

Pan-cancer analysis of transmembrane protease serine 2 and cathepsin L that mediate cellular SARS-CoV-2 infection leading to COVID-19

PERIKLIS KATOPODIS^{1,2}, VLADIMIR ANIKIN¹⁻³, HARPAL S. RANDEVA⁴⁻⁶, DEMETRIOS A. SPANDIDOS⁷, KAMALJIT CHATHA⁸, IOANNIS KYROU^{4-6*} and EMMANOUIL KARTERIS^{1*}

¹Biosciences, College of Health and Life Sciences, Brunel University London, Uxbridge UB8 3PH;
²Division of Thoracic Surgery, The Royal Brompton and Harefield NHS Foundation Trust, Harefield Hospital, London UB9 6JH, UK; ³Department of Oncology and Reconstructive Surgery, Sechenov First Moscow State Medical University, 119146 Moscow, Russia; ⁴Warwickshire Institute for the Study of Diabetes, Endocrinology and Metabolism (WISDEM), University Hospitals Coventry and Warwickshire NHS Trust, Coventry CV2 2DX; ⁵Aston Medical Research Institute, Aston Medical School, Aston University, Birmingham B4 7ET; ⁶Warwick Medical School, University of Warwick, Coventry CV4 7AL, UK; ⁷Laboratory of Clinical Virology, School of Medicine, University of Crete, 71003 Heraklion, Greece; ⁸Department of Biochemistry and Immunology, University Hospitals Coventry and Warwickshire NHS Trust, Coventry CV2 2DX, UK

Received May 11, 2020; Accepted May 26, 2020

DOI: 10.3892/ijo.2020.5071

Abstract. Severe acute respiratory syndrome (SARS) coronavirus-2 (SARS-CoV2) is the cause of a new disease (COVID-19) which has evolved into a pandemic during the first half of 2020. Older age, male sex and certain underlying diseases, including cancer, appear to significantly increase the risk for severe COVID-19. SARS-CoV-2 infection of host cells is facilitated by the angiotensin-converting enzyme 2 (ACE-2), and by transmembrane protease serine 2 (TMPRSS2) and other host cell proteases such as cathepsin L (CTSL). With the exception of ACE-2, a systematic analysis of these two other SARS-CoV2 infection mediators in malignancies is lacking. Here, we analysed genetic alteration, RNA expression, and DNA methylation of TMPRSS2 and CTSL across a wide spectrum of tumors and controls. TMPRSS2 was overexpressed in cervical squamous cell carcinoma and endocervical adenocarcinoma, colon adenocarcinoma, prostate adenocarcinoma (PRAD), rectum adenocarcinoma

(READ), uterine corpus endometrial carcinoma and uterine carcinosarcoma, with PRAD and READ exhibiting the highest expression of all cancers. CTSL was upregulated in lymphoid neoplasm diffuse large B-cell lymphoma, oesophageal carcinoma, glioblastoma multiforme, head and neck squamous cell carcinoma, lower grade glioma, pancreatic adenocarcinoma, skin cutaneous melanoma, stomach adenocarcinoma, and thymoma. Hypo-methylation of both genes was evident in most cases where they have been highly upregulated. We have expanded on our observations by including data relating to mutations and copy number alterations at pan-cancer level. The novel hypotheses that are stemming out of these data need to be further investigated and validated in large clinical studies.

Introduction

Severe acute respiratory syndrome (SARS) coronavirus-2 (SARS-CoV2) is the cause of COVID-19, which was first identified at the end of 2019 and has evolved into a pandemic during the following months (1). Based on increasing data, older age and male sex predispose to severe COVID-19, whilst a number of underlying diseases/conditions are also directly related with significantly higher risk for adverse clinical outcomes from COVID-19 (1). The latter include diabetes, hypertension, obesity, immunosuppression, asthma, and chronic obstructive pulmonary disease (1).

