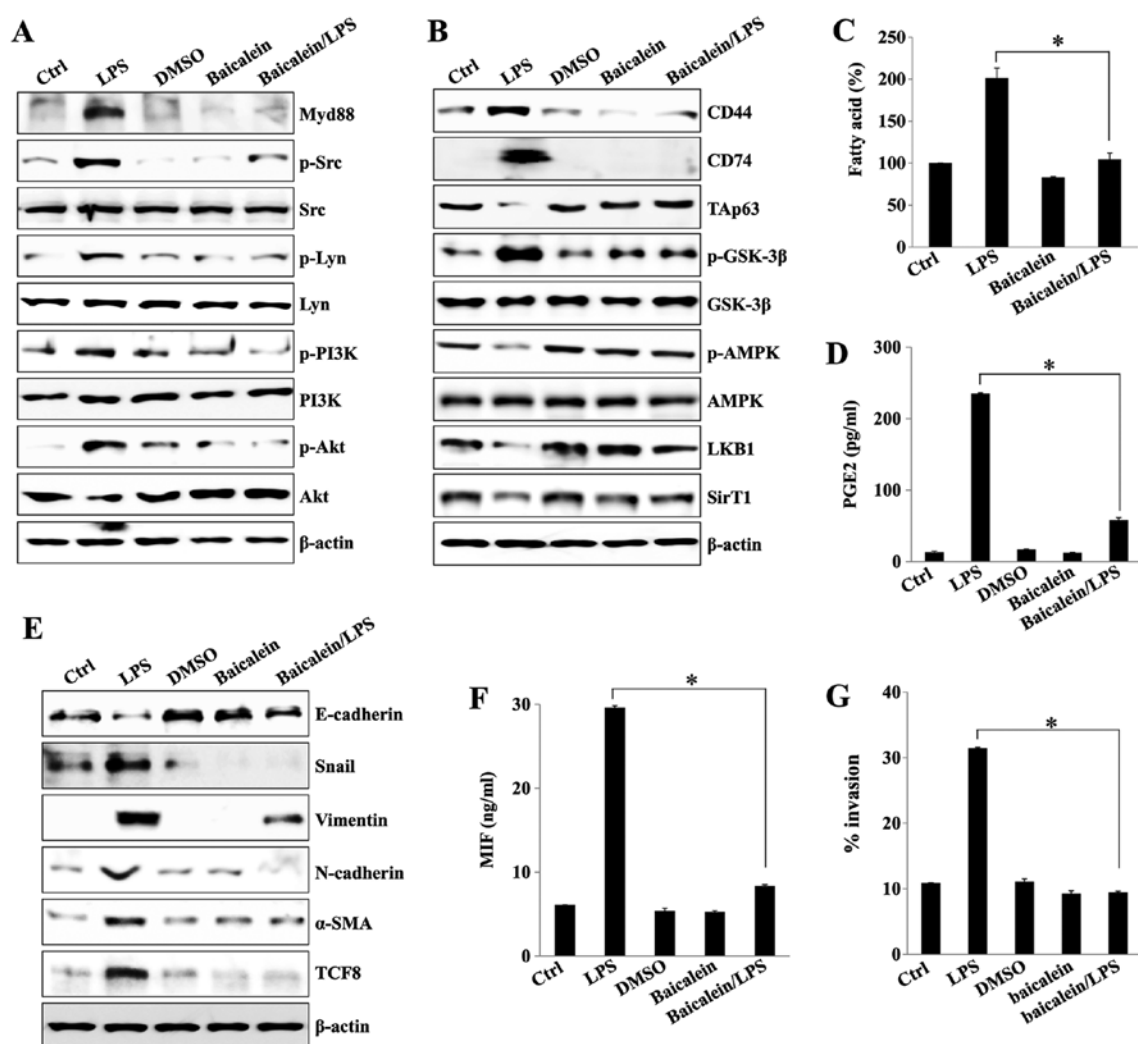


Figure 6. Baicalein inhibits migratory activity of LPS-treated colon cancer cells by modulation of GSK-3 β . HCT-116 cells were preincubated with 20 μ M baicalein for 2 h and then treated with 500 ng/ml LPS for 24 h. (A, B and E) Total cell lysates were subjected to western blotting with antibodies against (A) Myd88, p-Src, Src, p-Lyn, Lyn, p-PI3K, PI3K, p-Akt, Akt, (B) CD44, CD74, TAp63, p-GSK-3 β , GSK-3 β , p-AMPK, AMPK, LKB1, SirT1, and (E) E-cadherin, N-cadherin, vimentin, Snail, α -SMA or TCF8/Zeb1. β -actin served as an internal control. (C) Production of intracellular long chain fatty acids in LPS-stimulated colon cancer cells was determined using a free fatty acid quantification kit and compared to that of non-treated control cells. Data from at least three independent experiments performed in triplicate are represented as a percentage of individual values in nmol/ μ l compared to control values (set at 100%) and are shown as mean \pm SD. * P <0.05. (D and F) Concentrations of PGE2 (D) and MIF (F) in culture supernatants of HCT-116 cells were quantified by ELISA. Cells were seeded onto 6-well plates (2×10^5 /well) and treated with 500 ng/ml LPS for 24 h. * P <0.005. Data are presented as the mean of three independent experiments and error bars represent the SD of the mean. (G) Baicalein inhibited invasiveness following TLR stimulation, as detected by a BME cell invasion assay kit as described in Materials and methods. * P <0.005. Each value is the mean \pm standard deviation of 3 determinations.

