

New insights into the physiological, pathological and pharmacological roles of voltage-gated potassium channel Kv10.1 in cancer (Review)

YUXIAO ZHEN^{1*}, WEIBO HU^{2*}, SHANSHAN DONG¹, BOCHANG WANG³, JIAN SHEN⁴,
LEYUAN DING⁵, LIFENG LI⁶, HAILONG AN¹, XUZHAO WANG⁷ and YAFEI CHEN¹

¹Key Laboratory of Molecular Biophysics of Hebei Province, Institute of Biophysics, School of Health Sciences and Biomedical Engineering, Hebei University of Technology, Tianjin 300401, P.R. China; ²Department of Oncology, The People's Hospital of Leling, Leling, Shandong 253600, P.R. China; ³Department of Breast Cancer, Tianjin Cancer Hospital Airport Hospital, National Clinical Research Center for Cancer, Tianjin 300308, P.R. China; ⁴Department of Bioengineering, School of Chemical Engineering, Hebei University of Technology, Tianjin 300401, P.R. China; ⁵Department of Applied Statistics, School of Science, Hebei University of Technology, Tianjin 300401, P.R. China; ⁶Department of Otolaryngology-Head and Neck Surgery, Beijing Tongren Hospital, Capital Medical University, Beijing 100730, P.R. China; ⁷School of Medicine, Hebei University of Engineering, Handan, Hebei 056038, P.R. China

Received October 25, 2025; Accepted February 5, 2026

DOI: 10.3892/ijo.2026.5911

Abstract. Kv10.1, also known as Eag1 or KCNH1, is a voltage-gated potassium ion channel, which exists in cell membrane and is closely associated with cancer and multiple precancerous lesions. Emerging experimental evidence shows that Kv10.1 is essential for the occurrence, growth, metastasis, proliferation and death of various malignant tumors. The pathogenesis of Kv10.1 and the signal pathways involved in its regulation are different in different cancers. The present review explored the origin, structure, distribution in normal and tumor cells, physiological and pathological characteristics, roles in cancer and tumor regulation mechanisms of Kv10.1. Finally, Kv10.1 related signaling pathways and its current use as a pharmacological modulator are summarized, aiming to provide new insights into the pharmacological research of Kv10.1 in cancer.

Contents

1. Introduction to tumor related potassium channels
2. Kv10.1 potassium channel: An overview
3. Physiological and pathological functions of Kv10.1
4. Tumor regulation mechanisms of Kv10.1.
5. Signaling pathway
6. Regulators and inhibitors
7. Clinical status
8. Summary

1. Introduction to tumor related potassium channels

Ion channels are pore-forming transmembrane proteins, which regulate the life activities of organisms by controlling the ion transport, not only participating in the remodeling of cytoskeleton and the interaction between cells, but also regulating the processes of cell migration and invasive growth. Moreover, it has been shown that abnormal expression or activity change of ion channels can regulate the proliferation and apoptosis of tumor cells (1).

Voltage-gated potassium channels (Kv) are a type of protein channels formed on the cell membrane, which can be turned on or off according to the voltage change of the cell membrane. They can regulate the transmembrane flow of potassium ions, thus maintaining the normal physiological functions of cells, such as nerve conduction, muscle contraction and cell excitability (2). The human genome contains 40 voltage-gated potassium channels, which are involved in a number of physiological processes, including the repolarization of neuron or cardiac action potential, the regulation of calcium signal and cell volume and the promotion of cell proliferation and migration. The Kv channel has shown promising research potential

Correspondence to: Dr Xuzhao Wang, School of Medicine, Hebei University of Engineering, 19 Taiji Road, Economic and Technological Development District, Handan, Hebei 056038, P.R. China

E-mail: wangxuzhao@hebeu.edu.cn

Professor Yafei Chen, Key Laboratory of Molecular Biophysics of Hebei Province, Institute of Biophysics, School of Health Sciences and Biomedical Engineering, Hebei University of Technology, 5340 Xiping Road, Beichen, Tianjin 300401, P.R. China

E-mail: chenyafei@hebut.edu.cn

*Contributed equally

Key words: Kv10.1, tumors, inhibitor, signaling pathway

in cancer, autoimmune diseases, metabolic, neurological and cardiovascular diseases. It has been found that several voltage-gated potassium channels are closely associated with tumors: KCa3.1 (KCNN4) (3), Kv10.1 (EAG1, KCNH1) (4), Kv1.3 (KCNA3) (5) and HERG (Kv11.1, KCNH2) (6).

Among the potassium ion channels, Kv10.1 has become a focus of current anti-cancer research due to its high expression characteristics in tumors and close association with tumor occurrence and development.

2. Kv10.1 potassium channel: An overview

Kv10.1 (potassium voltage-gated channel subfamily H member 1/KCNH1, known as Ether-à-go-go-1 or EAG1) is a voltage-gated potassium channel (4). Kv10.1 has been found to be expressed in a number of different cancer cell lines, including gastric cancer (7), liver cancer (8) and lung cancer (9), but its expression in normal tissues is limited (10,11). It has been proved that it is highly expressed in a number of tumors and plays an important role, which makes it a potential marker and target for tumor diagnosis and treatment (7-9,12-16).

The origin of Kv10.1. The discovery of Kv10.1 channel can be traced back to the study of voltage-gated potassium ion channels. Kv10.1 was first discovered by Kaplan *et al.* (2) in 1969 in the mutant X chromosome of *Drosophila*. Its name 'ether-a-go-go' comes from the irregular leg tremor phenotype exhibited by *Drosophila* mutants under ether anesthesia, which is caused by a functional defect in voltage-gated potassium channels due to gene locus mutations (17). Studies have shown that molecular cloning technology confirms that the protein encoded by the *Eag* gene belongs to the Kv superfamily and reveals its direct association with the regulation of neural excitability in *Drosophila* (17). Kv10.1 is located in chromosome 1, band q32.1-32.3 and comprises 457,343 bases and 12 exons (18).

The molecular structure of Kv10.1. Kv10.1 is encoded by *KCNH1* gene, which encodes a 989 amino acid protein with an estimated molecular weight of 111,423 Daltons. The overall structure of Kv10.1 channel is similar to these of the other Kvs. The core region contains six transmembrane domains (transmembrane helices S1-S6), including voltage sensor domains (transmembrane helices S1-S4) and potassium ion selective permeability channels (transmembrane helices S5, pore helices S6) and the N-terminal and C-terminal of this channel protein (Fig. 1). The single-hole channels of these six transmembrane regions (S1-S6) are activated by depolarization (19-21). The N-terminal contains Per-Arnt-Sim (PAS) domain. C-terminal contains cyclic nucleotide binding homology domain (CNBHD), C-linker, ciliary localization signal and tetrameric coiled helix domain (19,22).

Although the structure of Kv10.1 has a number of similarities with other Kvs, based on the three-dimensional structure of the rat Kv10.1 obtained by single-particle cryoelectron microscopy, it was found that Kv10.1 has a number of unique structural features compared with other Kv channels. The S2-S3 connector of Kv10.1 extending into cytoplasm is different from those of other voltage-gated potassium channels. This connector is a conservative feature of Kv family and the only

other Kv subfamily member containing the similar connectors is Kv7 (19,23). In Kv10.1, S4 region is the main positive charge concentration region, usually with 5-7 positive charges distributed. As a voltage receptor, this region can respond to the electric field force when the membrane potential changes and drive the conformational change of the channel. In addition, other transmembrane regions (such as S1, S2 and S3) may also contain a small amount of positively charged amino acid residues, but the positive charge density is relatively low and the sensitivity to voltage change is not as significant as that of S4 region. S1-S4 constitute positively charged amino acids in the field of voltage sensors, which move membrane potential with the depolarization of cerebral cortex (24-26). The S4-S5 linker in Kv10.1 is a short loop composed of five amino acid residues, which contrasts with the helical structure formed by 15 residues parallel to the membrane in domain exchange potassium channels (such as Kv1.2) (19,27). The segments S5-S6 form the pore domain of the channel, while the ring P (between S5-S6) is responsible for the K⁺ selectivity. These local structural features determine that Kv10.1 has a different voltage gating process compared with other types of Kv channels. S4 in Kv10.1 directly interacts with the carbon terminal C-linker to close the channel, that is, the channel can realize voltage-dependent activation gating without covalent connection between S4 and S5 (27,28).

Kv10.1 also has unique N-terminal and C-terminal domains, which are necessary for subfamily-specific channel assembly (17,29,30). Kv10.1 has a large N-terminal and a C-terminal in cells, accounting for ~70% of its molecular weight. In addition, there are several regulatory domains at the ends of cells. N-terminal contains a PAS domain, which is usually involved in the detection of redox reaction. It is considered as an oxygen sensor, which can induce the production of hypoxia inducible factor hypoxia-inducible factor 1 (HIF-1) in hypoxic environment, thus causing the increase of glycolysis rate and angiogenesis, which is beneficial to the growth of tumor cells in hypoxic environment. The C-terminal contains a CNBHD, which is a calmodulin binding site (31). The fragment between CNBHD and S6 is called C-linker. S6 spirally extends to the intracellular region and is connected to C-linker, forming an intracellular ring above CNBHD. The C-linker couples the movements of S6 and CNBHD and CNBHD is connected to the pore region through C-linker (32). As with other ion channels, Kv10.1 undergoes nitrogen chain glycosylation at N388 and N406 sites, which is very important for the correct transport of the channel to the membrane. Three-dimensional structural analysis by cryo-electron microscopy showed that the PAS domain of Kv10.1 channel in rats was located in the intracellular region and mainly interacted with CNBHD, a neighboring subunit. In addition, Kv10.1 channel contain amino acid residues carrying glycosylation regions of polymers, among which glycosylation of N406 is a necessary condition to maintain the normal structure and function of Kv10.1 and deglycosylation will lead to weakening of channel current and slow activation (19,33).

The distribution of Kv10.1

The distribution of Kv10.1 in normal tissues. Kv10.1 channel is a protein embedded in plasma membrane, but it can also be found in subcellular structural membrane including

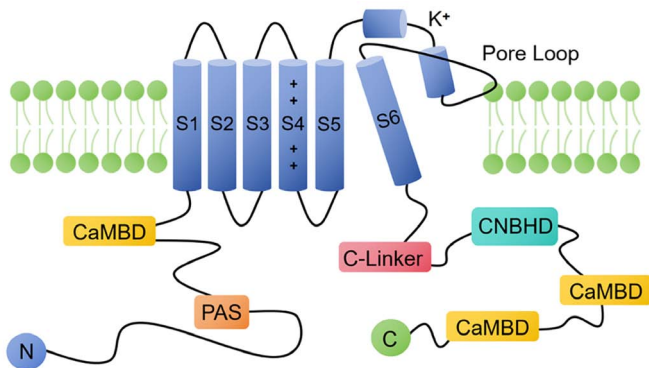


Figure 1. Structure of Kv10.1. Kv10.1 comprises six transmembrane domains (transmembrane helices S1-S6), including a voltage-sensing domain (formed by transmembrane helices S1-S4, with S4 bearing positive charges) and a potassium ion-selective permeation pathway (formed by transmembrane helix S5 and the pore helix S6), along with the N-terminus and C-terminus of the channel protein. The N-terminus contains a PAS domain. The C-terminus is connected to the CNBHD regulatory domain via a C-Linker and multiple CaMBDs are involved in the regulation of channel activity mediated by calcium signaling. PAS, Per-Arnt-Sim; CNBHD, cyclic nucleotide-binding homology domain; CaMBDs, calmodulin-binding domains.

the inner nuclear membrane, intracellular vesicles and close to the primary cilia. Under normal physiological conditions, Kv10.1 is almost undetected in peripheral tissues, except for a few peripheral tissues, such as pre fusion myoblasts (34) and Kv10.1 shows abundant expression in specific brain regions. Previous studies have shown that Kv10.1 mainly exists in the brain, locates at the presynaptic terminal and regulates the release of neurotransmitters, which can be used as a regulator of local action potential (35).

