

Cancer stem cell and mesenchymal cell cooperative actions in metastasis progression and hormone resistance in prostate cancer: Potential role of androgen and gonadotropin-releasing hormone receptors (Review)

HÉCTOR R. CONTRERAS, FERNANDA LÓPEZ-MONCADA and ENRIQUE A. CASTELLÓN

Laboratory of Cellular and Molecular Oncology, Department of Basic and Clinical Oncology,
Faculty of Medicine, University of Chile, Santiago 8380453, Chile

Received July 18, 2019; Accepted January 9, 2020

DOI: 10.3892/ijo.2020.5008

Abstract. Prostate cancer (PCa) is the leading cause of male cancer-associated mortality worldwide. Mortality is associated with metastasis and hormone resistance. Cellular, genetic and molecular mechanisms underlying metastatic progression and hormone resistance are poorly understood. Studies have investigated the local effects of gonadotropin-releasing hormone (GnRH) analogs (used for androgen deprivation treatments) and the presence of the GnRH receptor (GnRH-R) on PCa cells. Furthermore, cell subpopulations with stem-like properties, or cancer stem cells, have been isolated and characterized using a cell culture system derived from explants of human prostate tumors. In addition, the development of preclinical orthotopic models of human PCa in a nonobese diabetic/severe combined immunodeficiency mouse model of compromised immunity has enabled the establishment of a reproducible system of metastatic progression *in vivo*. There is increasing evidence that metastasis is a complex process involving the cooperative actions of different cancer cell subpopulations, in which cancer stem-like cells would be responsible for the final step of colonizing premetastatic niches. It has been hypothesized that PCa cells with stemness and mesenchymal signatures act cooperatively in metastatic progression and the inhibition of stemness genes, and that overexpression of androgen receptor (AR) and GnRH-R decreases the rate the metastasis and sensitizes tumors to hormone therapy. The aim of the present review is to analyze the evidence regarding this cooperative process and

the possible influence of stem-like cell phenotypes, AR and GnRH-R in metastatic progression and hormone resistance. These aspects may represent an important contribution in the understanding of the mechanisms underlying metastasis and hormone resistance in PCa, and potential routes to blocking these processes, enabling the development of novel therapies that would be particularly relevant for patients with metastatic and castration-resistant PCa.

Contents

1. Introduction
2. Androgen deprivation and local effects of GnRH analogs in prostate cancer
3. Epithelial-mesenchymal transition and cancer stem cells in prostate cancer
4. Tumor cell phenotypic heterogeneity and metastatic processes
5. Different malignant cell types in a solid tumor may collaborate to produce distant metastasis
6. Orthotopic model for the study of human prostate cancer metastasis
7. Conclusions

1. Introduction

Prostate cancer (PCa) is one of the major causes of male cancer-associated death worldwide (1). Over the last few decades, screening programs have increased early diagnosis and identified treatments with the potential to cure the disease (2-5). However, the high rates of recurrence and metastasis remain major challenges in treating PCa (6-12). During a long period of the disease, PCa can become sensitive to androgen treatment (13,14). Testosterone controls cell proliferation, tumor growth and, potentially, dissemination (15-17), which is an advantage in treatments that involve androgen deprivation (AD), when curative surgery cannot be performed (18,19). Pharmacological castration using gonadotropin-releasing hormone (GnRH) analogs to

Correspondence to: Dr Enrique A. Castellón, Laboratory of Cellular and Molecular Oncology, Department of Basic and Clinical Oncology, Faculty of Medicine, University of Chile, Independencia 1027, Santiago 8380453, Chile
E-mail: ecastell@med.uchile.cl

Key words: cancer stem cells, epithelial-mesenchymal transition, hormone resistance, androgen receptor, gonadotrophin-releasing hormone receptor, metastatic cooperation

block the hypothalamus-hypophysis-testicular axis provides the first-line treatment for disseminated PCa (20-22). However, during AD therapy, PCa cells frequently become androgen-resistant, resulting in a castration-resistant form of the disease with a poor prognosis (23-25). The genetic and molecular mechanisms underlying androgen resistance remain poorly understood (26-28). Research suggests that, in certain cases, the androgen receptor (AR) is involved in this resistance (29-31). On the other hand, recurrence and metastasis progression are complex processes that involve several mechanisms and genomic modifications of malignant cells (32,33). It is well-known that epithelial-mesenchymal transition (EMT) is the main pathway via which malignant epithelial cells from carcinomas alter their gene expression profile to display a mesenchymal phenotype, acquiring, among other features, one of the hallmarks of cancer cells: Invasive behavior (34-38). However, increasing evidence indicates that tumors contain a phenotypically heterogeneous cell population, and that the cooperative action of these different types of malignant cells is potentially required to accomplish a successful metastatic process (39-42). In the past few years, a small subpopulation of malignant cells with stem-like properties has been identified in numerous types of cancer, including PCa (43,44). These cells have been termed tumor-initiating cells (TICs) or cancer stem cells (CSCs), and are hypothesized to be responsible for recurrence and metastasis (45-48).

2. Androgen deprivation and local effects of GnRH analogs in PCa

As aforementioned, GnRH analog therapy is the gold standard to treat disseminated PCa (20,49). This treatment induces AD by blocking the hypothalamus-hypophysis-testicular axis, resulting in pharmacological castration. This type of therapy is very efficient at delaying tumor growth until PCa becomes castration-resistant (20). Gene amplification, mutations and other alterations in the AR gene have been identified (29,50-52). In addition, overexpression or constitutive activation of other proliferation signaling pathways that overcome androgen control have been reported (53,54). In addition, alterations in androgen metabolism within the prostate gland have been associated with androgen sensitivity (55,56). It is postulated that castration-resistant PCa arises from a combination of these different mechanisms. Our previous research, as well as other studies, have reported the presence of GnRH receptor (GnRH-R) in PCa cells (57-59). Furthermore, it has been observed that GnRH analogs induce proliferation arrest and apoptosis in PCa cells in a primary culture system (57,58). GnRH-R expression increases from benign prostatic hyperplasia to medium histological grade (Gleason score 6-7), and subsequently decreases in samples from patients with higher Gleason scores (60). Local cellular effects of GnRH analogs may be of clinical relevance, as these effects remain despite cell androgen insensitivity (58,60). Concentrations >20 ng/ml are required to obtain significant *in vitro* apoptotic effects (61); however, the plasma concentrations in patients receiving AD treatment are below this level (62). This problem may be solved via intraprostatic administration of GnRH analogs. Unfortunately, patients who are castration-resistant often have a higher Gleason score and, as aforementioned, GnRH-R expression decreases with

higher Gleason scores. There is evidence that GnRH-R in PCa, specifically in the gonadotropic cells, is retained primarily in the endoplasmic reticulum, where it can be moved to the plasma membrane using peptide-mimetic compounds called pharmacoperones (pharmacological chaperones) (60). Using this strategy, it is possible to increase GnRH-R expression in cultured PCa cells and sensitize them to the apoptotic effects of GnRH analogs (Fig. 1).

