

2-Methoxyestradiol enhances radiosensitivity in radioresistant melanoma MDA-MB-435R cells by regulating glycolysis via HIF-1 α /PDK1 axis

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Abstract. HIF-1 α overexpression is associated with radioresistance of various cancers. A radioresistant human melanoma cell model MDA-MB-435R (435R) was established by us previously. Compared with the parental cells MDA-MB-435 (435S), an elevated level of HIF-1 α expression in 435R cells was demonstrated in our recent experiments. Therefore, in the current study, we sought to determine whether selective HIF-1 α inhibitors could radiosensitize the 435R cells to X-ray, and to identify the potential mechanisms. Our data demonstrated that inhibition of HIF-1 α with 2-methoxyestradiol (2-MeOE2) significantly enhanced radiosensitivity of 435R cells. 2-MeOE2 increased DNA damage and ratio of apoptosis cells induced by irradiation. Whereas, cell proliferation and the expression of pyruvate dehydrogenase kinase 1 (PDK1) were decreased after 2-MeOE2 treatment. The change of expression of GLUT1, LDHA and the cellular ATP level and extracellular lactate production indicates that 2-MeOE2 suppressed glycolytic state of 435R cells. In addition, the radioresistance, glycolytic state and cell proliferation of 435R cells were also decreased after inhibiting pyruvate dehydrogenase kinase 1 (PDK1) with dichloroacetate (DCA). DCA could also increase DNA damage and ratio of apoptotic cells induced by irradiation. These results also suggest that inhibition of HIF-1 α with

2-MeOE2 sensitizes radioresistant melanoma cells 435R to X-ray irradiation through targeting the glycolysis that is regulated by PDK1. Selective inhibitors of HIF-1 α and glycolysis are potential drugs to enhance radiosensitivity of melanoma cells.

Introduction

Hypoxia-inducible factor-1 α (HIF-1 α) plays a crucial role in the adaptation of cancer cells to hypoxia by activating transcription of target genes that regulate several biological processes, including cell proliferation, glucose metabolism and pH regulation (1). Slominski *et al* found that HIF-1 α was upregulated in advanced malignant melanoma compared with melanocytic nevi or thin melanomas localized to the skin (1). High expression level of HIF-1 α is an independent predictor of poor prognosis after radiotherapy (2,3). 2-Methoxyestradiol (2-MeOE2) is a special inhibitor that suppresses HIF-1 α protein levels and its transcriptional activity. It was shown to inhibit the expression of HIF-1 α in a dose-dependent manner in cancer cells by depolymerising microtubules and blocking HIF-1 α nuclear accumulation (4). Activation of glycolytic genes by HIF-1 α is considered to be a very important factor for metabolic adaptation to hypoxia, with increased conversion of glucose to pyruvate and subsequently to lactate (5). Many studies demonstrated that the expression and activity of glycolytic enzymes and the lactic acid concentration were reduced by inhibiting HIF-1 α (6,7).

Kim *et al* found that HIF-1 suppressed glucose metabolism through the tricarboxylic acid cycle (TCA) by directly transactivating the gene encoding pyruvate dehydrogenase kinase 1 (PDK1). PDK1 inactivated the TCA cycle enzyme and pyruvate dehydrogenase (PDH), which converted pyruvate to acetyl-CoA, and rescued these cells from hypoxia-induced apoptosis (8). HIF-1 α causes an increase in pyruvate dehydrogenase kinase 1 (PDK1), which acts to limit the amount of pyruvate entering the citric acid cycle, leading to decreased mitochondrial oxygen consumption. PDK downregulates the activity of PDH-E1 α , decreases the oxidation of pyruvate in mitochondria, and increases

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the conversion of pyruvate to lactate in the cytosol. Dichloroacetate (DCA), as an inhibitor of pyruvate dehydrogenase kinase (PDK), decreases the glycolysis state of cells by leading to the reactivation of pyruvate dehydrogenase (PDH) and shifts glucose metabolism from glycolysis to mitochondrial oxidation (9).

The reprogramming of metabolism, especially the glucose metabolism is one of the hallmarks of cancer (10). Cancer cells have generally higher level of glucose uptake and lactate secretion, regardless of oxygen content. This phenomenon is called 'aerobic glycolysis' or the 'Warburg effect' (11,12). Metabolic studies supported the metabolic switch toward aerobic glycolysis in melanoma cells (13,14). Recently, some studies revealed that elevated glycolysis of cancer cells will not only provide a growth advantage but also involves in resistance to chemotherapy and ionizing radiation resistance (15,16). High glycolytic states of tumor cells are known to correlate strongly with radioresistance (17-21).

In our previous study, radiosensitive/radioresistant human melanoma cell model MDA-MB-435/MDA-MB-435R was established (22). An elevated level of HIF-1 α expression in radioresistant melanoma cells was also demonstrated in our recent experiments. Therefore, we aimed to investigate the effect of HIF-1 α on glycolysis and radioresistance in the 435R cells. Since PDK1 is a key regulator of glycolysis and it can be downregulated by inhibition of HIF-1 α , DCA was used in the recent study to elucidate the possible underlying mechanisms of 2-MeOE2 radiosensitizing to radioresistant melanoma cells, especially the HIF-1 α /PDK1-mediated glycolysis.

Materials and methods

Cells, cell culture and reagents. Human melanoma cell line MDA-MB-435S was purchased from the Cell Bank of Type Culture Collection of Chinese Academy of Sciences (Shanghai, China). Cell lines were cultured in DMEM growth media (Life Technologies, Carlsbad, CA, USA) which was supplemented with 10% fetal bovine serum (FBS, Life Technologies) and maintained at 37°C in a humidified atmosphere at 5% CO₂. DCA and 2-MeOE2 were purchased from Sigma-Aldrich (St. Louis, MO, USA).

X-ray irradiation. Radioresistant cell model of MDA-MB-435S were established by irradiation with X-ray. All the cells were first grown to approximately 90% confluence then irradiated by a Siemens Primus Accelerator at an average dose rate of 2 Gy/fraction, total dose was 60 Gy.

Colony formation assay. The standard clonogenic assay was performed as previously described (23,24). Cells were digested with trypsin enzyme at room temperature for 30-60 sec and then the clumped cells were pipetted. The single cell suspension was adjusted and seeded into typical 6-well plates. Then, cells were left to settle overnight, and exposed to irradiation at room temperature with the dose of 0, 2, 4, 6, 8 and 10 Gy, then incubated at 37°C, 5% CO₂ for 14 days. After fixation and staining, colonies of >50 cells were scored. Surviving rates were evaluated relative to 0 Gy radiation treated controls. A single-hit multi-target model was used to analyze the data (25). The survival curve of each group was plotted as the log of the

survival fraction vs. the radiation dose using GraphPad Prism 5.0 software.