The distribution of Kv10.1 in malignant tumors and precancerous lesions. Kv10.1 potassium channel is the first voltage-dependent potassium channel proved to be closely associated with tumor growth (36-39). In recent years, a large number of studies have shown that Kv10.1 potassium channels are abnormally expressed in malignant tumors and precancerous lesions (7,40-55). Kv10.1 was highly expressed in the primary tumor of head and neck squamous cell carcinoma and was detected in 10 of 12 cell lines derived from head and neck squamous cell carcinoma (40,41). Kv10.1 was positively expressed in breast cancer cell lines, such as MCF-7 (12), T-47D (12) and MDA-MB-231 (42). Kv10.1 is highly expressed in lung cancer. A549 and NCI-H1975 are common lung cancer cell lines (9). In liver cancer, Kv10.1 inhibitor can inhibit the proliferation and migration of liver cancer cells HuH-7 cells and HepG2 cells (13). By immunohistochemical methods, >70% colon cancer tissues showed positive expression of Kv10.1, which was markedly higher than that of normal tissues adjacent to cancer. Kv10.1 was also highly expressed in colon cancer cell line SW480 (14). The primary tissue samples of gastric cancer were detected by immunohistochemistry and it was found that 70.5% of tumor tissues showed positive expression of Kv10.1 protein, which was markedly higher than that of normal tissues adjacent to cancer. Compared with normal gastric mucosal cells, the expression of Kv10.1 in gastric cancer cell lines (SGC-7901, BGC-823) is also higher (7,43). The expression of Kv10.1 in cervical cancer tissue was markedly higher than that in normal cervical tissue and the expression

content of Kv10.1 in cervical cancer cells HeLa, Siha and Casci was higher than that in normal cervical cells (15,44). Studies have shown that Kv10.1 can be expressed in cervical intraepithelial neoplasia and the expression level is associated with the level of cervical intraepithelial neoplasia (16,45,46). Kv10.1 is highly expressed in ovarian cancer and atypical adenoma hyperplasia of precancerous lesions (16). The high expression of Kv10.1 may be a potential prognostic marker of prostate cancer, which is associated with the poor prognosis of patients. Kv10.1 is highly expressed in prostate cancer tissues and tumor cell lines, but almost not in normal prostate tissues (47-49). The abnormally high expression of Kv10.1 in neuroblastoma is not only driven by the transcription level, but also depends on the fine regulation of post-translational modifications (such as ubiquitination and phosphorylation) (50). Kv10.1 is also expressed in melanoma, which also affects the proliferation of melanoma cells (51-53). Kv10.1 is highly expressed in bone marrow primordial cells of acute myeloid leukemia patients, but hardly expressed in normal hematopoietic stem cells and benign blood diseases (54). The expression level of Kv10.1 in patients with soft tissue sarcoma has been detected by immunohistochemistry. Kv10.1 was expressed in 71% tumors and the frequency ranged from 56% in liposarcoma to 82% in rhabdomyosarcoma (55). Kv10.1 is also expressed in osteosarcoma, which also affects the proliferation of osteosarcoma cells (56). The aforementioned results are listed in Table I. The abnormal expression of Kv10.1 in a number of malignant tumors and precancerous lesions will provide a certain molecular basis for the diagnosis and treatment of corresponding diseases. Different experimental techniques are used to analyze its mechanism of action from multiple levels in studying the expression and function of Kv10.1. Fresh patient samples can be quantified for protein and mRNA expression levels through western blotting and quantitative (q) PCR. In cell lines in good condition, channel current characteristics can be directly measured through electrophysiological recording, while wax blocks can be used for immunohistochemistry and *in situ* localization of protein expression. Nevertheless, each method has its limitations. Western blotting involves semi-quantitative detection following protein separation by electrophoresis, allowing precise measurement of protein molecular weight and relative expression level. However, it cannot provide protein localization information and requires tedious experimental steps, with strict control of parameters such as sample loading volume and antibody concentration to avoid errors. qPCR reflects gene expression levels by detecting mRNA levels, which is simple to operate and has high throughput. However, it cannot directly reflect protein function and results may be affected by factors such as RNA quality and primer specificity. Inappropriate primer design may lead to PCR reactions with low specificity or efficiency. Immunohistochemistry localizes target proteins in tissue sections through antigen-antibody reactions, visually displaying their distribution in the cell membrane, cytoplasm, or nucleus. However, staining intensity is easily affected by factors such as antibody concentration and incubation time. Antibodies from different manufacturers have varying binding abilities to target proteins, leading to differences in results. Electrophysiological experiments directly measure the current characteristics of ion channels to reflect their functional activity. However, they require extremely high cell states and

Table I. Cell lines with high expression of Kv10.1.

Cancer type	Cell line	(Refs.)
Head and neck cancer	SCC29, SCC40, SCC42B, SCC040, SCC041, SCC078, SCC094, SCC096A, SCC120, SCC147	(33,36)
Breast cancer	MCF-7, T-47D	(12)
	MDA-MB-231	(42)
Lung cancer	A549, NCI-H1975	(9)
Liver cancer	HuH-7, HepG2	(13)
Colon cancer	SW480	(14)
Gastric cancer	SGC-7901, BGC-823	(7,43)
Cervical cancer	HeLa, SiHa, CaSki	(15,46)
Ovarian cancer	SKOV3, OVCAR3	(16)
Prostatic cancer	DU-145, PC-3	(47-49)
Neuroblastoma	SHSY-5Y	(50)
Melanoma	IGR1, IPC298, IGR39, A375	(51-53)
Acute Myeloid Leukemia	HL-60, K562, PLB-985, HEL, CMK, KASUMI, UT-7	(54)
Rhabdomyosarcoma	TE-671, A-204	(55)
Fibrosarcoma	HT-1080, Hs633t	(55)
Osteosarcoma	SaOS-2, MG-63	(56)

have long experimental cycles, making them difficult to apply on a large scale. In practical applications, selection should be based on research objectives and combined with the biological characteristics of the samples.

In addition, the ectopic expression of Kv10.1 is associated with the phenotype of chemotherapy-resistant cells. Therefore, the inhibition of Kv10.1 channel can improve the response of cells to commonly used therapeutic drugs in chemotherapy. *In vitro* experiments on ovarian cancer cells showed that the expression of Kv10.1 was associated with the chemotherapy resistance to cisplatin. Compared with cells treated with cisplatin alone, the combination of down-regulation of Kv10.1 and cisplatin increased the apoptosis of ovarian cancer cells (44).

3. Physiological and pathological functions of Kv10.1

Physiological characteristics of Kv10.1. Kv10.1 is a voltage-gated potassium channel, which is highly expressed in nervous system and some cancer cells. It has unique electrophysiological characteristics and plays an important role in cell excitability regulation, tumor occurrence and development.

Kv10.1 channel is voltage-dependent and its activation degree changes with the membrane potential. This characteristic makes Kv10.1 channel play an important role in regulating cell excitability. In addition, the gating characteristics of Kv10.1 channel are also different from other potassium channels, with a slow activation and deactivation process. The activation characteristics of Kv10.1 can be associated with Cole-Moore shift observed in squid axons published by Cole KS and Moore JW in 1960 (57). This effect was originally used to describe the potassium current in squid axons, which is characterized by a significant delay in current activation when hyperpolarized resting membrane potential channels are activated (58,59). The Cole-Moore shift of Kv10.1 is ~10s of milliseconds, while that of other members of the voltage-gated potassium channel

family is close to 1 msec and the Cole-Moore shift of Kv10.1 is steeper than that of Kv10.2 (60). This electrophysiological feature can be used as a marker to identify the expression of Kv10.1 in cells and ectopic expression. The activation of Kv10.1 depends on cell membrane resting potential and extracellular Mg^{2+} concentration and low extracellular pH can slow its activation (61,62). Cole-Moore shift can be enhanced by increasing extracellular Mg^{2+} concentration in physiological range (63). The Cole-Moore shift of Kv10.1 can be inhibited by using amiodarone and mibefradil (64,65).

Kv10.1 is mainly expressed in the nervous system and its electrophysiological characteristics are closely associated with the function of the nervous system. In adult rats, the mRNA of Kv10.1 can be mainly detected in olfactory bulb, cerebral cortex, hippocampus and cerebellum and it is consistent with its protein expression level (66). Further research found that Kv10.1 was mostly expressed on dopaminergic cells in physiological state, which may be associated with its electrophysiological function (67). In the nervous system, Kv10.1 can be used as a regulator of local action potential, especially when other potassium channels suffer from high-frequency bursts of cumulative inactivation. Part of the function adjustment of Kv10.1 channel involves its detergent resistant membrane part (also called fat raft) (68,69).

The pathological function of Kv10.1 in cancer. Kv10.1 channel is highly expressed in 70% of tumors and its expression level is closely associated with the malignant degree, metastatic ability and clinical prognosis of tumors. Kv10.1 is highly expressed in head and neck cancer, breast cancer and acute myeloid cells (54,70-72) and inhibiting or knocking out Kv10.1 can reduce tumor growth (73-76). Therefore, Kv10.1 is considered as a potential target for cancer treatment.

Kv10.1 channel affects the production of resting potential and action potential by regulating the potassium ion

permeability of cell membrane. This electrophysiological change may further affect the intracellular signal transduction pathway, thus regulating the cell proliferation process. In tumor cells, the high expression of Kv10.1 channel may lead to uncontrolled cell proliferation and promote tumor growth. In addition to regulating cell proliferation, Kv10.1 channel may also participate in tumor metastasis by regulating cell migration and invasion.

Kv10.1 is very important for the proliferation of tumor cells. In 1997, Brüggemann *et al* first discovered the function of Kv10.1 in cell proliferation in *Xenopus* oocytes (77). Kv10.1 leads to abnormal proliferation of cells, including enhanced metabolic activity and reduced dependence of cells transfected with Kv10.1 on growth factors in the medium. Cells transfected with Kv10.1 can continue to grow in the medium with low concentration of serum, even without matrix indicating the loss of contact inhibition characteristics (17). *In vitro*, inhibiting the expression of Kv10.1 can reduce the cell proliferation, migration and invasion of cancer cells. *In vitro* experiments, injecting exogenous CHO cells expressing Kv10.1 into immunosuppressed mice induced the occurrence of invasive tumors (78), while using specific monoclonal antibodies to inhibit Kv10.1 can inhibit the growth of tumors *in vitro* (79) and similar effects were also observed in animal experiments (80-85). In recent years, the closed state wild-type channel model of cryoelectron microscope structure based on HERG and Kv10.1 channels has been constructed by using Rosetta software and cryoelectron microscope structure. These models were then used in molecular docking studies to explore the mechanism of drug channel interaction (86-89).

Generally, Kv10.1 channel plays an important role in the occurrence and development of cancer and its high expression is closely associated with the clinical prognosis of a number of tumors. In-depth study on the pathological function and regulation mechanism of Kv10.1 channel is expected to provide new theoretical basis and therapeutic strategies for cancer prevention and treatment.

4. Tumor regulation mechanisms of Kv10.1

In the process of carcinogenesis, Kv10.1 may play a role through various mechanisms. First, as a potassium ion channel, Kv10.1 can regulate the potassium ion permeability of cell membrane, thus affecting the generation of cell resting potential and action potential. This electrophysiological change may further affect the signal transduction pathway in cells and then regulate the gene expression associated with carcinogenesis. Second, Kv10.1 may also interact with other signal pathways to jointly promote the occurrence and development of tumors. Kv10.1 is highly expressed in a number of tumor cells and tissues, but its specific mechanism of action is not very clear. At present, there are only some explorations about Kv10.1 in cell cycle regulation, cell hyperpolarization, signaling pathway, epigenetic regulation mechanism.

Cell cycle regulation. In 2006, it was found that the expression activity of Kv10.1 was associated with the cell cycle (90). Cell cycle regulates the progress of mitosis. During mitosis, if progesterone or mitotic promoter exists, the activity of Kv10.1

channel will be inhibited, which also proves its cell cycle sensitivity (91).

The research after 2016 gave more specific results. That is, Kv10.1 is located in centrosome and primary cilia (92) and promotes the decomposition of primary cilia in G₂/M phase, which is beneficial to the progress of cell cycle (93). Following Kv10.1 knock-out, ciliary decomposition is inhibited and proliferation is delayed. Therefore, the regulation of Kv10.1 on ciliary development can explain the influence of Kv10.1 expression on normal cell proliferation and may be a main mechanism of its tumorigenesis (94). Kv10.1 affects Ca²⁺ entry through hyperpolarization and Ca²⁺ inhibits EAG1 function through calmodulin (3). Ca²⁺ is associated with the cell cycle and cell replication. Calcium influx is considered necessary for the transition from G₁ phase to S phase in the cell cycle. This hyperpolarization also facilitates the entry of a large amount of nutrients into the cell, thereby promoting cell proliferation (10,95).

Similar research also support the aforementioned view. Urrego *et al* (96) reported that Kv10.1 could be detected only in G₂/M phase. As the E2F transcription factor 1 (E2F1) coordinates cell division and induces the expression of Kv10.1 in G₂ phase, Kv10.1 can only be detected in G₂/M phase and downregulation of Kv10.1 will prolong the duration of G₂/M phase. However, Kv10.1 will be diluted in G₀/G₁ phase, so it cannot be detected in other stages of cell cycle (97). Some studies also hypothesize that the strong influence of Kv10.1 channel in cell cycle progress is also associated with ciliary disintegration, which usually provides the signal transduction needed to start proliferation (96,97). Cyclic expression of Kv10.1 has been proved to exist at the base of primary cilia of hTERRPE-1 cells, which may lead to significant disposability of microtubules in the subsequent process (10).

Regarding microtubule activity, the upregulation of Kv10.1 is closely associated with a significant increase in microtubule dynamics, characterized by assembly and disassembly rates. Therefore, there is an important association between Kv10.1 overexpression and certain processes including cellular variability (96,97). It has been proved that the mobility of MDA-MB-231 cells decreased markedly when Kv10.1 was exposed to a specific blocking agent such as astemizole. Some hypotheses suggest that cell migration caused by Kv10.1 channel is mainly due to the change of microtubule dynamics caused by hyperpolarization of action potential, which has been proved to induce calcium to enter through Ca²⁺ release and activates ORAI calcium release-activated calcium modulator 1 (ORAI1). ORAI1 association with Ca²⁺ ATPase seems to be associated with the proliferation and survival of cancer cells (98).

