

# Characterizing key nucleotide polymorphisms of hepatitis C virus-disease associations via mass-spectrometric genotyping

YUTA HORIUCHI<sup>1\*</sup>, JASON LIN<sup>2\*</sup>, YUI SHINOJIMA<sup>3</sup>, KYOKO FUJIWARA<sup>3,4</sup>,  
MITSUHIKO MORIYAMA<sup>1</sup> and HIROKI NAGASE<sup>2</sup>

<sup>1</sup>Institute of Gastroenterology, Nihon University School of Medicine, Itabashi-ku, Tokyo 173-8610, Japan;

<sup>2</sup>Laboratory of Cancer Genetics, Chiba Cancer Center Research Institute, Chuo-ku, Chiba 260-8717, Japan;

<sup>3</sup>Department of Cancer Genetics, <sup>4</sup>Division of General Medicine, Department of Medicine,  
Nihon University School of Medicine, Itabashi-ku, Tokyo 173-8610, Japan

Received June 21, 2017; Accepted October 16, 2017

DOI: 10.3892/ijo.2017.4209

**Abstract.** As more than 80% of hepatocellular carcinoma patients in Japan also suffer from hepatitis C virus infections some time in their medical history, identifying genetic aberrations associated to hepatitis C virulence in these individuals remains a high priority in the diagnosis and treatment of hepatocellular carcinoma. From the BioBank Japan Project, we acquired 480 subjects of hepatocellular carcinoma, chronic hepatitis and liver cirrhosis, and genotyped 131 clinically relevant host single nucleotide polymorphisms to survey the potential association between certain risk alleles and genes to a patient's predisposition to hepatitis C and liver cancer. Among those polymorphisms, we found 12 candidates with statistical significance to support association with hepatitis C virus susceptibility and genetic predisposition to hepatocellular

carcinoma. SNPs in genes such as *XPC*, *FANCA*, *KDR* and *BRCA2* also suggested likely connections between hepatitis C virus susceptibility and the contraction of liver diseases. Single nucleotide polymorphisms reported here provided suggestions for genes as biomarkers and elucidated insights briefing the linkage of hepatitis C virulence to the alteration of healthy liver genomic landscape as well as liver disease progression.

## Introduction

The seriousness of hepatitis C virus (HCV) infections lies in their elevation of risks pertaining to various life-threatening liver conditions such as liver cirrhosis (LC), chronic hepatitis (CH) as well as hepatocellular carcinoma (HCC). For instance, the estimated risk of hepatocellular carcinoma has been reported to be often 15 to 20 times higher in subjects infected with HCV (1). Statistics also suggest that 75-85% of people infected will become chronic carriers, and 10-15% of HCV infection cases will advance to cirrhosis within the first 20 years (2), along with increased risks of developing HCC. In Japan, ~80% of patients suffering from HCC is caused by prior or concurrent HCV (3), suggesting potential links between possible genetic predisposition and infection incidence. While genetic aberrations can direct and drive carcinogenesis, exact details, especially with the ethnic Japanese population, linking HCV virulence to HCC susceptibility (and to a lesser extent LC and CH) remain opaque. Preventative treatment options for HCV are also lacking in a similar regard, although a meta-analysis by Colombo and Iavarone had suggested the possibility of reducing HCC risks in a small fraction of cases with interferon administration (4). To-date, while various studies have sought to characterize the effect of genotypic alterations in liver diseases, the role of SNPs in liver diseases still remain poorly understood (5-11). Additionally, while genes such as IFN $\gamma$  and IL-28B have been reported to harbor potential associations to HCV infections leading to conditions such as fibrosis and jaundice (12-14), no clinical screening and characterization studies have been conducted between HCV and severe liver conditions such as CH, LC and HCC.

With an estimated 143 million people infected with hepatitis C worldwide (15), early genetic screening can have a beneficial role in greatly improving the quality of life for those

**Correspondence to:** Professor Mitsuhiro Moriyama, Institute of Gastroenterology, Nihon University School of Medicine, 30-1 Oyaguchikami-cho, Itabashi-ku, Tokyo 173-8610, Japan  
E-mail: moriyama.mitsuhiro@nihon-u.ac.jp

Professor Hiroki Nagase, Laboratory of Cancer Genetics, Chiba Cancer Center Research Institute, 666-2 Nitona-cho, Chuo-ku, Chiba 260-8717, Japan  
E-mail: hnagase@chiba-cc.jp

\*Contributed equally

**Abbreviations:** HCC, hepatocellular carcinoma; SNP, single nucleotide polymorphisms; HCV, hepatitis C virus; CH, chronic hepatitis; LC, liver cirrhosis; PCR, polymerase chain reaction; DNA, deoxyribose nucleic acid; JSNP, Japanese Single Nucleotide Polymorphisms databank; TMM, Tohoku Medical Megabank; AMACR,  $\alpha$ -methylacyl-CoA racemase; ARHGAP, Rho GTPase-activating proteins; ADAMTS, a disintegrin and metalloproteinase with thrombospondin motifs; G1000, the 1000 Genomes Project

**Key words:** glioblastoma, microRNA-485, p21 activated kinases 4, proliferation, migration, invasion, apoptosis

afflicted with liver diseases. With the establishment of the BioBank Japan Project that stores and maintains a number of annotated liver disease cases, we decide to explore the possibility, if any, that certain genetic risk factors are associated with the prevalence of HCV infections to hepatocellular carcinoma in the Japanese population by MassARRAY genotyping, a MALDI-TOF mass spectrometry-based approach well demonstrated in medium- to large-scale cohort studies (16) to analyze a set of specific polymorphisms. We herein present one of the earliest studies attempting to assess several clinically relevant marker gene and variant candidates linking HCV to liver cancer, as well as CH and LC for the purpose of comparison, in the Japanese population using DNA extracted from blood specimens. For each candidate, we then assessed its significance by comparing genotypic frequencies against records from the Japan Single Nucleotide Polymorphisms databank (JSNP), NCBI dbSNP and the Tohoku Medical Megabank (TMM) datasets (17-20) to elucidate their potential roles as risk factors in HCV-induced liver diseases and their pertinence in other ethnic groups.

## Material and methods

**SNP selection.** Among published SNPs and polymorphisms in genes relevant to cancer in recent literature, we generated a preliminary list of SNP candidates for screening. To broaden the scope, this list was later expanded to include a small number of SNPs that had few reports of clinical significance to-date, but were well-conserved, disease-associated and believed to impact protein structure by SNPs3D searches (21). In all, 131 SNPs over 4 primer sets were found to meet the MassARRAY prerequisites and selected for analysis (tabulated data of SNPs and corresponding primers available upon request).

**Specimen collection and subject demographics.** Blood DNA samples from a collection of CH (200), LC (80) and HCC (200) subjects were obtained from the BioBank Japan Project (22). Hepatitis subjects had a gender makeup of 103/97 male/female, with 102 over the age of 50 (mean 54±15). Samples were confirmed to be HCV-negative and cancer-free. The Cirrhosis test group consisted of 45 males and 35 females, with 71 people over the age of 50 (mean 65±11) and similarly confirmed to be HCV-negative and cancer-free. Among HCC patients of 103 men and 97 women, 194 were over 50 (mean 69±8) and all cases were confirmed to be HCV-positive. No other medical history (including history of hepatitis virus B infection) or personal information was obtained. For the purpose of statistical assessment, a simulated control ('healthy') set based on JSNP (last updated May, 2014) and NCBI data (build 142, October, 2014) was used. In cases where multiple genotypes exist from both JSNP and NCBI, prevailing JSNP results were used for the control set. The TMM 2KJPN database (release June, 2016) was also consulted for comparative analysis of variant allele frequencies for the Japanese population.