RNA extraction and quantitative reverse transcription PCR. Total RNA was isolated using TRIzol reagent (Invitrogen) following the manufacturer's instructions. First-strand cDNA was obtained using the RevertAid™ First-Strand cDNA Synthesis kit (Fermentas International, Inc., Burlington, ON, Canada). For the quantitative analysis of HIF-1 α mRNA, the human β -actin gene was used as an internal control. Primer sequences were designed as follows: HIF-1 α sense, TGCAA CATGGAAGGTATTGC, and antisense, TTCACAAATCAG CACCAAGC; β -actin sense, TGC GTTACACCCTTTCT TGA, and antisense, CACCTTCACCGTTCCAGTTT. The Threshold cycle (Ct) values were measured by SYBR-Green PCR Master mix (Takara, Shiga, Japan) with Mx3000P (Stratagene, La Jolla, CA, USA). The Mx 3000P analysis program was used to analyze the results.

Cellular ATP content and extracellular lactate level. Cells (1x10⁶/ml) were seeded into 6-well plates. The extracellular lactate level was detected by Lactic acid assay kit (Jiancheng, Nanjing, China) following the manufacturer's instructions. The cellular ATP content was determined by measuring the luminescence with an ATP-dependent bioluminescence assay kit (Jiancheng). Briefly, the cells (1x10⁶/ml) were seeded into 6-well plates, lysed with lysis on ice, collected into 1.5 ml tubes, and centrifuged for 5 min at 10,000 x g; the Absorbance Microplate Reader was used to determine the OD of the supernatant (BioTek Instruments Inc., Winooski, VT, USA) at 630 nm (A630). The ATP content was calculated according to the formula provided by manufacturer's instructions. The above procedures were performed three times under the same condition.

Mitochondrial membrane potential and apoptosis analysis. The mitochondrial membrane potential was detected by the JC-1 kit (Beyotime Biotechnology, Haimen, China). Cell suspensions at 1x10⁵/ml were grown on glass coverslips with/without 2-MeOE2 for 2 h before irradiation treatment. At 30 min after irradiation, the cells were incubated with DPBS with 1 μ l JC-1 reagent at 37°C for 15 min. Then, cells were washed three times. The mitochondrial membrane potential was detected under the fluorescence microscope. The increase of green fluorescence suggests the decrease of the mitochondrial membrane potential.

For detection of apoptosis, 200,000 cells were seeded in a 6-well plate, pre-incubated with/without 2-MeOE2 for 2 h before irradiation treatment. The cells were harvested 48 h after irradiation. The cells were re-suspended in 100 μ l staining buffer with 5 μ l Annexin V- FITC (BD Pharmingen, San Jose, CA, USA) and stained at room temperature for 15 min followed by a quick staining with 1 μ g/ml propidium iodide (PI). The samples were analyzed on LSR II flow cytometer (Cytomics FC 500; Beckman Coulter, Fullerton, CA, USA), and the data were analyzed using Flow Jo software (FlowJo, Ashlan, OR, USA). Cells were stained with Annexin V-fluorescein isothiocyanate and PI and classified into 4 subpopulations as follows: viable cells (Annexin V and PI double-negative), apoptotic cells (Annexin V-positive),

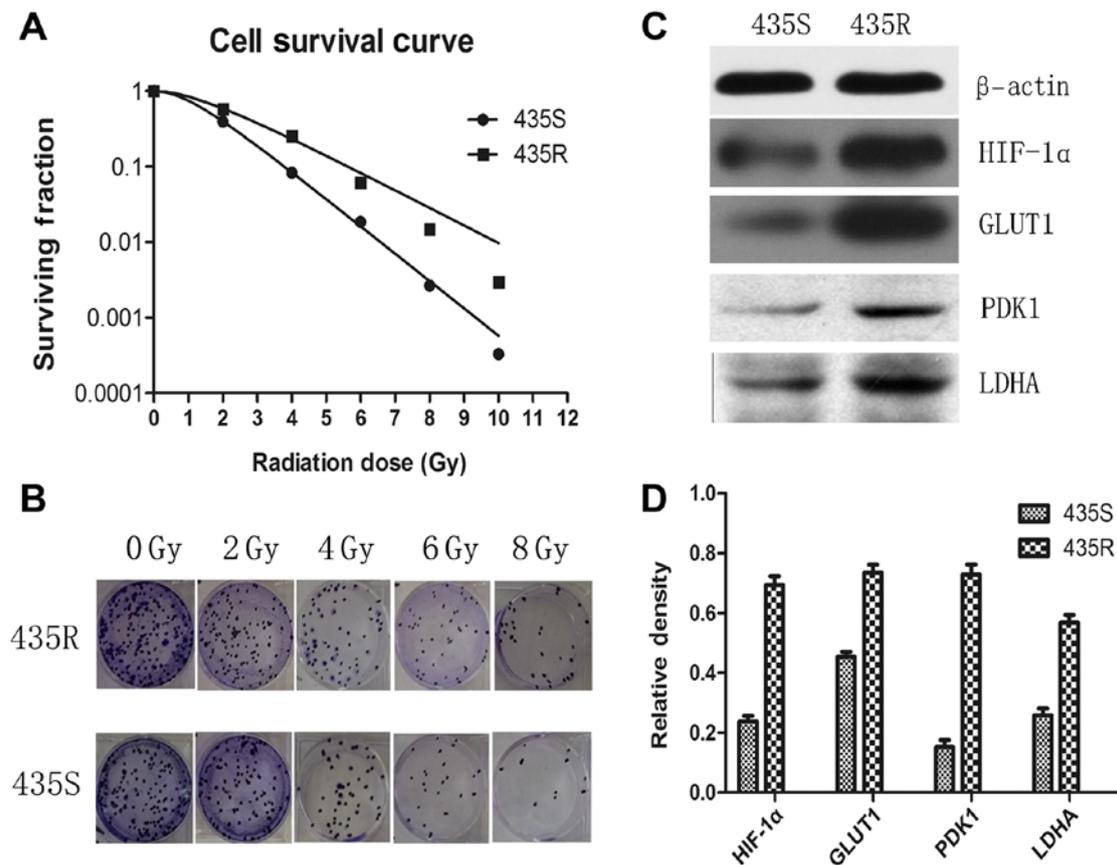


Figure 1. Expression of HIF-1 α and glycolysis-related proteins in radioresistant 435R and the parental 435S cells. (A) Radiosensitivity of 435R cells and 435S cells detected by colony formation assay. (B) Colony formation ability at the different irradiation doses of 435R cells and 435S cells. (C) Western blot analysis of HIF-1 α , PDK1 and glycolysis-related proteins GLUT1, LDHA. (D) Relative density of HIF-1 α , PDK1 and glycolysis-related proteins GLUT1, and LDHA normalized to their internal protein.

early dead cells (Annexin V and PI double-positive) and dead cells (PI-positive).