Hyperpolarization mechanism. The outflow of K⁺ through Kv10.1 channel makes cells hyperpolarized, thus promoting the influx of Ca²⁺, leading to the proliferation of non-invasive cells (10,99,100). However, the relationship between Kv10.1 and intracellular Ca²⁺ is complicated. Although Kv10.1 is beneficial to Ca²⁺ influx under hyperpolarization, high intracellular Ca²⁺ reversibly inhibits the overexpression of Kv10.1 in a number of tumors (101,102). Overexpression of Kv10.1 can lead to membrane potential hyperpolarization and affect cell cycle progression. Potential changes in the

cell cycle are much slower, smoother and less pronounced than rapid action potentials. Due to membrane hyperpolarization, the driving force for calcium influx is enhanced. Ca^{2+} is a major second messenger involved in cell proliferation, migration, survival and apoptosis (103). Depletion of extracellular calcium ions can arrest cells in the early G_1 and G_1/S phase transition. Normal cell proliferation relies on intracellular and extracellular calcium ions, especially during the G_1 phase. However, cancer cells can bypass this requirement and continue to proliferate in calcium-deficient media (104,105). This mechanism exhibits high similarity across various types of cancer. Calcineurin is one of the main regulators of intracellular Ca^{2+} signaling. Calcineurin inhibits the degradation of cyclin D1 through dephosphorylation of T286 residue, promoting cell cycle progression (106). Ca^{2+} can also form a dynamic interaction network with the Hippo pathway, jointly regulating cell proliferation, differentiation and tissue homeostasis (107). Although there are currently no specific studies directly elucidating the role of Kv10.1 in the interaction with Ca^{2+} regulation of cyclin D1 and the Hippo pathway, based on the existing understanding of the complex regulatory relationship between Ca^{2+} and cyclin D1 and the Hippo pathway, it is reasonable to speculate that Kv10.1 may play a role in this regulatory network. Cell hyperpolarization is very sensitive to the activation of Kv10.1, which can regulate cell hyperpolarization through the interaction between N-terminal of PAS domain and S4-S5 connector (28).

When the cell membrane is hyperpolarized, the Kv10.1 channel can sense this potential change and open, allowing potassium ions to flow from the inside of the cell to the outside (17). Therefore, drug development for Kv10.1 channel has become a potential cancer treatment strategy, aiming at inhibiting the growth and spread of tumor by blocking its hyperpolarization mechanism.

Epigenetic regulation mechanism. Epigenetic regulation mechanism is a mechanism that changes biological phenotype without involving DNA sequence changes, but including DNA methylation, histone modification and non-coding RNA-mediated regulation. These modifications can affect the transcription activity of genes, thus regulating gene expression (108). For example, the DNA methylation status in the promoter region of Kv10.1 gene may affect its transcription activity, thus regulating the expression level of Kv10.1. In addition, histone modification may also regulate the transcription process of Kv10.1 gene by affecting the position and arrangement of nucleosomes. Studies have shown that several abnormally methylated genes, including Kv10.1, have been found in gastric cancer tissue and the methylation level of Kv10.1 in the samples is more than three times higher, suggesting that the hypermethylation of Kv10.1 may play a role in the occurrence and development of gastric cancer (17,35). Some have suggested that epigenetic changes in early life may alter lung cell function and lead to asthma risk (109). In mouse models, early exposure to house dust mite allergens alters DNA methylation and the expression of different genes detected, up to three consecutive generations and is associated with airway hyperresponsiveness and inflammation (109). In mice exposed to allergens, the *Kcnh1* gene was hydroxymethylated

and upregulated, indicating a susceptibility to asthma (109). In head and neck squamous cell carcinoma (HNSCC), histone acetylation, rather than DNA methylation, is hypothesized to be involved in the regulation of Kv10.1 (40).

Reversal of drug resistance. Simultaneous drug resistance to multiple drugs with different chemical structures and targets is the main obstacle to effective cancer treatment (110-114). Multidrug resistance (MDR) is an acquired resistance of microorganisms and tumor cells to chemotherapeutic drugs, which is characterized by different chemical structures and mechanisms of action. MDR is the result of overexpression of a number of protein and these protein drugs squeeze chemotherapy drugs out of cells to make their concentration lower than the effective concentration (115). MDR in cancer treatment causes tens of thousands of deaths every year and it can be endowed by a number of transporters that pump drugs out of cells. They can transport various substrates, including amino acids, peptides, ions, sugars, toxins, lipids and drugs and are associated with several serious human diseases (116). First-line therapy is usually followed by the proliferation of a small number of surviving cancer cells, which leads to the development of secondary tumors, which are insensitive to initial drugs. This may lead to tumor progression after stabilization or significant regression, because successful chemotherapy in the first stage becomes ineffective (117,118).

Platinum drugs (such as cisplatin and carboplatin) are one of the commonly used chemotherapy drugs in clinic and are widely used to treat various tumors. However, the resistance of tumor cells to platinum drugs limits their clinical application. As aforementioned, the ectopic expression of Kv10.1 is not only associated with the proliferation of tumor cells, but also associated with the phenotype of chemotherapy-resistant cells. *In vitro* experiments in ovarian cancer cells showed that the expression of Kv10.1 was associated with the chemotherapy resistance of cisplatin. Compared with the cells treated with cisplatin alone, the downregulation of Kv10.1 combined with cisplatin increased the apoptosis of ovarian cancer cells (16). Similar results were also observed in the chemotherapy-resistant glioblastoma cell line U251AR, which showed a high level of Kv10.1 (mRNA and protein). Notably, when the expression of Kv10.1 is downregulated, U251AR cells are more sensitive to chemotherapy drugs (119). Therefore, the inhibition of Kv10.1 channel can improve the response of cells to commonly used therapeutic drugs in chemotherapy.

Other mechanisms. The tumorigenic mechanism of Kv10.1 is associated with the fact that calcium-dependent calmodulin kinase-II is always activated, which can cause imbalance of cell proliferation and apoptosis even at low calcium ion concentration (120). Kv10.1 also promotes tumor progression by increasing the activity of hypoxia sensor hypoxia-inducible factor-1 alpha (HIF-1 α), stimulating the secretion of vascular endothelial growth factor (VEGF) and increasing angiogenesis under hypoxia (78). The tumor microenvironment usually shows changes in extracellular matrix (ECM). ECM products, such as type 1 collagen and fibronectin, promote tumor progression by increasing cell movement (121).

Ion channels are highly fine-tuned protein necessary for cell physiology. Therefore, the synthesis and degradation of

ion channels should be a quality control process, which is necessary for correct cell function. However, the knowledge about the synthesis and degradation of Kv10.1 is limited. A study has shown that protein degradation of Kv10.1 depends on the protein ligase cullin 7 (Cul 7) (122). Cul7 degrades Kv10.1 protein on plasma membrane through proteasome and lysosomal pathways respectively (122).

5. Signaling pathway

In the field of tumor biology, abnormal regulation of cell signaling pathway is recognized as the core mechanism driving tumor occurrence, progress and metastasis. In recent years, because of its unique molecular structure, specific expression in tumor tissues and its regulation on cell proliferation, migration and invasion, Kv10.1 has become a typical example in the study of connecting ion channels with tumor signal pathways (Fig. 2).

p53-miR34-E2F1-hEAG1 signaling pathway. Studies have shown that the tumor suppressor gene regulatory network includes the p53-miR-34 pathway and E2F1 transcription factor, which can regulate the expression of Kv10.1 (96,123). The expression of Kv10.1 may be regulated by the negative feedback of this signal pathway (124,125). This negative regulation may be achieved through two mechanisms: direct inhibition of Kv10.1 at the post-transcriptional level and negative regulation of Kv10.1 through negative feedback mechanism through the p53-miR-34-E2F1 pathway. Studies have also shown that estrogen and human papillomavirus (HPV) regulate the level of Kv10.1 protein (126). As estrogen receptor interacts with p53 (125), HPV inhibits p53 and activates E2F1 (127), suggesting that EAG1 may affect downstream gene transcription by regulating E2F1. In addition, insulin-like growth factor 1 can also increase the expression of EAG1 by activating kinase AKT1 (128). Studies have shown that AKT1 can overcome the apoptosis-inducing effect of p53 (129), suggesting that p53 and Kv10.1 can act in opposite directions along the axis of apoptosis and proliferation (130).

HIF-1 α /signal transducer and activator of transcription 3 (STAT3)/VEGF signaling pathway. The influence mechanism of silent Kv10.1 on HIF-1 α /STAT3/VEGF pathway is a multi-level and multi-step process. As a potassium channel, the abnormal expression of Kv10.1 may regulate the activity of intracellular signal transduction pathway by affecting the balance of potassium concentration inside and outside the cell. Kv10.1 can upregulate the expression of HIF-1 α by interacting with intracellular hypoxia homeostasis system. HIF-1 α is a key transcription factor to regulate the expression of VEGF, which promotes the transcription of VEGF under hypoxia, thus inducing angiogenesis (131). In osteosarcoma cells, the phosphorylation level of STAT3 decreased markedly after Kv10.1 silencing, which indicated that Kv10.1 might regulate the expression of STAT3-VEGF downstream by influencing its activation state (84,132).

PI3K/AKT signaling pathway. The PI3K/AKT signaling pathway is an important signal transduction pathway with multiple biological functions mediated by enzyme-linked

receptors in mammals. The PI3K/AKT pathway plays an important role in the occurrence and development of tumors. When stimulated by upstream signals, PI3K activates AKT, which further activates downstream signal molecules and regulates cancer cell proliferation, invasion and metastasis, angiogenesis and carbohydrate metabolism. In osteosarcoma, tumor growth and angiogenesis in osteosarcoma can be inhibited by downregulating VEGF/PI3K/AKT signaling pathway to silence Kv10.1 (133). The Kv10.1 channel and PI3K/AKT signaling pathway form a cancer-promoting signal axis through functional correlation and Kv10.1 is the key target of Nutlin-3 regulating PI3K/AKT through p53 (81). Therefore, it is of great significance to study PI3K/AKT signaling pathway and develop targeted drugs for tumor treatment.

The mitogen-activated protein kinase (MAPK) signaling pathway. The nuclear localization signal at the C-terminal of Kv10.1 potassium channel can activate the MAPK pathway to cause changes in cell morphology (134). The Kv10.1 channel plays the role of oncogene, such as promoting the proliferation of human osteosarcoma cells. The high expression of Kv10.1 in osteosarcoma cells is regulated by p38MAPK/p53 pathway (135). Inhibition of p38MAPK activation by p38 MAPK inhibitor SB203580 or short interfering RNA also decreased the level of Kv10.1 protein, but increased the level of p53 protein. In addition, the activation of p53 will lead to the growth stagnation of osteosarcoma cells and decrease the level of Kv10.1 protein, while the inactivation of p53 will promote the cell growth and increase the expression of Kv10.1 protein. This suggests that Kv10.1 may affect the proliferation of osteosarcoma cells by regulating p53 (136).

Cancer type specificities in the anti-tumor mechanisms of Kv10.1. Although Kv10.1 exhibits mechanistic similarities across various malignancies, namely, influencing tumor proliferation through hyperpolarization, exerting effects via the PI3K/AKT signaling pathway and inhibiting apoptosis (81), its downstream signaling network still demonstrates significant cancer type specificity. For instance, the expression of Kv10.1 in endometrial cancer tissues is markedly higher than that in normal endometrial tissues. Kv10.1 can be regulated by cyclin D1, p53 and other factors, thereby affecting the proliferation, migration and invasion of tumor cells (137). Kv10.1 may form different complexes or interaction networks with other molecules in various cancers, thereby influencing its function. For example, in osteosarcoma and hepatocellular carcinoma, Kv10.1 enhances tumor angiogenesis by upregulating the expression of hypoxia-inducible factor HIF-1 α and promoting the secretion of VEGF. This effect is not dependent on its ion transport function but is achieved through interaction with the cellular hypoxia homeostasis system (133,138). Different mechanisms predominate in various cancers; for instance, in breast cancer, the PI3K/AKT pathway is central, driving cell proliferation and metabolic reprogramming; in hepatocellular carcinoma, the HIF-1 α /VEGF pathway is predominant, promoting angiogenesis and tumor progression; in endometrial cancer and head and neck cancer, it may affect tumor behavior through cell cycle regulation and cytoskeleton dynamics (137-139). Drug targets are critical in drug discovery and therapy, as their identification is essential for achieving

