

Cancer-associated peritoneal mesothelial cells lead the formation of pancreatic cancer peritoneal dissemination

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Abstract. The interaction between the cancer cells and the peritoneal mesothelial cells (PMCs) plays an important role in the peritoneal dissemination in several types of cancer. However, the role of PMCs in the peritoneal dissemination of pancreatic cancer remains unclear. In the present study, we investigated the interaction between the pancreatic cancer cells (PCCs) and the PMCs in the formation of peritoneal dissemination *in vitro* and *in vivo*. The tumor-stromal interaction of PCCs and PMCs significantly enhanced their mobility and invasiveness and enhanced the proliferation and anoikis resistance of PCCs. In a 3D organotypic culture model of peritoneal dissemination, co-culture of PCCs and PMCs significantly increased the cells invading into the collagen gel layer compared with mono-culture of PCCs. PMCs pre-invaded into the collagen gel, remodeled collagen fibers, and increased parallel fiber orientation along the direction of cell invasion. In the tissues of peritoneal dissemination of the KPC (LSL-Kras^{G12D/+};

LSL-Trp53^{R172H/+};Pdx-1-Cre) transgenic mouse, the monolayer of PMCs was preserved in tumor-free areas, whereas PMCs around the invasive front of peritoneal dissemination proliferated and invaded into the muscle layer. *In vivo*, intraperitoneal injection of PCCs with PMCs significantly promoted peritoneal dissemination compared with PCCs alone. The present data suggest that the cancer-associated PMCs have important promoting roles in the peritoneal dissemination of PCCs. Therapy targeting cancer-associated PMCs may improve the prognosis of patients with pancreatic cancer.

Introduction

Pancreatic cancer is the fourth leading cause of cancer-related death in the United States, with a 5-year overall survival rate of 3-5% (1). Most pancreatic cancer patients have distant metastasis at diagnosis, and the prognosis of patients with peritoneal dissemination is extremely poor (2,3). Furthermore, peritoneal dissemination induces bowel obstruction and formation of malignant ascites, which leads to poor performance status (3). Therefore, it is important to determine a mechanism to control the peritoneal dissemination of pancreatic cancer.

Pancreatic cancer is characterized by excessive desmoplasia, which plays a crucial role in its aggressive behavior through the tumor-stroma interaction (4). In the process of peritoneal dissemination, cancer cells detached from the primary tumor are transported by peritoneal fluid and disseminate in the peritoneum (5). The peritoneum consists of a monolayer of peritoneal mesothelial cells (PMCs) supported by a basement membrane that rest on a layer of connective tissue with fibroblasts, mast cells, macrophages and vessels (6). Therefore, the tumor-stroma interaction is important in the process of peritoneal dissemination (7). We recently investigated the tumor-stroma interaction in peritoneal dissemination in pancreatic cancer and found that peritoneal myofibroblasts contributed to the promotion of peritoneal dissemination in pancreatic cancer (8). Previous studies reported that myofibroblasts were derived from resident peritoneal fibroblasts, bone marrow progenitor cells, the primary tumor itself or PMCs (9-12). However, the origin of peritoneal myofibroblasts in pancreatic cancer is not yet clearly understood.

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Abbreviations: PMCs, peritoneal mesothelial cells; PCCs, pancreatic cancer cells; KPC, (LSL-Kras^{G12D/+};LSL-Trp53^{R172H/+};Pdx-1-Cre) mouse model; MMT, mesothelial-to-mesenchymal transition; CAFs, cancer associated fibroblasts; DMEM, Dulbecco's modified Eagle's medium; FBS, fetal bovine serum; TGF- β 1, transforming growth factor- β 1; aPMCs, activated PMCs; H&E, hematoxylin and eosin; α -SMA, α -smooth muscle actin; CM, conditioned medium; PBS, phosphate-buffered saline; FN1, fibronectin 1; qRT-PCR, real-time quantitative reverse transcription polymerase chain reaction; PSCs, pancreatic stellate cells

Key words: peritoneal mesothelial cells, pancreatic cancer, peritoneal dissemination, tumor-stromal interaction, remodeling

PMCs generally act as a passive barrier and play important roles in the response to wound healing and infection (13). Other reports also showed that PMCs are converted into myofibroblasts via mesothelial-to-mesenchymal transition (MMT) by peritoneal dialysis (9) and in ovarian and gastric cancers (14,15). PMCs that are converted into myofibroblasts via MMT affect cancer cells as cancer associated fibroblasts (CAFs) in ovarian and gastric cancers (14,15). CAFs support the malignant progression of tumors by promoting growth, survival, angiogenesis, inflammation, drug resistance and invasion and metastasis of tumors (16). A previous study showed that heat-shock-factor-1 was one of the potent activator of CAFs promoting malignancy (17). On the other hand, the interaction between PMCs and cancer cells promoted the proliferation and invasiveness of cancer cells in ovarian (14,18,19) and gastric (10,20) cancer. Moreover, adhesive interaction between cancer cells and PMCs played an important role in peritoneal dissemination (21). A recent study revealed molecular mechanisms of peritoneal dissemination in gastric cancer (22). However, the role of PMCs in the peritoneal dissemination of pancreatic cancer remains unclear.

In the present study, we revealed the interaction between the pancreatic cancer cells (PCCs) and PMCs in the process of peritoneal dissemination *in vitro* and *in vivo*.

Materials and methods

Cell isolation and culture conditions. PMCs were isolated from ascites recovered from patients with no evidence of distant metastasis who underwent curative resection for pancreatic cancer, as previously described (9,23–25). Briefly, after ascites were centrifuged at 1,500 rpm for 10 min, the cell pellets were resuspended in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% fetal bovine serum (FBS), penicillin and streptomycin and cultured in Collagen type I coated dishes (Iwaki, Co., Ltd., Tokyo, Japan). The isolated cells were identified as PMCs by their polygonal morphology and the expression of calretinin and cells between passages 2 and 5 were used for assays. We confirmed that there was no contamination with fibroblasts, endothelial cells or malignant cells. PMCs were activated by transforming growth factor- β 1 (TGF- β 1) administration and these cells were regarded as activated PMCs (aPMCs), as previously described (9,10). Briefly, PMCs were incubated with or without 10 ng/ml TGF- β 1 (R&D Systems, Oxon, UK) in DMEM with 1% FBS for 48 h. We also used the following three PCC lines: SUIT-2 (Health Science Research Bank, Osaka, Japan), AsPC-1 (American Type Culture Collection, Manassas, VA, USA) and PANC-1 (RIKEN BioResource Center, Tsukuba, Japan). These cells were maintained as previously described (26).

Establishment of immortalized PMCs. We cloned the DNA encoding hTERT and SV40 Large T in the pLVSIIN vector. We used these vectors to construct lentiviral particles for infection of PMCs, followed by G418 selection.