CCK-8 assay for cell proliferation. Cell proliferation was determined using a Cell Counting Kit-8 (CCK-8; Dojindo Kumamoto, Japan). Cells were seeded in 96-well plates, serum starved, and then treated with the experimental reagents as described (26). At the end of the incubation, CCK-8 solution was added and 3 h later, the absorbance at 450 nm ($A_{450\text{nm}}$) was measured with a microplate reader (Sunrise). Cell proliferation rates were calculated and normalized with the OD value of the first day.

Immunofluorescence. The immunofluorescence of γ -H2AX foci was used to determine the residual DNA double-strand breaks (DSBs). Melanoma cells were grown on glass coverslips with/without 2-MeOE2 for 2 h before irradiation, 30 min after irradiation, cells were fixed in 4% paraformaldehyde for 15 min before staining overnight with rabbit anti- γ -H2AX (ser139, Millipore Corp., Billerica, MA, USA) diluted in PBS 1:400. After staining for 60 min with goat anti-rabbit Cy3 (Millipore Corp.) diluted 1:100, the coverslips were mounted in Prolong Gold mounting medium containing DAPI (Life Technologies). Fluorescent images were obtained using Zeiss LSM710 confocal microscope (Leica Microsystems GmbH, Wetzlar, Germany) equipped with Plan-Apochromat X63/1.4

Oil DICII objective and analyzed using the ZEN2011 software and Adobe Photoshop CS5.

Protein isolation and western blot analysis. Briefly, 10×10^5 cells were seeded in 60 mm² plates and cultured in the presence or absence of 5 μ M 2-MeOE2 for 24 h. Following treatment, cells were lysed on ice with lysis buffer (Cell Signaling Technology, Danvers, MA, USA) supplemented with 1 mM phenylmethylsulfonyl fluoride (PMSF). Samples were centrifuged at 12,000 \times g for 10 min at 4°C, and supernatant was aliquoted and stored at -80°C for future use.

Total protein (30 μ g) was resolved in a 10% polyacrylamide gel using SDS-PAGE then transferred to a polyvinylidene difluoride (PVDF) membrane. Following transfer, membrane was blocked for 2 h at room temperature in 5% non-fat dry milk diluted in 0.1% Tween-20 in TBS (TBST) followed by an overnight incubation in blocking solution with primary antibodies. After washing, membranes were incubated for 2 h at room temperature with the appropriate peroxidase-conjugated secondary antibody, washed, and subjected to chemiluminescent substrate (Luminata Forte; Millipore Corp.). Membranes were imaged using the ChemiDoc XRS+ system (Bio-Rad, Hercules, CA, USA) and densitometry was performed using Image Lab software (Bio-Rad). Every target protein of our experiment was calculated by gray scanning using ImageJ (free software from NIH website), the relative