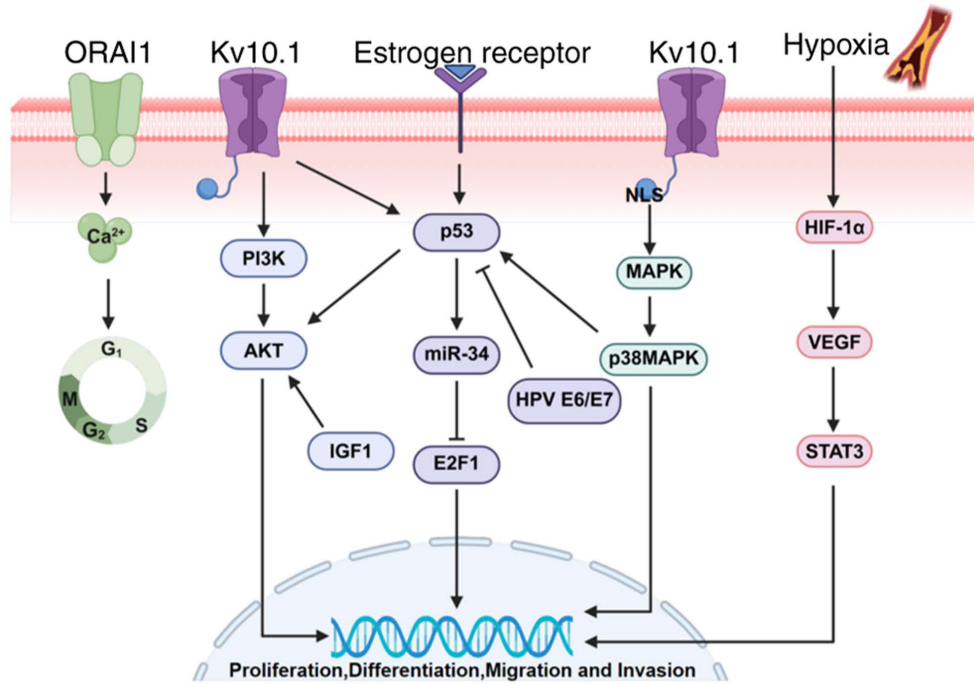


Figure 2. Kv10.1 activates multiple signaling pathways. Kv10.1 has become a typical example of the connection between ion channels and tumor signaling pathways due to its unique molecular structure, specific expression in tumor tissues and regulation of cell proliferation, migration and invasion. Kv10.1 is negatively regulated by the negative feedback mechanism of p53-miR-34-E2F1 pathway. P53 is regulated by a number of factors, such as estrogen receptor, HPV and E6/E7. Kv10.1 can upregulate the expression of HIF-1 α by interacting with the intracellular hypoxia homeostasis system and promote the transcription of VEGF under hypoxia. In osteosarcoma cells, Kv10.1 regulates the expression of STAT3-VEGF by affecting the activation of stat3-vegf downstream. Kv10.1 channel and PI3K/AKT signaling pathway form cancer promoting signal axis through functional correlation and regulate cancer cell proliferation, invasion and metastasis, angiogenesis and glucose metabolism. The NLS in the C-terminal domain of Kv10.1 activates the MAPK signaling pathway. For example, the high expression of Kv10.1 in osteosarcoma cells is regulated by p38MAPK/p53 pathway. The increase of ORAI1 in plasma membrane can regulate intracellular calcium ion and mediate cell cycle. miR, microRNA; E2F1, E2F transcription factor 1; HPV, human papillomavirus; HIF-1 α , hypoxia-inducible factor-1 alpha; VEGF, vascular endothelial growth factor; STAT3, signal transducer and activator of transcription 3; NLS, nuclear localization signal; MAPK, mitogen-activated protein kinase; ORAI1, ORAI calcium release-activated calcium modulator 1.

precision treatment (140). Therefore, Kv10.1 is considered a potential therapeutic target in various cancers.

6. Regulators and inhibitors

According to the activation mechanism of Kv10.1, its regulators are mainly divided into two categories: activators and inhibitors. Validation of Kv10.1 as a therapeutic target requires specific modulators and can be used as a basic tool for understanding channel pharmacology. Therefore, it is necessary to find more selective and effective modulators, especially inhibitors. According to the structure of Kv10.1, this section introduces its regulators and inhibitors and their applications in cancer (Table II).

PAS domain. The Kv10.1 channel is formed by assembling tetramer subunits and each subunit contains N-terminal PAS domain and C-terminal cyclic nucleotide binding homology domain. Small molecular ligands can inhibit Kv10.1 channels by binding to their PAS domains. Deleting PAS domain can cancel the inhibitory effect of chlorpromazine (141). At low voltage, chlorpromazine can inhibit mouse Eag1 by binding with PAS domain, but when the channel is open at high voltage, it will be inhibited by blocking the channel (142). Chlorpromazine, a small molecule binding agent of PAS domain, changes the interaction between PAS and CNBH domain and reduces the coupling between tetramer ring and channel hole in cells, while

the coupling between PAS and voltage sensor domain (VSD) domain has little effect. In addition, the combination of chlorpromazine and PAS domain does not change the Cole-Moore shift characteristics of Kv10.1 channel, which further indicates that chlorpromazine has no effect on the movement of VSD from deep closed state to open state (143). Chlorpromazine inhibits the proliferation of oral cancer cells by regulating the PI3K/AKT/mTOR signaling pathway and effectively suppresses tumor growth in zebrafish and mouse models. Given that chlorpromazine can act on Kv10.1, it is hypothesized that it regulates tumor proliferation by modulating the PI3K/AKT signaling pathway through Kv10.1. Based on the mechanism of action of chlorpromazine on oral cancer cells and its potential association with Kv10.1, it is reasonable to hypothesize that chlorpromazine may regulate the PI3K/AKT signaling pathway through Kv10.1, thereby affecting tumor proliferation (144).

VSD. VSD has a small molecule binding site as a gating regulator and extracellular VSD allows molecules to regulate channels without transmembrane, as with toxins. Most of the known small molecules that regulate Kv10.1 activity first cross the cell membrane and bind to the inner side of the channel. Most of these compounds are physically closed pore blocker ion permeation pathways, such as mibefradil (65,145), Purpurealidin analog 5 (146), 20 (s)-Ginsenoside Rg3 (147,148), APETx4 (149) and Corydaline (150).

Table II. Kv10.1 inhibitors.

Compound name	Binding domain	Cell line	IC ₅₀	(Refs.)
Chlorpromazine	PAS	<i>Xenopus</i> oocytes	3.7±0.7 μM	(143)
Mibefradil	VSD	293	Kd 1.3 μM, nH0.8	(65,145)
Purpurealidin analog 5	VSD	<i>Xenopus</i> oocytes	7.7±1.0 μM	(50)
20(S)-ginsenoside Rg3	VSD	<i>Xenopus</i> oocytes	1.18 μM	(1547,148)
APETx4	VSD	<i>Xenopus</i> oocytes	1.1 μM	(149)
Corydaline	VSD	HepG2	11.3±0.6 μM	(150)
Tetrandrine	Selectivity filter	CHO	70±5.2 μM	(155)
ProcyanidinB1	Selectivity filter	293	10.38±0.87 μM	(13)
Tetraethylammonium	Pore domain	<i>Xenopus</i> oocytes	1.2±0.1 μM	(156)
Amiodarone	Pore domain	293	Kd 203 nM, nH0.9	(157)
Astemizole	Pore domain	293	196 nM	(157)
		<i>Xenopus</i> oocytes	2.8±0.1 μM	
Imipramine	Pore domain	<i>Xenopus</i> oocytes	40.2±0.3 μM	(157)
Dronedarone	Pore domain	293	Kd 9 μM, nH0.9	(76)
Quinidine	Pore domain	CHO	1.4±0.1 μM	(60)
Haloperidol	Pore domain	CHO	590±121 nM	(67)
κ-Hefutoxin 1	Pore domain	-	26±2 μM	(158)
Heme	C-linker	<i>Xenopus</i> oocytes	4nM	(161)

PAS, Per-Arnt-Sim; VSD, voltage sensor domain.

Mibefradil has been reported to alter the gating of Kv10.1 by binding to the VSD (65,145). When the hyperpolarizing potential acts outside the cell, mibefradil induces obvious open inactivation, but not from inside the cell. In addition, mibefradil also inhibited the Cole-Moore shift. It is also not used as a pore size blocker because it does not compete with the known pore size blocker, quinidine. Mibefradil 1 also inhibits other potassium channels and L-type and T-type calcium channels (65).

Purpurealidin is considered to be a gating modifier, which binds to the mibefradil binding site near the voltage sensor Kv10.1. Mibefradil is a Ca²⁺ channel antagonist and a Kv10.1 door control regulator. As with purpurealin, mibefradil moves the activation curve to the left. Studies have shown that the binding site of purpurealidin on Kv10.1 overlaps with that of mibefradil (65,145,146).

Ginsenoside is a steroidal glycoside that inhibits the KCNH family at sub μM concentrations. Its function is not limited to Kv channels. Studies have shown that ginsenosides can markedly inhibit the voltage dependent activation curves of Kv10.1 and HERG channels and increase their opening probability at more negative potentials (147,148). When combined with cisplatin, it can enhance chemotherapy sensitivity and reverse drug resistance by blocking the PI3K/AKT pathway (151).

Apetx4 is a new toxin isolated from sea anemones, which can inhibit the Kv10.1 channel. APETx4 inhibits its current by binding to Kv10.1 channel and the inhibitory effect is concentration dependent. APETx4 is a gating modifier that presumably binds to the S3b-E2-S4 region (voltage sensor paddle) of Kv10.1 (149). Corydaline is a novel natural product

that selectively inhibits Kv10.1 channels, while being insensitive to other KCNH channels. Corydaline can inhibit the proliferation and migration of liver cancer cells by targeting Kv10.1. This previously unidentified new site can specifically bind to the medicinal pocket of Corydaline, providing possibilities for drug screening against diseases associated with abnormal Kv10.1 channels (150).

Selectivity filter. The selectivity filter of Kv10.1 consists of a highly conserved amino acid sequence, forming a structure similar to the 'potassium channel signature sequence' (152). Through the spatial arrangement of carbonyl oxygen atoms and threonine hydroxyl oxygen atoms, this region mimics the hydration layer mechanism of water molecules surrounding K⁺, forming a series of K⁺ binding sites (153,154).

Tetrandrine, a natural compound used in traditional Chinese medicine, can inhibit Kv10.1 in a concentration dependent manner with an IC₅₀ of 69.97±5.2 μM (155). Tetrandrine specifically inhibited Kv10.1 channel, blocked potassium outflow, induced cell cycle arrest in G₁/S phase and enhanced cisplatin sensitivity by downregulating PI3K/AKT signaling pathway (44). Tetrandrine may also have the same binding site as procyanidin B1, a natural compound present in grape seeds, which inhibits Kv10.1 in a concentration dependent manner with an IC₅₀ value of 10.4±0.9 μM (13). Procyanidin B1 directly inhibits channel activity and blocks tumor cell proliferation by hydrogen bonding to i550, t552 and q557 amino acids in the filter. Similar to tetrandrine, 100 μM procyanidin B1 did not markedly inhibit Kv7.1, Kir2.1, or HERG (13).

Pore domain. The pore region of Kv10.1 is composed of S5 and S6 transmembrane helices and intermediate pore rings, which is the core domain of ion selective permeation. In recent years, in the development of targeted drugs for the Kv10.1 channel, the pore domain has become the core target in the design of small molecular inhibitors because of its unique structural characteristics and functional importance. Tetraethylammonium is a commonly used potassium channel blocker, which can physically block ion passage (156). Tetraethylammonium can combine with aromatic amino acid residues on the inner wall of the channel, which hinders the selective filtration function of potassium ions, thus inhibiting the channel current. The mechanism of quinidine and tetraethylammonium is similar and does not depend on the voltage gating state of the channel, so it belongs to non-competitive inhibition (60). The blocking effect of quinidine on Kv10.1 channel is concentration-dependent and the inhibitory effect is more obvious at high concentration. Astemizole competitively binds to Kv10.1 channel with tricyclic antidepressants imipramine and tetraethylammonium (157). The blocking effect of imipramine is voltage-dependent and can be antagonized by intracellular tetraethylammonium. Amiodarone and dronedarone both act through the pore region, but their blocking effects on Kv10.1 are different and dronedarone cannot inhibit the Cole-Molar displacement characteristics of Kv10.1 channel (76). Haloperidol also inhibited the Kv10.1 channel through the pore region and the inhibition was associated with the drug concentration and membrane potential (67). κ -Hefutuxin 1, as the first peptide inhibitor of Kv10.1 channel, inhibits its activity by binding to Kv10.1 channel (158). As the first peptide inhibitor of Kv10.1 channel, κ -hefutuxin 1 inhibits its activity by binding to Kv10.1 channel and may act on or near the pore area of Kv10.1 channel. Channel residues met397 and asp398 may be the anchors to stabilize its binding (158).

Studies have investigated the effects of Haloperidol on the PI3K/AKT signaling pathway in PC12 cells (a neuroblastoma cell line) (159). The research found that Haloperidol can induce the nuclear translocation of PI3K, generating phosphatidylinositol-3,4,5-trisphosphate (PIP3) in the nucleus, effectively inhibiting the phosphorylation of AKT, leading to a decrease in AKT activity. Furthermore, Haloperidol can effectively suppress the expression of Kv10.1 (159,160). Although no studies have yet explored the direct relationship between these factors, it is possible that Haloperidol exerts its anticancer effects by indirectly influencing the PI3K/AKT signaling pathway through the inhibition of Kv10.1 expression (159).

Other binding sites. At present, the mechanism of action of Kv10.1 inhibitors is complex and diverse and the binding sites of some inhibitors are not yet clear. Heme is an endogenous regulator of the Kv10.1 channel, which inhibits its activity by binding to the CxHxH motif in the C-Linker region (161). The aforementioned results are listed in Table II.