**SNP detection by single-base extension.** DNA specimens (5 µg) were diluted to the stock concentration of 10 ng/µl prior to PCR using the iPLEX Gold reagent kit (Sequenom, San Diego, CA, USA). Each specimen (1 µl) was mixed with the primer mix (500 nM) prior to SNP isolation, and

6/96 samples were then randomly selected to confirm the extent of PCR by gel electrophoresis. Four primer mixes, each containing 33 SNPs, were prepared to the same final composition per Sequenom's instructions. PCR products underwent shrimp alkaline phosphatase (0.5 unit) treatment to dephosphorylate unincorporated deoxynucleotides at 37°C for 40 min followed by 85°C for 5 min. Extension reactions were performed following the iPLEX-Extend protocol by preparing each sample in a solution of 0.62 µl water, 0.2 µl 10X iPLEX buffer plus, 0.2 µl iPLEX termination mix, 0.94 µl extend primer mix, 0.04 µl iPLEX enzyme for reaction as follows: 94°C (step 1, 30 sec), 94°C (step 2, 5 sec), 52°C (step 3, 5 sec), 80°C (step 4, 5 sec), in which steps III and IV were repeated for 5 cycles followed by 40 cycles of steps II-IV before the final step at 72°C for 3 min. Following instructions from the SpectroChip Chip and Resin kit (Sequenom), samples were characterized by mass spectrometric analysis on a MassARRAY compact MALDI-TOF mass spectrometric analyzer. Oligonucleotides were binned with replicates per manufacturer's recommendations to minimize calling failures. Data were acquired and processed using EpiTYPER 1.0 in the Sequenom MassARRAY Workstation suite. For illustrative purposes, some data and cluster plot/spectrum snapshots were exported to XML and tab-delimited text files, respectively, for replotting and annotation with R 3.2 (available R-project.org).

**Statistical analysis.** Statistical analyses were performed using SPSS 15.0 (IBM Corporation, Armonk, NY, USA) and R. Allele and genotypic differences between healthy (see Specimen collection) and diseased subjects as well as ethnicity were evaluated by the  $\chi^2$  test of significance. Ancestral/reference and alternative allele were reordered for presentation according to dbSNP conventions; in certain cases where the ancestral allele was ambiguous, e.g. *PMS2* (rs1805321), the statistically dominant allele in dbSNP was selected as the ancestral allele (e.g. C for rs1805321). Comparisons to the TMM dataset were determined in R, but not used to determine significance as only allele frequencies were available. A predefined  $\alpha$ -level of 0.05 was used for statistical assessment. Per the exploratory and targeted nature of this study, P-values were not corrected for multiple comparisons; instead, the authors advised readers to use corrected significance levels, for instance the Dunn-Šidák corrected  $\alpha'=0.00039$  ( $m=131$ ) when cross-examining results here with other non-targeted genotyping association studies to account for potential consequences of multiple hypothesis testing.

**Changes to local protein structure.** As specimens obtained for this study came from a Japanese population, we consulted annotated records from TMM to determine potential changes to local protein structures. For non-synonymous SNPs meeting the significance level, such changes were predicted by evaluating the impact of amino acid substitutions by a composite probability from HumVar PolyPhen-2 (2.2.2r398) and Grantham scores (23,24). PolyPhen2 FPR thresholds were appraised at 10/20%. For the purpose of calculation, the Grantham value of all non-identical amino acid replacements were transformed to a kernel density function in R and re-evaluated as cumulative probabilities. Scores were averaged by the geometric mean (square root of the product of two probabilities).

**Interaction and gene enrichment analysis.** Potential interactions among the list of significant candidate genes were analyzed by GeneMANIA to identify the top 25 related genes. Interactions were selected to allow at most 25 genes with 25 attributes, and filtered by the presence of possible associations via co-expression, co-localization, genetic and physical interactions, pathway information as well as predicted interactions. Networks were analyzed and visualized in Cytoscape. Statistically enriched Reactome pathways (version 58) were identified by the Panther overrepresentation test with a predefined  $\alpha$ -level of 0.05 to assess individual pathway significance.

**Linkage disequilibrium analysis.** Potential linkage disequilibria were examined using LDproxy in the LDlink suite (25) to possible linkages among the significant SNPs, using the Japanese population genotype data from the Phase 3 of the 1000 Genomes Project as input. Proxy locus pairs with correlation coefficients ( $R^2$ )  $>0.80$ , and RegulomeDB scores of 1-5 (including subgroups of 1-3) were retained for further analysis. Variant pair associations, as a function of the correlation coefficient between loci pairs, were visually examined in Cytoscape as network diagrams to identify common functionally active proxy variants.

**Cohort allele frequency dilution.** VCF files containing individual variant data information from the Phase 3 1000 Genomes Project (26) were acquired via the Data Slicer (GRCh37 release 89, last accessed August, 2017) for each of the 12 candidate polymorphisms. To test the effect on changes in candidate allele frequencies if samples from this population were mixed with the cohort,  $n=0-480$  randomly selected genotypes from G1000 were mixed with the cohort, also randomly selected to make up a final sample size of 480, in order to examine the resultant reference (A) and alternative (B) allele counts as well as the mean allele frequencies (%B) as a function of  $n$ . In a separate validation, both the cohort and G1000 populations were randomly sampled (100 individuals/set) to evaluate the likelihood that both populations contained different mean allele frequencies. Hundred random trials were performed for both validations in R, with statistical significance by two-sample t-tests.

## Results

We found 12 out of the 131 SNPs investigated (Table I) to be potentially associated; among those, 10 SNPs, **1**, [ $\alpha$ -methylacyl-CoA racemase (*AMACR*) rs34677], **2** (*ARHGAP8*, rs2071762), **3** (*KDR*, rs2305948), **4** (*SELE*, rs5361), **5** (*XPC*, rs2228001), **6** (*FANCA*, rs2239359), **8** (*BRCA2*, rs766173), **9** (*PCDHGB7*, rs17208397), **11** (*PMS2*, rs1805321) and **12** (*ADAMTS16*, rs16875054) were non-synonymous; SNP **7** (*LRRN3*, rs17439799) was intronic and **10** (*ERCC4*, rs2020955) was synonymous. A full table containing genotypes, alleles and call rates for the 131 selected SNPs are available upon request and at [github.com/cccri-jlin/marray\\_si](https://github.com/cccri-jlin/marray_si). Discrepancies did exist in allele distribution between database annotations, e.g. *WRN* (rs1346044) contained T/C alleles in NCBI but A/G in JSNP.

We also evaluated the likelihood that these variants differed from corresponding records in the TMM data-

bank by their maximum  $\chi^2$  statistics between the JSNP and dbSNP datasets as a potential screen to confirm the ethnic specificity of those variants to the Japanese population, but did not assign significance based on these statistics as the TMM databank contained considerably less publicly available information than JSNP or dbSNP at the time of analysis. Most of the variants appeared to have allele frequencies that varied across different phenotypes, some with only one subgroup that strongly deviated from the other two, e.g. **3** in only HCC, and others in multiple subgroups, e.g. **4** and **8**. Generally, the HCC subgroup frequently exhibited allele frequencies that deviated from the other two phenotypes, for example candidates **5** and **6**. For SNPs with significant differences in genotype and allele frequency, all but **1** (*AMACR*) expressed differential compositions across different ethnic groups (Table II), suggesting that they may be specific to the Japanese population.

We also explored whether potential linkage disequilibria existed among candidates in Table I to ascertain the possibility of non-random association with other alleles as means to confirm their true phenotypic implications on the liver diseases. While some of those SNPs did have proxy pairs that would suggest some extent of linkage disequilibria (available upon request), we observed no common proxy variants among them (Fig. 1B and C). Annotations of those proxy variants also suggested either synonymous mutations or the lack of clear biological function. As not all variants in Table I were on the same chromosome, and most linked proxy pairs were outside the immediate proximity (only 12/170 were within 1,000 bp; data available upon request), potential associations on allele frequencies of SNPs in Table I by those proxy variants most likely had no effect on disease outcomes.