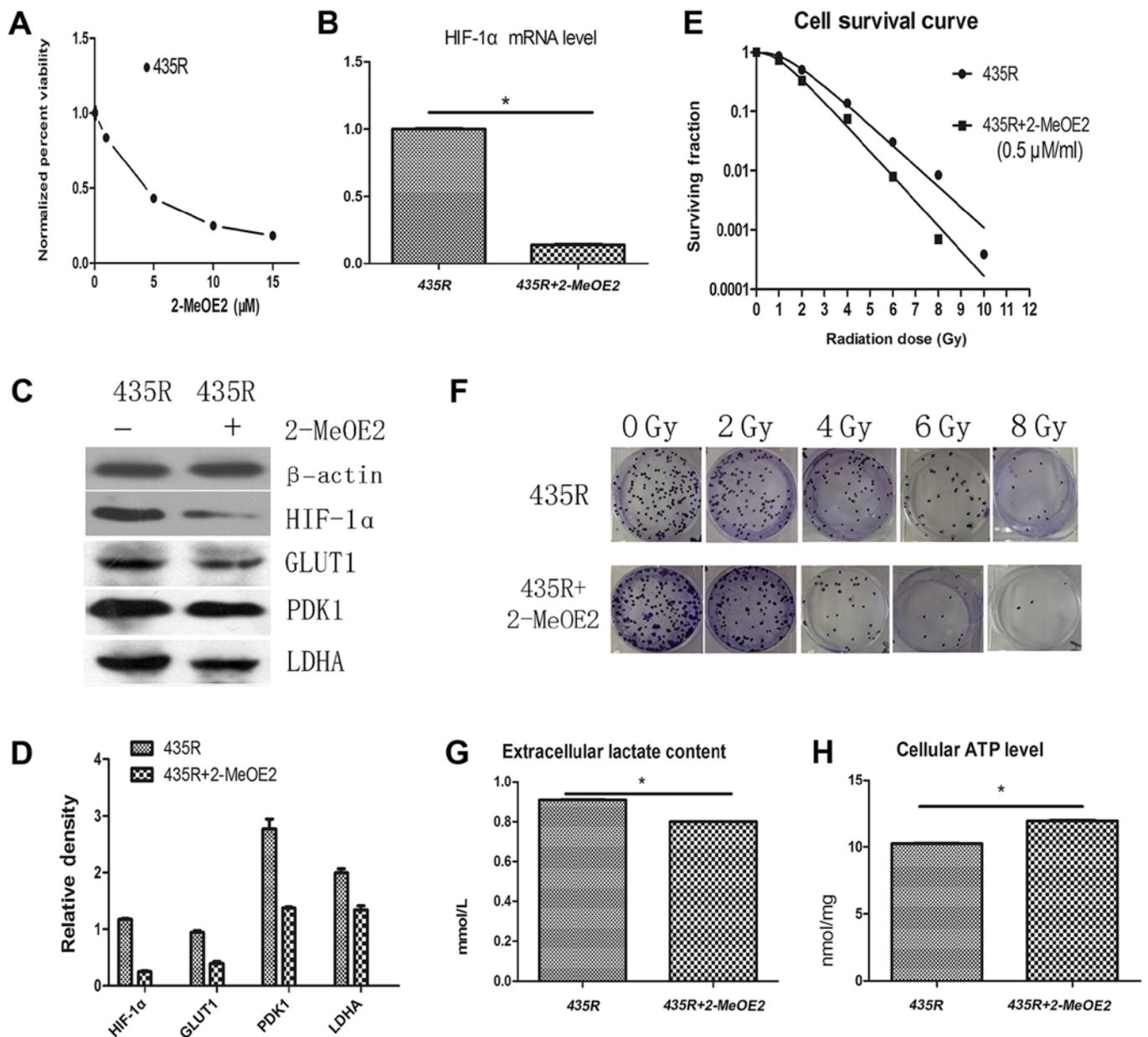


Figure 2. 2-MeOE2 decreases the radioresistance, PDK1 and glycolytic state of 435R cells. (A) The median IC_{50} of 2-MeOE2 for 435R cells. (B) The expression of HIF-1 α at mRNA level in 435R cells with/without 2-MeOE2, * $P < 0.0001$. (C) The expression of HIF-1 α , PDK1 and glycolysis-related proteins GLUT1, LDHA in 435R cells with/without 2-MeOE2. (D) Relative density of HIF-1 α , PDK1 and glycolysis-related proteins GLUT1, LDHA that normalized to its internal protein. (E) The change of radiosensitivity of 435R cells with 2-MeOE2 detected by colony formation assay. (F) Colony formation ability at the different irradiation doses of 435R cells with/without 2-MeOE2. (G) Extracellular lactate content in 435R cells with/without 2-MeOE2, * $P < 0.0001$. (H) Cellular ATP level in 435R cells with/without 2-MeOE2, * $P = 0.0125$.

density of target proteins were normalized to its marker internal protein. Primary antibodies used included rabbit anti-HIF1- α , β -actin, GLUT1, LDHA, PDK1. The quantification of band density was performed using ImageJ software.

Statistical analyses. All the experiments were repeated at least 3 times. Data are presented as the means \pm standard deviation (SD). Statistical analyses were performed using two-tailed Student's t-test (specified in the figure legend when paired analysis was used) and one way ANOVA using SPSS 19.0 and GraphPad Prism 5.0 software. The threshold for statistical significance was defined as $P \leq 0.05$.

Results

HIF-1 α and glycolysis-related proteins were highly expressed in radioresistant melanoma cells. In this study, we used a radioresistant cell model of melanoma cells to investigate radioresistance of melanoma cells more accurately. Radioresistant cell model MDA-MB-435R (435R) was established from melanoma cell line MDA-MB-435 (435S) through continuous X-ray radiation (22). The results of colony formation assay show that 435R cells displayed stronger radioresistance compared with its parental 435S cells (Fig. 1A and B), and demonstrate that we established the radioresistant cell model successfully.