Inhibitors with clear binding sites are beneficial for the development of new drugs for treating diseases, while inhibitors with unknown binding sites reveal the complexity and diversity of channel regulation. Future research should further strengthen the exploration of the mechanism of action of inhibitors with unknown binding sites and reveal the molecular mysteries of their interaction with Kv10.1 channels.

Meanwhile, based on the mechanism of action of inhibitors, more efficient and specific Kv10.1 inhibitors have been developed, providing new strategies and means for the treatment of related diseases.

7. Clinical status

From a clinical perspective, Kv10.1 is highly expressed in various malignant tumor tissues, markedly higher than in normal tissues, thus possessing the potential to become a diagnostic marker for tumors. Currently, little is known about the mechanism of Kv10.1 in tumorigenesis and most data on Kv10.1 inhibitors are derived from *in vitro* cell lines or simple animal models. This lack of accurate targeting in drug development makes it difficult to design highly effective and specific targeted drugs. The metabolic process of drugs in the body, potential toxic effects and how to effectively deliver drugs to tumor tissues have become major obstacles in the clinical translation of Kv10.1 targeted drugs.

The clinical value of Kv10.1 as a diagnostic marker. Kv10.1 is expressed in a variety of malignant tumors, including clear cell renal cell carcinoma, breast cancer, cervical cancer, gastric cancer, colon cancer and cervical intraepithelial neoplasia (14,42,43,162-165). According to clinical data analysis reported in relevant literature, a total of 68 clinical samples were tested in the study of esophageal squamous cell carcinoma. The results showed that 51 cases showed positive expression, with a positive rate of 75%. Further analysis showed that the positive expression was closely associated with the depth of tumor infiltration and the survival time of positive patients was generally shorter (166). In clinical data of ovarian cancer, there were 336 samples, of which 249 showed positive results, with a positivity rate of 65%. The expression of Kv10.1 is associated with tumor size, differentiation degree, staging and metastasis (167). In the clinical data of head and neck squamous cell carcinoma, there were a total of 54 clinical samples, of which 45 samples showed positive results, with a positive rate as high as 83.00%. It is worth noting that Kv10.1 plays a key role in this head and neck squamous cell carcinoma and has been proven to be a tumor marker and potential therapeutic target. The expression of Kv10.1 is associated with important clinical pathological features such as tumor size, differentiation degree, staging and metastasis (40).

A research group has conducted in-depth predictive analysis of the clinical correlation between cancer-related genes and renal cell carcinoma cell lines using the Cancer Genome Atlas database. The research results indicate that Kv10.1 is a key biomarker and potential therapeutic target in clear cell renal cell carcinoma (ccRCC) (162). The widespread abnormal expression of Kv10.1 in numerous tumors suggests that it may be intrinsically associated with the occurrence and development of tumors and has potential value as a tumor marker for tumor screening and diagnosis. However, most of the data on Kv10.1 inhibitors to date have been derived from *in vitro* cell lines or simple animal models and there is no systematic patient-derived xenografts (PDX) model data to clearly demonstrate the quantitative indicators of tumor inhibition rate and survival extension effect of representative inhibitors. Due to the lack of such data, it is difficult to directly support

the conclusion that Kv10.1 inhibitors have significant clinical translational value in PDX models. Kv10.1 is highly expressed in brain tissue and involved in regulating neuronal excitability, neurotransmitter release and synaptic plasticity. When treating brain tumors, Kv10.1 inhibitors need to penetrate the blood-brain barrier (BBB) to target tumor cells. At this time, increased permeability of the BBB (such as through hyperosmotic solutions or nanocarrier technology) may facilitate drug entry into the brain, but it also increases the risk of exposure to normal brain tissue. When treating non-brain tumors, the BBB can prevent Kv10.1 inhibitors from entering the brain tissue, thereby avoiding neurotoxicity. Studies have shown that knocking out Kv10.1 did not result in significant abnormalities in multiple behavioral tests in mice and its function may have a redundant compensation mechanism with other channels. Therefore, Kv10.1 inhibitors have less interference with normal tissue, especially brain tissue function (67). Therefore, at present, it can only be concluded that Kv10.1 is a potential therapeutic target.

Bottleneck of clinical transformation of targeted drugs. The BBB is a selective barrier between the central nervous system and the circulatory system. Its unique structural characteristics, such as tight junction, stability of cytoskeleton and active efflux of transmembrane proteins, together constitute the physical and chemical barrier for drugs to enter the brain tissue. Kv10.1 is highly expressed in brain tumors (such as glioma), but the existence of the blood-brain barrier makes it difficult for most drugs to penetrate effectively, resulting in low treatment efficiency of brain tumors. Even if drugs can break through the blood-brain barrier for a short time, its action time is often not enough to play a lasting effect (168,169).

The molecular weight of peptide inhibitors is relatively small and the structure is relatively simple. It is generally considered to have low immunogenicity potential and it is easy to be degraded and excreted in the body, reducing the contact time with the immune system, thus reducing the immunogenicity. In addition, in the production process of peptide inhibitors, some impurities may be introduced, such as pollutants, solvents, enzymes, impurities formed by amino acid side chain modification, truncation, duplication, oxidation, insertion or deletion in the active pharmaceutical ingredient sequence. These impurities may have potential immunogenicity and can trigger the immune response of the body, thus affecting the safety and effectiveness of drugs.

Kv10.1 combination therapy reverses drug resistance. Kv10.1 inhibitors combined with chemotherapeutic drugs can act on tumor cells simultaneously through different mechanisms to produce synergistic killing effect. This combined strategy is expected to improve the therapeutic effect and prolong the survival time of patients. For example, Kv10.1 can participate in metabolic adaptation to cancer cells by regulating mitochondrial dynamics and inhibit Kv10.1 expression or function, resulting in mitochondrial fragmentation, increased reactive oxygen species and increased autophagy. Kv10.1 endogenous overexpression cells were also more sensitive to mitochondrial metabolic inhibitors than low expression cells, indicating that they were more dependent on mitochondrial function. Therefore, the combined treatment of Kv10.1 functional

monoclonal antibody and mitochondrial metabolism inhibitor leads to the enhanced efficacy of the inhibitor (31). Kv10.1 specific scFv was fused with soluble TNF-related apoptosis inducing ligand. The combination of ligands and different chemotherapeutic drugs can overcome drug resistance and selectively induce apoptosis (170).

8. Summary

The Kv10.1 channel is overexpressed in 70% of tumor cells and has carcinogenic characteristics, regulates cell proliferation, survival, angiogenesis, migration and invasion and is associated with the formation and progress of invasive tumors. However, although Kv10.1, as a potential target for cancer treatment, has been confirmed *in vivo* and *in vitro* experiments and some carcinogenesis mechanisms have been proved, the specific mechanism of Kv10.1 participating in the occurrence and development of tumors is not completely clear.

However, several specific Kv10.1 inhibitors have been used in animal models and achieved good results, which can inhibit the proliferation of tumor cells, induce tumor cell apoptosis, preserve normal cells and make cancer cells more sensitive to chemotherapy drugs. If these specific Kv10.1 inhibitors are combined with the current routine clinical treatment, it will be a potential therapeutic strategy to make them really useful tumor targets.

Acknowledgements

Not applicable.

Funding

The present study was supported by the National Key Research and Development Plan of China (grant no. 2023YFF1205500), Central Government Guides Local Funds for Science and Technology Development for Hebei Province (grant nos. 246Z2701G and 254Z2702G), Science Research Project of Hebei Education Department (grant no. QN2025020) and Shijiazhuang Science and Technology Cooperation Special Project (grant no. SJZZXB24007).

Availability of data and materials

Not applicable.

Authors' contributions

Investigation was by YX, SD, BW, JS, LD, LL, XW and YC. Writing the original draft was by YX and WH. Data curation was by YX, SD, BW, JS, LD, XW and YC. Conceptualization was by YX. Writing, reviewing and editing was by WH, LL, HA, XW and YC. Methodology was by SD. Resources were provided by BW, HA, XW and YC. Supervision was by HA, XW and YC. Data authentication is not applicable. All authors read and approved the final manuscript.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

References

- Pardo LA and Stühmer W: Eag1: An emerging oncological target. *Cancer Res* 68: 1611-1613, 2008.
- Kaplan WD and Trout WE III: The behavior of four neurological mutants of *Drosophila*. *Genetics* 61: 399-409, 1969.
- Thi Hong Van N and Hyun Nam J: Intermediate conductance calcium-activated potassium channel (KCa3.1) in cancer: Emerging roles and therapeutic potentials. *Biochem Pharmacol* 230: 116573, 2024.
- Luis E, Anaya-Hernández A, León-Sánchez P and Durán-Pastén ML: The Kv10.1 Channel: A promising target in cancer. *Int J Mol Sci* 23: 8458, 2022.
- Cheng S, Jiang D, Lan X, Liu K and Fan C: Voltage-gated potassium channel 1.3: A promising molecular target in multiple disease therapy. *Biomed Pharmacother* 175: 116651, 2024.
- He S, Moutaoufik MT, Islam S, Persad A, Wu A, Aly KA, Fonge H, Babu M and Cayabyab FS: HERG channel and cancer: A mechanistic review of carcinogenic processes and therapeutic potential. *Biochim Biophys Acta Rev Cancer* 1873: 188355, 2020.
- Gao S, Wang W, Ye W and Wang K: The mechanism study of Eag1 potassium channel in gastric cancer. *Transl Cancer Res* 11: 3827-3840, 2022.
- Chen J, Xuan Z, Song W, Han W, Chen H, Du Y, Xie H, Zhao Y, Zheng S and Song P: EAG1 enhances hepatocellular carcinoma proliferation by modulating SKP2 and metastasis through pseudopod formation. *Oncogene* 40: 163-176, 2021.
- Chávez-López MG, Zúñiga-García V, Hernández-Gallegos E, Vera E, Chasiquiza-Anchahuña CA, Viteri-Yáñez M, Sanchez-Ramos J, Garrido E and Camacho J: The combination astemizole-gefitinib as a potential therapy for human lung cancer. *Onco Targets Ther* 10: 5795-5803, 2017.
- Pardo LA, Contreras-Jurado C, Zientkowska M, Alves F and Stühmer W: Role of voltage-gated potassium channels in cancer. *J Membr Biol* 205: 115-124, 2005.
- Cázares-Ordoñez V and Pardo LA: Kv10.1 potassium channel: From the brain to the tumors. *Biochem Cell Biol* 95: 531-536, 2017.
- Badaoui M, Mimsy-Julienne C, Saby C, Van Gulick L, Peretti M, Jeannesson P, Morjani H and Ouadid-Ahidouch H: Collagen type 1 promotes survival of human breast cancer cells by overexpressing Kv10.1 potassium and Orail1 calcium channels through DDR1-dependent pathway. *Oncotarget* 9: 24653-24671, 2017.
- Na W, Ma B, Shi S, Chen Y, Zhang H, Zhan Y and An H: Procyanidin B1, a novel and specific inhibitor of Kv10.1 channel, suppresses the evolution of hepatoma. *Biochem Pharmacol* 178: 114089, 2020.
- Ding XW, Yan JJ, An P, Lü P and Luo HS: Aberrant expression of ether à go-go potassium channel in colorectal cancer patients and cell lines. *World J Gastroenterol* 13: 1257-1261, 2007.
- de Guadalupe Chávez-López M, Hernández-Gallegos E, Vázquez-Sánchez AY, Gariglio P and Camacho J: Antiproliferative and proapoptotic effects of astemizole on cervical cancer cells. *Int J Gynecol Cancer* 24: 824-828, 2014.
- Hui C, Lan Z, Yue-Li L, Li-Lin H and Li-Lin H: Knockdown of Eag1 expression by RNA interference increases chemosensitivity to cisplatin in ovarian cancer cells. *Reprod Sci* 22: 1618-1626, 2015.
- Guy HR, Durell SR, Warmke J, Drysdale R and Ganetzky B: Similarities in amino acid sequences of *Drosophila* eag and cyclic nucleotide-gated channels. *Science* 254: 730, 1991.
- Occhiodoro T, Bernheim L, Liu JH, Bijlenga P, Sinnreich M, Bader CR and Fischer-Lougheed J: Cloning of a human Ether-a-Go-go potassium channel expressed in myoblasts at the onset of fusion. *FEBS Lett* 434: 177-182, 1998.
- Whicher JR and MacKinnon R: Structure of the Voltage-Gated K⁺ Channel Eag1 reveals an alternative voltage sensing mechanism. *Science* 353: 664-669, 2016.
- Barros F, de la Peña P, Domínguez P, Sierra LM and Pardo LA: The EAG Voltage-dependent K⁺ channel subfamily: Similarities and differences in structural organization and gating. *Front Pharmacol* 11: 411, 2020.
- Yellen G: The voltage-gated potassium channels and their relatives. *Nature* 419: 35-42, 2002.
- Morais Cabral JH, Lee A, Cohen SL, Chait BT, Li M and Mackinnon R: Crystal structure and functional analysis of the HERG potassium channel N terminus: A eukaryotic PAS domain. *Cell* 95: 649-655, 1998.
- Li S, Yang F, Sun D, Zhang Y, Zhang M, Liu S, Zhou P, Shi C, Zhang L and Tian C: Cryo-EM structure of the hyperpolarization-activated inwardly rectifying potassium channel KATI from *Arabidopsis*. *Cell Res* 30: 1049-1052, 2020.
- Ju M and Wray D: Molecular regions responsible for differences in activation between heag channels. *Biochem Biophys Res Commun* 342: 1088-1097, 2006.
- Terlau H, Heinemann SH, Stühmer W, Pongs O and Ludwig J: Amino terminal-dependent gating of the potassium channel rat eag is compensated by a mutation in the S4 segment. *J Physiol* 502: 537-543, 1997.
- Ludwig J, Terlau H, Wunder F, Brüggemann A, Pardo LA, Marquardt A, Stühmer W and Pongs O: Functional expression of a rat homologue of the voltage gated ether à go-go potassium channel reveals differences in selectivity and activation kinetics between the *Drosophila* channel and its mammalian counterpart. *EMBO J* 13: 4451-4458, 1994.
- Lorinczi E, Gómez-Posada JC, de la Peña P, Tomczak AP, Fernández-Trillo J, Leipscher U, Stühmer W, Barros F and Pardo LA: Voltage-dependent gating of KCNH potassium channels lacking a covalent link between voltage-sensing and pore domains. *Nat Commun* 6: 6672, 2015.
- Tomczak AP, Fernández-Trillo J, Bharill S, Papp F, Panyi G, Stühmer W, Isacoff EY and Pardo LA: A new mechanism of voltage-dependent gating exposed by KV10.1 channels interrupted between voltage sensor and pore. *J Gen Physiol* 149: 577-593, 2017.
- Fukai R, Saito H, Tsurusaki Y, Sakai Y, Haginoya K, Takahashi K, Hubshman MW, Okamoto N, Nakashima M, Tanaka F, *et al*: De novo KCNH1 mutations in four patients with syndromic developmental delay, hypotonia and seizures. *J Hum Genet* 61: 381-387, 2016.
- Lin TF, Lin IW, Chen SC, Wu HH, Yang CS, Fang HY, Chiu MM and Jeng CJ: The Subfamily-specific Assembly of Eag and ErgK⁺ Channels Is Determined by both the amino and the carboxyl recognition domains. *J Biol Chem* 289: 22815-22834, 2014.
- Schönherr R, Lober K and Heinemann SH: Inhibition of human ether à go-go potassium channels by Ca(2+)/calmodulin. *Embo J* 19: 3263-3271, 2000.
- Warmke J, Drysdale R and Ganetzky B: A distinct potassium channel polypeptide encoded by the *Drosophila* eag locus. *Science* 252: 1560-1562, 1991.
- Napp J, Monje F, Stühmer W and Pardo LA: Glycosylation of Eag1 (Kv10.1) potassium channels: Intracellular trafficking and functional consequences. *J Biol Chem* 280: 29506-29512, 2005.
- Pardo LA, Camino D, Sánchez A, Alves F, Brüggemann A, Beckh S and Stühmer W: Oncogenic potential of EAG K(+) channels. *EMBO J* 18: 5540-5547, 1999.
- Mortensen LS, Schmidt H, Farsi Z, Barrantes-Freer A, Rubio ME, Ufartes R, Eilers J, Sakaba T, Stühmer S, Tytgat J and Pardo LA: KV 10.1 opposes activity-dependent increase in Ca²⁺ influx into the presynaptic terminal of the parallel fibre-Purkinje cell synapse. *J Physiol* 593: 181-196, 2015.
- Hernández-Reséndiz I, Pacheu-Grau D, Sánchez A and Pardo LA: Inhibition of Kv10.1 channels sensitizes mitochondria of cancer cells to antimetabolic agents. *Cancers (Basel)* 12: 920, 2020.
- Gubič Š, Toplak Ž, Shi X, Dernovšek J, Hendrickx LA, Pinheiro-Junior EL, Peigneur S, Tytgat J, Pardo LA, Peterlin Mašič L and Tomašič T: New diarylamine KV10.1 inhibitors and their anticancer potential. *Pharmaceutics* 14: 1963, 2022.
- Girault A, Peretti M, Badaoui M, Hémon A, Morjani H and Ouadid-Ahidouch H: The N and C-termini of SPCA2 regulate differently Kv10.1 function: Role in the collagen 1-induced breast cancer cell survival. *Am J Cancer Res* 11: 251-263, 2021.
- Chávez-López MG, Zúñiga-García V, Pérez-Carreón JJ, Avalos-Fuentes A, Escobar Y and Camacho J: Eag1 channels as potential early-stage biomarkers of hepatocellular carcinoma. *Biologies* 10: 139-148, 2016.

