

Intraperitoneal neutrophils activated by KRAS-induced ovarian cancer exert antitumor effects by modulating adaptive immunity

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Abstract. Increased neutrophil counts are a hallmark of a poor prognosis for cancer. We previously reported that KRAS promoted tumorigenesis and increased neutrophil counts in a mouse peritoneal cancer model. In the current study, we evaluated the role of increased neutrophils in cancer progression, as well as their influence on the intraperitoneal microenvironment. A mouse peritoneal cancer model was established using the KRAS-transduced mouse ovarian cancer cell line, ID8-KRAS. Neutrophil function was assessed by neutrophil depletion in ID8-KRAS mice. Neutrophil depletion markedly accelerated tumor formation; this was accompanied by an increase in interleukin-6 concentrations in ascites. Neutrophil depletion significantly decreased the amount of local and systemic CD8⁺ T cells, while increasing the amount of local CD4⁺ T cells, accompanied by an increased amount of monocytic myeloid-derived suppressor cells (M-MDSCs) and regulatory T cells (Tregs) (P<0.05). The roles of peritoneal neutrophils (PENs) in CD8⁺ T cell activation were assessed *in vitro*. PENs of ID8-KRAS mice had a strong potential to enhance T cell proliferation with a higher expression of the T cell costimulatory molecules OX40 ligand (OX40L) and 4-1BB ligand (4-1BBL), as compared with peripheral blood neutrophils (PBNs). These findings suggest that neutrophils recruited into the KRAS-induced tumor microenvironment

(TME) have antitumor properties with the potential to modulate the numbers of M-MDSCs and Tregs and activate CD8⁺ T cells through T cell costimulatory molecules.

Introduction

Increased neutrophil counts can sometimes predict a poor prognosis among patients with advanced cancers (1-3). Pan-cancer analysis performed by Templeton *et al* aided in the identification of polymorphonuclear leukocyte signaling as the most potent predictor of a poor prognosis (4). Increased neutrophil/lymphocyte (N/L) ratios have recently been attracting interest as a suitable prognostic marker for cancer patients (4,5). Several reports have indicated that increased N/L ratios are a marker of a poor prognosis and a weak response to chemotherapy among patients with ovarian cancer (6,7). Although a number of studies have demonstrated the significance of increased neutrophil counts as a prognostic marker, an in-depth analysis has not yet been performed regarding its role in cancer progression, at least to the best of our knowledge.

Oncogenes are known to promote cell proliferation and prevent apoptosis in malignant cells. Although the association between oncogenes and tumor microenvironment (TME) modifications has not yet been sufficiently elucidated, recent studies have revealed that several oncogenes are associated with TME modulation (8,9). KRAS is a frequently mutated oncogene in cancer. In a previous study, we established a KRAS-transduced mouse ovarian cancer cell line, ID8-KRAS, and demonstrated that the oncogene KRAS promoted tumorigenesis and aggravated cancer-induced inflammation, accompanied by an increased number of neutrophils in ascites (8). RAS is considered to promote cancer progression by sustaining proliferation, metabolic reprogramming, anti-apoptosis, and remodeling of the TME (10,11). RAS followed by the activation of RAS-GTP and PI3K/AKT signaling may upregulate nuclear factor (NF)- κ B activity and lead to an inflammatory microenvironment (12). Similarly, several oncogenes may be associated with severe inflammation accompanied by an increased number of neutrophils.

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Although neutrophils have been regarded as a protumor marker, the role of tumor-associated neutrophils (TANs) in cancer progression has only recently become a focus area (13). There are two types of TANs: An interferon (IFN)- β -induced antitumor type (14) and a transforming growth factor (TGF)- β -induced protumor type (15). The majority of studies have focused on the protumor properties of neutrophils and have associated neutrophils with a poor prognosis (1,16,17). Recent findings have revealed that TANs exert antitumor effects in several cancer types through the stimulation of T cell activity (17,18). However, the role of neutrophils in the oncogene-induced TME has not yet been elucidated.

In the KRAS-transduced ovarian cancer model, a marked increase in the number of neutrophils was observed; however, it is still unclear whether the increased number of neutrophils exerts a pro-tumor or antitumor effect in this model. Therefore, in this study, we investigated the role of recruited neutrophils in KRAS-induced ovarian cancer progression, as well as their influence on the intraperitoneal microenvironment.

Materials and methods

Cell lines and establishment of oncogene-transduced ID8 cells. We established oncogene-transduced mouse ovarian cancer cell lines by transducing KRAS into the mouse ovarian epithelial immortalized cell line, ID8, which was established from C57BL/6 mice (8). The ID8 cells were a kindly gift from Dr Kathy Roby, Department of Anatomy and Cell Biology, University of Kansas Medical Center (Kansas City, KS, USA). An oncogenic mutant form of human KRAS (KRAS^{G12V}) was recombined into pDEST-CLXSN to generate pCLXSN-KRAS^{G12V}. Retrovirus packaging was performed as previously described (19). ID8-KRAS cells were established by infection of the LXSN-KRAS^{G12V} virus at a multiplicity of infection of 1, followed by G418 selection at a concentration of 800 μ g/ml for 1 week.

Mouse model. The ID8 and ID8-KRAS cells were cultured in Dulbecco's modified Eagle's medium (DMEM; Wako, Tokyo, Japan, 043-30085) containing 10% FBS, 100 U/ml penicillin, 0.1 mg/ml streptomycin, and 0.25 g/ml amphotericin B. The ID8 and ID8-KRAS cells (2×10^6) suspended in 1,000 μ l DMEM were injected into the peritoneal cavities of 8-week-old female C57/BL6 mice as previously described (8). The mice were obtained from Japan SLC and weighed 18–20 g. The animals were maintained at room temperature in a humidity-controlled room with a 12 h-light/12 h-dark cycle and were provided sterilized solid food and water ad libitum during the entire experimental period. Mice were sacrificed to minimize suffering when moribund behaviors were observed. In all cases, tissue collection procedures were initiated after animals had been euthanized by isoflurane overdose; mice were placed into a chamber filled with the vapor of the anesthetic isoflurane until respiration ceased (within 2 min) (20). For the mouse survival analysis, mice were sacrificed when their body weight (BW) exceeded 23 g after inoculation as in our previous study (8), we confirmed that that approximately 5 ml of ascites were accumulated with the formation of dissemination when the BW reached 23 g. BW and ascites weight were assessed at the time of sacrifice. The total number of mice used in this

study was 115 as follows: Experiment of the *in vitro* effects of neutrophils on CD8 T cell activation, 32 mice (no cancer mice, n=16; ID8 mice, n=8; ID8-KRAS mice, n=8); experiment of T cell costimulatory molecules on neutrophils, 83 mice (CD80: no cancer, n=8; ID8, n=5; ID8-KRAS, n=7; CD86: no cancer, n=6; ID8, n=6; ID8-KRAS, n=5; 4-1BBL: no cancer, n=11; ID8, n=5; ID8-KRAS, n=7; OX40L: no cancer, n=10; ID8, n=5; ID8-KRAS, n=8).