## Discussion

Roughly 10% of the SNPs genotyped in this study displayed critical roles in liver functions and various cancer phenotypes. For instance, *AMACR* is an ubiquitously expressed enzyme responsible for bile branched-chain fatty acid metabolism (27) that undergoes increases in mRNA and protein expressions in colorectal and prostate cancers as well as pre-cancerous hyperplastic polyps in the large intestine (28,29). Co-increases of COX-2 expression and *AMACR* could also potentially lead to immune suppression, neoplastic changes and tumor invasion (30). As polymorphisms in *AMACR* have been implicated in prostate and colorectal tumorigenesis (28), its possible contribution in HCV infection-induced disease phenotypes shall not be overlooked. Similarly, despite the lack of prior evidence suggesting direct association of *ARHGAP8* to HCV-induced HCC, the loss of Rho-suppressing *ARHGAP7* has been implicated in multiple cancers including HCC (31). Regulatory features of RhoGAP via Rho has been said to attribute to reductions in hepatic fibrosis, suggesting that a loss of function variant may induce liver cirrhosis in a manner independent from HCV infection. Adding to the ability for statins to attenuate liver fibrosis in chronic HCV infections (32), inter-regulatory feedback of Rho acylation by members of the ARHGAP family, for instance *ARHGAP8*, may well attribute to the aberrant dysregulation, via HCV infections, during HCC development (33).

Table I. Genotypes, alleles and impact on local protein structure of statistically significant SNPs and corresponding JSNP and dbSNP entries.

ID	Gene (dbSNP rsID)	A/B	DP	AA	AB	BB	Sum	A	B	ST	dbSNP $\chi^2$	dbSNP P-value	dbSNP ss#	JSNP $\chi^2$	JSNP P-value	TMM A/B	TMM $\chi^2$	TMM P-value	Mut	PV	GV	PG	GMI
1	AMACR (34677)	T/G	HCC	4	22	149	175	30	320	GT	41.7253	0.0000	68931918	8.1811	0.0167	C/A (3717/379)	0.1793	0.6720	Q>H	0.320	24	0.047	0.123
			LC	3	6	56	65	12	118	GT	18.0383	0.0001		12.0369	0.0024		0.0001	0.9932					
			CH	1	17	158	176	19	333	GT	49.3707	0.0000		11.4293	0.0033		13.2155	0.0003					
2	ARHGAP8 (2071762)	T/C	HCC	38	73	78	189	149	229	N/S			69271108			C/T (2456/1580)			P>L	0.018	98	0.521	0.097
			LC	13	29	36	78	55	101	N/S													
			CH	20	94	81	195	134	256	GT	8.5177	0.0141		7.0856	0.0289		3.4369	0.0638					
3	KDR (2305948)	T/C	HCC	0	25	158	183	25	341	N/S				6.2351	0.0443	C/T (3662/436)	5.2632	0.0218	V>I	0.999	29	0.079	0.281
			LC	0	9	57	66	9	123	N/S													
			CH	2	27	148	177	31	323	N/S													
4	SELE (5361)	A/C	HCC	180	2	0	182	362	2	AL				8.0262	0.0046	T/G (3953/145)	9.3738	0.0022	S>R	0.944	110	0.637	0.775
			LC	74	2	0	76	150	2	N/S													
			CH	180	2	0	182	362	2	AL				8.0262	0.0046		9.3738	0.0022					
5	XPC (2228001)	A/C	HCC	90	80	15	185	260	110	GT				13.1123	0.0014	G/T (1694/2398)	19.1813	0.0000	Q>K	0.000	53	0.179	0.000
			LC	25	35	28	78	85	91	N/S													
			CH	63	89	21	173	215	131	N/S													
6	FANCA (2239359)	T/C	HCC	105	49	8	162	259	65	GT				7.0776	0.0290	C/T (652/3396)	3.4226	0.0643	G>S	0.000	56	0.200	0.000
			LC	51	51	0	69	153	51	N/S													
			CH	138	40	4	182	316	48	N/S													
7	LRRN3 (17439799)	T/C	HCC	120	55	10	185	295	75	GT				6.6997	0.0351	T/C (3113/981)	2.5606	0.1096	Int				
			LC	27	1	12	40	55	25	GT				65.0216	0.0000		2.2784	0.1312					
			CH	114	43	13	170	271	69	GT				14.8948	0.0006		2.3367	0.1264					
8	BRCA2 (766173)	T/G	HCC	140	47	1	188	327	49	GT	10.5751	0.0051	66862596 69130493			A/C (3569/521)	0.0266	0.8703	N>H	0.034	68	0.279	0.097
			LC	47	24	0	71	118	24	AL	15.4693	0.0001					2.1199	0.1454					
			CH	144	47	1	192	335	49	GT	10.0898	0.0064					0.0002	0.9901					
9	PCDHGB7 (17208397)	G/C	HCC	152	6	0	158	310	6	AL	5.0817	0.0242	24521861 68951534			G/C (3955/129)	1.5656	0.2109	V>L	0.261	32	0.089	0.153
			LC	70	2	0	72	142	2	AL	4.0500	0.0442					1.4511	0.2284					
			CH	107	1	0	108	215	1	AL	9.7059	0.0018					5.0848	0.0241					

Table I. Continued.

ID	Gene (dbSNP rsID)	A/B	DP	AA	AB	BB	Sum	A	B	ST	dbSNP $\chi^2$	dbSNP P-value	dbSNP ss#	JSNP $\chi^2$	JSNP P-value	TMM A/B	TMM $\chi^2$	TMM P-value	Mut	PV	GV	PG	GMI
10	ERCC4 (1799801)	T/C	HCC	120	61	11	192	301	83	GT	7.2055	0.0272	52072800 66861998			T/C (3060/1032)	2.4396	0.1183	Syn				
11	PMS2 (1805321)	T/C	HCC	13	50	74	137	76	198	GT	56.2044	0.0000	5259684			N/A			P>S	0.001	74	0.295	0.017
12	ADAMTS16 (16875054)	A/G	HCC	9	11	105	125	29	221	GT	16.3429	0.0003	48414945 66315014 68925406			G/A (3559/537)	0.5531	0.4570	A>T	0.590	58	0.211	0.352
																	9.1853	0.0024					

Maximum  $\chi^2$  statistics against either curated JSNP or dbSNP genotype or allele counts and corresponding P-values. ID, SNP index to be used throughout this article. ST, significance type (statistical significance against control genotype 'GT' or allele 'A' frequencies, respectively); A/B, A and B alleles; DP, disease phenotype; Mut, mutation type (Int, intron; Syn, synonymous); PD, PolyPhen2 HumDiv score; PV, PolyPhen2 HumVar scores; GV, Grantham value; PG, Grantham probability of impact from amino acid substitution; GMI, geometric mean impact on protein structure.

While mutations in DNA mismatch or strand-break repairs have always been intimately associated with tumor development, the presence of risk alleles in *XPC*, a gene critical in nucleotide excision repair (34), further highlights the importance of exogenous genomic editing by HCV in HCC. *XPC* has been said to elevate the rise of skin cancer 1,000-fold and escalating organ neoplasms as much as 10-fold (35) through its role in xeroderma pigmentosum, a pre-cancerous condition. From this, it is likely that defects in *XPC* may also increase one's susceptibility to HCC. Additionally, aberrations in *FANCA*, a known tumor suppressor (36) as well as *BRCA2*, a DNA repair gene, can have serious repercussions leading to oncogenesis, most notably breast cancer, both in Japanese and other ethnic cohorts (37-40). Among these reports, candidate **8** also belongs to the group of 25 breast cancer potential risk alleles, sufficiently justifying the need for further investigation into its mechanistic role in HCV-induced HCC. Likewise, mutations in *PMS2* may be similarly implicated in response to HCV virulence as the gene was most remarkably known for its role in DNA mismatch repair and links to microsatellite instability as in hereditary nonpolyposis colorectal cancer (41-43).