40. Menéndez ST, Villaronga MA, Rodrigo JP, Alvarez-Teijeiro S, García-Carracedo D, Urdinguio RG, Fraga MF, Pardo LA, Vilorio CG, Suárez C and García-Pedrero JM: Frequent aberrant expression of the human ether à go-go (hEAG1) potassium channel in head and neck cancer: Pathobiological mechanisms and clinical implications. *J Mol Med (Berl)* 90: 1173-1184, 2012.
41. Kang C, Li X, Yang X, Cheng X, Zhang D and Wei X: Voltage-gated potassium channels associated with head and neck cancer. *Biochim Biophys Acta Rev Cancer* 1880: 189340, 2025.
42. Canella R, Brugnoli F, Gallo M, Keillor JW, Terrazzan A, Ferrari E, Grassilli S, Gates EWJ, Volinia S, Bertagnolo V, *et al*: A multidisciplinary approach establishes a link between transglutaminase 2 and the Kv10.1 Voltage-dependent K⁺ channel in breast cancer. *Cancers (Basel)* 15: 178, 2022.
43. Ding XW, Luo HS, Jin X, Yan JJ and Ai YW: Aberrant expression of Eag1 potassium channels in gastric cancer patients and cell lines. *Med Oncol* 24: 345-350, 2007.
44. Wang BC and Liu WX: Study on the effect and mechanism of tetrandrine combined with cisplatin in inhibiting the growth of cervical cancer cells. *Modern Oncol* 32: 2364-2370, 2024.
45. Qiu S, Wang Q, Jiang H and Feng L: Immunohistochemistry staining of Eag1 and p16/Ki-67 can help improve the management of patients with cervical intraepithelial Neoplasia after cold knife conversion. *Diagn Pathol* 19: 97, 2024.
46. Ortiz CS, Montante-Montes D, Saqui-Salces M, Hinojosa LM, Gamboa-Dominguez A, Hernández-Gallegos E, Martínez-Benítez B, Del Rosario Solís-Pancoatl M, García-Villa E, Ramírez A, *et al*: Eag1 potassium channels as markers of cervical dysplasia. *Oncol Rep* 26: 1377-1383, 2011.
47. Hartung F, Krüwel T, Shi X, Pfizenmaier K, Kontermann R, Chames P, Alves F and Pardo LA: A Novel Anti-Kv10.1 nanobody fused to Single-Chain TRAIL enhances apoptosis induction in cancer cells. *Front Pharmacol* 11: 686, 2020.
48. Bernal-Ramos G, Hernández-Gallegos E, Vera E, Chávez-López MG, Zúñiga-García V, Sánchez-Pérez Y, Garrido E and Camacho J: Astemizole inhibits cell proliferation in human prostate tumorigenic cells expressing ether à go-go-1 potassium channels. *Cell Mol Biol (Noisy-le-grand)* 63: 11-13, 2017.
49. Söğüt F, Çömelekoğlu Ü, Dervişoğlu H, Eroğlu P, Yalin S and Yılmaz NŞ: Effect of imipramine on ether à go-go potassium channel (Kv1.10) expression in DU145 prostate cancer cells. *Andrologia* 54: e14291, 2022.
50. Guasti L, Crociani O, Redaelli E, Pillozzi S, Polvani S, Masselli M, Mello T, Galli A, Amedei A, Wymore RS, *et al*: Identification of a posttranslational mechanism for the regulation of hERG1 K⁺ channel expression and hERG1 current density in tumor cells. *Mol Cell Biol* 28: 5043-5060, 2008.
51. Gavrilova-Ruch O, Schönherr K, Gessner G, Schönherr R, Klapperstück T, Wohlrab W and Heinemann SH: Effects of imipramine on ion channels and proliferation of IGR1 melanoma cells. *J Membr Biol* 188: 137-149, 2002.
52. Meyer R, Schönherr R, Gavrilova-Ruch O, Wohlrab W and Heinemann SH: Identification of ether à go-go and calcium-activated potassium channels in human melanoma cells. *J Membr Biol* 171: 107-115, 1999.
53. Ma B, Shi S, Guo W, Zhang H, Zhao Z and An H: Liensinine, a novel and Food-derived compound, exerts potent antihepatoma efficacy via inhibiting the Kv10.1 channel. *J Agric Food Chem* 72: 4689-4702, 2024.
54. Agarwal JR, Griesinger F, Stühmer W and Pardo LA: The potassium channel Ether à go-go is a novel prognostic factor with functional relevance in acute myeloid leukemia. *Mol Cancer* 9: 18, 2010.
55. Mello de Queiroz F, Suarez-Kurtz G, Stühmer W and Pardo LA: Ether à go-go potassium channel expression in soft tissue sarcoma patients. *Mol Cancer* 5: 42, 2006.
56. Mészáros B, Csoti A, Szanto TG, Telek A, Kovács K, Toth A, Volkó J and Panyi G: The hEag1 K⁺ channel inhibitor astemizole stimulates Ca²⁺ Deposition in SaOS-2 and MG-63 osteosarcoma cultures. *Int J Mol Sci* 23: 10533, 2022.
57. Cole KS and Moore JW: Liquid junction and membrane potentials of the squid giant axon. *J Gen Physiol* 43: 971-980, 1960.
58. Cole KS and Moore JW: Ionic current measurements in the squid giant axon membrane. *J Gen Physiol* 44: 123-167, 1960.
59. Cole KS and Moore JW: Potassium ion current in the squid giant axon: Dynamic characteristic. *Biophys J* 1: 1-14, 1960.
60. Schönherr R, Gessner G, Löber K and Heinemann SH: Functional distinction of human EAG1 and EAG2 potassium channels. *FEBS Lett* 514: 204-208, 2002.
61. Terlau H, Ludwig J, Steffan R, Pongs O, Stühmer W and Heinemann SH: Extracellular Mg²⁺ regulates activation of rat eag potassium channel. *Pflugers Arch* 432: 301-312, 1996.
62. Kazmierczak M, Zhang X, Chen B, Mulkey DK, Shi Y, Wagner PG, Pivaroff-Ward K, Sassic JK, Bayliss DA and Jegla T: External pH modulates EAG superfamily K⁺ channels through EAG-specific acidic residues in the voltage sensor. *J Gen Physiol* 141: 721-735, 2013.
63. Brelidze TI, Carlson AE and Zagotta WN: Absence of direct cyclic nucleotide modulation of mEAG1 and hERG1 channels revealed with fluorescence and electrophysiological methods. *J Biol Chem* 284: 27989-27997, 2009.
64. Barriga-Montoya C, Huanosta-Gutiérrez A, Reyes-Vaca A, Hernández-Cruz A, Picones A and Gómez-Lagunas F: Inhibition of the K⁺ conductance and Cole-Moore shift of the oncogenic Kv10.1 channel by amiodarone. *Pflugers Arch* 470: 491-503, 2018.
65. Gómez-Lagunas F and Barriga-Montoya C: Mibefradil inhibition of the Cole-Moore shift and K⁺-conductance of the tumor-related Kv10.1 channel. *Channels (Austin)* 11: 373-376, 2017.
66. Martin S, Lino de Oliveira C, Mello de Queiroz F, Pardo LA, Stühmer W and Del Bel E: Eag1 potassium channel immunohistochemistry in the CNS of adult rat and selected regions of human brain. *Neuroscience* 155: 833-844, 2008.
67. Ufartes R, Schneider T, Mortensen LS, de Juan Romero C, Hentrich K, Knoetgen H, Beilinson V, Moebius W, Tarabykin V, Alves F, *et al*: Behavioural and functional characterization of Kv10.1 (Eag1) knockout mice. *Hum Mol Genet* 22: 2247-2262, 2013.
68. Kohl T, Lörinczi E, Pardo LA and Stühmer W: Rapid internalization of the oncogenic K⁺ channel K(V)10.1. *PLoS One* 6: e26329, 2011.
69. Pardo LA and Stühmer W: The roles of K(+) channels in cancer. *Nat Rev Cancer* 14: 39-48, 2014.
70. Del-Río-Ibáñez N, Granda-Díaz R, Rodrigo JP, Menéndez ST and García-Pedrero JM: Ion Channel dysregulation in head and neck cancers: Perspectives for clinical application. *Rev Physiol Biochem Pharmacol* 181: 375-427, 2021.
71. Li Z, Zhu K, Gong X, Vasilescu S, Sun Y, Hong K, Li H, Li L and Shan Y: Inducing polyclonal Eag1-Specific antibodies by vaccination with a linear epitope immunogen and its relation to breast tumorigenesis. *Pathol Oncol Res* 23: 761-767, 2017.
72. García-Quiroz J, González-González ME, Díaz L, Ordaz-Rosado D, Segovia-Mendoza M, Prado-García H, Larrea F and García-Becerra R: Astemizole, an Inhibitor of Ether-à go-go-1 potassium channel, increases the activity of the tyrosine kinase inhibitor gefitinib in breast cancer cells. *Rev Invest Clin* 71: 186-194, 2019.
73. Haas B, Roth I, Säcker L, Wos-Maganga M, Beltzig L and Kaina B: Apoptotic and senolytic effects of hERG/Eag1 channel blockers in combination with temozolomide in human glioblastoma cells. *Naunyn Schmiedebergs Arch Pharmacol* 398: 12267-12278, 2025.
74. Rashno Z, Rismani E, Ghasemi JB, Mansouri M, Shabani M, Afsar A, Dabiri S, Rezaei Makhouri F, Hatami A and Harandi MF: Design of ion channel blocking, toxin-like Kunitz inhibitor peptides from the tapeworm, *Echinococcus granulosus*, with potential anti-cancer activity. *Sci Rep* 13: 11465, 2023.
75. Toplak Ž, Hendrickx LA, Gubič Š, Možina Š, Žegura B, Štern A, Novak M, Shi X, Peigneur S, Tytgat J, *et al*: 3D Pharmacophore-based discovery of novel KV10.1 inhibitors with antiproliferative activity. *Cancers (Basel)* 13: 1244, 2021.
76. Meléndez TA, Huanosta-Gutiérrez A, Barriga-Montoya C, González-Andrade M and Gómez-Lagunas F: Dronedarone blockage of the tumor-related Kv10.1 channel: A comparison with amiodarone. *Pflugers Arch* 472: 75-87, 2020.
77. Brüggemann A, Stühmer W and Pardo LA: Mitosis-promoting factor-mediated suppression of a cloned delayed rectifier potassium channel expressed in *Xenopus* oocytes. *Proc Natl Acad Sci USA* 94: 537-542, 1997.
78. Downie BR, Sánchez A, Knötgen H, Contreras-Jurado C, Gymnopoulos M, Weber C, Stühmer W and Pardo LA: Eag1 expression interferes with hypoxia homeostasis and induces angiogenesis in tumors. *J Biol Chem* 283: 36234-36240, 2008.
79. Gómez-Varela D, Zwick-Wallasch E, Knötgen H, Sánchez A, Hettmann T, Ossipov D, Weseloh R, Contreras-Jurado C, Rothe M, Stühmer W and Pardo LA: Monoclonal antibody blockade of the human Eag1 potassium channel function exerts antitumor activity. *Cancer Res* 67: 7343-7349, 2007.