Neutrophil depletion. Neutrophils were depleted using anti-Ly6G mAb (mouse) (Nimp-R14, AdipoGen Life Sciences, San Diego, CA, USA, AG-20B-0043-C100). This mAb has been reported to selectively deplete neutrophils *in vivo* (21,22). The mice were injected intraperitoneally with 250 μ g anti-Ly6G mAb or matched isotype (rat IgG2b isotype control, Bio X Cell, West Lebanon, NH, USA, BE0090) in 0.5 ml of PBS or with PBS alone every 3 days by the following two medication methods: Method A, from day 7 after inoculation of ID8-KRAS cells. Mice were sacrificed when their body weight exceeded 23 g (isotype, n=8; and anti-Ly6G, n=9). Method B, from day 10 after inoculation of ID8-KRAS cells. Mice were sacrificed on day 16 (isotype, n=15; and anti-Ly6G, n=16). As a control, we used mice with no cancer which were not injected with any substance (method A, n=6; method B, n=6).

Isolation of myelocyte peritoneal ascites. Mice were peritoneally inoculated with ID8-KRAS cells by the described medication described above in method B. Following the injection of 3 ml of PBS, peritoneal myelocytes were recovered from peritoneal cavities of no cancer mice (n=6) and ID8-KRAS mice treated with anti-Ly6G or isotype mAb. Peritoneal myelocytes were centrifuged at 300 x g at room temperature for 5 min. Red blood cells were removed by RBC lysed.

Isolation of neutrophils from blood and peritoneal ascites. After euthanized by isoflurane overdose, peritoneal ascites and approximately 1.5 ml of blood by cardiac puncture were obtained from ID8 and ID8-KRAS mice. Neutrophils were isolated from the collected peritoneal ascites and blood using a magnetic cell sorting kit (Neutrophil Isolation kit, MACS KK; Miltenyi Biotec, Cologne, Germany) according to the manufacturer's instructions.

Isolation of lymphocytes from ascites and splenocytes. Mice were peritoneally inoculated with ID8-KRAS cells by the described medication method B. Peritoneal cells were obtained from peritoneal cavities of no cancer mice and ID8-KRAS mice treated with anti-Ly6G or isotype mAb after injection of 3 ml of PBS. Splenocytes were prepared as follows: spleens obtained from no cancer mice and ID8-KRAS mice treated with anti-Ly6G or isotype mAb were homogenized between glass slides, RBC lysed, washed twice, and filtered with 100 μ m filter. Peritoneal cells and splenocytes were centrifuged (300 x g, 15 min) to pellet them. The pellet was resuspended in 3 ml of PBS and then layered onto a discontinuous Percoll gradient (44%/70%; Percoll, GE Healthcare, Chicago, IL, USA) followed by centrifugation at 400 x g for 18 min. The band between the

Table I. Antibodies used in flow cytometry.

Antibody (clone, company, cat. no.)	Isotype (clone, company, cat. no.)
FITC-anti-mouse CD45 (B3821F4A/N901/UCHT1, Beckman Coulter, Tokyo, Japan, CO6607071)	FITC-rat IgG, 2b, k isotype (eBRG1, eBioscience, 11-4031-82)
FITC-anti-mouse c-mesenchymal-epithelial transition (eBioclone7, eBioscience, Tokyo, Japan, 11-8854-80)	FITC-rat IgG1, k isotype (eBRG1, eBioscience, 11-4301-82)
FITC-anti-mouse CD80 (16-10A1, BioLegend, Tokyo, Japan, 104705)	FITC-Arm Hamster, IgG isotype (eBio299Arm, eBioscience, 11-4888-81)
FITC-anti-mouse CD86 (GL1, BioLegend, 105005)	FITC-rat IgG, 2b, k isotype (eB149/10H5, eBioscience, 11-4031-82)
PE-anti-mouse 4-1BB ligand (TKS-1, BioLegend, 107105)	PE-mouse IgG, 2a, k isotype (MOPC-173, BioLegend, 400211)
FITC-anti-mouse CD11b (M1/70, eBioscience, 11-0122-85)	FITC-rat IgG, 2b, k isotype (eBRG1, eBioscience, 11-4031-82)
APC-anti-mouse Ly6G (RB6-8C5, eBioscience, 17-5931-82)	APC-Rat, IgG, 2a, k isotype (eBR2a, eBioscience, 17-4321-81)
PE-anti-mouse Ly6G (RB6-8C5, eBioscience, 12-5931-82)	PE-mouse IgG, 2a, k isotype (MOPC-173, BioLegend, 40021)
APC-anti-mouse Ly6C (HK1.4, BioLegend, 128015)	APC-Rat, IgG, 2c, k isotype (RTK4174, BioLegend, 400713)
PE-Cyanine5-anti-mouse F4/80 (BM8, BioLegend, 123111)	PE-Cyanine5-Rat, IgG, 2a, k isotype (eBR2a, eBioscience, 35-4321-80)
FITC-anti-mouse CD8a (53-6.7, eBioscience, 11-0081-81)	FITC-Rat, IgG, 2a, k isotype (eBR2a, eBioscience, 11-4321-82)
PE-Cyanine5-anti-mouse CD3e (145-2C11, eBioscience, 15-0031-82)	PE-Cyanine5-Arm Hamster, IgG isotype (eBio299Arm, eBioscience, 15-4888-82)
APC-anti-mouse CD4 (RM4-5, eBioscience, 17-0042-81)	APC-Rat, IgG, 2a, k isotype (eBR2a, BioLegend, 400512)
FITC-anti-mouse CD25 (PC61.5, BD, 558689)	FITC-mouse IgG1, λ isotype (G0114F7, BioLegend, 401913)
PE-anti-mouse FoxP3 (NRRF-30, eBioscience, 12-4771-80)	PE-mouse IgG, 2a, k isotype (MOPC-173, BioLegend, 400211)

FITC, fluorescein isothiocyanate; APC, allophycocyanin; PE, phycoerythrin.

44 and 70% layers was pipetted into another round-bottom centrifuge tube, diluted with 10 ml of PBS, and centrifuged at 300 x g for 5 min to remove the Percoll or small particulates. The collected pellet was then used for flow cytometric analysis.

Isolation of naïve CD8⁺ T cells from splenocytes. Splenocytes were obtained from the spleens of no cancer mice. Naïve CD8⁺ T cells from splenocytes were isolated using a magnetic cell sorting kit (Naïve CD8a⁺ T Cell Isolation Kit, mouse, MACS KK; Miltenyi Biotec) according to the manufacturer's instructions.

Flow cytometry. Samples (2x10⁶ cells/ml) were suspended in 500 μ l of PBS/1% BSA and analyzed by flow cytometry (FACSCalibur flow cytometer; Becton-Dickinson, Mountain

View, CA, USA). Matched isotype antibodies were used as controls. T cells and neutrophils were stained with the antibodies listed in Table I.

ELISA. Cytokine levels [interleukin (IL)-6 and IFN- γ] were measured in ascites or culture media with a specific ELISA kit (DuoSet ELISA; R&D Systems, Minneapolis, MN, USA) according to the manufacturer's instructions.