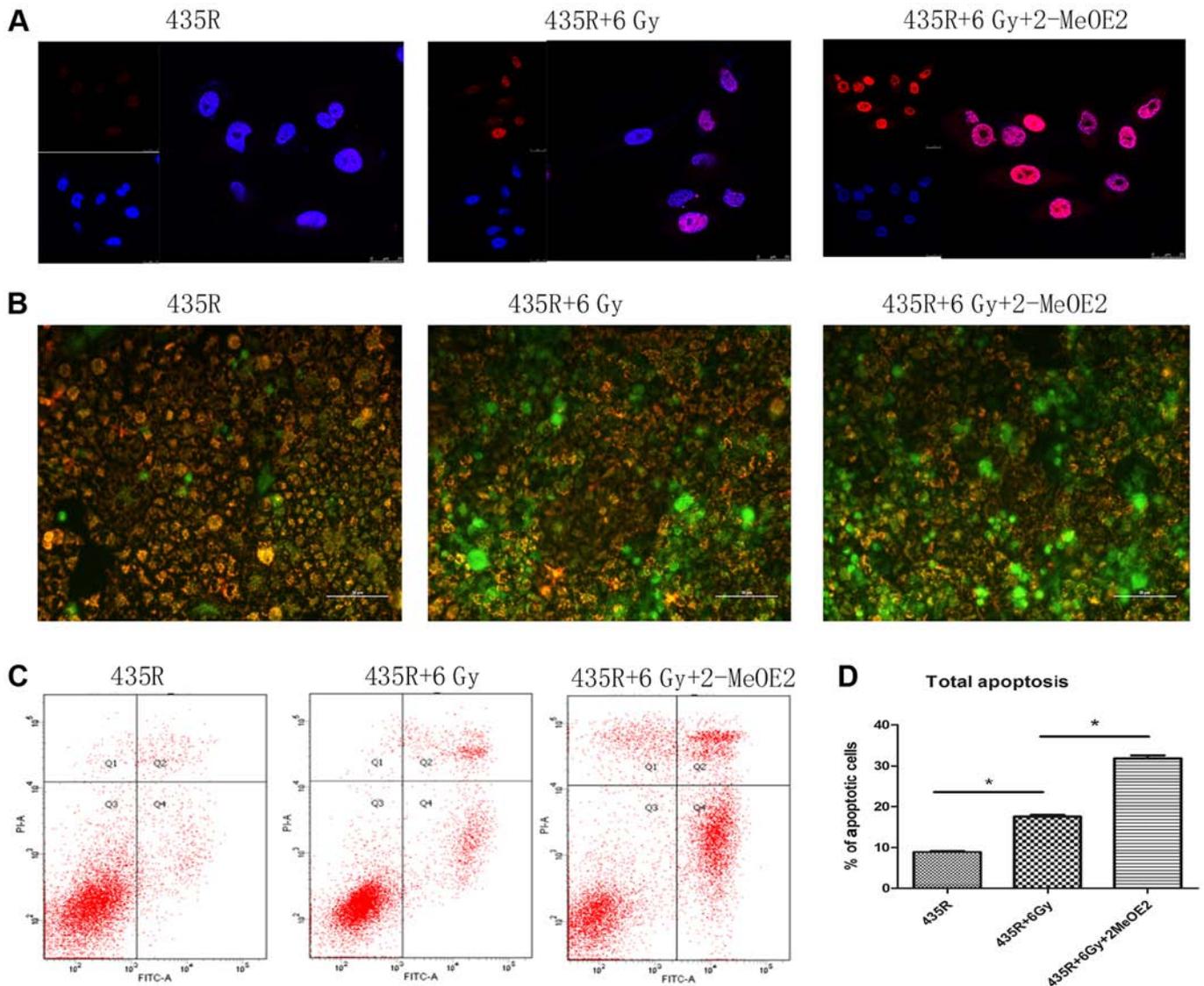


Figure 3. 2-MeOE2 increases DNA damage and apoptosis of 435R cells after irradiation. (A) γ -H2AX foci in combination treatment with 2-MeOE2 and RT or RT alone in 435R cells. Upper left corner of each image is the γ -H2AX foci with the red fluorescence in the nuclei. Lower left corner of each image is nuclei stained blue with DAPI. On the right side is the merged image of the left two images. (B) The change of mitochondrial membrane potential in combination treatment with 2-MeOE2 and RT or RT alone in 435R cells. The images demonstrate depolarization of mitochondrial membrane potential indicated by decrease of the red fluorescence combined with increase of the green fluorescence. (C) Apoptosis analyzed by flow cytometry in combination treatment with 2-MeOE2 and RT or RT alone in 435R cells. (D) The percentage of apoptosis on 435R cells with different treatment, $P < 0.001$.

Our data demonstrated that the expression of HIF-1 α in the 435R cells was higher than the parental cells 435S (Fig. 1C and D). Expression of the glycolysis-related proteins such as PDK1, GLUT1 and LDHA in the 435R cells were higher than the 435S cells (Fig. 1C and D).

2-MeOE2 decreases radioresistance, PDK1 and glycolytic state of 435R cells. The median IC50 of 2-MeOE2 was 4.3 μ M (48 h) for 435R cells (Fig. 2A). Our data indicated that 2-MeOE2 at the concentration $\leq 0.5 \mu$ M had no significant cytotoxic effect on 435R cells. Thus, we chose the 2-MeOE2 with 0.5 μ M (incubated with cells 6 h before irradiation) for our experiments. The expression of HIF-1 α at the mRNA and protein levels in the radioresistant cells were decreased after using the 2-MeOE2 (Fig. 2B and C). The results of colony formation assay show that survival fractions of 435R

cells with 2-MeOE2 decreased significantly at different irradiation doses compared with the cells without 2-MeOE2 (Fig. 2E and F).

Decrease of PDK1 expression of 435R cells was detected after inhibiting HIF-1 α with 2-MeOE2. Extracellular lactate content and the expression of glycolysis-related proteins GLUT1 and LDHA of the 435R cells were decreased (Fig. 2C, D and G), and the cellular ATP level of 435R cells was increased after the 2-MeOE2 treatment (Fig. 2H). The results indicated that inhibition of HIF-1 α decreased the glycolytic states of 435R cells.

2-MeOE2 increases DNA damage and apoptosis of 435R cells after irradiation. H2AX plays an essential role in the cellular response to DSBs of DNA induced by irradiation, the number of γ -H2AX foci is proportional to the amount of DSBs. As

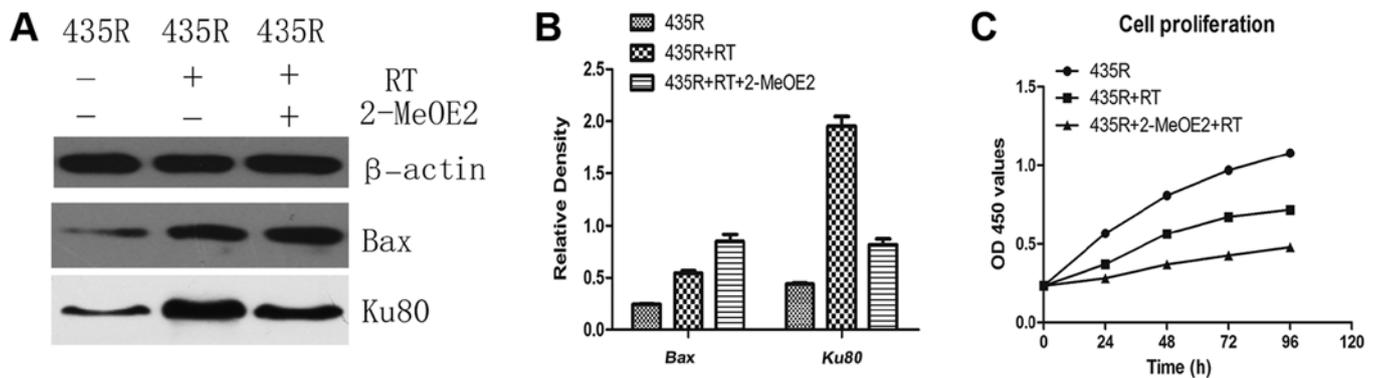


Figure 4. 2-MeOE2 decreases DNA damage repair and cell proliferation of 435R cells after irradiation. (A) Western blot analysis of Bax and Ku80 in 435R cells in combination treatment with 2-MeOE2 and RT or RT alone. (B) Relative density of Bax and Ku80 normalized to its internal protein. (C) Cell proliferation was detected using CCK-8 assays in 435R cells in combination treatment with 2-MeOE2 and RT or RT alone.

shown in Fig. 3A, The number of γ -H2AX foci in the 435R cells treated with 2-MeOE2 significantly increased compared with 435R without this after the same dose of irradiation.

It is well known that irradiation could cause DNA damage. If DNA damage can not be adequately repaired after irradiation, cells may progress towards apoptosis and/or necrosis. The decrease of mitochondrial membrane potential is an important factor in the early stage of apoptosis (27). The decrease of red fluorescence combined with the increase of green fluorescence suggested the loss of the mitochondrial membrane potential as shown in Fig. 3B. The mitochondrial membrane potential of 435R cells treated with 2-MeOE2 decreased to a larger extent than the parental cells after irradiation (Fig. 3B). The results of flow cytometry showed that inhibition of HIF-1 α increased the apoptotic percentages of radioresistant cells after irradiation (Fig. 3C and D). The results of western blot analysis showed the expression of Bax was increased after irradiation treated with 2-MeOE2 (Fig. 4A and B).

2-MeOE2 decreases DNA damage repair and cell proliferation of 435R cells after irradiation. The results of western blot analysis showed expression of DNA damage repair-related protein Ku80 in the 435R cells with 2-MeOE2 was decreased compared the 435R cells without treatment after irradiation (Fig. 4A and B). Cell proliferation was detected using CCK-8 assays. The result showed that inhibition of HIF-1 α with 2-MeOE2 can enhance suppression of cell proliferation of 435R cells induced by irradiation (Fig. 4C).