Stable luciferase-expressing SUIT-2 cells. The firefly luciferase (GenTarget, #LVP326) expression vector was transfected into SUIT-2 cells according to the manufacturer's instructions. Blastidine S hydrochloride (#15205; Sigma-Aldrich) was used

for selection for more than 3 weeks. Luciferase expression was confirmed by a significant increase in emission after adding 150 μ g/ml D-Luciferin potassium salt (#LK10000; OZ Biosciences, Marseille, France) as compared with wild-type SUIT-2 cells.

Immunohistochemical analysis. Tissues of peritoneal dissemination of pancreatic cancer in KPC (LSL-Kras^{G12D/+}; LSL-Trp53^{R172H/+}; Pdx-1-Cre) transgenic mice (27) were evaluated by hematoxylin and eosin (H&E), CK19, calretinin and α -smooth muscle actin (α -SMA) immunohistochemical staining. Tissues were sliced to a thickness of 4 μ m and incubated with rabbit anti-CK19 antibody (#133496, 1:100; Abcam), mouse anti-calretinin antibody (#M7245, 1:25; Dako) or mouse anti- α -SMA antibody (#M0851, 1:100; Dako) overnight at 4°C. The staining was performed using serial sections.

Conditioned medium of PMCs. To obtain conditioned medium of PMCs (PMCs-CM), PMCs were seeded and cultured until subconfluent. The medium was then replaced with DMEM serum-free medium and the supernatants were collected after 48 h of incubation.

Matrigel invasion and migration assays. The invasiveness and migration capacities of PCCs and PMCs were assessed by determining the number of cells that invaded or migrated across Transwell chambers, as previously described (26). For co-cultures, PMCs (4.0×10^4 /well) or aPMCs (4.0×10^4 /well) in 750 μ l of DMEM supplemented with 10% FBS or medium alone were seeded in the lower chambers for 24 h, and PCCs (4.0×10^4 /well) in 250 μ l of DMEM supplemented with 10% FBS were placed in the upper Transwell chamber (8 μ m pore size; Becton-Dickinson, Franklin Lakes, NJ, USA) containing 100 ml of reconstituted Matrigel-coated membrane (20 mg/well; BD Biosciences, Bedford, MA, USA). After incubation for 48 h, cell invasion was evaluated by counting the number of cells that invaded through the Transwell chambers. Cell migration was assessed after incubation for 24 h using uncoated Transwell chambers. To assess the invasiveness of PMCs, each lower well was seeded with PCCs (4.0×10^4 /well) in 750 μ l of DMEM supplemented with 10% FBS or medium alone and incubated for 24 h. PMCs (4.0×10^4 /well) in 250 μ l of DMEM supplemented with 10% FBS were seeded in each upper well and incubated for 24 h for the invasion assay and 12 h for the migration assay. In both assays and at each time-point, invading or migrated cells at the bottom of the chamber were fixed with 70% ethanol and stained with H&E, and five random fields at $\times 200$ magnification were counted under a light microscope. Each experiment was performed in triplicate and repeated at least three times.

Adhesion assay. The adhesion ability of PCCs was determined as previously described (28). Briefly, PMCs and aPMCs (8.0×10^4 /well) were cultured in a monolayer in 96-well Collagen I coated plates overnight. Collagen I was used as the principal extracellular matrix molecule. PCCs were labeled with CellTracker™ Green CMFDA (Life Technologies, Eugene OR, USA). PCCs (4.0×10^4 /well) were added to the 96-well Collagen I coated plates containing confluent PMCs

or aPMCs or without cells, and cells were incubated for 3 h at 37°C. The plates were then washed three times with 200 μ l of phosphate-buffered saline (PBS) to remove the non-adherent tumor cells. The number of adhered PCCs was determined in five random fields at x200 magnification using a fluorescent microscope (BZ-9000; Keyence Corp., Osaka, Japan). Each experiment was performed in triplicate and repeated at least three times.

Cell viability assay (adhered and non-adhered conditions). PCCs (1.0×10^3 /well) were seeded (Greiner Bio-One GmbH, Frickenhausen, Germany) and cell viability examined using the CellTiter-Glo[®] Luminescent Cell Viability assay kit (G7570; Promega, Madison, WI, USA) after culture for 24, 48 and 72 h following the manufacturer's instructions. PCCs (1.0×10^3 /well) were incubated with or without PMC-CM in 10% FBS/DMEM in 96-well plates. In non-adhered conditions, PCCs (1.0×10^3 /well) were cultured in a 96-well ultra-low adherence plate (Corning Costar CLS7007). Background was subtracted using values from wells containing only culture medium. Each experiment was performed in triplicate and repeated at least three times.

Apoptosis assay. PCCs (1.0×10^6 /well) were incubated with or without PMCs-CM in DMEM serum-free medium in a 90 mm ultra-low adherence plate (Nunclon Sphera Dishes; Thermo Fisher Scientific) for 72 h and the cells were lysed and subjected to western blotting to determine the expression of apoptosis regulators.

3D organotypic culture model. To assess the process of invasiveness of PCCs and PMCs, the 3D organotypic culture model was set up as previously described (20) with minor modifications. Briefly, 1000 μ l of the gel containing 2 mg/ml Collagen I (BD Biosciences) and 2.5 mg/ml Matrigel was laid onto the upper Transwell chambers (6-well). PMCs (6.0×10^5 /well) were added on the gels in DMEM containing 1% FBS. After 6 h, PCCs (6.0×10^5 /well) were added on the monolayer of PMCs. The bottom well was filled with DMEM containing 10% FBS. SUIT-2 cells were labeled with CellTracker[™] Green and PMCs were labeled with CellTracker[™] Red CMTPX (Life Technologies). A laser-scanning confocal fluorescence microscope (A1R; Nikon) was used for immunofluorescence microphotography. After incubation for 10 days, the gels were fixed in 4% paraformaldehyde and sections were cut into 4 μ m sections for H&E staining and incubated with rabbit Ki-67 antibody (#16667, 1:100; Abcam) or rabbit anti-CEA (#RB-368-A, 1:250; Thermo Fisher Scientific) overnight at 4°C. The Ki-67 labeling index was calculated by dividing the number of Ki-67 positive nuclei by the total number of nuclei in five random fields at x200 magnification.

Analysis of collagen fiber orientation. The orientation of the 3D gel collagen fibers was analyzed using OrientationJ (an ImageJ-plugin) (v.1.48u; National Institute of Health, Bethesda, MD, USA) (29). Counts of the total fibers as well as each orientation angle were measured, and the angles were determined by approximating the relative angle every 10°. To allow for comparison of acquired images, we set the direction of the invading cells to 0° in the invading area and the

tangential direction of the initial cell cluster surface to 0° in the non-invading area.