Entry of SARS-CoV2 into its host cells is facilitated by its spike proteins which bind to the angiotensin-converting enzyme 2 (ACE-2) (2). Moreover, the spike viral proteins are primed by the transmembrane protease serine 2 (TMPRSS2) (2). As such, SARS-CoV2 infection of host cells is

Correspondence to: Dr Emmanouil Karteris, Biosciences, College of Health and Life Sciences, Brunel University London, Uxbridge UB8 3PH, UK
E-mail: emmanouil.karteris@brunel.ac.uk

*Joint senior authorship

Key words: cathepsin L (CTSL), transmembrane protease serine 2 (TMPRSS2), pan-cancer, DNA methylation, COVID-19, angiotensin-converting enzyme 2

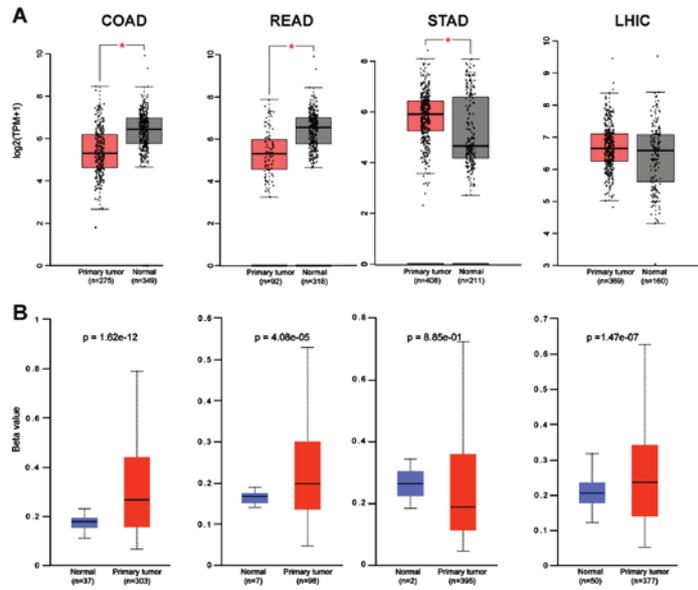


Figure 4. Correlation of CTSL expression with methylation status. COAD and READ had lower expression in tumors (A, red) due to higher methylation status (B, red). The opposite happens on STAD with the lower methylation, to allow higher CTSL expression. In LHIC there was no significant difference in normal to tumor expression even if the methylation is higher in the cancer samples ($P=1.47e-07$). CTSL, cathepsin L; COAD, colon adenocarcinoma; READ, rectum adenocarcinoma; STAD, stomach adenocarcinoma; LHIC, liver hepatocellular carcinoma.

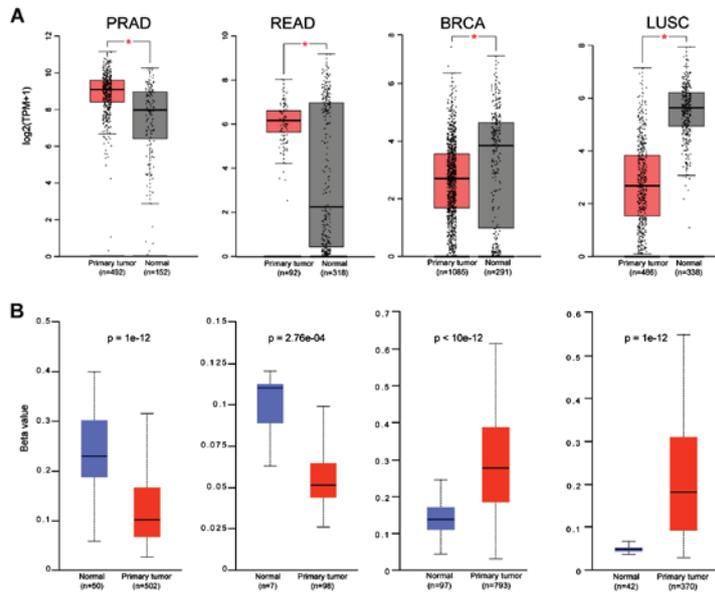


Figure 5. Correlation of TMPRSS2 expression with methylation status. PRAD and READ had higher expression in tumors (A, red) due to lower methylation status (B, red). The opposite happens on BRCA and LUSC with the higher methylation, to decrease the expression of TMPRSS2 in tumors. TMPRSS2, transmembrane protease serine 2; PRAD, prostate adenocarcinoma; READ, rectum adenocarcinoma; BRCA, breast invasive carcinoma; LUSC, lung squamous cell carcinoma.

facilitated by the cleavage of the spike proteins by TMPRSS2 and host cell proteases, such as cathepsin L (CTSL) (Fig. 1) (3).