LPS treatment triggered the stimulation of intracellular lipogenic enzymes, fatty acid synthesis and invasion activity of CRCs. Moreover, TAp63 overexpression suppressed GSK-3 β -mediated lipogenesis and cancer metastasis in LPS-activated colon cancer cells. These results suggest that TLR4-mediated TAp63 expression is associated with GSK-3 β phosphorylation and lipogenesis-related cancer progression.

CD74 is usually expressed at high levels by antigen-presenting cells (APCs), including B cells, monocytes, macrophages and dendritic cells in normal tissues (44). The TLR4 agonist LPS, a bacterial cell wall component, regulates CD44 expression in human monocytes (45). More recently, CD74 expression on epithelial cells has been shown to be related to chronic inflammation and carcinogenesis in the gastrointestinal tract (46). We observed that the levels of TAp63 and phosphorylated GSK-3 β of LPS-stimulated

colon cancer cells were expressed in an opposite manner. As expected, targeted inhibition of GSK-3 β with SB216763 prevented the induction of CD74, CD44 and MIF expression in LPS-activated CRCs. Moreover, overexpression of TAp63 failed to inhibit CD74/CD44 expression, but significantly suppressed intracellular lipogenesis. Based on these results, we investigated the association between CD74/CD44 and TAp63 as well as the role of CD74/CD44 on TAp63/GSK-3 β -mediated lipogenesis. Although gene silencing of CD44 or CD74 with siRNA reduced the expression of FASN and fatty acid production in addition to blocking invasion activity, downregulation of CD74 and CD44 failed to reduce the expression of TAp63 and related metabolic regulators, including AMPK, LKB1 and SirT1, in LPS-stimulated HCT-116 cells. These results suggest that GSK-3 β -dependent CD74/CD44 expression plays a role in downregulation of TAp63 expression and indirectly controls

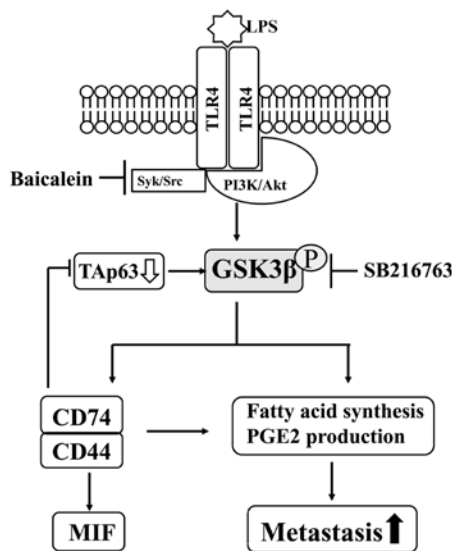


Figure 7. Schematic diagram of intracellular signaling mechanisms during LPS-induced TLR4 activation in human colorectal cancer cells. Interaction of LPS with TLR4 on colorectal cancer cells (HCT-116, HCT-8 and HT-29) transduces cellular signals through a common pathway that leads to activation of Syk/Src and PI3K/Akt. Subsequent downregulation of TAp63 by stimulation of TLR4 expression results in phosphorylation of GSK-3 β , upregulation of the MIF/CD74/CD44 signaling pathway, increased fatty acid synthesis, and PGE2 production, leading to metastasis of colon cancer cells. Treatment of colon cancer cells with baicalein rescues the induction of metastasis and GSK-3 β -mediated fatty acid synthesis by LPS-stimulated TLR activation. Our findings suggest that metastasis of colon cancer cells may be caused by decreased TAp63 expression and increased phosphorylation of GSK-3 β , upregulation of CD74/CD44 and production of PGE2 and MIF.

GSK-3 β -mediated intracellular lipid synthesis and EMT processes of LPS-activated colon cancer cells (Fig. 7).

Treatment of B-cell lymphoma cells with baicalein increases the expression of TAp63 to induce apoptosis and concomitantly reduces the levels of CD74 and CD44 (31). Baicalein treatment of LPS-stimulated HCT-116 cells also reversed the level of TAp63 and expression of related metabolic regulators. We also observed that baicalein reduced the activity of GSK-3 β -dependent lipogenic enzymes, CD74/CD44 expression, and fatty acid-related migratory activity in LPS-treated CRCs. These results suggest baicalein as a new candidate therapeutic drug for the control of lipogenesis-related colon cancer metastasis. TLR4 agonist, LPS derived from intestinal microflora, influences the growth of cancer cells (47). Conventional chemotherapy against colon cancer enhances the expression of TLR4 (48). Our results suggest that inhibition of the expression of CD44 or blocking CD44-mediated signaling pathway is critical for the conservation of TAp63 after treatment with anticancer drug. Based on these results, targeting inhibition of CD44 using specific agent or preventing the activation of CD44 using a newly developed drug, such as baicalein is important to control the metastasis of colon cancer *in vivo* animal model. Further studies are needed to examine these possibilities.

In conclusion, the present study demonstrates that the TAp63/GSK-3 β signaling pathway might be a critical regulator of lipogenesis-related cancer metastasis and that regulation of CD74/CD44 expression in colon cancer cells could be a promising target for TAp63-mediated cancer metastasis.

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