Real-time quantitative reverse transcription polymerase chain reaction (qRT-PCR). qRT-PCR was performed using the iTaq Universal SYBR-Green One-Step kit and CFX96 Touch Real-Time PCR Detection systems (Bio-Rad Laboratories). The primers were purchased from Takara Bio (Kusatsu, Japan). Human GAPDH gene was used as the endogenous control gene. The following primers were used in the present study: E-cadherin, forward, 5'-AAGTGCTGCAGCCAAAGACAGA-3' and reverse, 5'-AAATTGCCAGGCTCAATGACAAG-3'; fibronectin 1 (FN1), forward, 5'-ACAGAACTATGATGCCGACCAGAAG-3' and reverse, 5'-ACTGATCTCCAATGCGGTACATGA-3'; GAPDH, forward, 5'-GCACCGTCAAGGCTGAGAAC-3' and reverse, 5'-TGGTGAAGACGCCAGTGGGA-3'.

Western blot analysis. Whole cell lysates were prepared in PRO-PREP solution (iNtRON Biotechnology, Seongnam, Korea) from PMCs, aPMCs and PCCs. Proteins from cell lysates were fractioned by SDS-PAGE and transferred to a polyvinylidene difluoride membrane (Bio-Rad Laboratories). The membrane was incubated overnight at 4°C with the following antibodies: anti-E-cadherin (#3195, 1:1,000; Cell Signaling Technology, Danvers, MA, USA), anti-vimentin (#5741, 1:1,000; Cell Signaling Technology), anti- α -SMA (#M0851, 1:500; Dako), anti-FN1 (#sc-9068, 1:200; Santa Cruz Biotechnology), anti-calretinin (#92341, 1:1,000; Abcam), anti-CEA (#RB-368-A, 1:200; Thermo Fisher Scientific), anti-cleaved caspase-3 (#9661, 1:1,000; Cell Signaling Technology) and anti- β -actin (#8227, 1:5,000; Abcam). The membrane was then probed with horseradish-peroxidase-conjugated secondary antibodies (Cell Signaling Technology). Immunoblots were detected by enhanced chemiluminescence.

Animal experiments. All experiments with mice were conducted and approved by the Ethics Committee of Kyushu University. The model of intraperitoneal injection of BALB/c nu/nu mice (5-6 weeks of age; Kyudo Co.) was used to analyze the dissemination activity of PCCs alone and PCCs with PMCs. All animals were bred in laminar-flow cabinets under specific pathogen-free conditions. The mice were anesthetized with ether and suspensions of luciferase-expressing SUIT-2 cells (2×10^5) alone in 200 μ l PBS or with PMCs (1×10^6) in 200 μ l PBS were injected into the peritoneal cavity of groups of 6 mice. Luciferin luminescence was measured at 21 days after intraperitoneal injection using IVIS Spectrum (Caliper Life Sciences, Waltham, MA, USA) after injecting 150 mg D-Luciferin (#LK10000; OZ Biosciences) into the intraperitoneal cavity of anesthetized mice. Luminescence was quantified using Living Image software version 4.4 (Summit Pharmaceuticals International Corp., Tokyo, Japan). All mice were sacrificed at 21 days after the evaluation of luciferin luminescence and the disseminated nodules >3 mm in size were counted.

Statistical analysis. Results are presented as means \pm SD. Comparisons between the two groups were conducted using the Student's t-test. Tumor volume and disseminated nodules *in vivo*

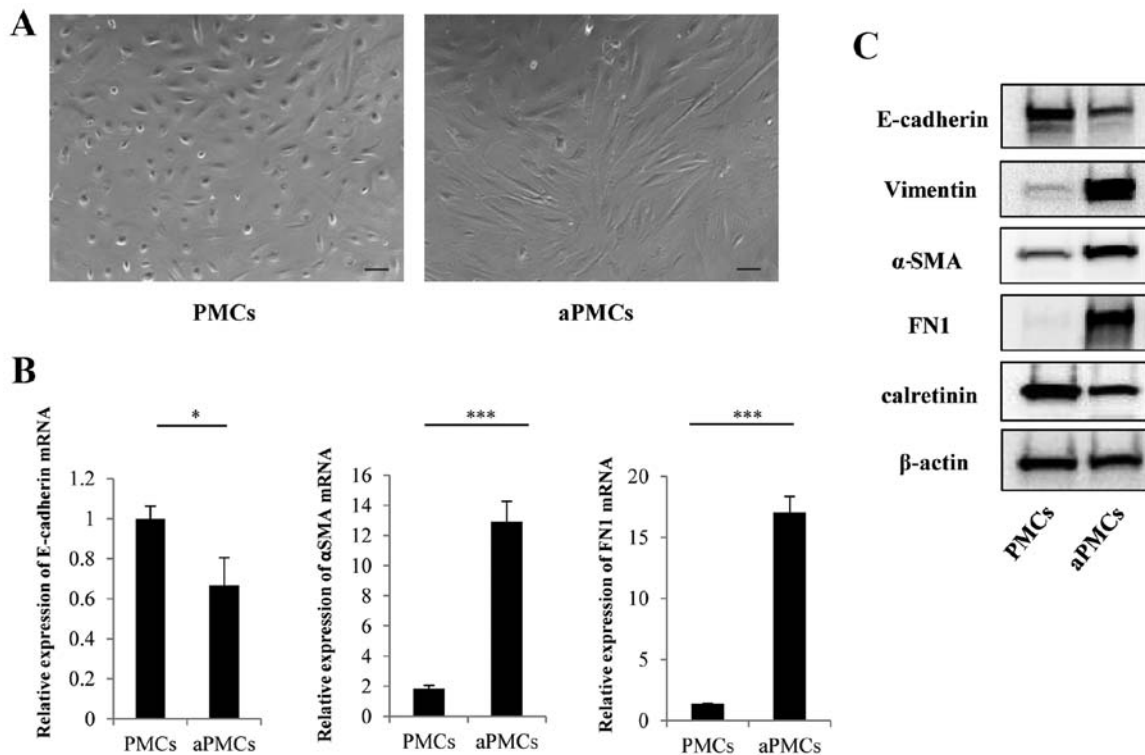


Figure 1. TGF- β 1 induces mesothelial-to-mesenchymal transition (MMT) in PMCs. (A) Representative microphotographs of PMCs and activated PMCs (aPMCs) that were incubated with TGF- β 1 (10 ng/ml) for 48 h. Original magnification, $\times 100$. Scale bars, 100 μ m. (B) qRT-PCR analysis and (C) western blot analysis of PMCs and aPMCs for the indicated molecular factors. * $P < 0.05$; *** $P < 0.001$.

were analyzed using the Wilcoxon test. Statistical significance was defined as $P < 0.05$. All statistical analyses were performed using JMP 11 software (SAS Institute, Cary, NC, USA).