3. EMT and CSCs in PCa

EMT is a process in which an epithelial genetic program switches to a mesenchymal program; as a result, an epithelial cell loses its polarity, proliferation, and differentiation control and positioning, changing to a mesenchymal phenotype (63-65). This is a physiologically normal process occurring primarily during embryonic development (66). During carcinogenesis, similar genetic changes occur in carcinomas that transform a malignant epithelial cell into a highly proliferative and invasive mesenchymal-like cell (65,67). Epithelial malignant cells progressively lose adhesion molecules, such as E-cadherin, syndecans and tight junction molecules, whereas gene-regulating factors, including Snail family transcriptional repressor SNAI1, SNAI2, zinc finger E-box-binding homeobox 1/2 and TWIST increase their expression, together with mesenchymal markers such as vimentin, N-cadherin and metalloproteinases, resulting in an invasive cell phenotype (35,38,68-70).

In PCa, syndecans are associated with Gleason score and EMT markers (71-73). It is hypothesized that this mesenchymal and invasive phenotype is responsible for the metastatic process (68,74). However, there is no direct evidence that these mesenchymal cells (MCs) also have colonizing abilities. Conversely, increasing evidence suggests that a small population of malignant cells present in most types of tumor, CSCs, may be responsible for the final step in recurrence and metastasis (75-77). Our previous study identified and characterized a CSC population in PCa samples and determined their molecular stem signature (CD133⁺/CD44⁺/ABCG2⁺/CD24⁻) (78). In addition, proliferative, migratory, invasive and clonogenic abilities have been evaluated in this cell population (79). It is possible to separate this CSC population from mesenchymal-like cells by changing culture conditions, followed by magnetic-associated cells sorting (MACS) (78). In adherent conditions, most cells remain in a mesenchymal differentiate state, which has been determined from using specific markers and functional assays. However, in non-adherent conditions, most mesenchymal adherent cells die by anoikis (anchorage-dependent apoptosis), whereas a few cells survive, and rapidly form spheres that grow and remain for several weeks (78). Following MACS, separated sphere-forming cells represent an enriched CSC population (78). These CSCs exhibit a low proliferation rate, increased resistance to apoptosis and drug treatments, reduced invasive properties and a high clonogenic capacity compared with that in mesenchymal-adherent cells (79). In addition, these PCa CSCs have no expression of GnRH-R or AR, nor of numerous differentiation markers (79). Preliminary experiments within our laboratory using CSCs with stable expression of AR and GnRH-R via lentiviral transduction suggest that these cells can become sensitive to androgens and GnRH analogs (unpublished data) (Fig. 2).

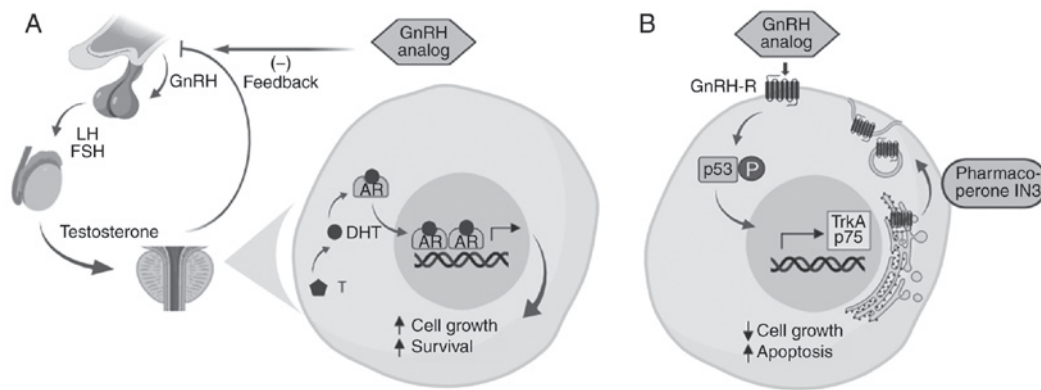


Figure 1. Comparison of systemic and local effects of GnRH analogs. (A) Systemic delivery of GnRH analogs blocks the hypothalamus-hypophysis-testicular axis, producing pharmacological castration that inhibits the AR-induced cell growth and survival of PCa cells. (B) Locally, in PCa cells, GnRH analogs activate GnRH-R, inducing phosphorylation of p53, and resulting in increased expression of TrkA and p75, which inhibits cell growth and stimulates the apoptosis of PCa cells. These effects can be potentiated with pharmacoperone IN3 by increasing GnRH-R availability in the cell membrane. AR, androgen receptor; DHT, dihydrotestosterone; FSH, follicle-stimulating hormone; GnRH, gonadotropin-releasing hormone; GnRH-R, GnRH receptor; LH, luteinizing hormone; PCa, prostate cancer; TrkA, tropomyosin receptor kinase A.

4. Tumor cell phenotypic heterogeneity and metastatic processes

It is becoming apparent that tumors present a significant degree of cell heterogeneity (80,81). Tumor heterogeneity may be understood at the phenotypic and genetic level (82). Tumor cell phenotypic heterogeneity will specifically be discussed. Cellular and molecular mechanisms responsible for this heterogeneous cell population remain poorly understood. There remains controversy regarding the origin of CSCs, and several hypotheses have been suggested (76,83-85). However, regardless of the origin of CSCs, the relevant point, particularly for clinical application, is that such a population is present in the majority of cancers studied. The multifocal origin of cancer cells within the organ and the distinct differentiation fate during EMT process may explain, in part, this phenomenon (75,86). As with the process of microevolution, cancer cells adapt to different microenvironments, first within the tumor niche and subsequently in potential metastatic niches (87,88). Within the tumor, it is possible to find a hypoxic microenvironment, for instance in the center of a solid tumor, whereas in the periphery, where neoangiogenesis is occurring, a more oxygenated milieu is more prevalent (89,90); cancer cells adapt differently to these distinct microenvironments. Therefore, it is possible that during EMT progression, certain cells express a stem gene program, forming a stable CSC population within a tumor (91-93).

Metastasis is an inefficient process; it is estimated that <2% of total cancer cells entering the blood stream from a solid tumor will be able to colonize a premetastatic niche (94). Furthermore, <0.02% will be able to survive in that niche and support sustained growth to give rise to clinically evident metastatic foci (94). Evidence suggests that this is not a stochastic process, indicating that not all malignant cells are able to sustain metastasis (94). Furthermore, very few cells have the ability to colonize, survive and grow in a tissue or organ different to the one from which it originated (95). The majority of researchers investigating CSCs have concluded that these metastatic cells express stemness genes and exhibit little invasive capacity (96). Previous results from our laboratory in CSCs from PCa are

consistent with this hypothesis (78). Instead, PCa CSCs, as with other CSCs, have a low proliferation rate, high resistance to drugs and apoptosis (particularly anoikis), sphere-growing ability and a high clonogenic capacity (97-99). Determining how these CSCs, with little invasive activity, can leave the tumor and colonize premetastatic niches will be subsequently addressed.