80. Sales TT, Resende FF, Chaves NL, Titze-De-Almeida SS, Bão SN, Brettas ML and Titze-De-Almeida R: Suppression of the Eag1 potassium channel sensitizes glioblastoma cells to injury caused by temozolomide. *Oncol Lett* 12: 2581-2589, 2016.
81. Wang X, Chen Y, Liu H, Guo S, Hu Y, Zhan Y and An H: A novel anti-cancer mechanism of Nutlin-3 through downregulation of Eag1 channel and PI3K/AKT pathway. *Biochem Biophys Res Commun* 517: 445-451, 2019.
82. García-Quiroz J, García-Becerra R, Santos-Martínez N, Barrera D, Ordaz-Rosado D, Avila E, Halhali A, Villanueva O, Ibarra-Sánchez MJ, Esparza-López J, *et al*: In vivo dual targeting of the oncogenic Ether-à-go-go-I potassium channel by calcitriol and astemizole results in enhanced antineoplastic effects in breast tumors. *BMC Cancer* 14: 745, 2014.
83. Shen C, Kuang Y, Xu S, Li R, Wang J, Zou Y, Wang C, Xu S, Liang L, Lin C, *et al*: Nitidine chloride inhibits fibroblast like synoviocytes-mediated rheumatoid synovial inflammation and joint destruction by targeting KCNH1. *Int Immunopharmacol* 101: 108273, 2021.
84. Chen ZD, Liu QJ, Zeng WR, Wu XY, Lin B and Wu J: Expression of Ether à go-go 1 and its molecular mechanism of regulating the malignant phenotype of osteosarcoma. *Zhonghua Zhong Liu Za Zhi* 38: 818-825, 2016 (In Chinese).
85. Valdés-Abadía B, Morán-Zendejas R, Rangel-Flores JM and Rodríguez-Menchaca AA: Chloroquine inhibits tumor-related Kv10.1 channel and decreases migration of MDA-MB-231 breast cancer cells in vitro. *Eur J Pharmacol* 855: 262-266, 2019.
86. Emigh Cortez AM, DeMarco KR, Furutani K, Bekker S, Sack JT, Wulff H, Clancy CE, Vorobyov I and Yarov-Yarovsky V: Structural modeling of hERG channel-drug interactions using Rosetta. *Front Pharmacol* 14: 1244166, 2023.
87. Abdelaziz R, Tomczak AP, Neef A and Pardo LA: Revealing a hidden conducting state by manipulating the intracellular domains in KV10.1 exposes the coupling between two gating mechanisms. *Elife* 12: RP91420, 2024.
88. Wei AD, Burgraff NJ, Oliveira LM, Moreira TS and Ramirez JM: Fentanyl blockade of K⁺ channels contributes to wooden chest syndrome. *J Physiol* 604: 582-604, 2026.
89. Wang ZJ, Ghorbani M, Chen X, Tiwari PB, Klauda JB and Brelidze TI: Molecular mechanism of EAG1 channel inhibition by imipramine binding to the PAS domain. *J Biol Chem* 299: 105391, 2023.
90. Camacho J: Ether à go-go potassium channels and cancer. *Cancer Lett* 233: 1-9, 2006.
91. Herrmann S, Ninkovic M, Kohl T, Lörinczi É and Pardo LA: Cortactin controls surface expression of the voltage-gated potassium channel K(V)10.1. *J Biol Chem* 287: 44151-44163, 2012.
92. Sánchez A, Urrego D and Pardo LA: Cyclic expression of the voltage-gated potassium channel KV10.1 promotes disassembly of the primary cilium. *EMBO Rep* 17: 708-723, 2016.
93. Urrego D, Sánchez A, Tomczak AP and Pardo LA: The electric fence to cell-cycle progression: Do local changes in membrane potential facilitate disassembly of the primary cilium?: Timely and localized expression of a potassium channel may set the conditions that allow retraction of the primary cilium. *Bioessays* 39: 1600190, 2017.
94. Martínez N, Boire A and Deangelis LM: Molecular interactions in the development of brain metastases. *Int J Mol Sci* 14: 17157-17167, 2013.
95. Rosendo-Pineda MJ, Moreno CM and Vaca L: Role of ion channels during cell division. *Cell Calcium* 91: 102258, 2020.
96. Urrego D, Movsisyan N, Ufartes R and Pardo LA: Periodic expression of Kv10.1 driven by pRb/E2F1 contributes to G2/M progression of cancer and non-transformed cells. *Cell Cycle* 15: 799-811, 2016.
97. Movsisyan N and Pardo LA: Kv10.1 regulates microtubule dynamics during mitosis. *Cancers (Basel)* 12: 2409, 2020.
98. Peretti M, Badaoui M, Girault A, Van Gulick L, Mabile MP, Tebbakha R, Sevestre H, Morjani H and Ouadid-Ahidouch H: Original association of ion transporters mediates the ECM-induced breast cancer cell survival: Kv10.1-Orail-SPCA2 partnership. *Sci Rep* 9: 1175, 2019.
99. Ouadid-Ahidouch H, Le Bourhis X, Roudbaraki M, Toillon RA, Delcourt P and Prevarskaya N: Changes in the K⁺ current-density of MCF-7 cells during progression through the cell cycle: Possible involvement of a h-ether.a-gogo K⁺ channel. *Recept Channels* 7: 345-356, 2001.
100. Ouadid-Ahidouch H and Ahidouch A: K⁺ channel expression in human breast cancer cells: Involvement in cell cycle regulation and carcinogenesis. *J Membr Biol* 221: 1-6, 2008.
101. Stansfeld CE, Röper J, Ludwig J, Weseloh RM, Marsh SJ, Brown DA and Pongs O: Elevation of intracellular calcium by muscarinic receptor activation induces a block of voltage-activated rat ether-à-go-go channels in a stably transfected cell line. *Proc Natl Acad Sci USA* 93: 9910-9914, 1996.
102. Bronk P, Kuklin EA, Gorur-Shandilya S, Liu C, Wiggin TD, Reed ML, Marder E and Griffith LC: Regulation of Eag by Ca²⁺/calmodulin controls presynaptic excitability in *Drosophila*. *J Neurophysiol* 119: 1665-1680, 2018.
103. Huang X and Jan LY: Targeting potassium channels in cancer. *J Cell Biol* 206: 151-162, 2014.
104. Cook SJ and Lockyer PJ: Recent advances in Ca(2+)-dependent Ras regulation and cell proliferation. *Cell Calcium* 39: 101-112, 2006.
105. Monteith GR, McAndrew D, Faddy HM and Roberts-Thomson SJ: Calcium and cancer: Targeting Ca²⁺ transport. *Nat Rev Cancer* 7: 519-530, 2007.
106. Goshima T, Habara M, Maeda K, Hanaki S, Kato Y and Shimada M: Calcineurin regulates cyclin D1 stability through dephosphorylation at T286. *Sci Rep* 9: 12779, 2019.
107. Sayedyahosseini S, Thines L and Sacks DB: Ca²⁺ signaling and the Hippo pathway: Intersections in cellular regulation. *Cell Signal* 110: 110846, 2023.
108. Cavalli G and Heard E: Advances in epigenetics link genetics to the environment and disease. *Nature* 571: 489-499, 2019.
109. Pulczynski JC, Shang Y, Dao T, Limjunyawong N, Sun Q, Mitzner W, Cheng RY and Tang WY: Multigenerational epigenetic regulation of allergic diseases: Utilizing an experimental dust Mite-induced asthma model. *Front Genet* 12: 624561, 2021.
110. Wang J, Seebacher N, Shi H, Kan Q and Duan Z: Novel strategies to prevent the development of multidrug resistance (MDR) in cancer. *Oncotarget* 8: 84559-84571, 2017.
111. Iacopetta D, Rosano C, Sirignano M, Mariconda A, Ceramella J, Ponassi M, Saturnino C, Sinicropi MS and Longo P: Is the way to fight cancer paved with gold? Metal-based carbene complexes with multiple and fascinating biological features. *Pharmaceuticals (Basel)* 13: 91, 2020.
112. Kulkarni P, Ramisetty S, Bruno D, Tan T, Merla A and Salgia R: Phenotypic plasticity, Non-genetic mechanisms, and immune drug resistance in cancer. *Cancer Treat Res* 129: 309-324, 2025.
113. Wei JR, Lu MY, Wei TH, Fleishman JS, Yu H, Chen XL, Kong XT, Sun SL, Li NG, Yang Y and Ni HW: Overcoming cancer therapy resistance: From drug innovation to therapeutics. *Drug Resist Updat* 81: 101229, 2025.
114. Liu S, Jiang A, Tang F, Duan M and Li B: Drug-induced tolerant persisters in tumor: Mechanism, vulnerability and perspective implication for clinical treatment. *Mol Cancer* 24: 150, 2025.
115. Madrid MF, Mendoza EN, Padilla AL, Choquenaira-Quispe C, de Jesus Guimarães C, de Melo Pereira JV, Barros-Nepomuceno FWA, Lopes Dos Santos I, Pessoa C, de Moraes Filho Mo, *et al*: In vitro models to evaluate multidrug resistance in cancer cells: Biochemical and morphological techniques and pharmacological strategies. *J Toxicol Environ Health B Crit Rev* 28: 1-27, 2025.
116. Thomas C and Tampe R: Structural and mechanistic principles of ABC transporters. *Annu Rev Biochem* 89: 605-636, 2020.
117. Mansoori B, Mohammadi A, Davudian S, Shirjang S and Baradaran B: The different mechanisms of cancer drug resistance: A brief review. *Adv Pharm Bull* 7: 339-348, 2017.
118. Zheng HC: The molecular mechanisms of chemoresistance in cancers. *Oncotarget* 8: 59950-59964, 2017.
119. Bai Y, Liao H, Liu T, Zeng X, Xiao F, Luo L, Guo H and Guo L: MiR-296-3p regulates cell growth and multi-drug resistance of human glioblastoma by targeting ether-à-go-go (EAG1). *Eur J Cancer* 49: 710-724, 2013.
120. Sun XX, Hodge JJ, Zhou Y, Nguyen M and Griffith LC: The eag potassium channel binds and locally activates calcium/calmodulin-dependent protein kinase II. *J Biol Chem* 279: 10206-10214, 2004.
121. Huttenlocher A and Horwitz AR: Integrins in cell migration. *Cold Spring Harb Perspect Biol* 3: a005074, 2011.
122. Hsu PH, Ma YT, Fang YC, Huang JJ, Gan YL, Chang PT, Jow GM, Tang CY and Jeng CJ: Cullin 7 mediates proteasomal and lysosomal degradations of rat Eag1 potassium channels. *Sci Rep* 7: 40825, 2017.
123. Horst CH, Titze-de-Almeida R and Titze-de-Almeida SS: The involvement of Eag1 potassium channels and miR-34a in rotenone-induced death of dopaminergic SH-SY5Y cells. *Mol Med Rep* 15: 1479-1488, 2017.