Results

Verification of PMCs and aPMCs. The isolated cells were identified as PMCs by their polygonal morphology and the expression of calretinin (Fig. 1A and C). Representative microphotographs of aPMCs showed spindle-like morphology compared with PMCs (Fig. 1A). To verify the MMT of PMCs, the expressions of MMT markers were analyzed by qRT-PCR and western blotting. The gene and protein expression of E-cadherin was decreased in PMCs, whereas the expression levels of α -SMA, FN1 and vimentin were increased (Fig. 1B and C) confirming MMT of PMCs.

The tumor-stromal interaction of PCCs and PMCs significantly enhances their migration and invasiveness in indirect co-culture. To investigate the tumor-stromal interaction between PMCs and PCCs, we evaluated the effect of PMCs on PCC migration and invasiveness using indirect co-culture. We found that PMCs significantly enhanced the migration (Fig. 2A) and the invasiveness (Fig. 2B) of PCCs in indirect co-culture (both $P < 0.001$). Furthermore, co-culture with aPMCs significantly enhanced the migration (Fig. 2A) and invasiveness (Fig. 2B) of PCCs compared with the co-culture with PMCs (both $P < 0.001$). We also investigated the effect of co-cultured PCCs on the mobility and invasiveness of PMCs and found that PCCs significantly enhanced the migration

(Fig. 2C) and the invasiveness (Fig. 2D) of PMCs (both $P < 0.001$).

Adhesion ability of PCCs to PMCs, aPMCs or Collagen I. Next, we assessed the adhesion ability of PCCs to PMCs, aPMCs or Collagen I and found that the adhesion ability of PCCs to PMCs was significantly decreased compared with the adhesion to Collagen I, whereas the adhesion ability of PCCs to aPMCs was significantly increased compared with the adhesion to PMCs (Fig. 3; $P < 0.001$).

PMCs enhance proliferation and anoikis resistance of PCCs. We found that PMCs significantly enhanced the proliferation of PCCs compared with controls both in adhered (Fig. 4A; $P < 0.001$) and non-adhered conditions (Fig. 4B; SUIT-2 cells, $P < 0.05$; AsPC-1 cells; $P < 0.001$). Furthermore, addition of PMCs-CM decreased the expression of cleaved caspase-3 of PCCs compared with controls (Fig. 4C).

PMCs enhance invasiveness and proliferation of PCCs and remodel collagen fibers in the 3D organotypic culture model. We evaluated the invasion process of PCCs and PMCs using a 3D organotypic culture model and confocal fluorescence microscopy. We found that the invasion distance of PMCs was significantly longer than that of SUIT-2 cells (Fig. 5A and B; $P < 0.001$). H&E sections showed that the co-culture of SUIT-2 cells and PMCs showed a significant increase in the number of cells invading into the collagen gel layer compared with mono-culture of SUIT-2 cells, which did not invade into the collagen gel layer (Fig. 5C and D; $P < 0.001$). To distinguish

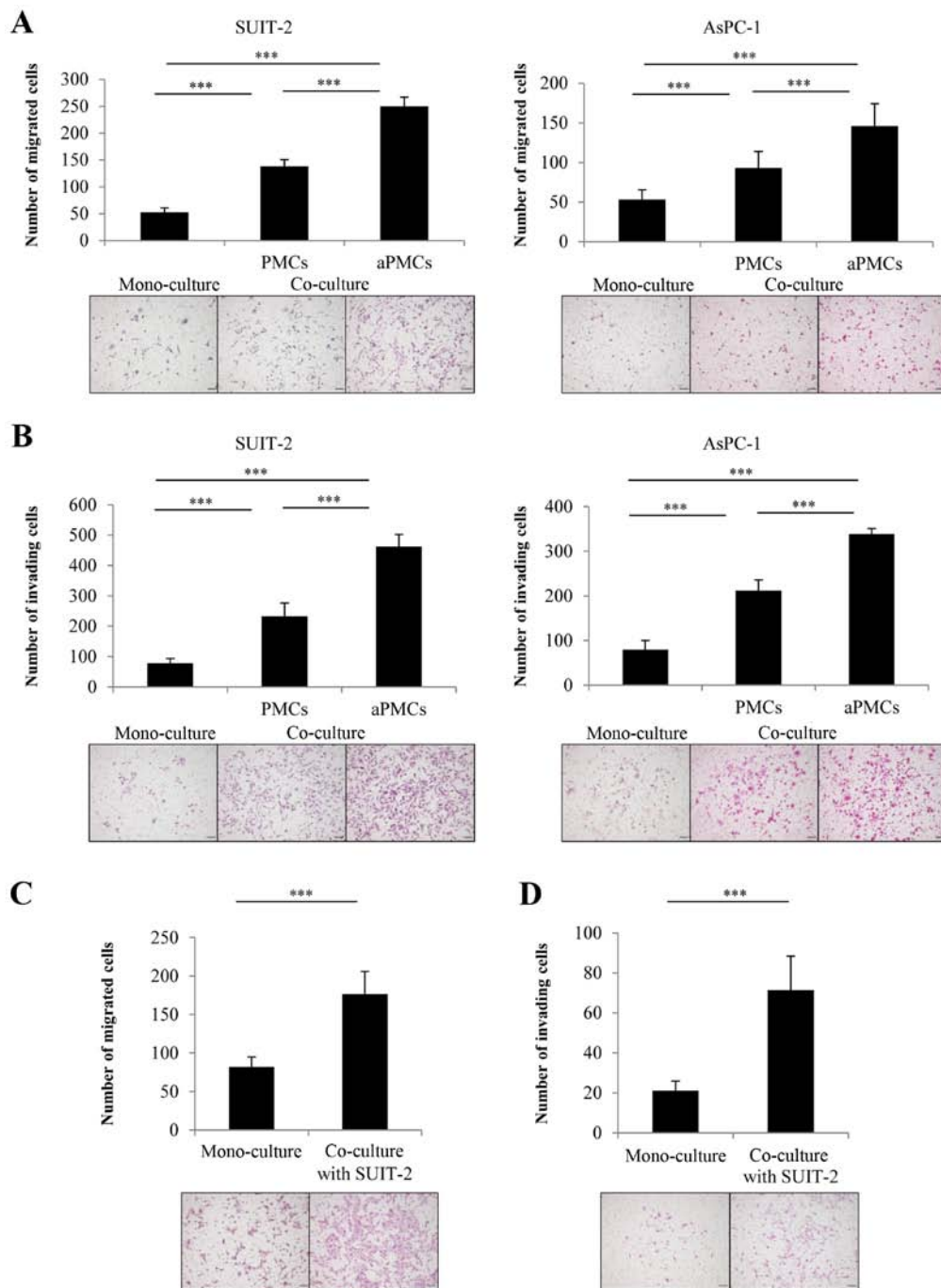


Figure 2. The tumor-stromal interaction of PCCs and PMCs significantly promoted their migration and invasiveness in indirect co-culture. (A) Cell migration assays and (B) cell invasion assays of PCCs alone (first column) and PCCs co-cultured with PMCs (second column) or aPMCs (third column). The images show representative H&E stainings. (C and D) The migration and invasiveness of PMCs were enhanced when co-cultured with PCCs. H&E staining; original magnification, x100. ***P<0.001.

between PCCs and PMCs, the expression of CEA was assessed by western blotting and was positive only in SUIT-2 cells (Fig. 5E). Immunohistochemical staining showed that SUIT-2 cells expressing CEA invaded into the collagen gel layer upon co-culture with PMCs (Fig. 5F). Immunohistochemical staining of Ki-67 showed that co-culture of SUIT-2 cells and PMCs significantly enhanced proliferation compared with monoculture of SUIT-2 cells (Fig. 5G and H; P<0.001). To investigate the mechanism of PMC-induced invasiveness of PCCs, we focused on the collagen fiber orientation. We found that the collagen fibers in the co-cultures of SUIT-2 cells and

PMCs or the monocultures of PMCs displayed an organized parallel orientation along the cells invading into collagen gel layer compared with the random fiber arrangement detected in the monocultures of SUIT-2 cells (Fig. 5I and J).