Metastasis is a complex process. Premetastatic niches are developed in advance by several signals originating from the initiating tumor determining the tissue tropism of the future metastatic foci (100,101). It is proposed that, once in the blood stream, CSCs are guided by homing signals from these premetastatic niches (102,103). Once colonizing a metastatic site has begun, CSCs can be induced by niche milieu factors to survive and proliferate, or to become quiescent (104,105). In the event of quiescence, future microenvironmental changes can subsequently induce cell proliferation and tumor growth, resulting in relapse, even if curative surgery was performed to remove the primary tumor (104). In human PCa, bone is one of the main sites of distant metastasis (106). Stromal-cell-derived-factor 1, acting through C-X-C chemokine receptor 4 on malignant cells, is hypothesized to promote cell survival in the niche (106). Secretion of several interleukins, tumor necrosis factor- α and other factors by cancer cells stimulates secretion of the receptor activator of NF- κ B ligand (RANKL), which in turn stimulates osteoclast differentiation (107). Increased osteoclast activity releases bone matrix and growth factors that promote CSC survival and growth for metastatic progression (106-108). Exosomes secreted by CSCs and bulk cancer cell cultures derived from PCa contain various microRNAs (miRNAs/miRs). Comparing those miRNAs using next-generation sequencing followed by bioinformatics analysis, specific miRNAs, such as miR-100-5p, miR-21-5p and miR-139-5p were found to be overexpressed and, analyzed in an *in vitro* system, they increased the expression of metalloproteinases-2, -9 and -13, and RANKL, as well as fibroblast migration, supporting the idea that the different PCa cells contribute cooperatively to prepare the premetastatic niche (100).

Considering that CSCs appear to be the only cells within a tumor with the ability to form metastasis, it is reasonable to propose that any increase in circulating CSCs will raise the risk

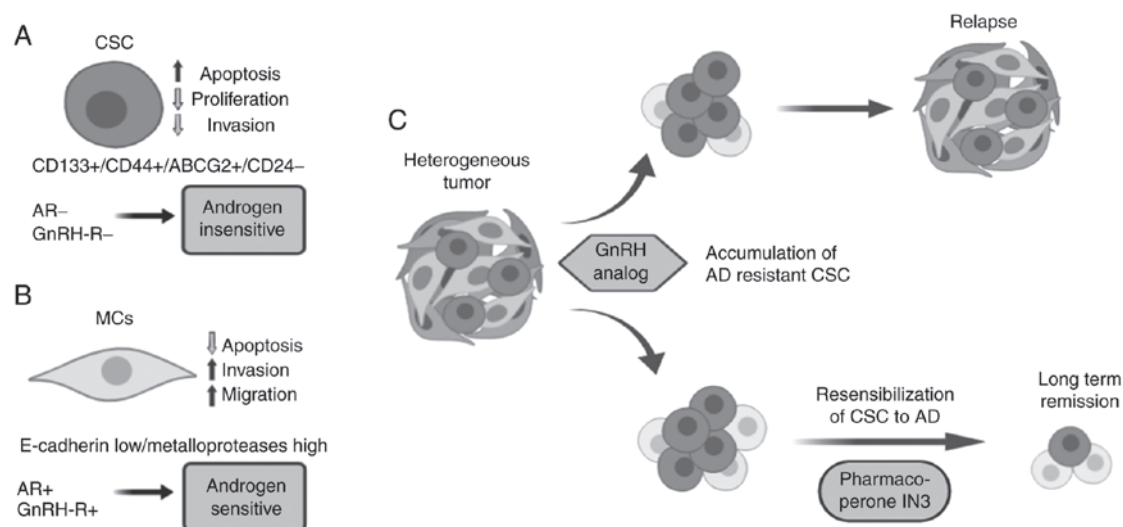


Figure 2. Resensibilization of CSCs to AD therapy could achieve long-term remission of metastatic CRPC. (A) CSCs are characterized by high resistance to apoptosis, low proliferation rates and low invasive capacities; as they do not express AR and GnRH-R, they are not responsive to AD therapies. (B) MCs have less resistance to apoptosis, and high invasive and migratory capacities; as they express AR and GnRH-R, they are responsive to AD therapies. (C) In a PCa tumor, characterized by heterogeneous subpopulations, MCs represent the bulk of the tumor. CSCs and MCs cooperate to form metastases. MCs migrate and invade within the tumor stroma, allowing CSCs to escape to the circulation and grow in the metastatic niche, where they are able to grow and generate the full heterogeneity of the original tumor. After AD therapy with GnRH analogs, androgen-responsive MCs are in remission, but androgen-insensitive CSCs accumulate and the tumor returns. Resensibilization of CSCs to GnRH analogs using pharmacoperones or lentiviral transduction could lead to long-term remission of metastatic CRPC. AD, androgen deprivation; AR, androgen receptor; CRPC, castration-resistant PCa; CSC, cancer stem cell; MC, mesenchymal cell; GnRH, gonadotropin-releasing hormone; GnRH-R, GnRH receptor; PCa, prostate cancer.

of metastasis or recurrence (88,109-111). Our previous study investigated the expression of stem signatures in PCa samples of different histological grades, using a tissue microarray and quantitative immunohistochemistry (78). It was observed that the number of cells expressing stem markers increases with Gleason grade, reaching maximal levels at medium Gleason, and decreasing thereafter in high-Gleason grade, lymph node and bone metastatic samples (78). Considering that malignant cells begin to enter the blood stream shortly after the tumor becomes locally invasive (low-to-medium histological grade), it is possible that a patient with a localized tumor with a medium Gleason score will contain the maximal number of CSCs potentially leaving the tumor and spreading throughout blood stream. At this stage, the indicated therapy is surgical removal of prostate gland (112). However, if CSCs already released from the tumor have seeded the metastatic niches, recurrence risk would be high. This is an important point to consider, particularly in patients with localized tumors of low Gleason grade where the therapeutic recommendation is active surveillance (5,113). Therefore, identifying and quantifying CSCs in PCa biopsies may be a valuable prognostic factor for relapse.

5. Different malignant cell types in a solid tumor may collaborate to produce distant metastasis

Reanalyzing the problem of how CSCs with little invasive activity can leave the tumor and colonize premetastatic niches, it is reasonable to suggest that some type of collaboration with highly invasive mesenchymal-like cells occurs (45,96). Previously, Celià-Terrassa *et al* (114) provided evidence regarding this potential cooperative action. Using commercial cell lines derived from PCa (PC3) and bladder cancer (TSU-Pr1), these were enriched with metastatic TICs, a cell population with a strong

epithelial profile. In turn, they deprived TICs, a cell population with a mesenchymal profile. Overexpression of mesenchymal genes in the former cell population (epithelial phenotype) decreased its TIC ability, whereas knockdown of these genes in the latter cell population (mesenchymal phenotype) enhanced its TIC capacity (114). Using immunocompromised nonobese diabetic/severe combined immunodeficiency (NOD/SCID) mice, it was observed that, injected in combination, mesenchymal-like cells increased the metastatic potential of epithelial TIC-enriched cell populations, suggesting a cooperative action between both cell types (114). Subsequently, the same research group described that secreted protein acidic and rich in cysteine (SPARC) mediates the metastatic cooperation between CSC and non-CSC cell subpopulations (39).