Cancer has been identified as a risk factor for severe COVID-19, as many patients can be immunocompromised (4). In a very recent robust systematic analysis, Chai *et al* curated a pan-cancer analysis of ACE-2 detailing the expression and mutations across a wide spectrum of tumors (5). Following the same approach with this elegant study, here we expanded on these excellent observations by curating data for the other two mediators of SARS-CoV-2 infection, namely CTSL and TMPRSS2.

Materials and methods

Bioinformatic analysis. TMPRSS2 and CTSL were validated in The Cancer Genome Atlas (TCGA), GEPIA (<http://gepia.cancer-pku.cn/>), UALCAN (<http://ualcan.path.uab.edu/cgi-bin/ualcan-res.pl>). The pan-cancer cohort of TCGA was downloaded through cBioPortal (<https://www.cbioportal.org/>). The datasets used for the two genes were: ACC, adrenocortical carcinoma; BLCA, bladder urothelial carcinoma; BRCA, breast invasive carcinoma; CESC, cervical squamous cell carcinoma and endocervical adenocarcinoma;

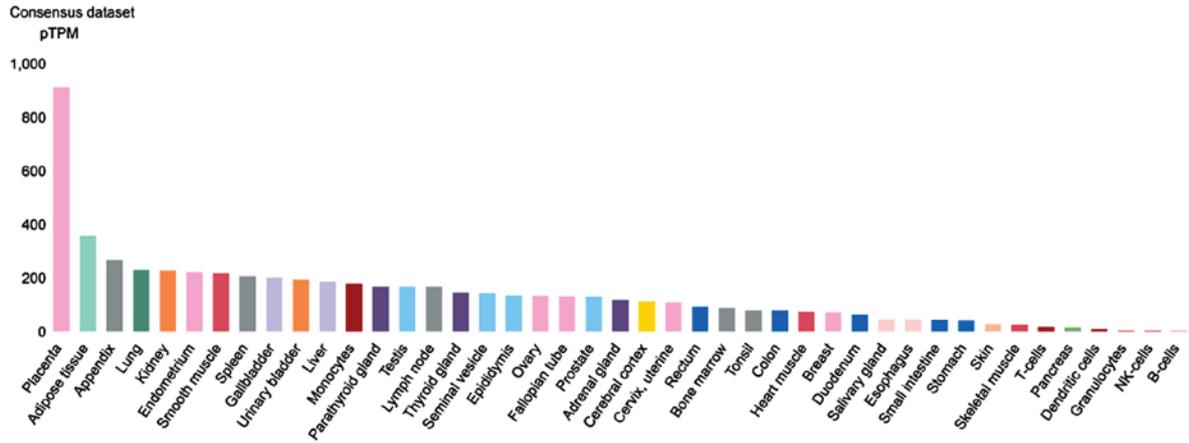


Figure 6. CTSL RNA expression in tissues. Consensus transcript expression levels for each gene were summarized in 74 human tissues based on transcriptomics data from three sources: HPA, GTEx and FANTOM5. The consensus NX value for each gene and organ/tissue represents the maximum NX value in the three data sources. Color-coding is based on tissue groups, each consisting of tissues with functional features in common. The data include 10,953 patients/10,967 samples in 32 cancer datasets. CTSL, cathepsin L; HPA, Human Protein Atlas; GTEx, Genotype-Tissue Expression; FANTOM5, the Functional Annotation of Mammalian Genomes 5; NX, normalized expression.