PMCs exist in the invasive front of peritoneal dissemination of pancreatic cancer derived from KPC mice. To assess the role of PMC in peritoneal dissemination of pancreatic cancer in the KPC mouse, we focused on the invasive front of peritoneal dissemination. We performed H&E staining and immunohistochemical staining of CK19 as a PCC marker,

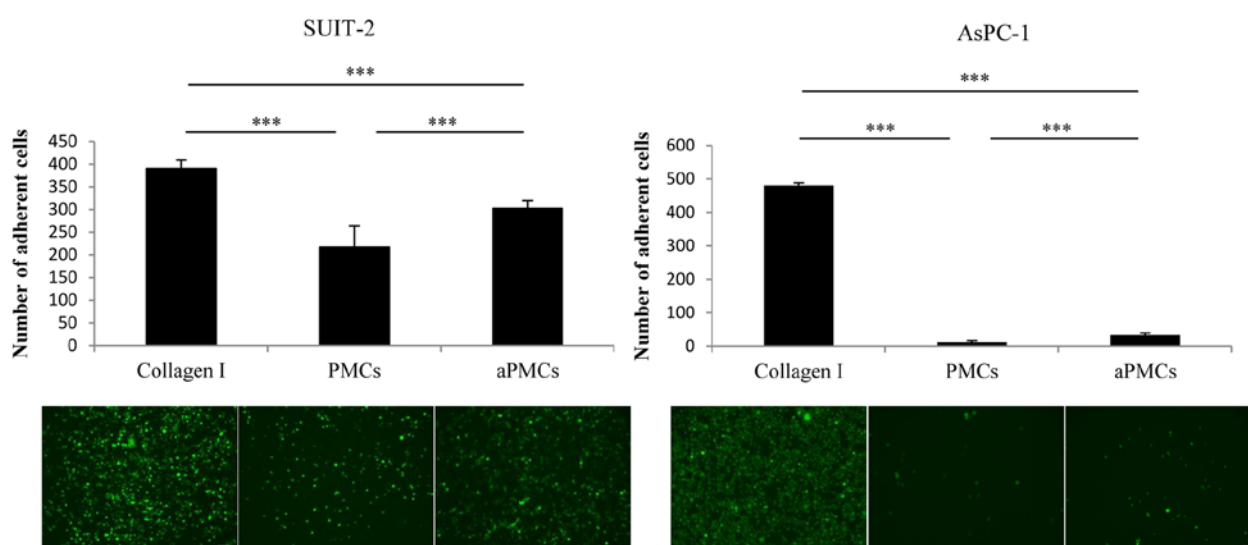


Figure 3. Adhesion ability of PCCs to collagen I, PMCs and aPMCs. PCCs were examined for adhesion ability with collagen I, PMCs or aPMCs. The top panel shows quantification of cell numbers. The bottom panel shows images by fluorescent microscope. Original magnification, x100; *** $P < 0.001$.

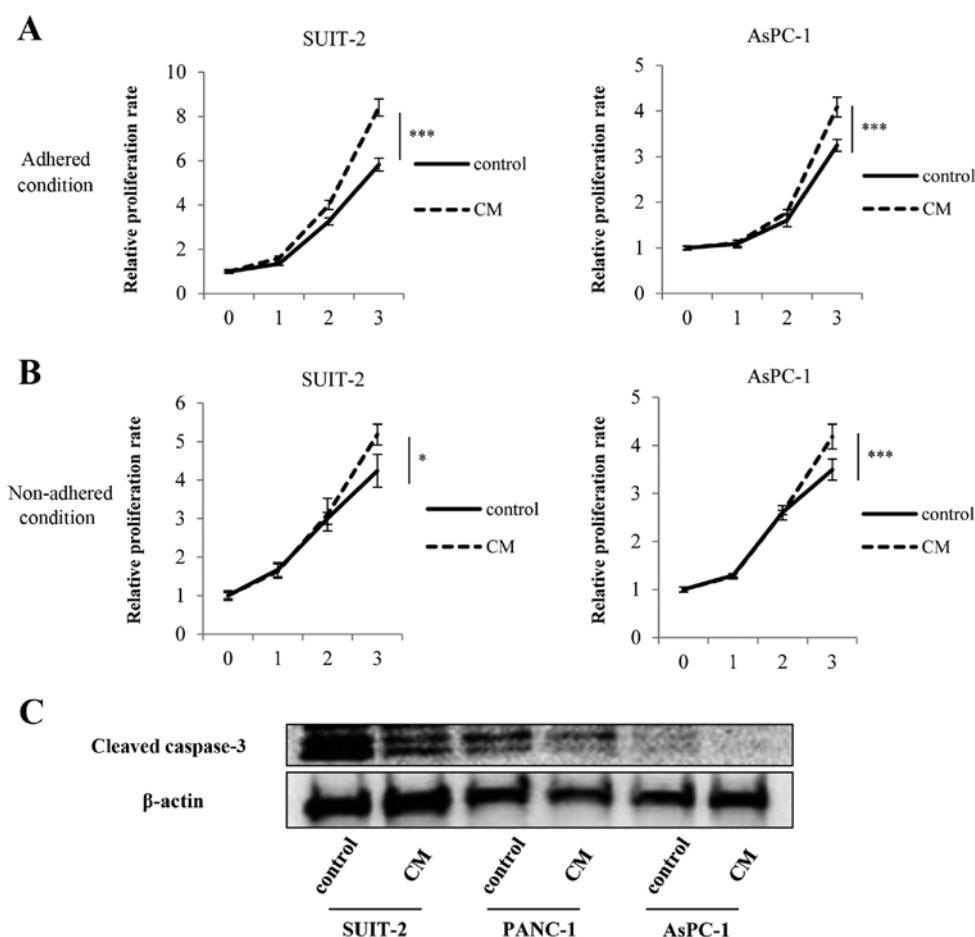


Figure 4. PMCs enhance proliferation and anoikis resistance of PCCs. PCCs were incubated with or without PMC-CM in (A) adhered conditions (96-well plates) or (B) non-adhered conditions (96-well ultra-low adherence plate) and cell viability was evaluated. (C) Western blot analysis of cleaved caspase-3 of PCCs in non-adhered conditions and in the presence of PMC-CM. * $P < 0.05$, *** $P < 0.001$.

calretinin as a PMC marker and α -SMA as a myofibroblast marker. The monolayer of PMCs was preserved in tumor-free areas, whereas in micro-peritoneal dissemination, multiple layers of PMCs were observed and a subset of PMCs invaded

into the muscle layer (Fig. 6A). In macro-peritoneal dissemination, PMCs existed in the invasive front and pre-invaded into the muscle layer, whereas α -SMA-positive and calretinin-negative stromal cells existed in the tumor (Fig. 6B).