Recently, it was reported that SPARC induced EMT, increasing the invasive capacities of PCa cells (115). Collectively, these findings support the hypothesis that within a tumor, MCs become the predominant population via EMT, increasing the invasive capacity of the tumor. However, it has been proposed that a small cell population that expresses a stem-like program (CSCs) remains in the tumor and can escape passively with the bulk of MCs. Once in the metastatic niche, it is hypothesized that CSCs proliferate and produce progenitor cells that may further differentiate to an epithelial-like phenotype. This may explain certain findings revealing that in metastatic PCa samples, an increase in epithelial markers and a decrease in mesenchymal markers is observed, which has been called mesenchymal-epithelial transition (116,117). It is postulated that the metastatic foci will generate the full heterogeneity of the original tumor, in which epithelial-like cells will undergo EMT again, whilst a small number of CSCs are retained in the tumor. On the other hand, tumor cell plasticity influences the phenotypic heterogeneity of tumor cells,

with the varied cell abilities enabling cooperation to promote cancer progression and metastasis. Differential cell distribution within the tumor, and spatial and temporal patterns during EMT-stemness processes may influence cell frequencies and the results of the proposed cell cooperation (118). This may contribute to why different patients with PCa at the same stage may have different outcomes.

Personalized medicine should take into consideration this evidence to develop novel and innovative therapeutic strategies. In this context, resensibilization of PCa cells (including CSCs) to GnRH analogs using pharmacoperones or lentiviral transduction may provide an effective treatment against metastatic castration-resistant PCa. It is necessary to validate this hypothesis using CSCs and MCs derived from the tumors of various patients. Metastasis is, by definition, a process that occurs in a living organism. Therefore, there are no *in vitro* models for investigating this complex pathological process. In previous years, several *in vivo* models have been developed (119-124). The majority of these use immunocompromised mice, and several mouse strains have been obtained, a number of them via transgenic manipulation (124-126). One of the most used models, at present, is the NOD/SCID mouse (127).

6. Orthotopic model for the study of human PCa metastasis

The NOD/SCID mouse has been widely used to investigate the metastasis of several types of human cancer (128). A critical issue is the type of injection used to introduce human cancer cells. Numerous researchers use subcutaneous, intravenous or intracardiac administration, with varying results (114,129). Additionally, orthotopic models have been developed (injection in the same mouse organ or tissue from which human cells were derived). This model mimics the metastatic process more precisely (129). Reports of orthotopic models for human PCa have been published (130-132). A modification of the orthotopic model for PCa using a cell injection in one of the anterior lobes of the NOD/SCID mouse prostate has been developed by our laboratory (133,134). This orthotopic injection results in consistent and reproducible metastatic progression. First, a fraction of tumor cells injected in the mouse prostate survives and generates a tumor derived from surviving injected cells (transduced with luciferase and red fluorescent protein genes). The fluorescence allows the tracking of metastatic progression *in vivo* using *in vivo* imaging equipment. In a chronological sequence, metastatic foci begin to appear in the liver, lungs and the kidneys. Injection of cells into the anterior lobe, instead of the ventral prostate, has the advantage that it is possible to surgically remove the prostate tumor to evaluate the progression of metastasis, with or without the primary prostate tumor. In this orthotopic model, the utility of prostatectomy during metastasis progression has been demonstrated (134), as has the effect of knocking down the stemness gene Sox2 on metastasis (unpublished data). In current studies, progression towards a castration-resistant PCa mouse model using surgical castration as an AD strategy is being established.

7. Conclusions

In conclusion, it is proposed that there is cooperation between CSCs and MCs during metastatic progression. Further

development of preclinical orthotopic models of PCa may provide additional evidence supporting this hypothesis. In addition, the role of stem genes, as well as AR, GnRH-R and differentiation genes, in metastasis progression and hormone resistance may have critical relevance. Further investigation of these aspects will contribute to the understanding of the cellular and molecular mechanisms of metastasis, recurrence and hormone resistance in PCa, which remain major challenges for the treatment of this disease. It is predicted that evidence obtained using preclinical models, will be beneficial for clinical purposes in the near future, identifying novel prognostic factors and therapeutic targets.

Acknowledgements

The authors thank the professional contribution of Mrs Graciela Caroca (Laboratory of Cellular and Molecular Oncology, Department of Basic and Clinical Oncology, Faculty of Medicine, University of Chile) for assistance in the laboratory.

Funding

The present study was funded by Fondecyt (grant nos. 1140417 and 1151214), ENLACE-VID (grant nos. ENL-22/19 and ENL-23/19) and URedes URC (grant no. 007/17).

Availability of data and materials

All data generated or analyzed during this study are included in this published article.

Authors' contributions

HRC and FLM contributed to reviewing and discussing the literature, and selecting relevant studies. EAC analyzed the subject and wrote the review.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

References

1. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA and Jemal A: Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 68: 394-424, 2018.
2. Rodgers L, Peer CJ and Figg WD: Diagnosis, staging and risk stratification in prostate cancer: Utilizing diagnostic tools to avoid unnecessary therapies and side effects. *Cancer Biol Ther* 18: 470-472, 2017.
3. Shah RB and Zhou M: Recent advances in prostate cancer pathology: Gleason grading and beyond. *Pathol Int* 66: 260-272, 2016.