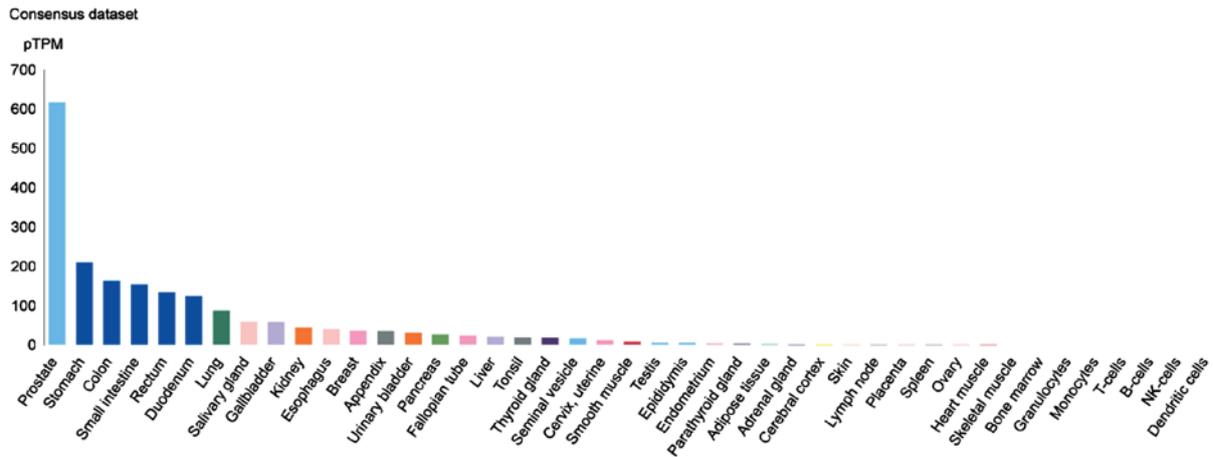


Figure 7. TMPRSS2 RNA expression in tissues. Consensus transcript expression levels for each gene were summarized in 74 human tissues based on transcriptomics data from three sources: HPA, GTEx and FANTOM5. The consensus normalized expression (NX) value for each gene and organ/tissue represents the maximum NX value in the three data sources. Color-coding is based on tissue groups, each consisting of tissues with functional features in common. The data include 10,953 patients/10,967 samples in 32 studies. HPA, Human Protein Atlas; GTEx, Genotype-Tissue Expression; FANTOM5, the Functional Annotation of Mammalian Genomes 5; NX, normalized expression.

CHOL, cholangio carcinoma; COAD, colon adenocarcinoma; DLBC, lymphoid neoplasm diffuse large B-cell lymphoma; ESCA, esophageal carcinoma; GBM, glioblastoma multiforme; HNSC, head and neck squamous cell carcinoma; KICH, kidney chromophobe; KIRC, kidney renal clear cell carcinoma; KIRP, kidney renal papillary cell carcinoma; LAML, acute myeloid leukemia; LGG, brain lower grade glioma; LIHC, liver hepatocellular carcinoma; LUAD, lung adenocarcinoma; LUSC, lung squamous cell carcinoma; MESO, mesothelioma; OV, ovarian serous cystadenocarcinoma; PAAD, pancreatic adenocarcinoma; PCPG, pheochromocytoma and paraganglioma; PRAD, prostate adenocarcinoma; READ, rectum adenocarcinoma; SARC, sarcoma; SKCM, skin cutaneous melanoma; STAD, stomach adenocarcinoma; TGCT, testicular germ cell tumors; THCA, thyroid carcinoma; THYM, thymoma; UCEC, uterine corpus endometrial carcinoma; UCS, uterine carcinosarcoma and UVM, uveal melanoma.

Results and Discussion

In TCGA datasets for all cancers, CTSL was upregulated in DLBC, ESCA, GBM, HNSC, LGG, PADD, SKCM, STAD and THYM, while it has lower expression than normal in COAD, LAML and READ (Fig. 2A and B). On the other hand, TMPRSS2 was upregulated in CESC, COAD, KICH, PRAD, READ, UCEC and UCS, with PRAD and READ exhibiting the highest expression of all cancers (Fig. 3A and B).