DCA decreases radioresistance and glycolytic state of 435R cells. In order to verify inhibition of HIF-1 α decrease the radioresistance of 435R cells via HIF-1 α /PDK1-mediated glycolysis, radioresistance of 435R cells were detected after inhibiting glycolytic state with DCA. The result of colony formation assay show that survival fractions of 435R cells with DCA decreased significantly at different irradiation doses compared with those without DCA (Fig. 5A and B). The expression of glycolysis related proteins GLUT1, LDHA and the extracellular lactate content of the 435R cells were decreased (Fig. 5C-E), cellular ATP level of the 435R cells was increased after the DCA treatment (Fig. 5F).

DCA increases DNA damage and apoptosis of 435R cells after irradiation. The number of γ -H2AX foci in the 435R cells treated with DCA significantly increased compared with 435R without it after the same dose of irradiation (Fig. 6A). The mitochondrial membrane potential of 435R cells treated with DCA decreased to a larger extent than the 435S cells after irradiation (Fig. 6B). The results of flow cytometry showed that DCA could increase the apoptotic percentage of radioresistant cells after irradiation (Fig. 6C and D). Furthermore, the expression of Bax was increased after irradiation when treated with DCA (Fig. 7A and B).

DCA decreases DNA damage repair and cell proliferation of 435R cells after irradiation. The expression of Ku80 in the 435R with DCA was decreased compared to those without DCA after irradiation (Fig. 7A and B). The result of cell proliferation assays showed that inhibition of PDK1 with DCA can enhance suppression of cell proliferation of 435R cells induced by irradiation (Fig. 7C).

Discussion

The existence of radioresistance limits the efficacy of radiotherapy for melanoma. However, recent studies suggest that under certain clinical circumstances, radiotherapy may play a significant role in the treatment of melanoma (28-31). Improving the radiosensitivity of melanoma cells may allow the expanded use of radiotherapy for melanoma. The results of our study indicated that the level of HIF-1 α and glycolysis in the radioresistant cells were higher than its parental cells. Inhibition of HIF-1 α sensitizes radioresistant melanoma cells 435R to X-ray irradiation by targeting the energy metabolism regulated by glycolysis.

HIF-1 α is an important regulator in the cellular and systemic homeostatic responses to hypoxia by activation of gene transcription (32). HIF-1 α overexpression is associated with radioresistance of several tumors. Therefore, selective HIF-1 α inhibitors are promising targeted compounds for adjuvant radiosensitizing therapy. In our study, we inhibited the expression of HIF-1 α in the radioresistant melanoma cells 435R with the special chemical molecule 2-MeOE2. 2-MeOE2 as a special inhibitor of HIF-1 α was shown to inhibit the expression

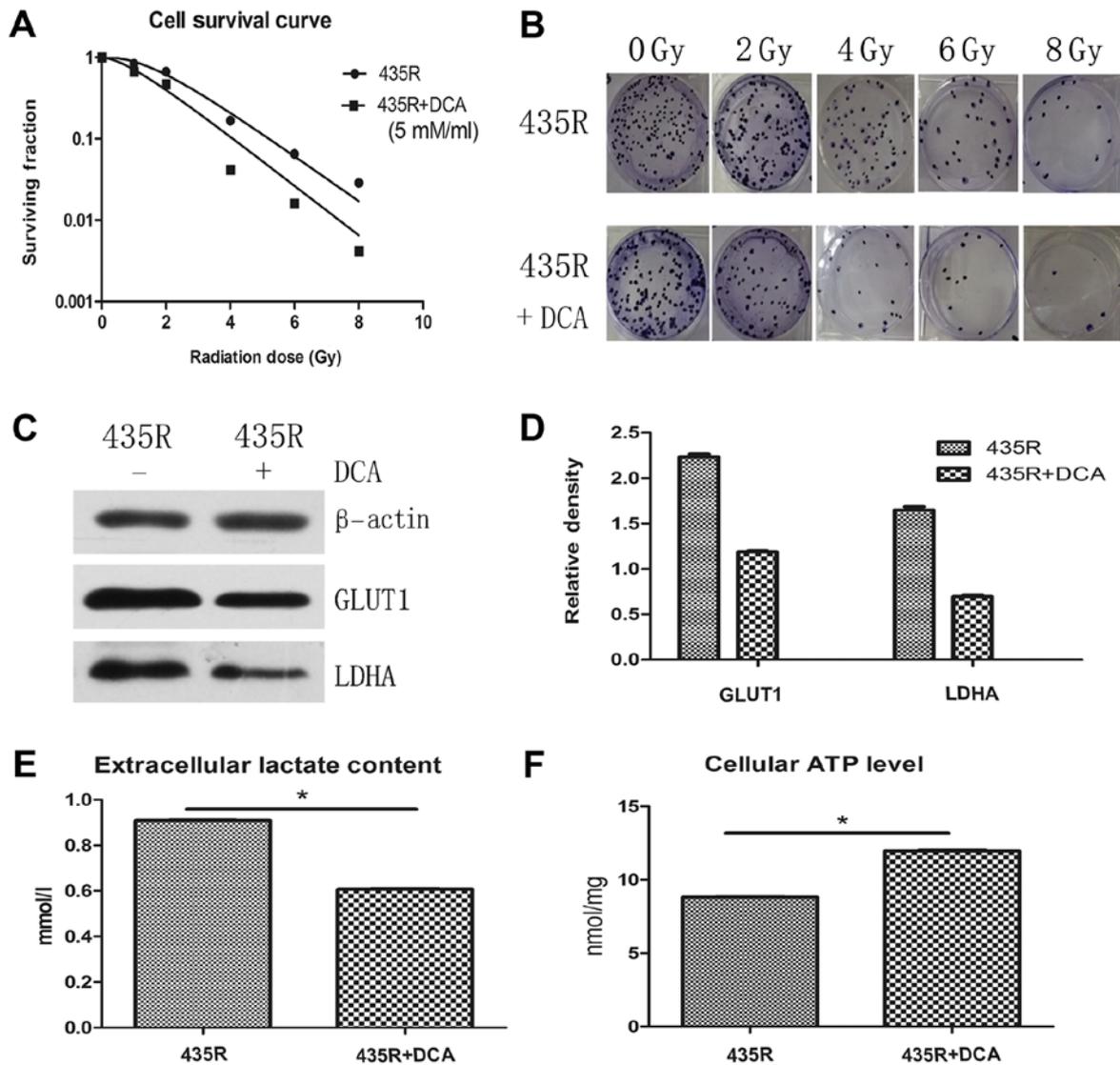


Figure 5. DCA decreases the radioresistance and glycolytic state of 435R cells. (A) The change of radiosensitivity of 435R cells with DCA detected by colony formation assay. (B) Colony formation ability at the different irradiation doses of 435R cells with/without DCA. (C) The expression of glycolysis-related proteins GLUT1, LDHA in 435R cells with/without DCA. (D) Relative density of GLUT1, LDHA that normalized to its internal protein. (E) Extracellular lactate content in 435R cells with/without DCA, *P<0.001. (F) Cellular ATP level in 435R cells with/without DCA, *P<0.001.