4. Heijnsdijk EAM, Bangma CH, Borràs JM, de Carvalho TM, Castells X, Eklund M, Espinàs JA, Graefen M, Grönberg H, Lansdorp-Vogelaar I, *et al*: Summary statement on screening for prostate cancer in Europe. *Int J Cancer* 142: 741-746, 2018.
5. Litwin MS and Tan HJ: The diagnosis and treatment of prostate cancer: A review. *J Am Med Assoc* 317: 2532-2542, 2017.
6. Ost P, Bossi A, Decaestecker K, De Meerleer G, Giannarini G, Karnes RJ, Roach M III and Briganti A: Metastasis-directed therapy of regional and distant recurrences after curative treatment of prostate cancer: A systematic review of the literature. *Eur Urol* 67: 852-863, 2015.
7. Fakhrejahani F, Madan RA and Dahut WL: Management options for biochemically recurrent prostate cancer. *Curr Treat Options Oncol* 18: 26, 2017.
8. Artibani W, Porcaro AB, De Marco V, Cerruto MA and Siracusano S: Management of biochemical recurrence after primary curative treatment for prostate cancer: A review. *Urol Int* 100: 251-262, 2018.
9. Sartor O and de Bono JS: Metastatic prostate cancer. *N Engl J Med* 378: 645-657, 2018.
10. Song C, Kang T, Yoo S, Jeong IG, Ro JY, Hong JH, Kim CS and Ahn H: Tumor volume, surgical margin, and the risk of biochemical recurrence in men with organ-confined prostate cancer. *Urol Oncol* 31: 168-174, 2013.
11. Suzman DL, Boikos SA and Carducci MA: Bone-targeting agents in prostate cancer. *Cancer Metastasis Rev* 33: 619-628, 2014.
12. Dong L, Zieren RC, Xue W, de Reijke TM and Pienta KJ: Metastatic prostate cancer remains incurable, why? *Asian J Urol* 6: 26-41, 2019.
13. Pelekanou V and Castanas E: Androgen control in prostate cancer. *J Cell Biochem* 2234: 2224-2234, 2016.
14. Tan MH, Li J, Xu HE, Melcher K and Yong EL: Androgen receptor: Structure, role in prostate cancer and drug discovery. *Acta Pharmacol Sin* 36: 3-23, 2014.
15. Fujita K and Nonomura N: Role of androgen receptor in prostate cancer: A review. *World J Mens Health* 36: 288-295, 2018.
16. Rodriguez KM, Pastuszak AW and Khera M: The role of testosterone therapy in the setting of prostate cancer. *Curr Urol Rep* 19: 67, 2018.
17. Obinata D, Takayama K, Takahashi S and Inoue S: Crosstalk of the androgen receptor with transcriptional collaborators: Potential therapeutic targets for castration-resistant prostate cancer. *Cancers (Basel)* 9: pii: E22, 2017.
18. Grossmann M, Cheung AS and Zajac JD: Androgens and prostate cancer; pathogenesis and deprivation therapy. *Best Pract Res Clin Endocrinol Metab* 27: 603-616, 2013.
19. Hahn AW, Hale P, Rathi N and Agarwal N: Novel androgen axis systemic therapies for metastatic hormone-sensitive prostate cancer. *Curr Opin Urol* 27: 559-565, 2017.
20. Shore ND, Abrahamsson P, Anderson J, Crawford ED and Lange P: New considerations for ADT in advanced prostate cancer and the emerging role of GnRH antagonists. *Prostate Cancer Prostatic Dis* 16: 7-15, 2013.
21. Lama G, Papi M, Angelucci C, Maulucci G, Sica G and De Spirito M: Leuporelin acetate long-lasting effects on GnRH receptors of prostate cancer cells: An atomic force microscopy study of agonist/receptor interaction. *PLoS One* 8: e52530, 2013.
22. Thomas BC and Neal DE: Androgen deprivation treatment in prostate cancer. *BMJ* 346: 1-5, 2013.
23. Katsogiannou M, Ziouziou H, Karaki S, Andrieu C, Henry de Villeneuve M and Rocchi P: The hallmarks of castration-resistant prostate cancers. *Cancer Treat Rev* 41: 588-597, 2015.
24. Yuan X, Cai C, Chen S, Chen S, Yu Z and Balk SP: Androgen receptor functions in castration-resistant prostate cancer and mechanisms of resistance to new agents targeting the androgen axis. *Oncogene* 33: 2815-2825, 2014.
25. Fujimoto N: Role of the androgen-androgen receptor axis in the treatment resistance of advanced prostate cancer: From androgen-dependent to castration resistant and further. *J UOEH* 38: 129-138, 2016.
26. Chandrasekar T, Yang JC, Gao AC and Evans CP: Mechanisms of resistance in castration-resistant prostate cancer (CRPC). *Transl Androl Urol* 4: 365-380, 2015.
27. Tilki D, Schaeffer EM and Evans CP: Understanding mechanisms of resistance in metastatic castration-resistant prostate cancer: The role of the androgen receptor. *Eur Urol Focus* 2: 499-505, 2019.
28. Huang Y, Jiang X, Liang X and Jiang G: Molecular and cellular mechanisms of castration resistant prostate cancer. *Oncol Lett* 15: 6063-6076, 2018.
29. Ho Y and Dehm SM: Androgen receptor rearrangement and splicing variants in resistance to endocrine therapies in prostate cancer. *Endocrinology* 158: 1533-1542, 2017.
30. Recouvreux MV, Wu JB, Gao AC, Zonis S, Chesnokova V, Bhowmick N, Chung LW and Melmed S: Androgen receptor regulation of local growth hormone in prostate cancer cells. *Endocrinology* 158: 2255-2568, 2017.
31. Stelloo S, Nevedomskaya E, van der Poel HG, de Jong J, van Leenders GJ, Jenster G, Wessels LF, Bergman AM and Zwart W: Androgen receptor profiling predicts prostate cancer outcome. *EMBO Mol Med* 7: 1450-1464, 2015.
32. Höti N, Shah P, Hu Y, Yang S and Zhang H: Proteomics analyses of prostate cancer cells reveal cellular pathways associated with androgen resistance. *Proteomics* 17, 2017 doi: 10.1002/pmic.201600228.
33. Van Den Eeden SK, Lu R, Zhang N, Quesenberry CP Jr, Shan J, Han JS, Tsiatis AC, Leimpeter AD, Lawrence HJ, Febbo PG and Presti JC: A Biopsy-based 17-gene genomic prostate score as a predictor of metastases and prostate cancer death in surgically treated men with clinically localized disease. *Eur Urol* 73: 129-138, 2017.
34. Hanahan D and Weinberg RA: Hallmarks of cancer: The next generation. *Cell* 144: 646-674, 2011.
35. Nieto MA, Huang RY, Jackson RA and Thiery JP: EMT: 2016. *Cell* 166: 21-45, 2016.
36. Hasegawa S, Nagano H, Konno M, Eguchi H, Tomokuni A, Tomimaru Y, Asaoka T, Wada H, Hama N, Kawamoto K, *et al*: A crucial epithelial to mesenchymal transition regulator, Sox4/Ezh2 axis is closely related to the clinical outcome in pancreatic cancer patients. *Int J Oncol* 48: 145-152, 2016.
37. Frisch SM, Schaller M and Cieply B: Mechanisms that link the oncogenic epithelial-mesenchymal transition to suppression of anoikis. *J Cell Sci* 126: 21-29, 2013.
38. Serrano-Gomez SJ, Maziveyi M and Alahari SK: Regulation of epithelial-mesenchymal transition through epigenetic and post-translational modifications. *Mol Cancer* 15: 18, 2016.
39. Mateo F, Meca-Cortés O, Celià-Terrassa T, Fernández Y, Abasolo I, Sánchez-Cid L, Bermudo R, Sagasta A, Rodríguez-Carunchio L, Pons M, *et al*: SPARC mediates metastatic cooperation between CSC and non-CSC prostate cancer cell subpopulations. *Mol Cancer* 13: 237, 2014.
40. Lin KC, Torga G, Sun Y, Axelrod R, Pienta KJ, Sturm JC and Austin RH: The role of heterogeneous environment and docetaxel gradient in the emergence of polyploid, mesenchymal and resistant prostate cancer cells. *Clin Exp Metastasis* 36: 97-108, 2019.
41. Bakker B, Taudt A, Belderbos ME, Porubsky D, Spierings DC, de Jong TV, Halsema N, Kazemier HG, Hoekstra-Wakker K, Bradley A, *et al*: Single-cell sequencing reveals karyotype heterogeneity in murine and human malignancies. *Genome Biol* 17: 115, 2016.
42. Chapman MP, Risom T, Aswani AJ, Langer EM, Sears RC and Tomlin CJ: Modeling differentiation-state transitions linked to therapeutic escape in triple-negative breast cancer. *PLoS Comput Biol* 15: e1006840, 2019.
43. Eun K, Ham SW and Kim H: Cancer stem cell heterogeneity: Origin and new perspectives on CSC targeting. *BMB Rep* 50: 117-125, 2017.
44. Adamowicz J, Pakravan K, Bakhshinejad B, Drewa T and Babashah S: Prostate cancer stem cells: From theory to practice. *Scand J Urol* 51: 95-106, 2017.
45. Shahriari K, Shen F, Worrede-Mahdi A, Liu Q, Gong Y, Garcia FU and Fatatis A: Cooperation among heterogeneous prostate cancer cells in the bone metastatic niche. *Oncogene* 36: 2846-2856, 2017.
46. Chang L, Graham P, Hao J, Ni J, Deng J, Bucci J, Malouf D, Gillatt D and Li Y: Cancer stem cells and signaling pathways in radioresistance. *Oncotarget* 7: 11002-11017, 2016.
47. Geng SQ, Alexandrou AT and Li JJ: Breast cancer stem cells: Multiple capacities in tumor metastasis. *Cancer Lett* 349: 1-7, 2014.
48. Leão R, Domingos C, Figueiredo A, Hamilton R, Tabori U and Castelo-Branco P: Cancer stem cells in prostate cancer: Implications for targeted therapy. *Urol Int* 99: 125-136, 2017.
49. Rosario DJ, Davey P, Green J, Greene D, Turner B, Payne H and Kirby M: The role of gonadotrophin-releasing hormone antagonists in the treatment of patients with advanced hormone-dependent prostate cancer in the UK. *World J Urol* 34: 1601-1609, 2016.