Our genotyping results also revealed some curiosities on the role of functional surrogates in HCV-induced oncogenesis, particularly through potential risk alleles in *ADAMTS16*. *ADAMTS16* belongs to a family of secreted metalloproteases that tends to be fairly tissue limited, and the discovery of its association to HCV was unexpected as *ADAMTS16* was primarily expressed only in the lung and brain; with that said, recent literature did suggest the possibility of high-quantity shuttling of *ADAMTS13* and *ADAMTS19* to unexpected destinations, such as tumors and nearby tissues in cases of osteosarcoma, melanoma and colon cancer (44). Surfacing of *ADAMTS16* in HCC samples could also infer the occurrence of a similar phenomenon by mutations or other mechanisms. Interestingly, while some in the ADAMTS family were implicated in diseases such as angio-inhibition (45) and Ehlers-Danlos syndrome (46), most remain relatively unknown and inconclusive (44,47,48). As such, risk alleles in *ADAMTS16* as well as the observation that these metalloproteases were typically downregulated in cancer as a consequence of promoter hypermethylation (49), would suggest that polymorphisms in *ADAMTS16* may potentially push the gene to displace and redistribute itself and other members in the family, possibly via extracellular matrix remodeling (50), and become oncogenic. Additionally, not all conditions were genetically predisposed to the same polymorphisms (Table III) or reacted identically to the virulence of HCV. For example, a mutation to 3 would more likely drive onset in HCC over LC or CH, while subjects with a mutation to 6 would perhaps present HCC phenotypes differently. Odds ratios (OR >1, P<0.05) also inferred differential susceptibility for subjects with mutations in *XPC* and *BRCA2*, as *XPC* showed elevated odds for CH and *BRCA* for all subgroups. Just as the *BRCA* genes helped physicians to assess breast cancer susceptibility, the combination of *BRCA* and perhaps a combination of several aforementioned SNPs could also help elucidate risks to HCC by the process of elimination.

In some situations nearby variants may act in concert and contribute jointly to particular phenotypes; to examine the possibility of these nonrandom associations, we next explored

Table II. Stratified comparisons of significant SNPs across different ethnic groups.

Cytoband	Rs# Symbol	NCBI SSID	Population	Groups	P-value	
					GT	AL
5p13.2	rs34677 AMACR	12675253	JPT, HCB, CEU, YRI	G1	0.096	0.089
		68931918			0.070	0.029
		98712196			**	**
22q13.31	rs2071762 ARHGAP8	12527661	JPT, HCB, CEU, YRI	G1	**	**
		69271108			**	**
		71647951			**	**
4q12	rs2305948 KDR	48430022	JPT, HCB, CEU, YRI	G1	**	**
		68899172			**	**
1q24.2	rs5361 SELE	48428825	JPT, HCB, CEU, YRI	G1	0.001	**
		68784305			**	**
3p25.1	rs2228001 XPC	44411054	JPT, HCB, CEU, YRI	G1	0.009	0.004
		66856826	JPT, HSP, CEU, AAM, CHB, YRI GENO PANEL	G2	0.001	0.002
		68853585	JPT, HCB, CEU, YRI	G1	0.009	0.007
16q24.3	rs2239359 FANCA	16702260	JPT, HCB, CEU, YRI	G1	**	**
		43872290	AoD Japanese, African American, Caucasian, Chinese	G3	N/A	**
		66860952	JPT, HSP, CEU, AAM, CHB, YRI GENO PANEL	G2	**	**
		69355410	JPT, HCB, CEU, YRI	G1	**	**
7q31.1	rs17439799 LRRN3	24455640	JPT, HCB, CEU, YRI	G1	**	**
13q13.1	rs766173 BRCA2	5586314	JPT, HCB, CEU, YRI	G1	**	**
		66862596	JPT, HSP, CEU, AAM, CHB, YRI GENO PANEL	G2	**	**
		69130493	JPT, HCB, CEU, YRI	G1	**	**
5q31.3	rs17208397 PCDHGB7	24621861	JPT, HCB, CEU, YRI	G1	**	**
		68951534			**	**
16p31.12	rs1799801 ERCC4	52072800	JPT, HCB, CEU, YRI	G1	**	**
		66861998	JPT, HSP, CEU, AAM, CHB, YRI GENO PANEL	G2	**	**
5p15.32	rs16875054 ADAMTS16	48414945	JPT, HCB, CEU, YRI	G1	**	**
		66315014			**	**
		68925406			**	**

Reference datasets from NCBI dbSNP with specified SSIDs. G1, Asian/European/Sub-Saharan African; G2, Asian/European/African American/Sub-Saharan African; G3, Asian/Caucasian/African American. HapMap groups: JPT, Japanese (Tokyo, Japan); HCB, Han Chinese (Beijing, China); CEU, Utah residents with ancestry from northern and western Europe; YRI, Yoruba (Ibadan, Nigeria); \*\*, P<0.001; N/A, data not available. GT, genotype; AL, allele.

possible interactions and the presence of common functionally active proxy variants associated with polymorphisms identified in this study. While the presence of a small interaction revealed the presence of common interacting partners with the 10 candidate genes, for instance *SMARCA4*, *PPP2CA* and *XPO1* (Fig. 1A), linkage disequilibrium analysis painted a slightly different picture, alternatively suggesting that these candidates were genotypically pairwise independent, and contributed cumulatively to the outcome of each disease.

Additionally, multivariate analysis across genders and ages revealed no strong covariances among the candidates (data not shown). Interestingly, enrichment analysis highlighted strong connections to DNA repair and post-translational modifications, two biologically orthogonal functions, for the candidate genes and their interacting partners (table available upon request). Enrichment in the two nonparallel pathways would thus posit an overall additive effect that reflected the candidates' genotypic independence. While defects in DNA repair

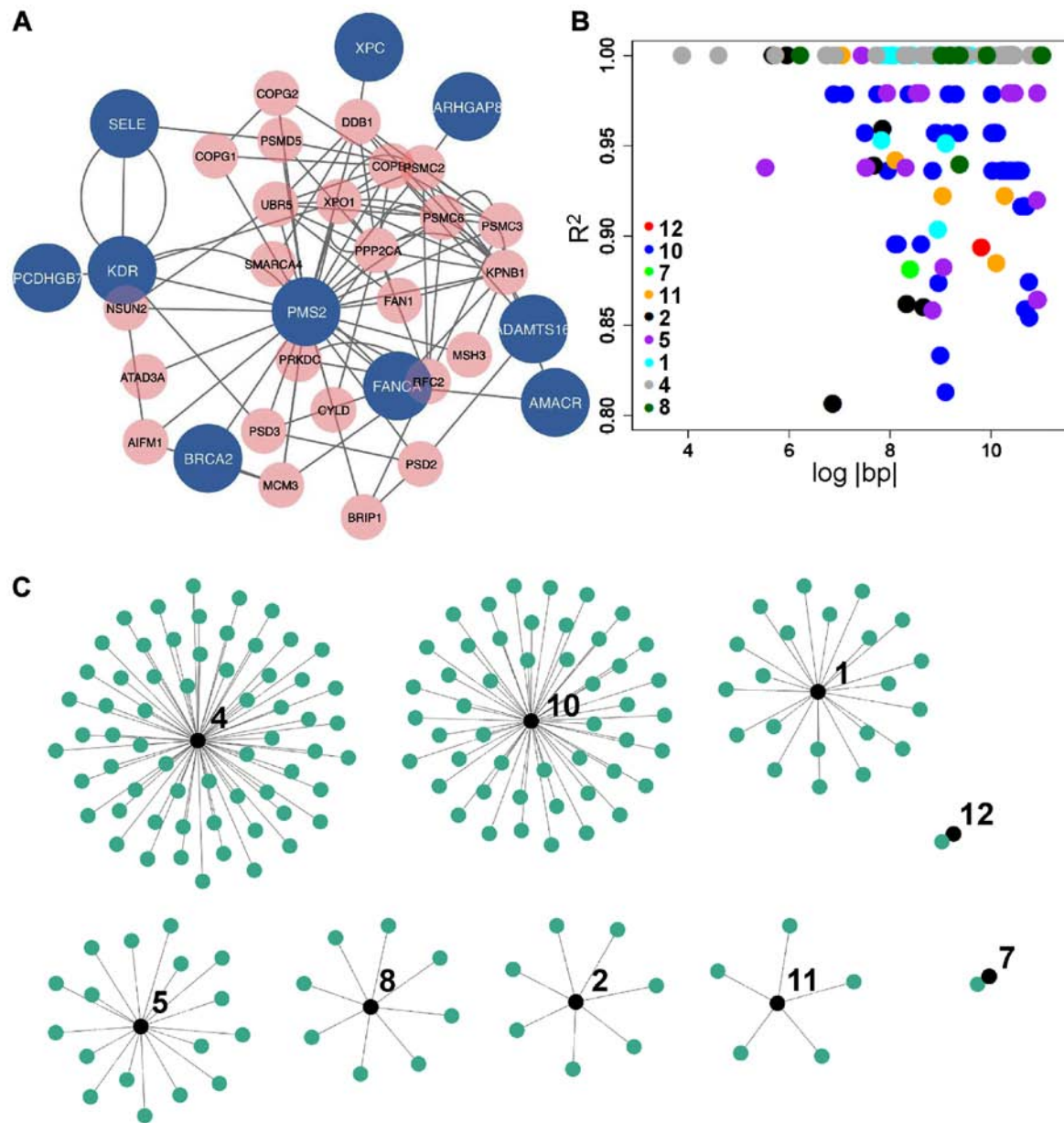
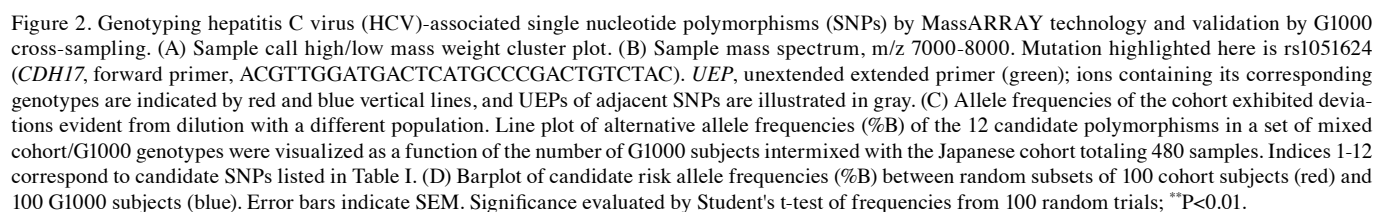