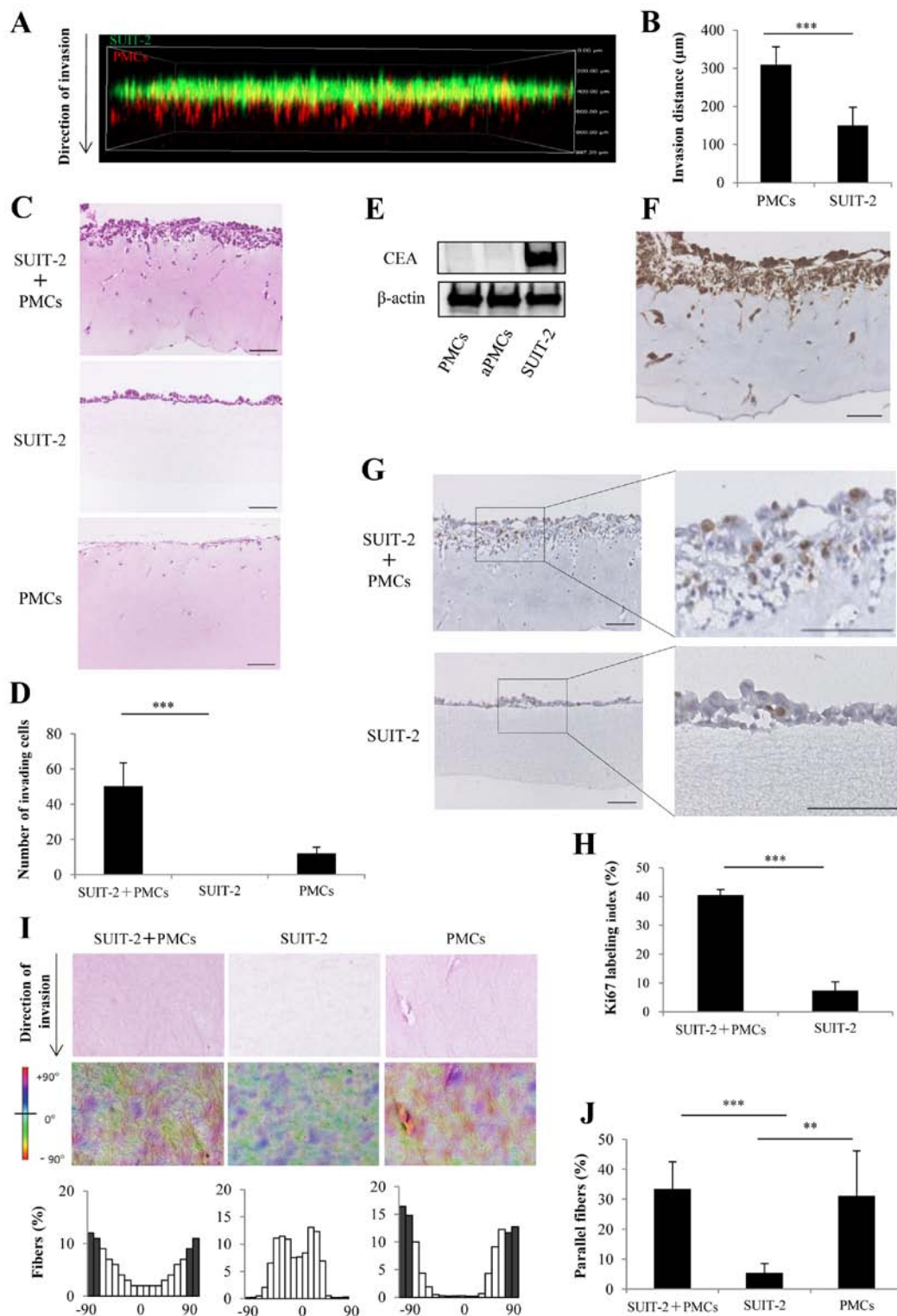


Figure 5. PMCs enhance invasiveness and proliferation of PCCs and remodel collagen fibers in the 3D organotypic culture model. (A) Representative confocal fluorescence images on day 5 showed that PMCs (red) pre-invaded into the collagen compared with SUI-2 cells (green). (B) Quantification analysis of invasion distance of PMCs and SUI-2 cells. (C) Representative image of H&E sections of collagen gel on day 10 and (D) quantification analysis of the number of invading cells. (E) Western blot analysis of CEA in PMCs, aPMCs and SUI-2 cells. (F) Representative images of immunohistochemical staining showed that SUI-2 cells expressing CEA invaded into the collagen gel layer when co-cultured with PMCs. (G) Representative images of immunohistochemical staining of Ki-67 and (H) Ki-67 labeling index showed that co-culture with SUI-2 cells and PMCs significantly enhanced proliferation compared with mono-culture of SUI-2. Original magnification, x200 (left panel), x600 (right panel). (I) Representative images of orientation of collagen fiber using OrientationJ. (J) The percentage of fibers in parallel orientation along the invasive direction (-90 to -70° , 70 to 90°) were calculated in five fields of the indicated matrices. Scale bars, 100 μm . ** $P < 0.01$, *** $P < 0.001$.