of HIF-1 α in a dose-dependent manner in cancer cells and blocking HIF-1 α nuclear accumulation by depolymerizing microtubules (4). When detected the expression of proteins in our study, use of β -tubulin was avoided as the internal reference to prevent the influence of change of level of microtubules. We also avoided use of GAPDH as the internal reference to prevent influence of the change of level of GAPDH caused by glycolytic activity. 2MeOE-2 is a naturally occurring derivative of estradiol, which has been shown to be orally active, well-tolerated small molecule that possesses antitumor and anti-angiogenic activity (33). There are several completed Phase I/II clinical trials for the effectiveness of 2MeOE-2 in various cancers (34-36).

Activation of glycolytic genes by HIF-1 α is considered to be a very important factor for metabolic adaptation to hypoxia, with increased conversion of glucose to pyruvate and subsequently to lactate. HIF-1 also suppressed glucose metabolism through the tricarboxylic acid cycle by directly transactivating PDK1. PDK1 is an important glucose metabolism enzyme

for the Warburg effect, promotes the conversion of pyruvate to lactic acid and ATP in the presence of oxygen (aerobic glycolysis), generating the necessary amount of energy needed for rapid cellular proliferation (10). Lactate dehydrogenase A (LDHA) is an enzyme which plays a critical role in the glucose metabolism. Expression and post-transcriptional modification of LDHA are regulated by several known oncogenes and deacetylases, such as MYC and HIF-1 α (37-39). GLUT1 is a facilitative glucose transporter which belongs to the solute-linked carrier gene family SLC2, and the elevated expression of GLUT1 has been documented in most cancers (40). We have detected the indicators of glycolysis in the radioresistant cell model and the parental cells, and the results show that the expression of PDK1, LDHA and GLUT1 in the radioresistant cells were higher than its parental cells. In addition, extracellular lactate production in the radioresistant cells were also higher than the parental cells. These results show evidence that highly level of HIF-1 α and glycolysis has an important relationship with the radioresistance of melanoma cells.

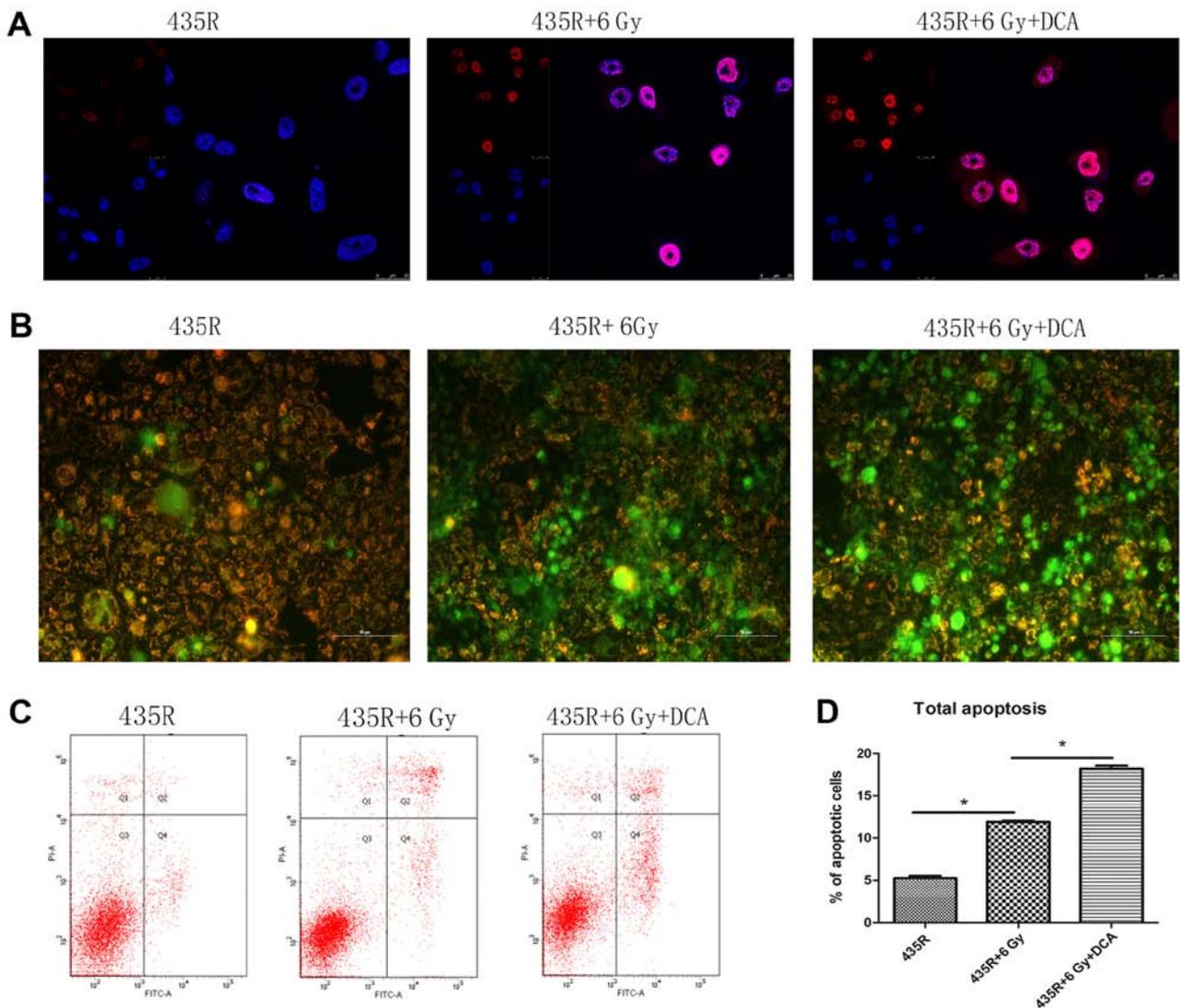


Figure 6. DCA increases DNA damage and apoptosis of 435R cells after irradiation. (A) γ -H2AX foci in combination treatment with DCA and RT or RT alone in 435R cells. Upper left corner of each image is the γ -H2AX foci with the red fluorescence in the nuclei. Lower left corner of each image is nuclei stained blue with DAPI. On the right of each image is the merged image of the left two images. (B) The change of mitochondrial membrane potential in combination treatment with DCA and RT or RT alone in 435R cells. The images demonstrate depolarization of mitochondrial membrane potential indicated by decrease of the red fluorescence combined with increase of the green fluorescence. (C) Apoptosis analyzed by flow cytometry in combination treatment with DCA and RT or RT alone in 435R cells. (D) The percentage of apoptosis on 435R cells with different treatment, *P<0.001.