Coculture of neutrophils and naïve CD8⁺ T cells. T cell proliferation induced by plate-bound anti-mouse CD3e (1 μ g/ml; BioLegend, San Diego, CA, USA; clone: 145-2C11) was assessed using standard carboxyfluorescein diacetate succinimidyl ester (CFSE; Cayman Chemical, Ann Arbor, MI, USA) dilution methods. Purified naïve CD8⁺ T cells (2.0x10⁶ cells/ml) were labeled with CFSE and cocultured

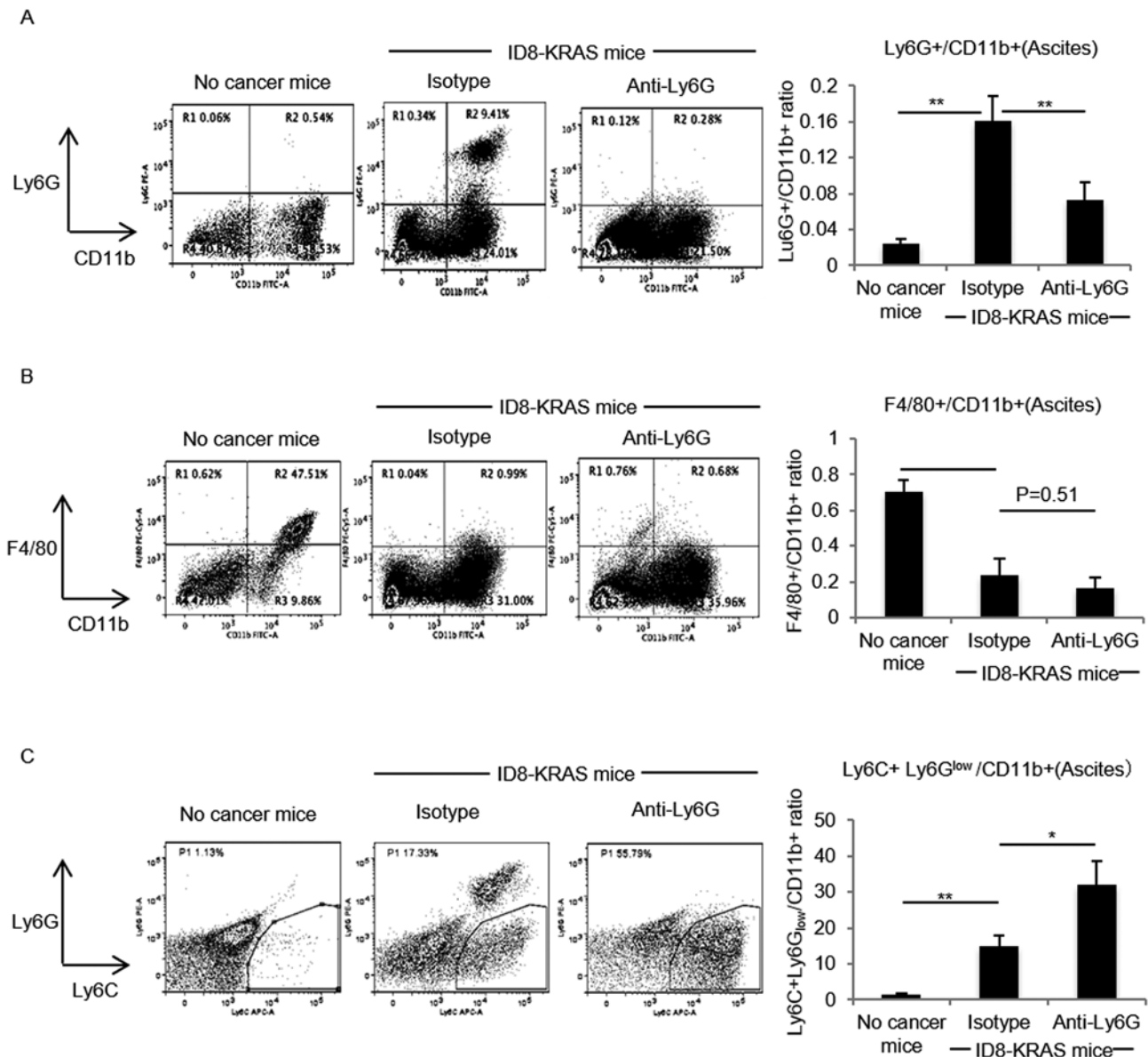


Figure 1. Effects of neutrophil depletion on peritoneal myelocytes. Murine ID8-KRAS cells (2×10^6) were intraperitoneally injected into 17 mice (isotype, $n=8$ and anti-Ly6G, $n=9$). The mice were intraperitoneally injected with $250 \mu\text{g}$ of anti-Ly6G mAb or matched isotype in 0.5 ml of PBS every 3 days from day 7 after the inoculation of ID8-KRAS cells. Peritoneal myelocytes were obtained from ascites of no cancer mice ($n=6$) and ID8-KRAS mice treated with anti-Ly6G or isotype mAb after washing with 3 ml of PBS. The depletion status was confirmed by assessing the proportion of (A) Ly6G⁺/CD11b⁺ cells, (B) F4/80/CD11b⁺ cells, and (C) Ly6C⁺Ly6G^{low} cells after gating for CD11b by flow cytometry. Peritoneal myelocytes were stained with the following antibodies: FITC-anti-mouse CD11b, PE-anti-mouse Ly6G, APC-anti-mouse Ly6C and PE-Cyanine5-anti-mouse F4/80. Error bars represent the means $\pm$ SEM. Statistical analysis was performed using the Student's t-test. The P-value was adjusted with the Holm's method (* $P<0.05$, ** $P<0.01$).

with neutrophils (2.0×10^6 cells/ml) obtained from the blood or ascites of each mouse (no-cancer control, ID8 mice, or ID8-KRAS mice) in CD3-coated plates for 3 days in complete cell culture medium. The complete cell culture medium was composed of RPMI-1640 medium (Wako) supplemented with 10% FBS, 100 U/ml penicillin, 0.1 mg/ml streptomycin and 0.25 g/ml amphotericin B. To establish whether T cell proliferation was induced by intercellular communication, we assessed cocultures using chambers. Five hundred microliters of neutrophils obtained from the ascites of ID8-KRAS mice were added to the upper compartments of the chambers at a concentration of 2.0×10^6 cells/ml. Five hundred microliters of purified naïve CD8⁺ T cells were seeded onto the bottom chamber at a concentration of 2.0×10^6 cells/ml. Following a

3-day incubation, cells on the upper chamber were completely removed. The CFSE signal was analyzed by flow cytometry.

Statistical analysis. Data are presented as the means $\pm$ SEM. The log-rank test was used to detect differences in animal survival (Kaplan-Meier survival curves). All other comparisons were performed with the two-tailed Student's t-test. For performing multiple comparisons, all the P-values were adjusted by the Holm's method using the Microsoft Office Excel 2015 (BellCurve, Tokyo, Japan). Other statistical analyses were conducted using JMP11 (SAS Institute Japan, Tokyo, Japan). Other statistical analyses were conducted using JMP11 (SAS Institute Japan, Tokyo, Japan). A value of $P<0.05$ was considered to indicate a statistically significant difference.