PMCs promote peritoneal dissemination of PCCs *in vivo*. To investigate the functional role of PMCs on the dissemination

of PCCs *in vivo*, we intraperitoneally injected luciferase-expressing SUI-2 cells alone or with PMCs in nude mice and

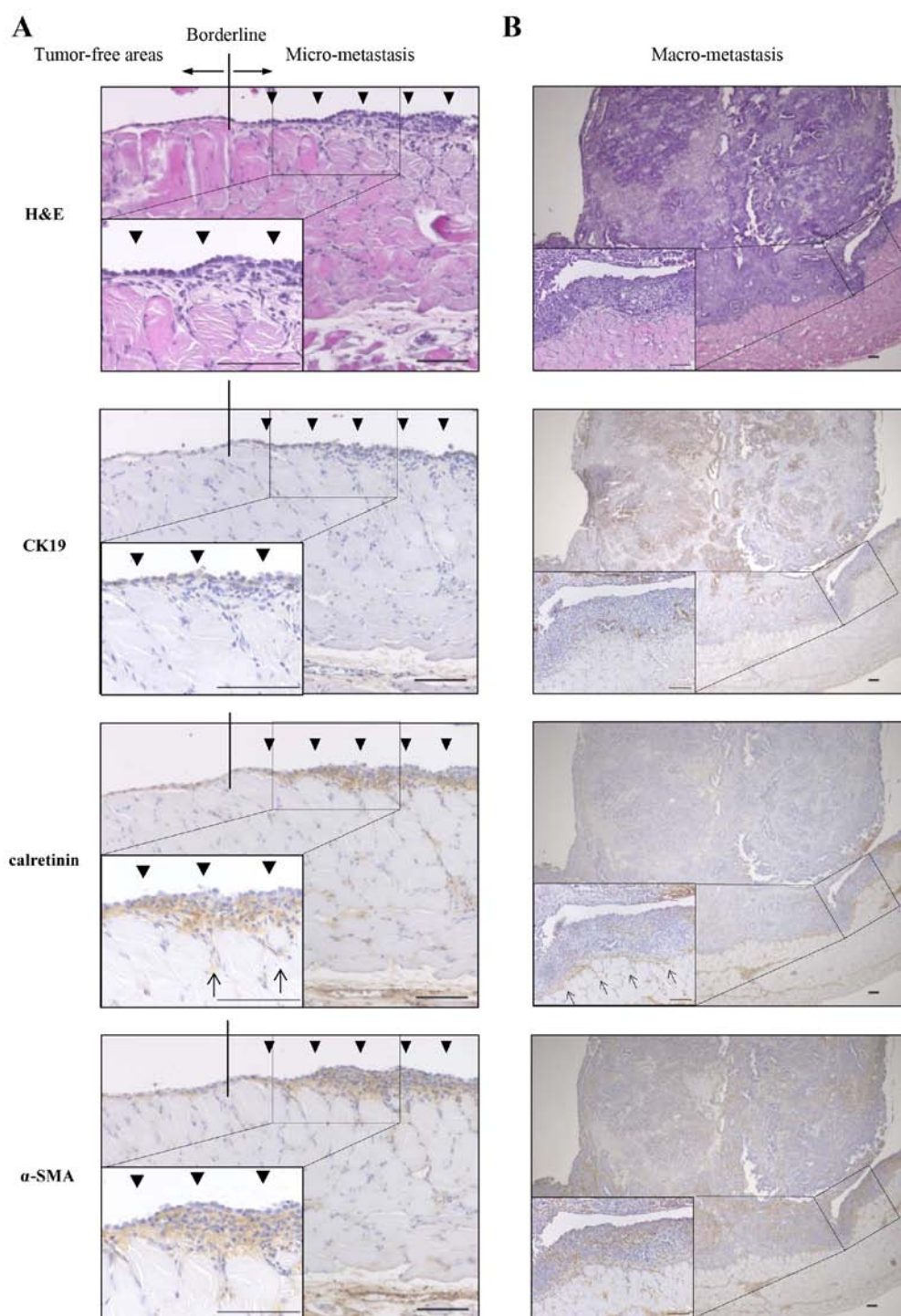


Figure 6. PMCs exist in the invasive front of peritoneal dissemination of pancreatic cancer derived from KPC transgenic mouse. Representative images of H&E staining and immunohistochemical staining for CK19, calretinin and α -SMA in peritoneal dissemination derived from the KPC mouse. (A) The monolayer of PMCs was preserved in tumor-free areas, whereas in micro-peritoneal dissemination, multiple layers of PMCs were observed and a subset of PMCs invaded into the muscle layer. (B) In macro-peritoneal dissemination, PMCs existed in the invasive front and pre-invaded into the muscle layer, whereas α -SMA-positive and calretinin-negative stromal cells existed in the tumor. The arrowheads indicate cancer cells and the arrow indicate PMCs. Scale bars, 100 μ m.

measured the luciferase luminescence to evaluate the growth of disseminated nodules of SUI-2 cells. We found that intraperitoneal injection of SUI-2 cells with PMCs significantly promoted peritoneal dissemination compared with SUI-2 cells alone (Fig. 7A and B; $P < 0.01$). Intraperitoneal injection of SUI-2 cells with PMCs (Fig. 7C, right panel) yielded a mean 29.3 ± 14.1 peritoneal disseminated nodules > 3 mm compared with 7.0 ± 4.8 nodules in mice injected with SUI-2 cells alone

(Fig. 7C, left panel), a difference that was statistically significant (Fig. 7D; $P < 0.01$).

Discussion

In the present study, we found that the tumor-stromal interaction of PCCs and PMCs significantly enhanced their migration and invasiveness and enhanced proliferation and