Figure 1. Single nucleotide polymorphisms (SNPs) potentially associated with hepatitis C virus-induced liver conditions in surveyed Japanese subjects, as identified by MassARRAY genotyping. (A) Interaction network of genes with candidate SNPs and their top 25 common interacting partners. Blue ovals, candidate genes; pink, interacting partners, with gray edges indicating possible associations. (B) Scatterplot of proxy variants to candidate SNPs and the linkage disequilibrium correlation coefficient. Dist, distance between a proxy variant to candidate SNP, expressed as  $\log_{10}$  of the absolute value in bp;  $R^2$ , the correlation coefficient of linkage disequilibrium. (C) Association networks of proxy variants (green) to candidate SNPs (black) as numbered in Table I; edges between nodes are illustrative only, with lengths not respective of association strengths. A fully annotated network diagram as vector graphics is available upon request.

were commonplace in cancer as a consequence of unregulated division, no mechanistically direct explanation could answer the functional enrichment in protein modification as a consequence; as such, the hypothesis that both pathways, via functional alteration by the candidates genotyped, acted in concert to achieve this additive effect reflected in the association of HCV to HCC, appeared to be a plausible one and hopefully could be further pondered.

We also analyzed changes in protein local secondary and quaternary structures in order to attribute the impact of non-synonymous mutations to the connection between HCV and HCC to ascertain the underlying routes to disease progression. Although DNA repair mechanisms may be biochemically similar across different types of

cancer, focused studies on *BRCA2* and *FANCA* still remain relatively unexplored in HCV-induced HCC. Especially since hepatic tissues are constantly under oxidative stress, genome repair during acute and chronic HCV infection and the oncogenesis of HCC are critical in understanding disease progression. We observed that *ADAMTS16*, *SELE* and *KDR* all carried polymorphisms with potentially damaging mutations; as a matter of fact, both *ADAMTS16* and *SELE* were associated with HCC and CH, suggesting a possible path of disease progression upon HCV infection from CH to HCC via these mutations (Table I). Nonetheless, the fact that *KDR* also manifested a damaging mutation and was associated with HCC would also suggest a possible role for the gene, as KDR-mediated mechanisms could support



A difficulty in establishing genetic connections between diseases and HCV infections is that the virus can remain dormant in the liver, leaving the patient with normal liver functions and no symptoms. As such, disease progression in early stages becomes retarded. These associated SNPs can, in theory, display no phenotypical effect during early onset, which can delay diagnosis and treatment altogether. As both acute and chronic HCV infections could transform healthy livers, the full and exact extent of the infection would be difficult to gauge. HCV carriers may present normal ALT levels from normal liver panel screenings and the amount of fat deposits in the tissue may also mask the presence of HCVs, liver biopsies are likely to be inconclusive in these cases for determining the stage of infection. As such, extended genotyping studies

Among the different genotyping technologies, MassARRAY proved to be a capable choice, demonstrating high sensitivities in various applications such as human papillomavirus genotyping for cervical cancer (52) or classification of ectopic crypts in colorectal cancers (53) via *KRAS* and *BRAF* mutations. Although we were unable to call every SNP on the initial test, we nonetheless obtained call rates surpassing 90% in duplicate typing of most SNPs, suggesting reasonably minimal genotyping errors (Fig. 2A and B). Mass spectrometry-based genotyping offers excellent sensitivity in analyte detection and is thus an ideal approach for characterizing specimens from tissue depositories, where amounts of DNA may be very limited. Extension (1-bp) coupled with mass

Table III. Stratified genotypic comparisons of significant SNPs among disease phenotypes.

ID Symbol dbSNP rsID	Alleles	Grouping (or JSNP/NCBI set ID)	Genotype			
			All P-value	HCC P-value	LC P-value	CH P-value
1 <i>AMACR</i> rs34677	T/G	HCC	-	-	0.511	0.259
		LC	-	-	-	0.092
		CH	-	-	-	-
		NCBI 12675253	<b>0.032</b>	0.106	0.147	<b>0.002</b>
		NCBI 68931918	<b>**</b>	<b>**</b>	<b>**</b>	<b>**</b>
		JSNP IMS-JST038996	<b>**</b>	<b>0.017</b>	<b>0.002</b>	<b>0.003</b>
2 <i>ARHGAP8</i> rs2071762	T/C	HCC	-	-	0.712	<b>0.017</b>
		LC	-	-	-	0.159
		CH	-	-	-	-
		NCBI 12527661	0.111	0.988	0.818	0.053
		NCBI 69271108	0.065	0.862	0.659	0.014
		NCBI 71647951	0.111	0.988	0.818	0.053
		JSNP IMS-JST007439	<b>0.012</b>	0.056	0.102	<b>0.029</b>
3 <i>KDR</i> rs2305948	T/C	HCC	-	-	0.59	0.316
		LC	-	-	-	0.646
		CH	-	-	-	-
		NCBI 48430022	0.647	0.374	0.438	0.603
		NCBI 68899172	0.651	0.401	0.461	0.599
		JSNP IMS-JST063410	0.158	<b>0.044</b>	0.302	0.389
4 <i>SELE</i> rs5361	A/C	HCC	-	-	0.338	0.689
		LC	-	-	-	0.338
		CH	-	-	-	-
		NCBI 48428825	0.188	0.091	0.413	0.091
		NCBI 68784305	0.201	0.095	0.424	0.095
		JSNP	<b>0.001</b>	<b>0.001</b>	0.145	<b>0.001</b>
5 <i>XPC</i> rs2228001	A/C	HCC	-	-	<b>0.001</b>	0.054
		LC	-	-	-	0.086
		CH	-	-	-	-
		NCBI 44411054	<b>0.011</b>	0.523	0.092	0.656
		NCBI 66856826	<b>0.01</b>	0.365	0.108	0.847
		NCBI 68853585	<b>0.012</b>	0.326	0.185	0.555
		JSNP IMS-JST086794	<b>0.003</b>	<b>0.001</b>	0.371	0.165
6 <i>FANCA</i> rs2239359	T/C	HCC	-	-	0.117	0.061
		LC	-	-	-	0.386
		CH	-	-	-	-
		NCBI 16702260	0.169	0.366	0.191	0.581
		NCBI 43872290	-	-	-	-
		NCBI 66860952	0.169	0.366	0.191	0.581
		NCBI 69355410	0.169	0.366	0.191	0.581
		JSNP IMS-JST010482	0.087	<b>0.029</b>	0.504	0.488
7 <i>LRRN3</i> rs17439799	T/C	HCC	-	-	<b>**</b>	0.5
		LC	-	-	-	<b>**</b>
		CH	-	-	-	-
		NCBI 24455640	<b>**</b>	0.845	<b>**</b>	0.926
		JSNP	<b>**</b>	<b>0.035</b>	<b>**</b>	<b>0.001</b>
8 <i>BRCA2</i> rs766173	T/G	HCC	-	-	0.313	0.993
		LC	-	-	-	0.274
		CH	-	-	-	-
		NCBI 5586314	<b>0.019</b>	<b>0.008</b>	<b>**</b>	<b>0.01</b>
		NCBI 66862596	<b>0.013</b>	<b>0.005</b>	<b>**</b>	<b>0.006</b>
		NCBI 69130493	<b>0.013</b>	<b>0.005</b>	<b>&lt;0.001</b>	<b>0.003</b>