50. Poelaert F, Kumps C, Lumen N, Verschuere S, Libbrecht L, Praet M, Rottey S, Claeys T, Ost P, Decaestecker K, *et al*: Androgen receptor gene copy number and protein expression in treatment-naïve prostate cancer. *Urol Int* 99: 222-228, 2017.
51. Prekovic S, van Royen ME, Voet AR, Geverts B, Houtman R, Melchers D, Zhang KY, Van den Broeck T, Smeets E, Spans L, *et al*: The effect of F877L and T878A mutations on androgen receptor response to enzalutamide. *Mol Cancer Ther* 15: 1702-1712, 2016.
52. Sutinen P, Malinen M, Heikkinen S and Palvimo JJ: SUMOylation modulates the transcriptional activity of androgen receptor in a target gene and pathway selective manner. *Nucleic Acids Res* 42: 8310-8319, 2014.
53. Rowlands MA, Holly JM, Hamdy F, Phillips J, Goodwin L, Marsden G, Gunnell D, Donovan J, Neal DE and Martin RM: Serum insulin-like growth factors and mortality in localised and advanced clinically detected prostate cancer. *Cancer Causes Control* 23: 347-354, 2012.
54. Lescarbeau RM, Seib FP, Prewitz M, Werner C and Kaplan DL: In vitro model of metastasis to bone marrow mediates prostate cancer castration resistant growth through paracrine and extracellular matrix factors. *PLoS One* 7: e40372, 2012.
55. Penning TM and Tamae D: Current advances in intratumoral androgen metabolism in castration-resistant prostate cancer. *Curr Opin Endocrinol Diabetes Obes* 23: 264-270, 2016.
56. Price DK, Chau CH, Till C, Goodman PJ, Leach RJ, Johnson-Pais TL, Hsing AW, Hoque A, Parnes HL, Schenk JM, *et al*: Association of androgen metabolism gene polymorphisms with prostate cancer risk and androgen concentrations: Results from the prostate cancer prevention trial. *Cancer* 122: 2332-2340, 2016.
57. Clementi M, Sánchez C, Benítez DA, Contreras HR, Huidobro C, Cabezas J, Acevedo C and Castellón EA: Gonadotropin releasing hormone analogs induce apoptosis by extrinsic pathway involving p53 phosphorylation in primary cell cultures of human prostatic adenocarcinomas. *Prostate* 69: 1025-1033, 2009.
58. Sánchez C, Clementi M, Benítez D, Contreras H, Huidobro C and Castellón E: Effect of GnRH analogs on the expression of TrkA and p75 neurotrophin receptors in primary cell cultures from human prostate adenocarcinoma. *Prostate* 65: 195-202, 2005.
59. Angelucci C, Lama G, Iacopino F, Ferracuti S, Bono AV, Millar RP and Sica G: GnRH receptor expression in human prostate cancer cells is affected by hormones and growth factors. *Endocrine* 36: 87-97, 2009.
60. Sánchez CA, Mercado AJ, Contreras HR, Cabezas JC, Huidobro CC and Castellón EA: Pharmacoperone IN3 enhances the apoptotic effect of leuprolide in prostate cancer cells by increasing the gonadotropin-releasing hormone receptor in the cell membrane. *Anticancer Drugs* 23: 959-969, 2012.
61. Castellón E, Clementi M, Hitschfeld C, Sánchez C, Benítez D, Sáenz L, Contreras H and Huidobro C: Effect of leuprolide and cetorelix on cell growth, apoptosis, and GnRH receptor expression in primary cell cultures from human prostate carcinoma. *Cancer Invest* 24: 261-268, 2006.
62. Saltzstein D, Shore ND, Moul JW, Chu F, Concepcion R, de la Motte S, McLane JA, Atkinson S, Yang A and Crawford ED: Pharmacokinetic and pharmacodynamic comparison of subcutaneous versus intramuscular leuprolide acetate formulations in male subjects. *Ther Adv Urol* 10: 43-50, 2017.
63. Nieto MA and Cano A: The epithelial-mesenchymal transition under control: Global programs to regulate epithelial plasticity. *Semin Cancer Biol* 22: 361-368, 2012.
64. García de Herreros A and Baulida J: Cooperation, amplification, and feed-back in epithelial-mesenchymal transition. *Biochim Biophys Acta* 1825: 223-228, 2012.
65. Savagner P: The epithelial-mesenchymal transition (EMT) phenomenon. *Ann Oncol* 21 (Suppl 7): vii89-vii92, 2010.
66. Chen T, You Y, Jiang H and Wang ZZ: Epithelial-mesenchymal transition (EMT): A biological process in the development, stem cell differentiation, and tumorigenesis. *J Cell Physiol* 232: 3261-3272, 2017.
67. Micalizzi DS, Farabaugh SM and Ford HL: Epithelial-mesenchymal transition in cancer: Parallels between normal development and tumor progression. *J Mammary Gland Biol Neoplasia* 15: 117-134, 2010.
68. De Craene B and Berx G: Regulatory networks defining EMT during cancer initiation and progression. *Nat Rev Cancer* 13: 97-110, 2013.
69. Osorio LA, Farfán NM, Castellón EA and Contreras HR: SNAIL transcription factor increases the motility and invasive capacity of prostate cancer cells. *Mol Med Rep* 13: 778-786, 2016.
70. Orellana-Serradell O, Herrera D, Castellón EA and Contreras HR: The transcription factor ZEB1 promotes an aggressive phenotype in prostate cancer cell lines. *Asian J Androl* 20: 294-299, 2018.
71. Contreras HR, Ledezma RA, Vergara J, Cifuentes F, Barra C, Cabello P, Gallegos I, Morales B, Huidobro C and Castellón EA: The expression of syndecan-1 and -2 is associated with Gleason score and epithelial-mesenchymal transition markers, E-cadherin and beta-catenin, in prostate cancer. *Urol Oncol* 28: 534-540, 2010.
72. Poblete CE, Fulla J, Gallardo M, Muñoz V, Castellón EA, Gallegos I and Contreras HR: Increased SNAIL expression and low syndecan levels are associated with high Gleason grade in prostate cancer. *Int J Oncol* 44: 647-654, 2014.
73. Farfán N, Ocaez N, Castellón EA, Mejía N, de Herreros AG and Contreras HR: The transcriptional factor ZEB1 represses Syndecan 1 expression in prostate cancer. *Sci Rep* 8: 11467, 2018.
74. Montanari M, Rossetti S, Cavaliere C, D'Aniello C, Malzone MG, Vanacore D, Di Franco R, La Mantia E, Iovane G, Piscitelli R, *et al*: Epithelial-mesenchymal transition in prostate cancer: An overview. *Oncotarget* 8: 35376-35389, 2017.
75. Mitra A, Mishra L and Li S: EMT, CTCs and CSCs in tumor relapse and drug-resistance. *Oncotarget* 6: 10699-10710, 2015.
76. Peitzsch C, Tyutyunnykova A, Pantel K and Dubrovskaya A: Cancer stem cells: The root of tumor recurrence and metastases. *Semin Cancer Biol* 44: 10-24, 2017.
77. Ajani JA, Song S, Hochster HS and Steinberg IB: Cancer stem cells: The promise and the potential. *Semin Oncol* 42 (Suppl 1): S3-S17, 2015.
78. Castellón EA, Valenzuela R, Lillo J, Castillo V, Contreras HR, Gallegos I, Mercado A and Huidobro C: Molecular signature of cancer stem cells isolated from prostate carcinoma and expression of stem markers in different Gleason grades and metastasis. *Biol Res* 45: 297-305, 2012.
79. Castillo V, Valenzuela R, Huidobro C, Contreras HR and Castellón EA: Functional characteristics of cancer stem cells and their role in drug resistance of prostate cancer. *Int J Oncol* 45: 985-994, 2014.
80. McGranahan N and Swanton C: Clonal heterogeneity and tumor evolution: Past, present, and the future. *Cell* 168: 613-628, 2017.
81. Bosman FT: Tumor heterogeneity: Will it change what pathologists do. *Pathobiology* 85: 18-22, 2018.
82. Jolly MK and Celià-Terrassa T: Dynamics of phenotypic heterogeneity during EMT and stemness in cancer progression. *J Clin Med* 8: pii: E1542, 2019.
83. Bu Y and Cao D: The origin of cancer stem cells. *Front Biosci (Schol Ed)* 4: 819-830, 2012.
84. Parsons BL: Multiclonal tumor origin: Evidence and implications. *Mutat Res* 777: 1-18, 2018.
85. Vicente-Dueñas C, Hauer J, Cobaleda C, Borkhardt A and Sánchez-García I: Epigenetic priming in cancer initiation. *Trends Cancer* 4: 408-417, 2018.
86. Ye X and Weinberg RA: Epithelial-mesenchymal plasticity: A central regulator of cancer progression. *Trends Cell Biol* 25: 675-686, 2015.
87. Graham TA and Sottoriva A: Measuring cancer evolution from the genome. *J Pathol* 241: 183-191, 2017.
88. Francart M, Lambert J, Vanwynsberghe AM, Thompson EW, Bourcy M, Polette M and Gilles C: Epithelial-mesenchymal plasticity and circulating tumor cells: Travel companions to metastases. *Dev Dyn* 247: 432-450, 2018.
89. Carnero A and Leonart M: The hypoxic microenvironment: A determinant of cancer stem cell evolution. *Bioessays* 38 (Suppl 1): S65-S74, 2016.
90. Yeo CD, Kang N, Choi SY, Kim BN, Park CK, Kim JW, Kim YK and Kim SJ: The role of hypoxia on the acquisition of epithelial-mesenchymal transition and cancer stemness: A possible link to epigenetic regulation. *Korean J Intern Med* 32: 589-599, 2017.
91. Rhim AD: Epithelial to mesenchymal transition and the generation of stem-like cells in pancreatic cancer. *Pancreatol* 13: 114-117, 2013.
92. Lan L, Luo Y, Cui D, Shi BY, Deng W, Huo LL, Chen HL, Zhang GY and Deng LL: Epithelial-mesenchymal transition triggers cancer stem cell generation in human thyroid cancer cells. *Int J Oncol* 43: 113-120, 2013.