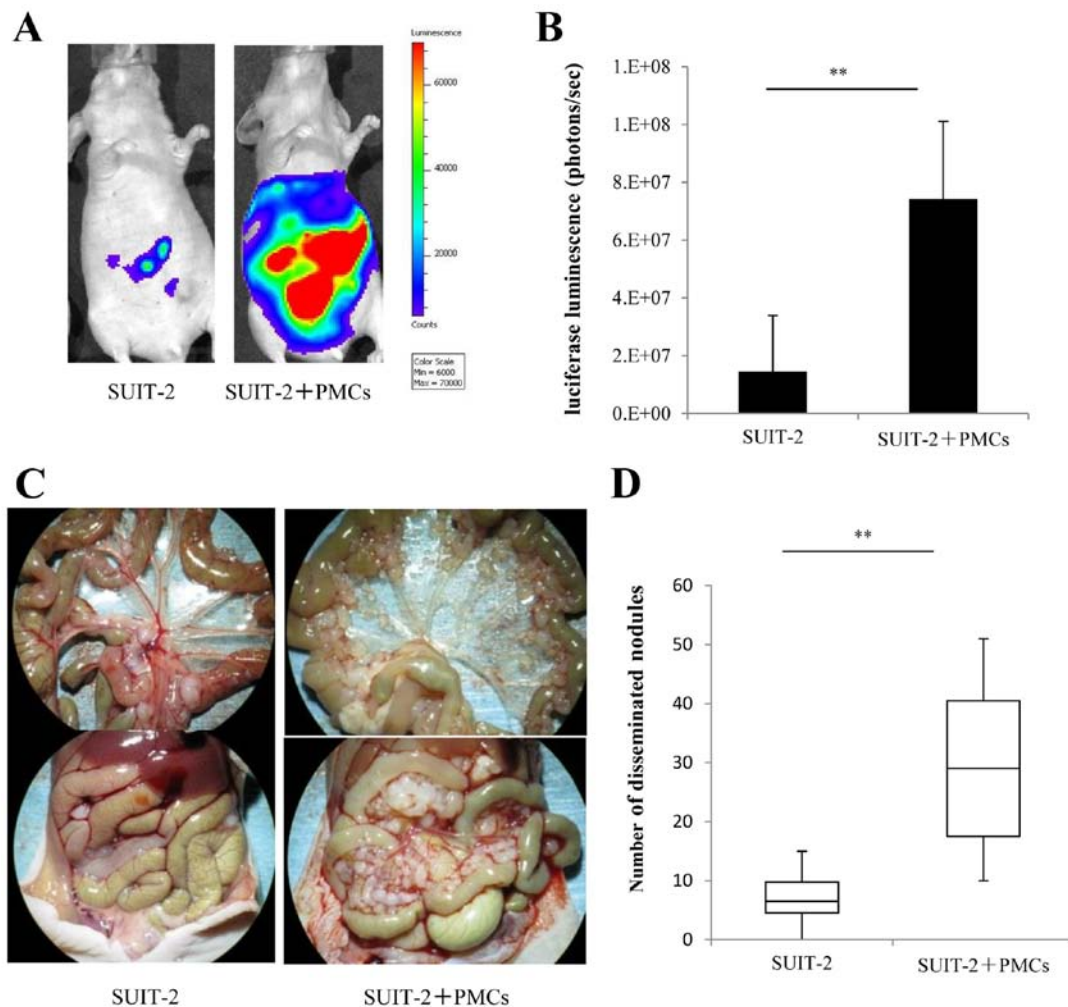


Figure 7. PMCs promote peritoneal dissemination of PCCs *in vivo*. (A) Representative photograph of IVIS imaging system in nude mice intraperitoneally injected luciferase-expressing SUIT-2 cells alone or with PMCs. (B) Quantification of luciferase luminescence in mice injected luciferase-expressing SUIT-2 cells alone or with PMCs; **P<0.01. (C) Representative photographs of peritoneal dissemination in nude mice intraperitoneally injected with SUIT-2 cells alone or with PMCs at 21 days. (D) Quantification of disseminated nodules in mice with intraperitoneal injection of SUIT-2 cells with PMCs or with SUIT-2 cells alone; **P<0.01.

anoikis resistance of PCCs. In the 3D organotypic culture model, we found that co-culture with PCCs and PMCs significantly increased the numbers of cells invading into the collagen gel layer compared with monoculture of PCCs. We also found that PMCs pre-invaded into the collagen gel, remodeled collagen fibers, and increased parallel fiber orientation along the direction of cell invasion. We recently reported that 3D matrices with the parallel fiber architecture derived from pancreatic stellate cells (PSCs) under hypoxia promoted cancer cell motility by inducing directional migration of PCCs (30). These findings suggest that PMCs might enhance the directional invasion of PCCs by increasing parallel fiber orientation besides growth factors associated with the tumor-stromal interaction.

Kasagi *et al* (31) showed that peritoneal lavage fluid that contained peritoneal collagen type IV and plasma fibronectin facilitated spheroid formation of colon cancer cells. Moreover, Condello *et al* (32) revealed that spheroid formation of cancer cells under non-adherent conditions served to protect cells from environmental-induced anoikis. We demonstrated that PMCs enhanced proliferation in non-adherent conditions and

anoikis resistance of PCCs. These findings suggest that PMCs promote spheroid formation of cancer cells and contribute to survival of cancer cells in the peritoneal fluid.

Next, we demonstrated that the adhesion ability of PCCs to PMCs was significantly decreased compared with Collagen I, whereas the adhesion ability of PCCs to aPMCs was significantly increased compared with PMCs. These findings were similar to previous reports in gastric cancer (33). A previous study also showed that the expression of FN1 in PMCs that was stimulated by ovarian cancer cells promoted the adhesion of cancer cells in ovarian cancer (33,34). In the present study, the expression of FN1 in PMCs with MMT was significantly increased compared with PMCs (Fig. 1B and C). These findings indicate that the expression of FN1 in PMCs with MMT might be involved in promoting the adhesion of PCCs.

We demonstrated that growth factors associated with tumor-stromal interaction of PCCs and PMCs mutually enhanced their migration and invasiveness in indirect co-culture. Moreover, aPMCs significantly enhanced the migration and invasiveness of PCCs. In the 3D organotypic culture model of peritoneal dissemination, co-culture of SUIT-2 and PMCs

significantly increased the number of cells invading into the collagen gel layer compared with monoculture of SUIT-2 cells, which did not invade into the collagen gel layer. The interaction of cancer cells and PMCs enhanced the invasiveness of cancer cells in ovarian (14) and gastric (20) cancer in the 3D organotypic culture model. Moreover Satoyoshi *et al* (20) revealed that Tks5 activation in PMCs created the invasion front of peritoneal metastasis, which guided invasiveness of cancer cells. In the present study, we revealed that PMCs pre-invaded into the collagen gel, remodeled collagen fibers, and increased parallel fiber orientation along the direction of cell invasion. Conklin *et al* (35) revealed that the presence of straightened collagen fibers is a predictor of breast cancer survival. Furthermore, we previously reported that 3D matrices with the parallel fiber architecture derived from PSCs under hypoxia promoted cancer cell motility by inducing directional migration of PCCs (30). These findings suggest that a subset of PMCs enhanced the directional invasion of PCCs by increasing parallel fiber orientation besides growth factors associated with tumor-stromal interaction.

In previous studies, the interaction of cancer cells and PMCs in the murine tissues of peritoneal dissemination was assessed *in vivo* using intraperitoneal injection of cancer cells into nude mice (14,20). However, these models might not reflect the spontaneous formation of peritoneal dissemination because peritoneal dissemination of these models was artificially created. Therefore, in the present study, we assessed the tissues of peritoneal dissemination that were spontaneously developed in the KPC mouse with pancreatic cancer, which histologically recapitulates the human tumors. Similar to our findings in the 3D organotypic culture model, the monolayer of PMCs was preserved in tumor-free areas, whereas PMCs were present in the invasive front of micro-peritoneal dissemination and proliferated there, and a subset of PMCs invaded into the muscle layer. These findings indicate that PMCs pre-invade, possibly to lead invasiveness of PCCs through the tumor-stroma interaction and the matrices remodeling.

We also showed that intraperitoneal injection of PCCs and PMCs significantly promoted peritoneal dissemination compared with PCCs alone *in vivo*. Similar results were reported in gastric (20) and ovarian cancer (19). In the present study, exogenous PMCs promoted peritoneal dissemination *in vivo* possibly by enhancing proliferation, anoikis resistance and invasiveness of PCCs.

In conclusion, our results suggest that a subset of PMCs promote the formation of peritoneal dissemination through the tumor-stromal interaction and the matrices remodeling although PMCs were generally thought to play a protective role for peritoneal dissemination. Therapy targeting this specific subset of PMCs may improve the prognosis of patients with pancreatic cancer.

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