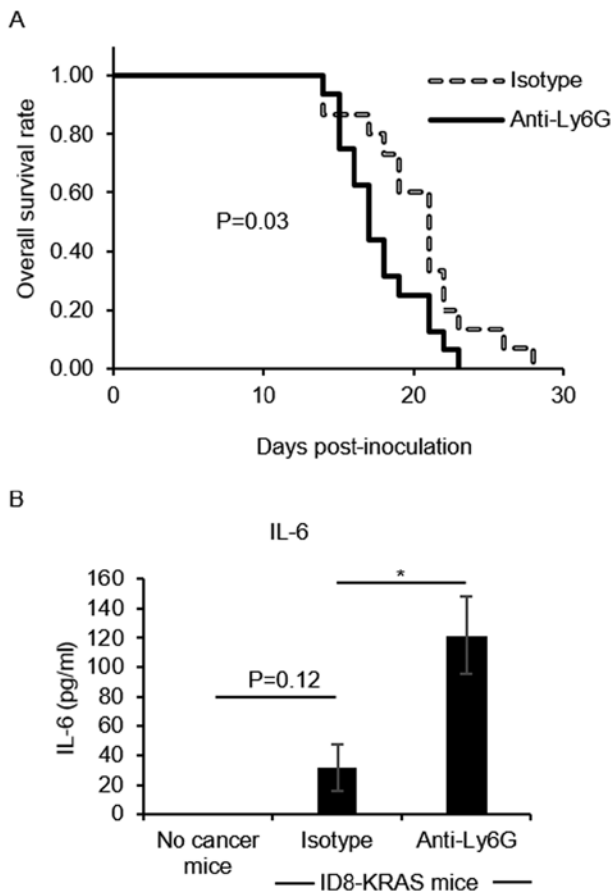


Figure 2. Effects of neutrophil depletion on cancer progression. (A) Murine ID8-KRAS cells (2×10^6) were intraperitoneally injected into 31 mice (isotype, $n=15$; and anti-Ly6G, $n=16$). The mice were intraperitoneally injected with $250 \mu\text{g}$ of anti-Ly6G mAb or matched isotype in 0.5 ml of PBS every 3 days from day 7 after inoculation of ID8-KRAS cells. For the mice survival analysis, the end point was determined as 'body weight (BW) = 23 g ', and mice were sacrificed when their BW exceeded 23 g after inoculation. The log-rank test was used to detect differences (Kaplan-Meier survival curves). (B) ID8-KRAS-injected mice were treated with anti-Ly6G mAb ($n=9$) or isotype control mAb ($n=8$) from day 10 after the inoculation of ID8-KRAS cells, and the ascites were obtained on day 16 after washing with 3 ml of PBS. The ascites from no cancer mice were also obtained after washing with 3 ml of PBS ($n=6$). The interleukin (IL)-6 concentration in ascites of the 3 groups (no cancer, ID8-KRAS treated with isotype mAb, and ID8-KRAS treated with anti-Ly6G mAb) was analyzed using specific ELISA kits. Error bars represent the means $\pm$ SEM. Statistical analysis was performed using the Student's t-test. The P-value was adjusted using the Holm's method ($P < 0.05$).

Study approval. Animal studies were approved by the University of Tokyo Animal Committee. Our IACUC permitted this study and provided this study the approval numbers P-14-027 and P15-060.

Results

Neutrophil depletion modulates the peritoneal microenvironment. We previously reported that the neutrophil count is markedly increased in ID8-KRAS-induced ascites (8), and demonstrated that neutrophil recruitment into the peritoneal cavity begins at around 10 days after the intraperitoneal injection. Therefore, in this study, we focused in particularly on neutrophil functions in the late phase and began the depletion of neutrophils 10 days after the injection. To investigate the role

of increased neutrophils in the peritoneal microenvironment, neutrophils were depleted with a specific antibody, anti-Ly6G mAb, and its isotype antibody was used as the control. Profiles of intraperitoneal myelocytes were assessed by flow cytometry. We first confirmed that the isotype antibody had no effect on the myelocyte population in ascites (data not shown). When compared with the isotype antibody, the anti-Ly6G antibody successfully decreased the $\text{Ly6G}^+/\text{CD11b}^+$ neutrophil numbers ($P < 0.01$, Fig. 1A); however, no significant difference was observed in the number of $\text{F4/80}^+/\text{CD11b}^+$ macrophages between the isotype and neutrophil-depleted groups ($P = 0.51$, Fig. 1B). Focusing on the profiles of myeloid-derived suppressor cells (MDSCs), the amount of $\text{Ly6C}^+/\text{Ly6G}^{\text{low}}/\text{CD11b}^+$ monocytic-MDSCs (M-MDSCs) was significantly increased between the isotype and neutrophil-depleted groups ($P < 0.05$, Fig. 1C).

Neutrophil depletion accelerates cancer progression and aggravates inflammation. We then investigated the effects of neutrophils on cancer progression. We first confirmed that the isotype antibody had no effect on the accumulation of ascites ($P = 0.18$, data not shown). When compared with the isotype group, the neutrophil-depleted group exhibited a more rapid accumulation of ascites ($P = 0.03$, Fig. 2A).

A higher level of IL-6 is a hallmark of ovarian cancer progression (23,24). In addition, increases in IL-6 production have been observed in KRAS-induced ovarian cancer (8); therefore, in this study, we measured the IL-6 concentration as a surrogate marker of cancer progression. Neutrophil depletion significantly increased the IL-6 concentration in ascites compared with that in the isotype group ($P < 0.05$, Fig. 2B).

Neutrophil depletion modulates the T cell profile in ascites and spleen. We examined the effects of neutrophils on the T lymphocyte population. Differences in the $\text{CD4}/\text{CD3}\epsilon$ ratio and $\text{CD8}/\text{CD3}\epsilon$ ratio in the spleen and ascites were compared between the isotype and neutrophil-depleted groups. The $\text{CD4}/\text{CD3}\epsilon$ ratio in the KRAS-induced ascites was significantly increased by neutrophil depletion ($P < 0.05$, Fig. 3A). On the contrary, the $\text{CD8}/\text{CD3}\epsilon$ ratio in the KRAS-induced ascites and spleen was reduced by neutrophil depletion ($P < 0.05$, Fig. 3B).

Neutrophil depletion increases the number of regulatory T cells in ascites. Previous studies have demonstrated that M-MDSCs induce regulatory T cell (Treg) differentiation (25-27). The data of this study indicated that an increased number of $\text{Ly6C}^+/\text{Ly6G}^{\text{low}}/\text{CD11b}^+$ M-MDSCs was accompanied by an increased number of CD4^+ T cells. Therefore, we hypothesized that in the neutrophil-depleted group, the number of Tregs would be increased. The number of Tregs was assessed by FoxP3-CD25-CD4 flow cytometry. The number of $\text{FoxP3}^+/\text{CD25}^+/\text{CD4}^+$ Tregs tended to be increased in the KRAS-induced ascites, and their number was markedly increased by neutrophil depletion ($P < 0.05$, Fig. 4A); however, their number did not significantly differ in the spleen (Fig. 4B).

Neutrophils from ID8-KRAS ascites stimulated CD8 T cell activity. The results of the *in vivo* assessment suggested that neutrophils also modulate CD8^+ T cell activity. Therefore,