Subsequently, the expression of these two genes from GEPIA were correlated with the methylation status using the Ualcan database. For most of the cancers there was a strong correlation of the gene expression with its promoter methylation status in agreement with the findings for ACE-2 (5). CTSL in COAD and READ has a higher beta value; hence, lower expression of the gene, while in STAD, the lower methylation of the promoter, leads to higher

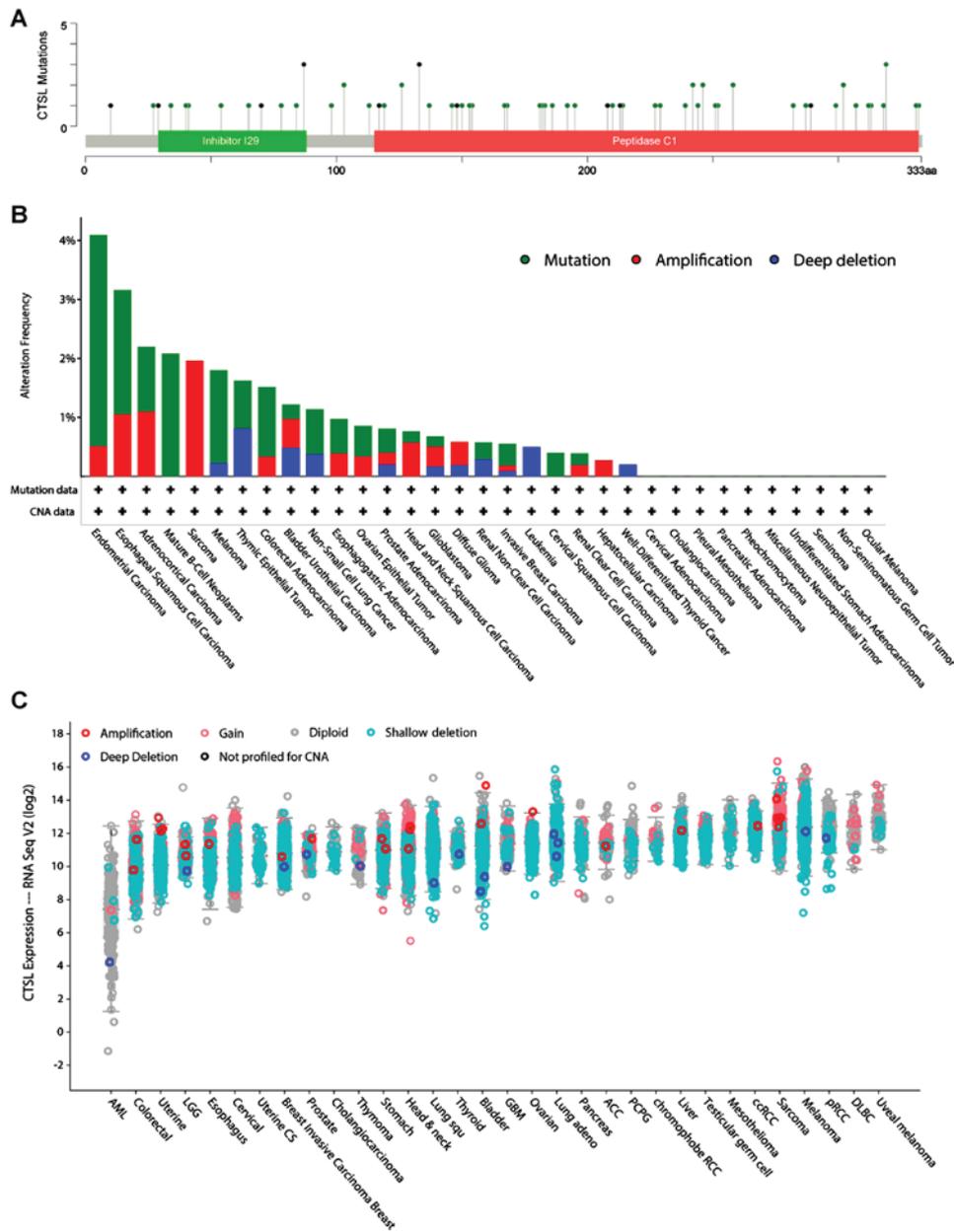


Figure 8. (A) CTSL mutations were distributed across the CTSL gene (cBioPortal) with 8 hot spot mutation sites in TCGA cohort using cBioPortal and 3d hotspots (12). Mutation diagram circles are colored with respect to the corresponding mutation types. In case of different mutation types at a single position, color of the circle is determined with respect to the most frequent mutation type. Mutation types and corresponding color codes are as follows: green: Missense Mutations; black: Truncating Mutations. (B) The relevance of genetic disorders and CTSL expression where it is shown that mutations were not relevant to RNA expression. (C) DNA copy variation were not statistically relevant to RNA CTSL expression. CTSL, cathepsin L; TCGA, The Cancer Genome Atlas.