Table III. Continued.

ID Symbol dbSNP rsID	Alleles	Grouping (or JSNP/NCBI set ID)	Genotype			
			All P-value	HCC P-value	LC P-value	CH P-value
9 <i>PCDHGB7</i> rs17208397	G/C	HCC	-	-	0.518	0.147
		LC	-	-	-	0.351
		CH	-	-	-	-
		NCBI 24621861	0.004	0.026	<b>0.04</b>	0.002
		NCBI 68951534	0.004	0.026	<b>0.04</b>	0.002
10 <i>ERCC4</i> rs1799801	T/C	HCC	-	-	0.354	0.573
		LC	-	-	-	0.815
		CH	-	-	-	-
		NCBI 52072800	0.156	<b>0.027</b>	0.237	0.07
		NCBI 66861998	0.156	<b>0.027</b>	0.237	0.07
11 <i>PMS2</i> rs1805321	T/C	HCC	-	-	0.011	0.061
		LC	-	-	-	0.388
		CH	-	-	-	-
		NCBI 5259684	**	**	**	**
12 <i>ADAMTS16</i> rs16875054	A/G	HCC	-	-	0.258	0.018
		LC	-	-	-	0.682
		CH	-	-	-	-
		NCBI 48414945	**	**	0.074	<b>0.007</b>
		NCBI 66315014	**	**	0.074	<b>0.007</b>
		NCBI 68925406	**	**	0.074	<b>0.007</b>

The full table including genotypic frequencies, allele frequencies, dominant (AA+AB/BB) and recessive allele models (AA/AB+BB), is available as Table S4 upon request. P, P-values of significance of a particular subgroup compared against all three disease phenotypes (All) or individual subgroups. Bold values indicate genes with call rate >80%. \*\*, P<0.001; -, not available.

spectrometric-based characterization by MassARRAY was a suitable choice in assessing candidate SNPs for studies of similar scale. We do, however, recognize that such a method is limited to the number of SNP candidates determined *a priori*, so the discovery of novel SNPs can be more challenging. Nonetheless, for cancers and precancerous conditions dependent on a small of oncogenic drivers, this method is a sensible and attractive option at the clinical setting.

The MassARRAY method allowed us to maintain a reasonable subject size of 480, although the relatively lower number of LC cases might have marginally reduced the statistical power in this study. While increasing the number of test subjects could certainly be beneficial in improving the statistical power, such an approach was not always possible in practice. In consideration for the potential loss in statistical power, we performed two validation simulations to see whether these differences in allele frequencies were still preserved if subsets of our cohort were 'diluted' with a different population sample. By taking genotypes from randomly selected G1000 individuals and mixing them with (also randomly selected) subjects in the cohort, we assembled subsets of 480 individuals and checked the frequencies of the candidate risk allele. Plotting such changes as a function of different numbers of G1000 individuals (Fig. 2C) revealed moderate frequency shifts for all of the candidate polymorphisms. Frequencies of these risk alleles thus appeared to

be condition-specific, as they were certainly susceptible to dilution with subjects from a different and considerably more heterogeneous population of various ethnicities and health states. Additionally, bootstrapping our cohorts to smaller subsets of 100 individuals for direct comparison to sets of equal sizes in the G1000 population also confirmed differences in allele frequencies (Fig. 2D). While such tests could not truly offset benefits of increasing cohort sizes, results nonetheless demonstrated a reasonable level of statistical robustness considering the size of our cohort.

Genotyping more candidates, e.g. Toll-like 1 protein (54), from overlooked biochemical pathways (for instance, stress responses, inflammation and cell cycles) could allow us to further understand the manner in which HCV infections evolve phenotypically. Characterizing changes in known and nearby polymorphisms in genes implicated in insulin biosynthesis, inflammation and adipose tissue remodeling, etc., would be highly useful in deciphering risk factors of obesity to HCV-induced HCC; in a similar vein, alcohol metabolism and various other pathways related to opioid metabolism would also be highly useful in risk assessment of substance abuse to HCC. Additionally, this type of candidate expansion would be particularly more beneficial in analyzing ethnically more heterogeneous populations. As most candidate SNPs, with the exception of *AMACR* (rs34677), exhibited pairwise independence across non-ethnically Japanese datasets, our results

hinted that HCV-induced liver disease progressed differently from other infection-driven oncogenesis such as the case with *Helicobacter pylori* and stomach adenocarcinoma. Based on the differences across different datasets, we would recommend increasing the number of candidates for genotyping to validate this hypothesis. Nonetheless, our approach here could still serve as a useful tool for early diagnosis based on liver disease examination.

An ongoing discussion in liver cancer is the possibility of oncogenic addiction, in which cancer progression is often controlled by a handful of driver genes subsequently leading to uncontrollable growth and eventual transformation to tumor. Horizontally integrating SNP-based oncogenesis information from one cancer type to another can be a useful approach in deciphering the puzzle. For liver diseases that may ultimately lead to cancer, identifying function-altering SNPs is a useful tool for facilitating earlier diagnosis and expediting treatment. Furthermore, for cancers which the theory of oncogenic addiction well applies, the ability to identify driver mutations quickly also has significant therapeutic implications. Based on our previous knowledge of SNPs in liver diseases and oncogenes, we utilized a targeted SNP-screening approach using a MassARRAY-based protocol from a set of 131 SNPs, and were able to identify 12 positions with implications ranging from disease onset, survival rates and in other metrics. Additionally, we were also able to highlight the significance of *BRCA2* and *FANCA* germ-line mutations and associate them to liver cancers. Most of these HCV-dependent mutation candidates were non-synonymous, and their association with other risk factors further suggested their potential roles as biomarkers for the liver conditions described in this study.