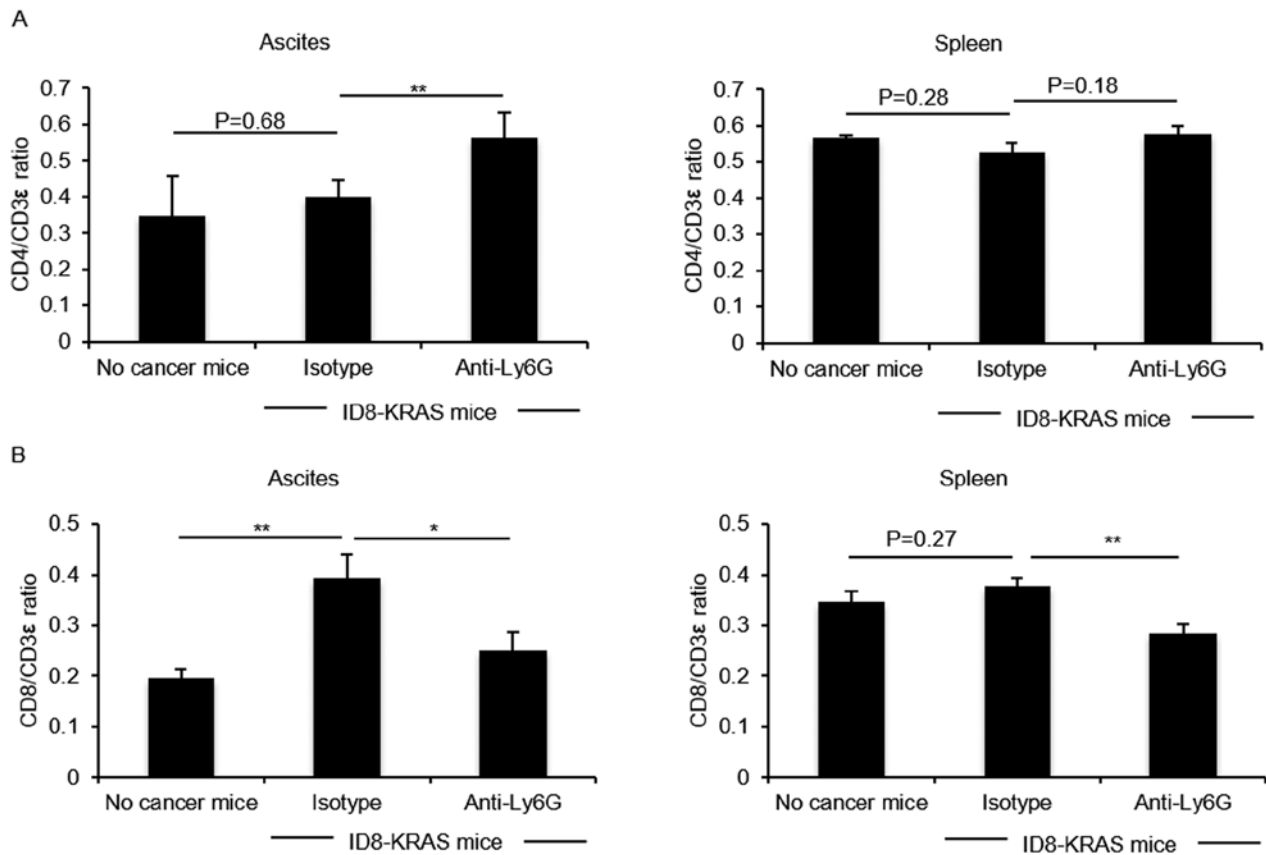


Figure 3. Effects of neutrophil depletion on the T cell subset. ID8-KRAS-injected mice were treated with anti-Ly6G mAb (n=9) or isotype control mAb (n=8) from day 10 after the inoculation of ID8-KRAS cells and sacrificed on day 16. No cancer mice were used as controls (n=6). Ascites and spleen were obtained from the no-cancer mice and ID8-KRAS mice treated with anti-Ly6G or isotype mAb after washing with 3 ml of PBS. T cells were collected by Percoll density gradient (44%/70%). T cells were stained with the following antibodies: PE-Cy5-anti-mouse CD3ε, FITC-anti-mouse CD8a, and APC-anti-mouse CD4. The (A) CD4/CD3ε ratio of ascites and spleen and (B) CD8/CD3ε ratio of ascites and spleen were analyzed by flow cytometry. Error bars represent the means $\pm$ SEM. Statistical analysis was performed using the Student's t-test. The P-value was adjusted using the Holm's method (* $P < 0.05$, ** $P < 0.01$).

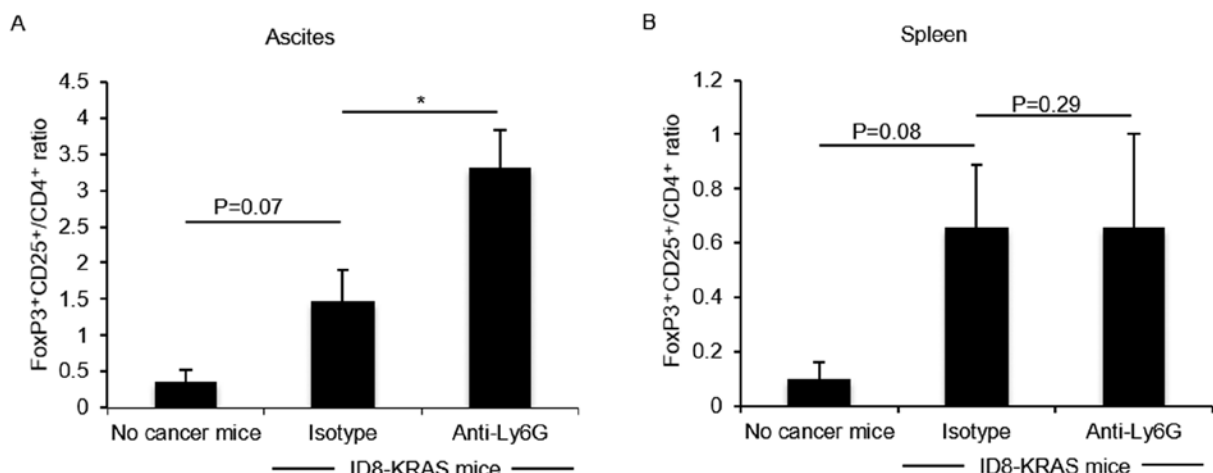


Figure 4. Effects of neutrophil depletion on Tregs. ID8-KRAS-injected mice were treated with anti-Ly6G mAb (n=9) or isotype control mAb (n=8) from day 10 after the inoculation of ID8-KRAS cells and sacrificed on day 16. No cancer mice were used as controls (n=6). Ascites and spleen were obtained from the no cancer mice and ID8-KRAS mice treated with anti-Ly6G or isotype mAb after washing with 3 ml of PBS. T cells were collected by Percoll density gradient (44%/70%). T cells were stained with the following antibodies: FITC-anti-mouse CD25, APC-anti-mouse CD4, and PE-anti-mouse FoxP3. The FoxP3+CD25+/CD4+ ratio of (A) ascites and (B) spleen was analyzed by flow cytometry. Error bars represent the means $\pm$ SEM. Statistical analysis was performed using the Student's t-test. The P-value was adjusted using the Holm's method (* $P < 0.05$).

we assessed the effects of neutrophils on CD8⁺ T cell activity *in vitro*. Naïve CD8⁺ T cells were cultured with anti-CD3 antibody alone or with peripheral blood neutrophils (PBNs)

or peritoneal neutrophils (PENs) from each mouse (no cancer mice and ID8-KRAS mice). The proliferation of naïve CD8⁺ T cells was assessed using CFSE staining. CD8⁺ T cell