93. Li N, Babaei-Jadidi R, Lorenzi F, Spencer-Dene B, Clarke P, Domingo E, Tulchinsky E, Vries RGJ, Kerr D, Pan Y, *et al*: An FBXW7-ZEB2 axis links EMT and tumour microenvironment to promote colorectal cancer stem cells and chemoresistance. *Oncogenesis* 8: 13, 2019.
94. Croker AK and Allan AL: Cancer stem cells: Implications for the progression and treatment of metastatic disease. *J Cell Mol Med* 12: 374-390, 2008.
95. Massagué J and Obenauf AC: Metastatic colonization by circulating tumour cells. *Nature* 529: 298-306, 2016.
96. Hayashida T, Jinno H, Kitagawa Y and Kitajima M: Cooperation of cancer stem cell properties and epithelial-mesenchymal transition in the establishment of breast cancer metastasis. *J Oncol* 2011: 591427, 2011.
97. Hsu CL, Chung FH, Chen CH, Hsu TT, Liu SM, Chung DS, Hsu YF, Chen CL, Ma N and Lee HC: Genotypes of cancer stem cells characterized by epithelial-to-mesenchymal transition and proliferation related functions. *Sci Rep* 6: 32523, 2016.
98. Yun EJ, Lo UG and Hsieh JT: The evolving landscape of prostate cancer stem cell: Therapeutic implications and future challenges. *Asian J Urol* 3: 203-210, 2016.
99. Lin CJ, Lo UG and Hsieh JT: The regulatory pathways leading to stem-like cells underlie prostate cancer progression. *Asian J Androl* 21: 233-240, 2019.
100. Sánchez CA, Andahur EI, Valenzuela R, Castellón EA, Fullá JA, Ramos CG and Triviño JC: Exosomes from bulk and stem cells from human prostate cancer have a differential microRNA content that contributes cooperatively over local and pre-metastatic niche. *Oncotarget* 7: 3993-4008, 2016.
101. Hoshino A, Costa-Silva B, Shen TL, Rodrigues G, Hashimoto A, Tesic Mark M, Molina H, Kohsaka S, Di Giannatale A, Ceder S, *et al*: Tumour exosome integrins determine organotropic metastasis. *Nature* 527: 329-335, 2015.
102. Langley RR and Fidler IJ: The seed and soil hypothesis revisited-The role of tumor-stroma interactions in metastasis to different organs. *Int J Cancer* 128: 2527-2535, 2011.
103. Miftakhova R, Hedblom A, Semenas J, Robinson B, Simoulis A, Malm J, Rizvanov A, Heery DM, Mongan NP, Maitland NJ, *et al*: Cyclin A1 and P450 aromatase promote metastatic homing and growth of stem-like prostate cancer cells in the bone marrow. *Cancer Res* 76: 2453-2464, 2016.
104. Shiozawa Y, Berry JE, Eber MR, Jung Y, Yumoto K, Cackowski FC, Yoon HJ, Parsana P, Mehra R, Wang J, *et al*: The marrow niche controls the cancer stem cell phenotype of disseminated prostate cancer. *Oncotarget* 7: 41217-41232, 2016.
105. Sharma S, Xing F, Liu Y, Wu K, Said N, Pochampally R, Shiozawa Y, Lin HK, Balaji KC and Watabe K: Secreted protein acidic and rich in cysteine (SPARC) mediates metastatic dormancy of prostate cancer in the bone. *J Biol Chem* 291: 19351-19363, 2016.
106. Jin J, Dayyani F and Gallick G: Steps in prostate cancer progression that lead to bone metastasis. *Int J Cancer* 128: 2545-2561, 2011.
107. Peyruchaud O, Leblanc R and David M: Pleiotropic activity of lysophosphatidic acid in bone metastasis. *Biochim Biophys Acta* 1831: 99-104, 2013.
108. Roodman GD: Genes associate with abnormal bone cell activity in bone metastasis. *Cancer Metastasis Rev* 31: 569-578, 2012.
109. Zhang T and Armstrong AJ: Clinical utility of circulating tumor cells in advanced prostate cancer. *Curr Oncol Rep* 18: 3, 2016.
110. Barriere G, Fici P, Gallerani G, Fabbri F, Zoli W and Rigaud M: Circulating tumor cells and epithelial, mesenchymal and stemness markers: Characterization of cell subpopulations. *Ann Transl Med* 2: 109, 2014.
111. Vogelzang NJ, Fizazi K, Burke JM, De Wit R, Bellmunt J, Hutson TE, Crane E, Berry WR, Doner K, Hainsworth JD, *et al*: Circulating tumor cells in a phase 3 study of docetaxel and prednisone with or without lenalidomide in metastatic Castration-resistant prostate cancer. *Eur Urol* 71: 168-171, 2017.
112. Srivatsa N, Nagaraja H, Shweta S and Raghunath S: Radical prostatectomy for locally advanced prostate cancers-review of literature. *Indian J Surg Oncol* 8: 175-180, 2017.
113. Wilt T, Brawe M, Jones K, Barry MJ, Aronson WJ, Fox S, Gingrich JR, Wei JT, Gilhooly P, Grob BM, *et al*: Radical prostatectomy versus observation for localized prostate cancer. *N Engl J Med* 367: 203-213, 2012.
114. Celià-Terrassa T, Meca-Cortés Ó, Mateo F, Martínez de Paz A, Rubio N, Arnal-Estapé A, Ell BJ, Bermudo R, Díaz A, Guerra-Rebollo M, *et al*: Epithelial-mesenchymal transition can suppress major attributes of human epithelial tumor-initiating cells. *J Clin Invest* 122: 1846-1868, 2012.