124. Lin H, Li Z, Chen C, Luo X, Xiao J, Dong D, Lu Y, Yang B and Wang Z: Transcriptional and post-transcriptional mechanisms for oncogenic overexpression of ether à go-go K⁺ channel. *PLoS One* 6: e20362, 2011.
125. Wu X, Zhong D, Gao Q, Zhai W, Ding Z and Wu J: MicroRNA-34a inhibits human osteosarcoma proliferation by downregulating ether à go-go 1 expression. *Int J Med Sci* 10: 676-682, 2013.
126. Diaz L, Ceja-Ochoa I, Restrepo-Angulo I, Larrea F, Avila-Chávez E, García-Becerra R, Borja-Cacho E, Barrera D, Ahumada E, Gariglio P, *et al*: Estrogens and human papilloma virus oncogenes regulate human ether-à-go-go-I potassium channel expression. *Cancer Res* 69: 3300-3307, 2009.
127. Scheffner M, Werness BA, Huijbregtse JM, Levine AJ and Howley PM: The E6 oncoprotein encoded by human papilloma-virus types 16 and 18 promotes the degradation of p53. *Cell* 63: 1129-1136, 1990.
128. Borowiec AS, Hague F, Harir N, Guénin S, Guerinéau F, Gouilleux F, Roudbaraki M, Lassoued K and Ouadid-Ahidouch H: IGF-1 activates hEAG K(+) channels through an Akt-dependent signaling pathway in breast cancer cells: Role in cell proliferation. *J Cell Physiol* 212: 690-701, 2007.
129. Sabbatini P and McCormick F: Phosphoinositide 3-OH kinase (PI3K) and PKB/Akt delay the onset of p53-mediated, transcriptionally dependent apoptosis. *J Biol Chem* 274: 24263-24269, 1999.
130. Yoon AR, Gao R, Kaul Z, Choi IK, Ryu J, Noble JR, Kato Y, Saito S, Hirano T, Ishii T, *et al*: MicroRNA-296 is enriched in cancer cells and downregulates p21WAF1 mRNA expression via interaction with its 3' untranslated region. *Nucleic Acids Res* 39: 8078-8091, 2011.
131. Song S, Zhang G, Chen X, Zheng J, Liu X, Wang Y, Chen Z, Wang Y, Song Y and Zhou Q: HIF-1 α increases the osteogenic capacity of ADSCs by coupling angiogenesis and osteogenesis via the HIF-1 α /VEGF/AKT/mTOR signaling pathway. *J Nanobiotechnology* 21: 257, 2023.
132. Wu X, Chen Z, Zeng W, Zhong Y, Liu Q and Wu J: Silencing of Eag1 gene inhibits osteosarcoma proliferation and migration by targeting STAT3-VEGF pathway. *Biomed Res Int* 2015: 617316, 2015.
133. Wu J, Wu X, Zhong D, Zhai W, Ding Z and Zhou Y: Short Hairpin RNA (shRNA) Ether à go-go 1 (Eag1) inhibition of human osteosarcoma angiogenesis via VEGF/PI3K/AKT signaling. *Int J Mol Sci* 13: 12573-12583, 2012.
134. Sun XX, Bostrom SL and Griffith LC: Alternative splicing of the eag potassium channel gene in *Drosophila* generates a novel signal transduction scaffolding protein. *Mol Cell Neurosci* 40: 338-343, 2009.
135. Wu J, Zhong D, Wei Y, Wu X, Kang L and Ding Z: Potassium channel ether à go-go1 is aberrantly expressed in human liposarcoma and promotes tumorigenesis. *Biomed Res Int* 2014: 345678, 2014.
136. Wu X, Zhong D, Lin B, Zhai W, Ding Z and Wu J: p38 MAPK regulates the expression of ether à go-go potassium channel in human osteosarcoma cells. *Radiol Oncol* 47: 42-49, 2013.
137. Hemmerlein B, Weseloh RM, Mello de Queiroz F, Knötgen H, Sánchez A, Rubio ME, Martin S, Schliephacke T, Jenke M, Heinz-Joachim-Radzun, Stühmer W and Pardo LA: Overexpression of Eag1 potassium channels in clinical tumours. *Mol Cancer* 5: 41, 2006.
138. Lei Z, Luo Y, Lu J, Fu Q, Wang C, Chen Q, Zhang Z and Zhang L: FBXO22 promotes HCC angiogenesis and metastasis via RPS5/AKT/HIF-1 α /VEGF-A signaling axis. *Cancer Gene Ther* 32: 198-213, 2025.
139. Zhang HP, Jiang RY, Zhu JY, Sun KN, Huang Y, Zhou HH, Zheng YB and Wang XJ: PI3K/AKT/mTOR signaling pathway: An important driver and therapeutic target in triple-negative breast cancer. *Breast Cancer* 31: 539-551, 2024.
140. Liao Y, Wei Z, Xu H, Zhang Z and Zhu F: Unlocking precision medicine: Innovative strategies for druggable target identification and therapeutic enhancement. *Precision Medication* 1: 100002, 2024.
141. Wang ZJ, Soohoo SM, Tiwari PB, Piszczek G and Brelidze TL: Chlorpromazine binding to the PAS domains uncovers the effect of ligand modulation on EAG channel activity. *J Biol Chem* 295: 4114-4123, 2020.
142. Toplak Z, Hendrickx LA, Abdelaziz R, Shi X, Peigneur S, Tomašič T, Tytgat J, Peterlin-Mašič L and Pardo LA: Overcoming challenges of HERG potassium channel liability through rational design: Eag1 inhibitors for cancer treatment. *Med Res Rev* 42: 183-226, 2022.
143. Ghorbani M, Wang ZJ, Chen X, Tiwari PB, Klauda JB and Brelidze TL: Chlorpromazine inhibits EAG1 channels by altering the coupling between the PAS, CNBH and pore domains. *bioRxiv*: Feb 26, 2024 doi: 10.1101/2024.02.23.581826.
144. Jhou AJ, Chang HC, Hung CC, Lin HC, Lee YC, Liu WT, Han KF, Lai YW, Lin MY and Lee CH: Chlorpromazine, an antipsychotic agent, induces G2/M phase arrest and apoptosis via regulation of the PI3K/AKT/mTOR-mediated autophagy pathways in human oral cancer. *Biochem Pharmacol* 184: 114403, 2021.
145. Gómez-Lagunas F, Carrillo E, Pardo LA and Stühmer W: Gating modulation of the Tumor-related Kv10.1 Channel by Mibefradil. *J Cell Physiol* 232: 2019-2032, 2017.
146. Moreels L, Bhat C, Voračová M, Peigneur S, Goovaerts H, Mäki-Lohiluoma E, Zahed F, Pardo LA, Yli-Kauhaluoma J, Kiuru P and Tytgat J: Synthesis of novel purpurealidin analogs and evaluation of their effect on the cancer-relevant potassium channel KV10.1. *PLoS One* 12: e0188811, 2017.
147. Gardner A, Wu W, Thomson S, Zangerl-Plessl EM, Stary-Weinzinger A and Sanguinetti MC: Molecular basis of altered hERG1 channel gating induced by ginsenoside Rg3. *Mol Pharmacol* 92: 437-450, 2017.
148. Wu W, Gardner A, Sachse FB and Sanguinetti MC: Ginsenoside Rg3, a gating modifier of EAG Family K⁺ channels. *Mol Pharmacol* 90: 469-482, 2016.
149. Moreels L, Peigneur S, Galan DT, De Pauw E, Béress L, Waelkens E, Pardo LA, Quinton L and Tytgat J: APETx4, a novel sea anemone toxin and a modulator of the cancer-relevant potassium channel KV10.1. *Mar Drugs* 15: 287, 2017.
150. Ma B, Shi S, Ren S, Qu C, Zhao Z and An H: Corydaline binds to a druggable pocket of hEAG1 channel and inhibits hepatic carcinoma cell viability. *Eur J Pharmacol* 962: 176240, 2024.
151. Wang JH, Nao JF, Zhang M and He P: 20(s)-ginsenoside Rg3 promotes apoptosis in human ovarian cancer HO-8910 cells through PI3K/Akt and XIAP pathways. *Tumour Biol* Dec 35: 11985-11994, 2014.
152. Doyle DA, Morais Cabral J, Pfuetzner RA, Kuo A, Gulbis JM, Cohen SL, Chait BT and MacKinnon R: The structure of the potassium channel: Molecular basis of K⁺ conduction and selectivity. *Science* 280: 69-77, 1998.
153. Bezanilla F and Armstrong CM: Negative conductance caused by entry of sodium and cesium ions into the potassium channels of squid axons. *J Gen Physiol* 60: 588-608, 1972.
154. Kim DM and Nimigeon CM: Voltage-gated potassium channels: A structural examination of selectivity and gating. *Cold Spring Harb Perspect Biol* 8: a029231, 2016.
155. Wang X, Chen Y, Li J, Guo S, Lin X, Zhang H, Zhan Y and An H: Tetrandrine, a novel inhibitor of ether-à-go-go-1 (Eag1), targeted to cervical cancer development. *J Cell Physiol* 234: 7161-7173, 2019.
156. Garg V, Sachse FB and Sanguinetti MC: Tuning of EAG K(+) channel inactivation: Molecular determinants of amplification by mutations and a small molecule. *J Gen Physiol* 140: 307-324, 2012.
157. García-Ferreiro RE, Kerschensteiner D, Major F, Monje F, Stühmer W and Pardo LA: Mechanism of block of hEag1 K⁺ channels by imipramine and astemizole. *J Gen Physiol* 124: 301-317, 2004.
158. Hernández-Meza JM, Mares-Sámamo S and Garduño-Juárez R: Insights into the molecular inhibition of the oncogenic channel KV10.1 by globular toxins. *J Chem Inf Model* 61: 2328-2340, 2021.
159. Dai Y, Wei Z, Sephton CF, Zhang D, Anderson DH and Mousseau DD: Haloperidol induces the nuclear translocation of phosphatidylinositol 3'-kinase to disrupt Akt phosphorylation in PC12 cells. *J Psychiatry Neurosci* 32: 323-330, 2007.
160. Bai YH, Yang X and Zeng ZW: Protective effect of haloperidol on serum-deprived PC12 cells and its underlying mechanism. *J Psychiatry* 34: 119-123, 2021.
161. Sahoo N, Yang K, Coburger I, Bernert A, Swain SM, Gessner G, Kappl R, Kühl T, Imhof D, Hoshi T, *et al*: Intracellular hemin is a potent inhibitor of the voltage-gated potassium channel Kv10.1. *Sci Rep* 12: 14645, 2022.
162. Ke Z, Tang Z, Shen D, Liu Y, Shu Y, Mu X, Li Z, Xiang P, Zhong B, Hu X, *et al*: Co-Highly expressed SLC17A9 and KCNH1 as potential prognostic biomarkers and therapeutic targets in clear cell renal cell carcinoma. *Front Biosci (Landmark Ed)* 30: 38061, 2025.
163. Cortes-Hernández U, Lizardi-Aguilera TM, Noriega-Mejía BJ, González-Macias J, García-Quiroz J, Díaz L, Larrea F and Avila E: Prostaglandin E2 suppresses KCNH1 gene expression and inhibits the proliferation of CaSki cervical cells through its four prostanoid PTGER subtypes. *Gene* 933: 148997, 2025.

164. Vargas-Castro R, García-Becerra R, Díaz L, Avila E, Ordaz-Rosado D, Bernadez-Vallejo SV, Cano-Colín S, Camacho J, Larrea F and García-Quiroz J: Enhancing tamoxifen therapy with α -Mangostin: Synergistic antiproliferative effects on breast cancer cells and potential reduced endometrial impact. *Pharmaceuticals (Basel)* 16: 1576, 2023.
165. Fan Z, Chen Y, Yan D and Li Q: Effects of differentially methylated CpG sites in enhancer and promoter regions on the chromatin structures of target lncRNAs in breast cancer. *Int J Mol Sci* 25: 11048, 2024.
166. Ding XW, Wang XG, Luo HS, Tan SY, Gao S, Luo B and Jiang H: Expression and prognostic roles of Eag1 in resected esophageal squamous cell carcinomas. *Dig Dis Sci* 53: 2039-2044, 2008.
167. Asher V, Khan R, Warren A, Shaw R, Schalkwyk GV, Bali A and Sowter HM: The Eag potassium channel as a new prognostic marker in ovarian cancer. *Diagn Pathol* 5: 78, 2010.
168. Martínez R, Stühmer W, Martin S, Schell J, Reichmann A, Rohde V and Pardo L: Analysis of the expression of Kv10.1 potassium channel in patients with brain metastases and glioblastoma multiforme: Impact on survival. *BMC Cancer* 15: 839, 2015.
169. Pardridge WM: The blood-brain barrier: Bottleneck in brain drug development. *NeuroRx* 2: 3-14, 2005.
170. Hartung F and Pardo LA: Guiding TRAIL to cancer cells through Kv10.1 potassium channel overcomes resistance to doxorubicin. *Eur Biophys J* 45: 709-719, 2016.



Copyright © 2026 Zhen et al. This work is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International (CC BY-NC-ND 4.0) License.