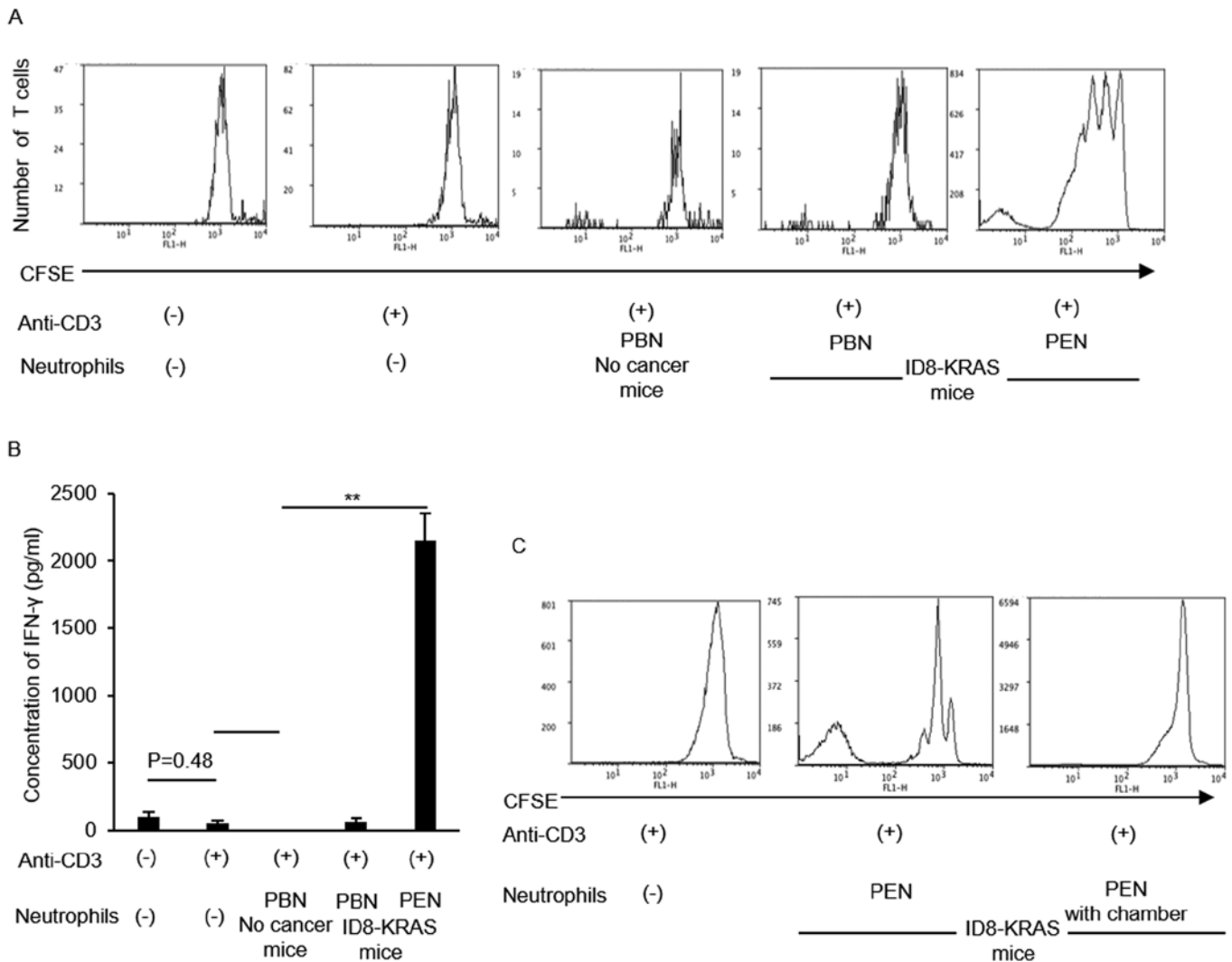


Figure 5. *In vitro* effects of neutrophils on CD8⁺ T cell activation. CFSE-labeled naïve CD8⁺ T cells were isolated from the spleen of no cancer mice (n=8). Naïve CD8⁺ T cells (2.0×10^6 cells/ml) were cocultured with neutrophils (2.0×10^6 cells/ml) from each group under stimulation of plate-bound anti-CD3 antibody for 3 days. Neutrophils were obtained from PBNs of no cancer mice or PBNs and PENs of ID8-KRAS mice (n=8). PBN and PEN indicate peripheral blood neutrophil and peritoneal neutrophils, respectively. (A) The CFSE signal was analyzed by flow cytometry. The data provided are representative results of 3 independent experiments. (B) The concentration of IFN- γ in the supernatant of each culture medium was analyzed by specific ELISA. The data provided are representative results of 3 independent experiments. Error bars represent the means $\pm$ SEM. Statistical analysis was performed using the Student's t-test. The P-value was adjusted using the Holm's method ($P < 0.05$, $^{**}P < 0.01$). (C) To assess the effects of cell-to-cell interactions between T cells and neutrophils, PENs of ID8-KRAS mice were added to the upper compartments of the chambers at a concentration of 2.0×10^6 cells/ml. Naïve CD8⁺ T cells were added to the bottom chamber at a concentration of 2.0×10^6 cells/ml and cocultured for 3 days in CD3-coated plates. The CFSE signal was analyzed by flow cytometry. The data provided are representative results of 3 independent experiments.

proliferation markedly increased in coculture with PENs from ID8-KRAS mice (Fig. 5A). IFN- γ production by CD8⁺ T cells was also assessed in each group using specific ELISA kits. When compared with the PBNs from no cancer mice or ID8-KRAS mice, coculture with PENs from ID8-KRAS mice markedly increased IFN- γ production by naïve CD8⁺ T cells (Fig. 5B). The T cell stimulatory effects of PENs from ID8-KRAS mice were reversed when PENs from ID8-KRAS mice were added to the coculture chamber, suggesting that cell-to-cell contact was indispensable for the T cell stimulatory effects of KRAS-related neutrophils (Fig. 5C).

T cell costimulatory molecules are expressed at high levels in neutrophils from ID8-KRAS ascites. To elucidate the mechanisms through which PENs from ID8-KRAS mice

strongly promote IFN- γ production and the proliferation of CD8⁺ T cells, T cell costimulatory molecules CD80, CD86, OX40 ligand (OX40L) and 4-1BB ligand (4-1BBL) on the surface of neutrophils from no cancer mice, ID8 mice and ID8-KRAS mice were assessed by flow cytometry. Almost no difference in CD80 and CD86 expression in the PBNs and PENs was observed among these 3 groups (Fig. 6A and B). Furthermore, no significant differences in CD80 and CD86 expression between the PBNs and PENs was noted in each mouse group (Fig. 6A and B). The ID8-KRAS mice expressed higher levels of 4-1BBL in both the PBNs and PENs (Fig. 6C) (PBNs: no cancer vs. ID8-KRAS, $P < 0.01$; ID8 vs. ID8-KRAS, $P = 0.05$; PENs: ID8 vs. ID8-KRAS, $P < 0.05$). No significant differences were observed between the PBNs and PENs in each mouse group (ID8 mice, $P = 0.20$; ID8-KRAS mice,