gene expression (Fig. 4A and B). Five cancers were found to have lower methylation than normal for the CTSL, and seven cancers for the TMPRSS2. In LUSC and BRCA, the very high methylation rate of the TMPRSS2 gene leads to a very low expression level (Fig. 5A and B), while the very low methylation rate in PRAD and READ lead to very high numbers of gene transcripts.

Moreover, using the Human Protein Atlas analysis, of the consensus transcript expression levels that combines the Human Protein Atlas (HPA), Genotype-Tissue Expression; (GTEx) and the Functional Annotation of Mammalian Genomes 5 (FANTOM5) datasets, the expression of these two genes in organ/tissue samples were examined. CTSL is highly expressed in the placenta, adipose, appendix and lung tissue

(Fig. 6). TMPRSS2 is highly expressed in prostate [indeed, TMPRSS2 and PCA are utilized as biomarkers for prostate cancer (6)], stomach, colon and small intestine tissues (Fig. 7).

Furthermore, using the cBioportal pan-cancer panel, the region and the types of mutations were identified which these two genes have in all the examined cancer types (Figs. 8A and 9A). Most of the CTSL mutations are lying on the peptidase region and are mostly found in CESC, ESCA, Mature B-cell Neoplasms, Melanoma and COAD (Fig. 8B). Of note, in most of the cancers the majority of the patients had deletions and partly some gains and amplifications (Fig. 8C). TMPRSS2 mutations were lying across the whole gene region and mostly consist of gene fusions (TMPRSS2-ERG) in prostate adenocarcinoma (Fig. 9B and C).