115. López-Moncada F, Torres MJ, Castellón EA and Contreras HR: Secreted protein acidic and rich in cysteine (SPARC) induces epithelial-mesenchymal transition, enhancing migration and invasion, and is associated with high Gleason score in prostate cancer. *Asian J Androl* 21: 557-564, 2019.
116. Gunasinghe NP, Wells A, Thompson EW and Hugo HJ: Mesenchymal-epithelial transition (MET) as a mechanism for metastatic colonisation in breast cancer. *Cancer Metastasis Rev* 31: 469-478, 2012.
117. Bullock MD, Sayan AE, Packham GK and Mirnezami AH: MicroRNAs: Critical regulators of epithelial to mesenchymal (EMT) and mesenchymal to epithelial transition (MET) in cancer progression. *Biol Cell* 104: 3-12, 2012.
118. Bocci F, Gearhart-Serna L, Boareto M, Ribeiro M, Ben-Jacob E, Devi GR, Levine H, Onuchic JN and Jolly MK: Toward understanding cancer stem cell heterogeneity in the tumor microenvironment. *Proc Natl Acad Sci USA* 116: 148-157, 2019.
119. Harris JE, Shin J, Lee B, Pelosky K, Hooker CM, Harbom K, Hulbert A, Zahnow C, Yang SC, Baylin S, *et al*: A murine xenograft model of spontaneous metastases of human lung adenocarcinoma. *J Surg Res* 171: e75-e79, 2011.
120. Rea D, Del Vecchio V, Palma G, Barbieri A, Falco M, Luciano A, De Biase D, Perdonà S, Facchini G and Arra C: Mouse models in prostate cancer translational research: From Xenograft to PDX. *Biomed Res Int* 2016: 11, 2016.
121. Daphu I, Sundström T, Horn S, Huszthy PC, Niclou SP, Sakariassen PØ, Immervoll H, Miletic H, Bjerkvig R and Thorsen F: In vivo animal models for studying brain metastasis: Value and limitations. *Clin Exp Metastasis* 30: 695-710, 2013.
122. Romano G, Chagani S and Kwong LN: The path to metastatic mouse models of colorectal cancer. *Oncogene* 37: 2481-2489, 2018.
123. Kahn J, Tofilon PJ and Camphausen K: Preclinical models in radiation oncology. *Radiat Oncol* 7: 223, 2012.
124. Loi M, Di Paolo D, Becherini P, Zorzoli A, Perri P, Carosio R, Cilli M, Ribatti D, Brignole C, Pagnan G, *et al*: The use of orthotopic models to validate antivasculature therapies for cancer. *Int J Dev Biol* 55: 547-555, 2011.
125. Grabowska MM, Degraff DJ, Yu X, Jin RJ, Chen Z, Borowsky AD and Matusik RJ: Mouse models of prostate cancer: Picking the best model for the question. *Cancer Metastasis Rev* 33: 377-397, 2014.
126. Usary J, Zhao W, Darr D, Roberts PJ, Liu M, Balletta L, Karginova O, Jordan J, Combet A, Bridges A, *et al*: Predicting drug responsiveness in human cancers using genetically engineered mice. *Clin Cancer Res* 19: 4889-4899, 2013.
127. Bastide C, Bagnis C, Mannoni P, Hassoun J and Bladou F: A Nod Scid mouse model to study human prostate cancer. *Prostate Cancer Prostatic Dis* 5: 311-315, 2002.
128. Hidalgo M, Amant F, Biankin AV, Budinská E, Byrne AT, Caldas C, Clarke RB, de Jong S, Jonkers J, Mølandsmo GM, *et al*: Patient derived xenograft models: An emerging platform for translational cancer research. *Cancer Discov* 4: 998-1013, 2014.
129. Dai J, Hensel J, Wang N, Kruithof-de Julio M and Shiozawa Y: Mouse models for studying prostate cancer bone metastasis. *Bonekey Rep* 5: 777, 2016.
130. Tumati V, Mathur S, Song K, Hsieh JT, Zhao D, Takahashi M, Dobin T, Gandee L, Solberg TD, Habib AA and Saha D: Development of a locally advanced orthotopic prostate tumor model in rats for assessment of combined modality therapy. *Int J Oncol* 42: 1613-1619, 2013.
131. Lee ST, Wong PF, He H, Hooper JD and Mustafa MR: Alpha-tomatine attenuation of in vivo growth of subcutaneous and orthotopic xenograft tumors of human prostate carcinoma PC-3 cells is accompanied by inactivation of nuclear factor-kappa B signaling. *PLoS One* 8: e57708, 2013.
132. Wang Y, Xue H, Cutz JC, Bayani J, Mawji NR, Chen WG, Goetz LJ, Hayward SW, Sadar MD, Gilks CB, *et al*: An orthotopic metastatic prostate cancer model in SCID mice via grafting of a transplantable human prostate tumor line. *Lab Invest* 85: 1392-1404, 2005.
133. Cifuentes FF, Valenzuela RH, Contreras HR and Castellón EA: Development of an orthotopic model of human metastatic prostate cancer in the NOD-SCIDγ mouse (Mus musculus) anterior prostate. *Oncol Lett* 10: 2142-2148, 2015.
134. Cifuentes FF, Valenzuela RH, Contreras HR and Castellón EA: Surgical cytoreduction of the primary tumor reduces metastatic progression in a mouse model of prostate cancer. *Oncol Rep* 34: 2837-2844, 2015.