It is however important to note that several factors such as age have not been well explored in this study, as illuminated by the relatively smaller sample sizes of subjects under 50 years old in all three disease groups; additionally, at 80 samples, the statistical power for SNPs associated to cirrhosis may also be lower than the other two groups. Other factors such as hepatitis B infection, substance use or obesity could also influence disease progression. It is now common knowledge that sharing syringe needles is a common way that HCV is transmitted between persons, and that alcohol abuse has been said to worsen chronic HCV progression (55). Obesity, mostly through non-alcoholic fatty liver disease and type 2 diabetes, can also have an effect on the outcome of liver cancer (56). In the Japanese population, hepatitis B infection attributed to ~16% of HCC cases (2), a fraction ~1/5 of HCV infections; while relatively minor compared to HCV, hepatitis B infections still present a great public health concern and future genotyping studies should also consider the inclusion of genes potentially implicated in these infections. Along this note, it is also important to note that the status of HCV infections such as virus titers, genotypes and history of prior treatments may also affect the progression of liver cancer and thus critical to be taken into consideration in the cohort selection. Nonetheless, the incorporation of *in silico* screening and whole-exome sequencing data to identify and refine the list of SNP candidates for future MassARRAY studies should provide definitive improvements for more reliable characterization of disease associations and medium-size clinical cohort studies.

## Acknowledgements

This study was supported by the Academic Frontier Project for Private Universities of 2006, Grant-in-Aid for Scientific Research (B) JP26290060 and Research on Innovative Areas (JP26112517, JP25134718 and JP16H01579) whilst being conducted as part of the BioBank Japan Project sponsored by the Ministry of Education, Culture, Sports, Science and Technology as well as the Japan Agency for Medical Research and Development.

## References

1. El-Serag HB: Hepatocellular carcinoma. *N Engl J Med* 365: 1118-1127, 2011.
2. Chen SL and Morgan TR: The natural history of hepatitis C virus (HCV) infection. *Int J Med Sci* 3: 47-52, 2006.
3. Yoshizawa H: Hepatocellular carcinoma associated with hepatitis C virus infection in Japan: Projection to other countries in the foreseeable future. *Oncology* 62 (Suppl 1): 8-17, 2002.
4. Colombo M and Iavarone M: Role of antiviral treatment for HCC prevention. *Best Pract Res Clin Gastroenterol* 28: 771-781, 2014.
5. Alavian SM and Haghbin H: Relative importance of hepatitis B and C viruses in hepatocellular carcinoma in EMRO countries and the Middle East: A systematic review. *Hepat Mon* 16: e35106, 2016.
6. Tang L, Marcell L and Kottitil S: Systemic manifestations of hepatitis C infection. *Infect Agent Cancer* 11: 29, 2016.
7. Kato N, Ji G, Wang Y, Baba M, Hoshida Y, Otsuka M, Taniguchi H, Moriyama M, Dharel N, Goto T, *et al*: Large-scale search of single nucleotide polymorphisms for hepatocellular carcinoma susceptibility genes in patients with hepatitis C. *Hepatology* 42: 846-853, 2005.
8. Gu X, Qi P, Zhou F, Ji Q, Wang H, Dou T, Zhao Y and Gao C: An intronic polymorphism in the corticotropin-releasing hormone receptor 2 gene increases susceptibility to HBV-related hepatocellular carcinoma in Chinese population. *Hum Genet* 127: 75-81, 2010.
9. Segat L, Milanese M, Pirulli D, Trevisiol C, Lupo F, Salizzoni M, Amoroso A and Crovella S: Secreted protein acidic and rich in cysteine (SPARC) gene polymorphism association with hepatocellular carcinoma in Italian patients. *J Gastroenterol Hepatol* 24: 1840-1846, 2009.
10. Tang KS, Lee CM, Teng HC, Huang MJ and Huang CS: UDP-glucuronosyltransferase 1A7 polymorphisms are associated with liver cirrhosis. *Biochem Biophys Res Commun* 366: 643-648, 2008.
11. Kim YS, Cheong JY, Cho SW, Lee KM, Hwang JC, Oh B, Kim K, Lee JA, Park BL, Cheong HS, *et al*: A functional SNP of the Interleukin-18 gene is associated with the presence of hepatocellular carcinoma in hepatitis B virus-infected patients. *Dig Dis Sci* 54: 2722-2728, 2009.
12. Tanaka Y, Nishida N, Sugiyama M, Kurosaki M, Matsuura K, Sakamoto N, Nakagawa M, Korenaga M, Hino K, Hige S, *et al*: Genome-wide association of IL28B with response to pegylated interferon-alpha and ribavirin therapy for chronic hepatitis C. *Nat Genet* 41: 1105-1109, 2009.
13. Nalpas B, Lavialle-Meziani R, Plancoulaine S, Jouanguy E, Nalpas A, Munteanu M, Charlotte F, Ranque B, Patin E, Heath S, *et al*: Interferon gamma receptor 2 gene variants are associated with liver fibrosis in patients with chronic hepatitis C infection. *Gut* 59: 1120-1126, 2010.
14. Tillmann HL, Thompson AJ, Patel K, Wiese M, Tenckhoff H, Nischalke HD, Lokhnygina Y, Kullig U, Göbel U, Capka E, *et al*: German Anti-D Study Group: A polymorphism near IL28B is associated with spontaneous clearance of acute hepatitis C virus and jaundice. *Gastroenterology* 139: 1586-1592, 1592.e1, 2010.
15. GBD 2015 Disease and Injury Incidence and Prevalence Collaborators: Global, regional, and national incidence, prevalence and years lived with disability for 310 disease and injuries, 1990-2015: a systematic analysis for the Global Burden of Disease Study 2015. *Lancet* 388: 1545-1602, 2016.
16. Clendenen TV, Rendleman J, Ge W, Koenig KL, Wirgin I, Currie D, Shore RE, Kirchhoff T and Zeleniuch-Jacquotte A: Genotyping of single nucleotide polymorphisms in DNA isolated from serum using sequenom MassARRAY technology. *PLoS One* 10: e0135943, 2015.