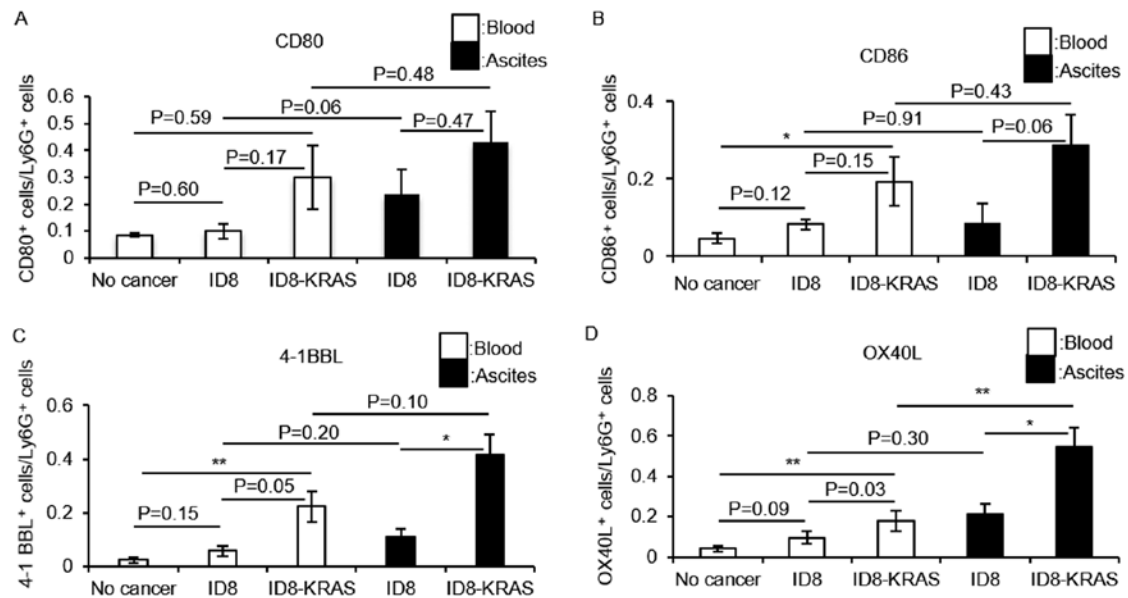


Figure 6. Expression of T cell costimulatory molecules on neutrophils. Murine ID8 cells (2×10^6) and ID8-KRAS cells (2×10^6) were injected into mice, and neutrophils were obtained from ascites or blood when the body weight exceeded 23 g. Neutrophils from no cancer mice were used as a control. (A-D) The expression of costimulatory molecules on the gated Ly6G⁺ neutrophils from ascites or blood was analyzed by flow cytometry. The number of mice used in each experiment was as follows: (A) CD80: no cancer, n=8; ID8, n=5; ID8-KRAS, n=7; (B) CD86: no cancer, n=6; ID8, n=6; ID8-KRAS, n=5; (C) 4-1BBL: no cancer, n=11; ID8, n=5; ID8-KRAS, n=7; (D) OX40L: no cancer, n=10; ID8, n=5; ID8-KRAS, n=8. Error bars represent the means $\pm$ SEM. Statistical analysis was performed using the Student's t-test. The P-value was adjusted using the Holm's method (* $P < 0.05$, ** $P < 0.01$). PBNs and PENs indicate peripheral blood neutrophils and peritoneal neutrophils, respectively.

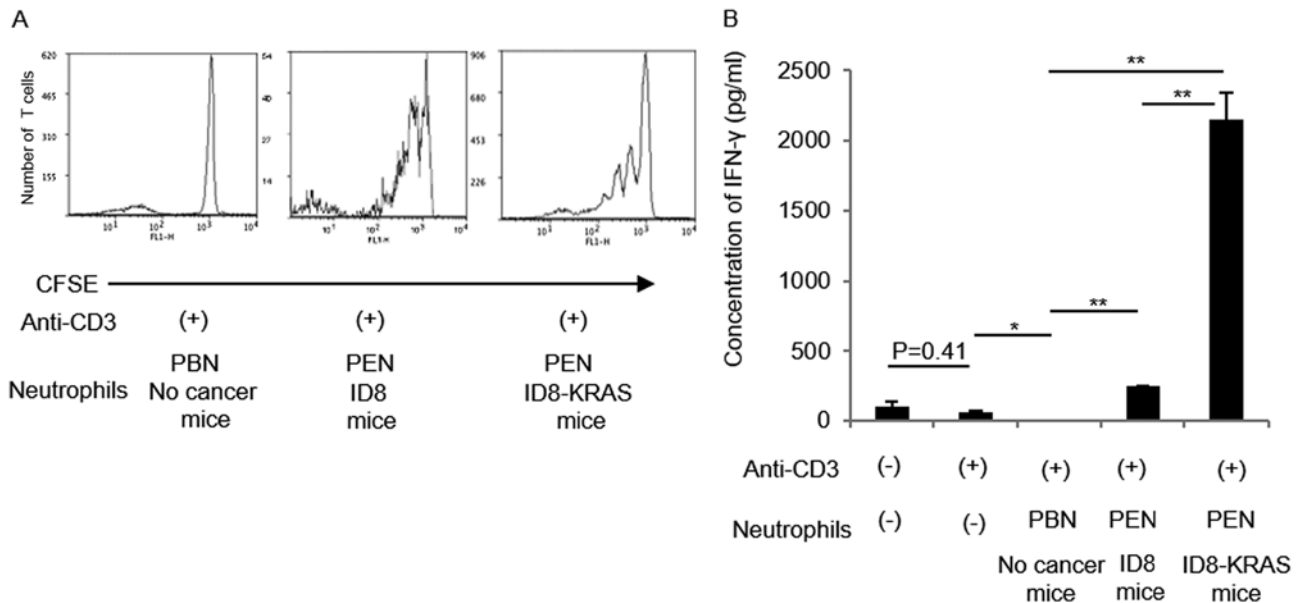


Figure 7. *In vitro* effects of neutrophils on activation of CD8⁺ T cells from ID8 mice. CFSE-labeled naïve CD8⁺ T cells were isolated from the spleen of no cancer mice (n=8). Naïve CD8⁺ T cells (2.0×10^6 cells/ml) were cocultured with neutrophils (2.0×10^6 cells/ml) from each group under stimulation of plate-bound anti-CD3 antibody for 3 days. Neutrophils were obtained from PBNs of no cancer mice or PENs of ID8 mice (n=8) and ID8-KRAS mice (n=8). PBNs and PENs indicate peripheral blood neutrophils and peritoneal neutrophils, respectively. (A) The CFSE signal was analyzed by flow cytometry. The data provided are representative results of 3 independent experiments. (B) The concentration of IFN- γ in the supernatant of each culture medium was analyzed by specific ELISA. The data provided are representative results of 3 independent experiments. Error bars represent the means $\pm$ SEM. Statistical analysis was performed using the Student's t-test. The P-value was adjusted using the Holm's method (* $P < 0.05$, ** $P < 0.01$).

$P = 0.10$, respectively). Marked differences were observed in the expression levels of OX40L. In both the PBNs and PENs, the OX40L expression levels were significantly higher in the ID8-KRAS mice than in the no cancer mice and ID8 mice (Fig. 6D) (PBNs: no cancer vs. ID8-KRAS, $P < 0.01$; ID8 vs.

ID8-KRAS, $P = 0.03$; PENs: ID8 vs. ID8-KRAS, $P < 0.05$). When the PBNs and PENs from each mouse were compared, the PENs from the ID8-KRAS mice exhibited a higher expression of OX40L than the PBNs (Fig. 6D) (ID8 mice, $P = 0.30$; ID8-KRAS mice, $P < 0.01$, respectively).

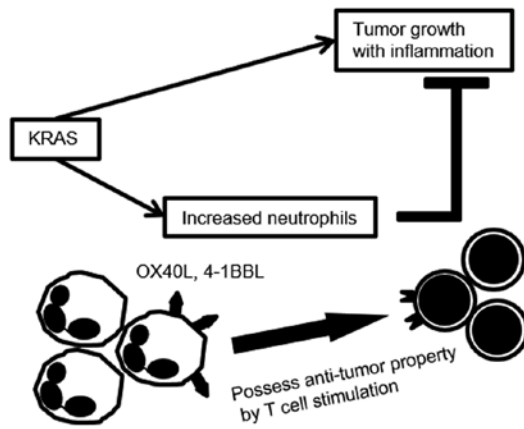


Figure 8. Schematic diagram illustrating intraperitoneal neutrophils activated by KRAS-induced ovarian cancer. The increased number of neutrophils in KRAS-induced ascites possessed antitumor properties by modulating the tumor microenvironment (TME).