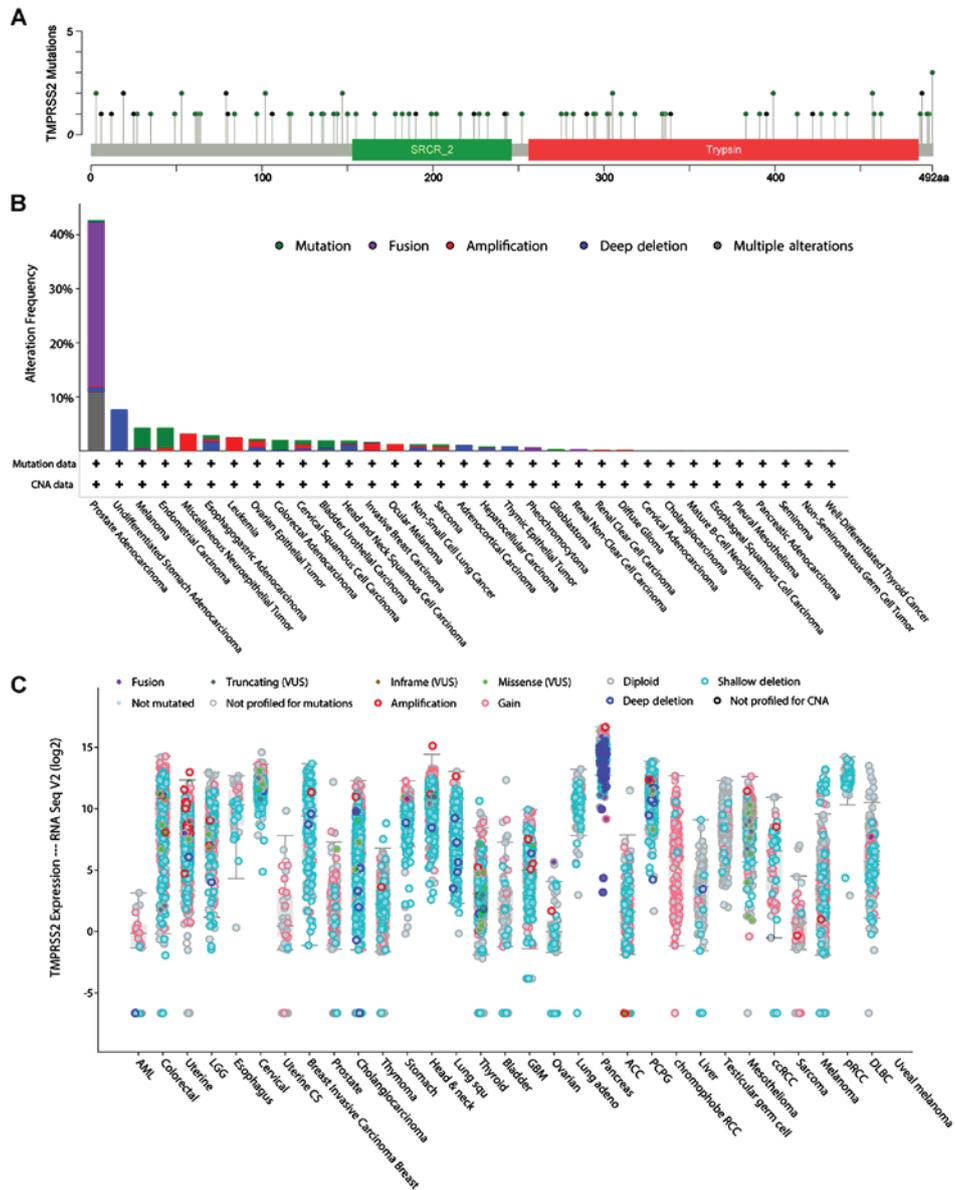


Figure 9. (A) TMPRSS2 mutations were distributed across the TMPRSS2 gene (cBioPortal) (12). Mutation diagram circles are colored with respect to the corresponding mutation types. In case of different mutation types at a single position, color of the circle is determined with respect to the most frequent mutation type. Mutation types and corresponding color codes are as follows: green: Missense mutations; black: Truncating mutations. (B) The relevance of genetic disorders and TMPRSS2 expression were as shown, in prostate cancer there is a 30% fusion of the gene and in the stomach adenocarcinoma 8% deep deletion. (C) DNA copy variation were not statistically relevant to RNA TMPRSS2 expression and as shown, many pancreatic cancer patients had deep deletions. CTSL, cathepsin L; TMPRSS2, transmembrane protease serine 2.

Finally, examination of the overall survival (OS) of all cancers, showed that CTSL low expression in KIRC (P=0.0001) has poor prediction for the patients, while low expression of CTSL in LUSC (P=0.0077) had better prognosis. High expression of TMPRSS2 in BRCA, SARC and UM had poor prediction for the patients, while gene expression was not statistically relevant for the patients of the other types of cancers (Figs. S1-S4).

Of note, neither of the two proteins were differentially regulated in LUAD; a comorbidity of severe COVID-19 contrary to ACE-2 (7). TMPRSS2 is significantly upregulated in prostate cancer, where it harbors gene fusion events, as documented in this study. While preparing this report, another interesting study was published which corroborated the expression of TMPRSS2 in the prostate. In this study, the authors have put

forward the question of whether the TMPRSS2 increased expression in prostate is involved with the documented sexual dimorphism of COVID-19 (8). This is an exciting hypothesis that needs to be studied further given that the clinical data indicate that male sex is among the risk factors for adverse COVID-19 related outcomes.