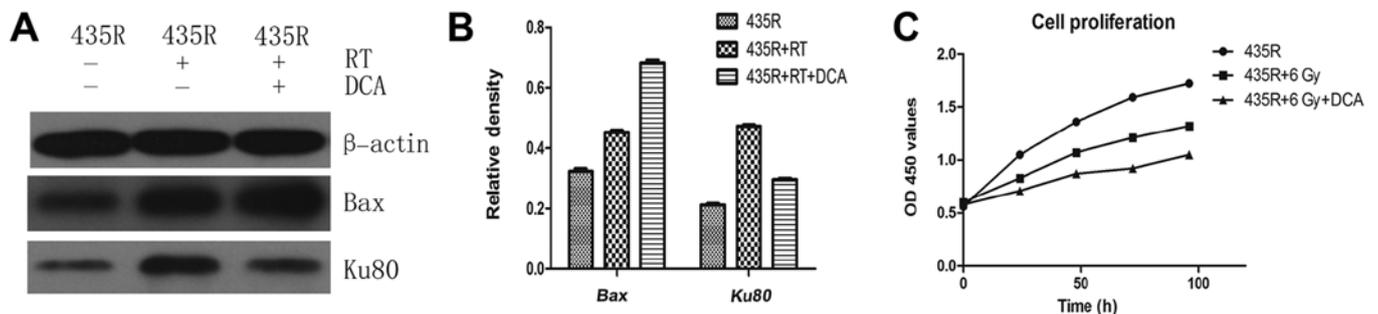


Figure 7. DCA decreases DNA damage repair and cell proliferation of 435R cells after irradiation. (A) Western blot analysis of Bax and Ku80 in 435R cells in combination treatment with DCA and RT or RT alone. (B) Relative density of Bax and Ku80 normalized to their internal protein. (C) Cell proliferation was detected using CCK-8 assays in 435R cells in combination treatment with DCA and RT or RT alone.

The elevated glycolytic state of cancer cells not only provides a growth advantage but also correlates strongly with radiore-

sistance (15,16). Therefore, inhibition of glycolytic metabolism during or before radiotherapy may be potentially exploited

for identifying new methods to overcome radioresistance. In our study, inhibition of HIF-1 α caused a decrease of PDK1 expression and radioresistance in the radioresistant cells. In order to verify whether inhibition of HIF-1 α decreased the radioresistance of 435R cells via glycolysis, radioresistance of 435R cells were detected after inhibiting glycolytic state with DCA. These results demonstrated that inhibition of HIF-1 α sensitizes 435R cells to X-ray irradiation by regulating PDK1-mediated glycolysis. Some special glycolysis-related enzymes which can regulate the activity of glycolysis can be the target of further investigation into combination with radiotherapy.

Radiation can induce single-strand or double-strand DNA breaks (SSBs and DSBs) by inducing the oxidation of DNA bases causing lesions in the DNA (41). As strong activators of apoptosis, DSBs have the most harmful effect on cell survival (42); cell death was induced by the persistence of DSBs if not repaired (43). Zou *et al* found that 2-MeOE2 could enhance the incidence of radiation-induced genomic damage (44). Ku80/XRCC5 is one of the XRCC (X-ray Repair cross-complementing) families which play a role in protecting mammalian cells from DNA damage caused by ionizing radiation and antitumor chemotherapy agents (45). Melanoma has been considered to be a highly radioresistant tumor due to its efficient DNA repair mechanism (46). Our results showed that the number of the foci of γ -H2AX in the radioresistant cells with inhibition of HIF-1 α after irradiation was higher than the cells without this. The expression of Ku80 in the radioresistant cells with 2-MeOE2 was lower than the cells without after the same dose of irradiation. It demonstrated that inhibition of HIF-1 α -mediated glycolysis could increase the DSBs and suppress the DNA damage repair of radioresistant melanoma cells after irradiation.

Inhibition of HIF-1 α can increase the expression of apoptosis promoting protein Bax and the apoptosis rate of 435R cells significantly after radiation than the cells after radiation without the inhibitor. The final results showed that inhibition of HIF-1 α could inhibit the cell proliferation of 435R after irradiation. Mueck *et al* observed that under a normoxic condition, 2-MeOE2 stimulated apoptosis, an effect probably due to Bcl-2 and Bcl-xL phosphorylation and the subsequent inhibition of the anti-apoptotic effects (47). The experiment of Long *et al* demonstrated that 2-MeOE2 effectively inhibited the protein expression of HIF-1 α , which significantly increased the late stage of radiation-induced apoptosis of keloid fibroblasts (48). The results of Aquino-Gálvez *et al* showed a dose-dependent inhibition of cell growth for 2-ME-treated normoxic cells (49).

In summary, our study detected high expression level of HIF-1 α and more glycolysis activity in the radioresistant melanoma 435R cell model than its parental cells. HIF-1 α , as a key regulator of PDK1, its inhibition can cause decrease of radioresistance and PDK1 of 435R cells. The possible mechanism of this effect might that inhibition of HIF-1 α suppresses the glycolysis by PDK1, then the balance of energy metabolism through glycolysis is disrupted. Since Warburg effect is the major source of energy in the highly glycolytic cancers (13), imbalanced metabolism may be the critical factor that influenced biological processes, including repair of DNA breaks, cell proliferation and other processes which can change the radiosensitivity of

cells. These observations have raised the possibility that targeting metabolic pathways that the cancer cell depends on may be a useful therapeutic strategy. Some special glycolysis-related enzymes could be new targets for further investigation in combination with radiotherapy, and more cell lines are needed in the future experiments.

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