CD8⁺ T cell stimulatory function on neutrophils is much stronger in ID8-KRAS mice than in ID8 mice. ID8-KRAS-associated neutrophils possessed T cell stimulatory function with higher expression of OX40L and 4-1BBL. We next evaluated whether T cell stimulatory properties of neutrophils were present in any cancer-related microenvironment. Therefore, we compared T cell stimulatory effects between neutrophils from ID8-induced ascites and those from ID8-KRAS-induced ascites. PENs from ID8-induced ascites slightly activated CD8⁺ T cells by inducing IFN- γ production, as well as stimulating CD8⁺ T cell proliferation, which was much weaker than that promoted by PENs from ID8-KRAS-induced ascites (Fig. 7).

A comparison of T cell costimulatory molecules between ID8 and ID8-KRAS revealed that the OX40L and 4-1BBL expression levels in PENs were significantly lower in ID8 mice than in ID8-KRAS mice (Fig. 8).

Discussion

In the current study, we found that neutrophil depletion in the ID8-KRAS model accelerated tumor formation and aggravated IL-6-associated inflammation. Neutrophil depletion increased the number of Ly6C⁺Ly6G^{low}/CD11b⁺ M-MDSCs accompanied with an increased number of Tregs in ascites. Furthermore, the neutrophil depletion decreased the CD8⁺ T cell population. PENs from ID8-KRAS mice exhibited strong potential to enhance T cell proliferation with a higher expression of T cell costimulatory molecules compared with the PENs from ID8 mice.

A number of studies have reported that an elevated neutrophil count or an increased N/L ratio in blood is a poor prognostic marker in several cancer types (28-31). The results of our previous study (8) also support the above finding that an increased neutrophil count, which may induce an increased N/L ratio, was observed in ID8-KRAS-induced rapid cancer progression. However, in our model, neutrophil depletion appeared to accelerate tumor formation. These findings suggest that the increase in TANs in our model had antitumor effects.

The results of this study revealed that neutrophil depletion increased the number of CD4⁺ T cells in ascites, particularly the Tregs fraction. Furthermore, we revealed that neutrophil depletion increased the M-MDSC population. MDSCs are subdivided into two major groups: Polymorphonuclear MDSCs (PMN-MDSCs) and M-MDSCs. PMN-MDSCs have a morphology similar to that of granulocytes, and M-MDSCs are morphologically similar to monocytes. In mice, PMN-MDSCs have a phenotype of CD11b⁺Ly6C⁺Ly6G^{high}, whereas M-MDSCs have a phenotype of CD11b⁺Ly6G⁺Ly6C^{high} (32,33). In our model, the MDSCs increased by neutrophil depletion were classified as M-MDSCs with the phenotype of CD11b⁺Ly6G⁺Ly6C^{high}. There is substantial variability in the M-MDSC proportion depending on the type of cancer. Patients with melanoma and prostate cancer have a substantially higher proportion of M-MDSCs in the peripheral blood than PMN-MDSCs (34). In ovarian cancer, M-MDSCs have been reported to be present in both the periphery and ascites. Ascite-derived IL-6 and IL-10 and their downstream signal transducer and activator of transcription 3 (STAT3) signal are critically responsible for the accumulation and suppressive activity of M-MDSCs (35). M-MDSCs potently suppress non-specific T cell responses, and on a per-cell basis, M-MDSCs have a high suppressive activity with a higher production of nitric oxide (NO), Arg1, TGF- β and immunosuppressive cytokines (36-40). Furthermore, it has been previously demonstrated that TGF- β induced Foxp3 gene expression in T cell receptor-challenged CD4⁺CD25⁻ peripheral naïve T cells, which were then transformed toward a Treg phenotype with potent immunosuppressive potential (41). In this model, M-MDSCs may contribute to cancer progression by inducing Treg differentiation and suppressing CD8 T cell activity.

The results of this study revealed that neutrophil depletion decreased the CD8⁺ T cell population in KRAS-induced ascites, suggesting that neutrophils promote CD8⁺ T cell proliferation or survival in the KRAS-induced environment. The finding that TANs in KRAS-induced ascites had a strong potential for promoting the proliferation of naïve CD8⁺ T cells *in vitro* supports the results obtained in these *in vivo* assessments. In lung cancer, T cell costimulatory molecules are responsible for T cell activation by TANs (18). We also confirmed that TANs in KRAS-induced ascites expressed OX40L and 4-1BBL at higher levels than neutrophils from blood or TANs in ID8-induced ascites. The T cell-stimulatory effects of TANs from KRAS-induced ascites were reversed by chamber separation, suggesting that cell-to-cell interactions are indispensable for T cell activation. T cell costimulatory molecules on neutrophils may be responsible for the involvement of TANs in KRAS-induced ascites in promoting T cell proliferation.

Although the function of neutrophils in promoting T cell activation has already been reported (39), the types of tumors that induce neutrophils with antitumor properties have not yet been identified. In this study, PENs from ID8-KRAS mice possessed markedly stronger potential for T cell activation than PENs of ID8 mice. A difference was also observed between the OX40L and 4-1BBL expression levels on the neutrophil surfaces. Our results suggest that the KRAS-induced environment generated neutrophils with potent antitumor properties. To the best of our knowledge, no study

to date has demonstrated an association between oncogenes and the properties of neutrophils. The cancer-specific modulation of neutrophils has not yet been reported; however, the antitumor or protumor function of neutrophils may be related to the activated oncogenes and their specific TME. Furthermore, given that KRAS-related neutrophils possessed antitumor properties with increased expression of OX40L and 4-1BBL, OX40L- or 4-1BBL-based immunotherapy may be a therapeutic strategy for KRAS mutated cancer.

In the current study, we used anti-Ly6G mAb to deplete neutrophils. It is possible that CD11b⁺Ly6C⁺Ly6G^{high} PMN-MDSCs were also depleted in our model as it is difficult to distinguish neutrophils from PMN-MDSCs. However, in our model, the depletion of Ly6G^{high} populations resulted in an enhanced cancer formation accompanied by the immunosuppressive microenvironment. Considering that MDSCs have an immunosuppressive function, it appeared that, in our model, a decreased number of neutrophils with immunostimulatory effects potentially affected the TME compared with a decreased number of PMN-MDSCs with immunosuppressive effects. In addition, we only used a mouse ovarian cancer model to elucidate the association between the oncogene KRAS and the properties of neutrophils. Further studies utilizing clinical samples are warranted for the confirmation of the association identified in this study.

We herein propose that the increased number of neutrophils in KRAS-induced ascites possessed antitumor properties by modulating the TME, and that this effect was more potent in KRAS-associated neutrophils. Knowledge regarding these characteristics of neutrophil modification may provide a better understanding of the cancer-induced microenvironment.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

MY, AT, KKa, KK, TN and KO were involved in the conception and design of the study. MY, AT, KKa, KK, MM, OWH and TK were involved in the development of methodology. TK and OWH were involved in the establishment of cell lines and technical support. MY, AT, KKa and KA were involved in the acquisition of data. MY, AT, KKa, JO, AK, HN, MS, AF, TI, KT, MM, TN, TA, KKo, OWH, KO, TK, YO and TF were involved in the analysis and interpretation of data. MY, AT, KKa and KA were involved in the writing, reviewing, and/or revision of the manuscript. MY, AT, KKa, JO, KA and AK

were involved in administrative, technical, or material support. MY, AT, KKa, MM, TN, TA, KKo, OWH, KO, TK, YO and TF supervised the study.

Ethics approval and consent to participate

The animal studies were approved by the University of Tokyo Animal Committee. Our IACUC permitted this study and gave this study the approval numbers P-14-027 and P15-060.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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