In our analysis we also demonstrate that the pancreas is riddled with deep deletions for TMPRSS2 where ACE-2 is co-expressed. Interestingly, Liu *et al* (9) have recently shown that ACE-2 expression in the pancreas may lead to pancreatic damage following SARS-CoV-2 infection. For CTSL, higher expression was noted in the human placenta where ACE-2 is also co-expressed (10). This finding warrants further investigation into the role of these viral entry mediators in placentation, vertical transmission of SARS-CoV-2 to the fetus, as well as

their potential involvement in maternal and neonatal complications (11). These hypotheses remain to be investigated in large clinical studies.

Acknowledgements

Not applicable.

Funding

No funding was received.

Availability of data and materials

All data generated or analysed during this study are included in this published article (and its supplementary information files).

Authors' contributions

PK generated the data, and produced the figures; VA and KC contributed to critical revision of the article; HSR, DAS contributed to the writing of the manuscript and final edits; IK and EK contributed equally to the conception of the work and data analysis and interpretation. All authors read and approved the final manuscript.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

DAS is the Editor-in-Chief for the journal, but had no personal involvement in the reviewing process, or any influence in terms of adjudicating on the final decision, for this article. The other authors declare that they have no competing interests.

References

1. Yuki K, Fujiogi M and Koutsogiannaki S: COVID-19 pathophysiology: A review. *Clin Immunol* 215: 108427, 2020.
2. Hoffmann M, Kleine-Weber H, Schroeder S, Krüger N, Herrler T, Erichsen S, Schiergens TS, Herrler G, Wu N-H, Nitsche A, *et al*: SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. *Cell* 181: 271-280.e8, 2020.
3. Smieszek SP, Przychodzen BP and Polymeropoulos MH: Amantadine disrupts lysosomal gene expression: A hypothesis for COVID19 treatment. *Int J Antimicrob Agents*: Apr 30, 2020 (Epub ahead of print).
4. Sidaway P: COVID-19 and cancer: What we know so far. *Nat Rev Clin Oncol*: Apr 7, 2020 (Epub ahead of print).
5. Chai P, Yu J, Ge S, Jia R and Fan X: Genetic alteration, RNA expression, and DNA methylation profiling of coronavirus disease 2019 (COVID-19) receptor ACE2 in malignancies: A pan-cancer analysis. *J Hematol Oncol* 13: 43, 2020.
6. Filella X and Foj L: Prostate Cancer Detection and Prognosis: From Prostate Specific Antigen (PSA) to Exosomal Biomarkers. *Int J Mol Sci* 17: 1784, 2016.
7. Kong Q, Xiang Z, Wu Y, Gu Y, Guo J and Geng F: Analysis of the susceptibility of lung cancer patients to SARS-CoV-2 infection. *Mol Cancer* 19: 80, 2020.
8. Stopsack KH, Mucci LA, Antonarakis ES, Nelson PS and Kantoff PW: *TMPRSS2* and COVID-19: Serendipity or opportunity for intervention? *Cancer Discov*: Apr 10, 2020 (Epub ahead of print).
9. Liu F, Long X, Zhang B, Zhang W, Chen X and Zhang Z: ACE2 expression in pancreas may cause pancreatic damage after SARS-CoV-2 infection. *Clin Gastroenterol Hepatol*: Apr 22, 2020 (Epub ahead of print).
10. Li M, Chen L, Zhang J, Xiong C and Li X: The SARS-CoV-2 receptor ACE2 expression of maternal-fetal interface and fetal organs by single-cell transcriptome study. *PLoS One* 15: e0230295, 2020.
11. Stumpfe FM, Titzmann A, Schneider MO, Stelzl P, Kehl S, Fasching PA, Beckmann MW and Ensser A: SARS-CoV-2 infection in pregnancy - a review of the current literature and possible impact on maternal and neonatal outcome. *Geburtshilfe Frauenheilkd* 80: 380-390, 2020 (In German).
12. Gao J, Aksoy BA, Dogrusoz U, Dresdner G, Gross B, Sumer SO, Sun Y, Jacobsen A, Sinha R, Larsson E, *et al*: Integrative analysis of complex cancer genomics and clinical profiles using the cBioPortal. *Sci Signal* 6: pii, 2013.



This work is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International (CC BY-NC-ND 4.0) License.