17. Hirakawa M, Tanaka T, Hashimoto Y, Kuroda M, Takagi T and Nakamura Y: JSNP: A database of common gene variations in the Japanese population. *Nucleic Acids Res* 30: 158-162, 2002.
18. Haga H, Yamada R, Ohnishi Y, Nakamura Y and Tanaka T: Gene-based SNP discovery as part of the Japanese Millennium Genome Project: Identification of 190,562 genetic variations in the human genome. Single-nucleotide polymorphism. *J Hum Genet* 47: 605-610, 2002.
19. Sherry ST, Ward MH, Kholodov M, Baker J, Phan L, Smigielski EM and Sirotkin K: dbSNP: The NCBI database of genetic variation. *Nucleic Acids Res* 29: 308-311, 2001.
20. Kuriyama S, Yaegashi N, Nagami F, Arai T, Kawaguchi Y, Osumi N, Sakaida M, Suzuki Y, Nakayama K, Hashizume H, *et al*: The Tohoku Medical Megabank Project: Design and Mission. *J Epidemiol* 26: 493-511, 2016.
21. Yue P, Melamud E and Moulton J: SNPs3D: Candidate gene and SNP selection for association studies. *BMC Bioinformatics* 7: 166, 2006.
22. Nakamura Y: The BioBank Japan Project. *Clin Adv Hematol Oncol* 5: 696-697, 2007.
23. Adzhubei IA, Schmidt S, Peshkin L, Ramensky VE, Gerasimova A, Bork P, Kondrashov AS and Sunyaev SR: A method and server for predicting damaging missense mutations. *Nat Methods* 7: 248-249, 2010.
24. Grantham R: Amino acid difference formula to help explain protein evolution. *Science* 185: 862-864, 1974.
25. Machiela MJ and Chanock SJ: LDlink: A web-based application for exploring population-specific haplotype structure and linking correlated alleles of possible functional variants. *Bioinformatics* 31: 3555-3557, 2015.
26. Auton A, Brooks LD, Durbin RM, Garrison EP, Kang HM, Korbel JO, Marchini JL, McCarthy S, McVean GA and Abecasis GR: 1000 Genomes Project Consortium: A global reference for human genetic variation. *Nature* 526: 68-74, 2015.
27. Levin AM, Zuhlke KA, Ray AM, Cooney KA and Douglas JA: Sequence variation in alpha-methylacyl-CoA racemase and risk of early-onset and familial prostate cancer. *Prostate* 67: 1507-1513, 2007.
28. Daugherty SE, Platz EA, Shugart YY, Fallin MD, Isaacs WB, Chatterjee N, Welch R, Huang WY and Hayes RB: Variants in the alpha-methylacyl-CoA racemase gene and the association with advanced distal colorectal adenoma. *Cancer Epidemiol Biomarkers Prev* 16: 1536-1542, 2007.
29. FitzGerald LM, Thomson R, Polanowski A, Patterson B, McKay JD, Stankovich J and Dickinson JL: Sequence variants of alpha-methylacyl-CoA racemase are associated with prostate cancer risk: A replication study in an ethnically homogeneous population. *Prostate* 68: 1373-1379, 2008.
30. Pruthi RS, Derksen E and Gaston K: Cyclooxygenase-2 as a potential target in the prevention and treatment of genitourinary tumors: A review. *J Urol* 169: 2352-2359, 2003.
31. Ng IO, Liang ZD, Cao L and Lee TK: DLC-1 is deleted in primary hepatocellular carcinoma and exerts inhibitory effects on the proliferation of hepatoma cell lines with deleted DLC-1. *Cancer Res* 60: 6581-6584, 2000.
32. Trebicka J and Schierwagen R: Statins, Rho GTPases and KLF2: New mechanistic insight into liver fibrosis and portal hypertension. *Gut* 64: 1349-1350, 2015.
33. Khan FS, Ali I, Afridi UK, Ishtiaq M and Mehmood R: Epigenetic mechanisms regulating the development of hepatocellular carcinoma and their promise for therapeutics. *Hepatol Int* 11: 45-53, 2017.
34. Haiman CA, Hsu C, de Bakker PI, Frasco M, Sheng X, Van Den Berg D, Casagrande JT, Kolonel LN, Le Marchand L, Hankinson SE, *et al*: Comprehensive association testing of common genetic variation in DNA repair pathway genes in relationship with breast cancer risk in multiple populations. *Hum Mol Genet* 17: 825-834, 2008.
35. de Boer J and Hoeijmakers JH: Nucleotide excision repair and human syndromes. *Carcinogenesis* 21: 453-460, 2000.
36. Krupa R, Sliwinski T, Morawiec Z, Pawlowska E, Zadrozny M and Blasiak J: Association between polymorphisms of the BRCA2 gene and clinical parameters in breast cancer. *Exp Oncol* 31: 250-251, 2009.
37. Carreira A, Hilario J, Amitani I, Baskin RJ, Shivji MK, Venkitaraman AR and Kowalczykowski SC: The BRC repeats of BRCA2 modulate the DNA-binding selectivity of RAD51. *Cell* 136: 1032-1043, 2009.
38. Johnson N, Fletcher O, Palles C, Rudd M, Webb E, Sellick G, dos Santos Silva I, McCormack V, Gibson L, Fraser A, *et al*: Counting potentially functional variants in BRCA1, BRCA2 and ATM predicts breast cancer susceptibility. *Hum Mol Genet* 16: 1051-1057, 2007.
39. Ishitobi M, Miyoshi Y, Ando A, Hasegawa S, Egawa C, Tamaki Y, Monden M and Noguchi S: Association of BRCA2 polymorphism at codon 784 (Met/Val) with breast cancer risk and prognosis. *Clin Cancer Res* 9: 1376-1380, 2003.
40. Sliwinski T, Krupa R, Majsterek I, Rykala J, Kolacinska A, Morawiec Z, Drzewoski J, Zadrozny M and Blasiak J: Polymorphisms of the BRCA2 and RAD51 genes in breast cancer. *Breast Cancer Res Treat* 94: 105-109, 2005.
41. Peltomäki P: Deficient DNA mismatch repair: A common etiologic factor for colon cancer. *Hum Mol Genet* 10: 735-740, 2001.
42. Yuan ZQ, Gottlieb B, Beitel LK, Wong N, Gordon PH, Wang Q, Puisieux A, Foulkes WD and Trifiro M: Polymorphisms and HNPCC: PMS2-MLH1 protein interactions diminished by single nucleotide polymorphisms. *Hum Mutat* 19: 108-113, 2002.
43. Umar A, Boland CR, Terdiman JP, Syngal S, de la Chapelle A, Rüschoff J, Fishel R, Lindor NM, Burgart LJ, Hamelin R, *et al*: Revised Bethesda Guidelines for hereditary nonpolyposis colorectal cancer (Lynch syndrome) and microsatellite instability. *J Natl Cancer Inst* 96: 261-268, 2004.
44. Cal S, Obaya AJ, Llamazares M, Garabaya C, Quesada V and López-Otin C: Cloning, expression analysis, and structural characterization of seven novel human ADAMTSs, a family of metalloproteinases with disintegrin and thrombospondin-1 domains. *Gene* 283: 49-62, 2002.
45. Vázquez F, Hastings G, Ortega MA, Lane TF, Oikemus S, Lombardo M and Iruela-Arispe ML: METH-1, a human ortholog of ADAMTS-1, and METH-2 are members of a new family of proteins with angio-inhibitory activity. *J Biol Chem* 274: 23349-23357, 1999.
46. Colige A, Sieron AL, Li SW, Schwarze U, Petty E, Wertschlecker W, Wilcox W, Krakow D, Cohn DH, Reardon W, *et al*: Human Ehlers-Danlos syndrome type VII C and bovine dermatosparaxis are caused by mutations in the procollagen I N-proteinase gene. *Am J Hum Genet* 65: 308-317, 1999.
47. Rodríguez-López J, Pombo-Suarez M, Loughlin J, Tsezou A, Blanco FJ, Meulenbelt I, Slagboom PE, Valdes AM, Spector TD, Gomez-Reino JJ, *et al*: Association of an sSNP in ADAMTS14 to some osteoarthritis phenotypes. *Osteoarthritis Cartilage* 17: 321-327, 2009.
48. Hu X, Chen H, Jin M, Wang X, Lee J, Xu W, Zhang R, Li S and Niu J: Molecular cytogenetic characterization of undifferentiated embryonal sarcoma of the liver: A case report and literature review. *Mol Cytogenet* 5: 26, 2012.
49. Kumar S, Rao N and Ge R: Emerging Roles of ADAMTSs in Angiogenesis and Cancer. *Cancers (Basel)* 4: 1252-1299, 2012.
50. Bonnans C, Chou J and Werb Z: Remodelling the extracellular matrix in development and disease. *Nat Rev Mol Cell Biol* 15: 786-801, 2014.
51. Goldman O, Han S, Sourisseau M, Dziedzic N, Hamou W, Corneo B, D'Souza S, Sato T, Kotton DN, Bissig KD, *et al*: KDR identifies a conserved human and murine hepatic progenitor and instructs early liver development. *Cell Stem Cell* 12: 748-760, 2013.
52. Basu P, Chandna P, Bamezai RNK, Siddiqi M, Saranath D, Lear A and Ratnam S: MassARRAY spectrometry is more sensitive than PreTect HPV-Proofer and consensus PCR for type-specific detection of high-risk oncogenic human papillomavirus genotypes in cervical cancer. *J Clin Microbiol* 49: 3537-3544, 2011.
53. Kim MJ, Lee EJ, Chun SM, Jang SJ, Kim DS, Lee DH and Youk EG: The significance of ectopic crypt formation in the differential diagnosis of colorectal polyps. *Diagn Pathol* 9: 212, 2014.
54. Matsuura K, Sawai H, Ikeo K, Ogawa S, Iio E, Isogawa M, Shimada N, Komori A, Toyoda H, Kumada T, *et al*: Japanese Genome-Wide Association Study Group for Viral Hepatitis: Genome-wide association study identifies TLL1 variant associated with development of hepatocellular carcinoma after eradication of hepatitis C virus infection. *Gastroenterology* 152: 1383-1394, 2017.
55. Peters MG and Terrault NA: Alcohol use and hepatitis C. *Hepatology* 36 (Suppl 1): S220-S225, 2002.
56. Aleksandrova K, Stelmach-Mardas M and Schlesinger S: Obesity and liver cancer. *Recent Results Cancer Res* 208: 177-198, 2016.