# Downregulation of expression of transporters associated with antigen processing 1 and 2 and human leukocyte antigen I and its effect on immunity in nasopharyngeal carcinoma patients

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Received April 10, 2013; Accepted July 31, 2013

DOI: 10.3892/mco.2013.194

Abstract. The human leukocyte antigen (HLA)-I and antigen-processing machinery (APM) are crucial in the anticancer immune response. The aim of this study was to assess the clinical significance of the APM components [transporters associated with antigen processing (TAP)-1 and -2 and HLA-I] in nasopharyngeal carcinoma (NPC). A total of 58 NPC specimens and 20 healthy specimens used as control were evaluated by semiquantitative immunohistochemistry for three APM components (TAP-1, TAP-2 and HLA-I). The expression of the APM components in NPC was downregulated. CD4+ and CD8<sup>+</sup> T cells were measured by flow cytometry and IL-10 was measured by ELISA. The number of CD8+ T cells and the expression of IL-10 were higher and the number of CD4+ T cells was lower in NPC, compared to the controls. The number of CD8+ T cells and the expression of IL-10 were negatively correlated with TAP-1, TAP-2 and HLA-I expression. The clinical phase, lymph node metastasis, distant metastasis, pathological type, TAP-1 expression, TAP-2 expression and HLA-I expression were identified as prognostic factors by the Kaplan-Meier analysis. A multivariate analysis using a Cox regression model indicated that distant metastasis and the downregulation of HLA-I expression were independent unfavorable prognostic factors. In conclusion, the lower expression

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of HLA-I induced immunosuppression in NPC patients and was associated with a poor prognosis.

## Introduction

Viruses and tumors evade cytotoxic T lymphocyte-mediated host immunity through the downregulation of antigen-presentation machineries. This may be achieved by either the downregulation of transcription of antigen presentation genes, or the post-translational inactivation of the proteins involved in antigen presentation (1). The optimal cell surface expression of human leukocyte antigen (HLA) molecules requires the coordinated expression of several genes, such as transporters associated with antigen processing (TAP)-1 and -2, low molecular weight peptide (LMP)-2 and -7 and tapasin, as well as HLA class I heavy chain and  $\beta_2$ -microglobulin ( $\beta_2$ M). In cases of concurrent tumorigenesis and viral infection, the expression of these genes and the function of the encoded proteins are often impaired.

Latent Epstein-Barr virus (EBV) infections are associated with lymphocyte and epithelial cell malignancies, with nasopharyngeal carcinoma (NPC) being the most frequent EBV-associated malignancy (2). The EBV-associated, undifferentiated form of NPC exhibits the most consistent association with EBV worldwide and is particularly common in China and Southeast Asia, reaching a peak incidence of ~20-30 cases per 100,000 individuals (3). In addition to genetic predisposition, EBV infection and environmental factors, such as dietary and geographic components, were considered to be important in the aetiology of NPC (4-6). Previous studies that used quantitative polymerase chain reaction to measure circulating tumor-derived EBV DNA in the blood of NPC patients demonstrated that the level of pre-treatment EBV DNA was significantly associated with overall survival and that post-treatment EBV DNA levels predicted progression-free and overall survival (7,8). Previous studies on normal nasopharyngeal tissue and premalignant biopsies indicated that genetic events occur early in the pathogenesis of NPC and they may predispose to subsequent EBV

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*Key words:* nasopharyngeal neoplasms, transporter associated with antigen processing 1, transporter associated with antigen processing 2, human leucocyte antigen I

infection. The EBV latent-gene expression in NPC is predominantly restricted to the Epstein-Barr nuclear antigen 1, the latent membrane proteins (LMP) 2A and 2B and the BamHI-A transcripts, with approximately 70% of the tumors also expressing the oncogenic LMP1 protein (9-11).

Currently, research is focused on the mechanisms underlying the escape of NPC from EBV-specific immune destruction and the development of novel strategies for immune intervention. BamHI-C fragment rightward reading frame 1 (BCRF-1) that is located in the EBV gene, is able to produce viral interleukin-10 (vIL-10), which is the homologue of the human IL-10 (hIL-10). In EBV-infected B lymphocytes, vIL-10 and hIL-10 are equally capable of downregulating TAP-1 expression, thereby interfering with the loading of major histocompatibility complex (MHC) class I molecules, resulting in empty and unstable MHC class I/ $\beta_2$ M complexes. The BCRF-1 gene and the vIL-10 protein are highly expressed in NPC tissues (data not shown); therefore, we aimed to assess whether the expression of TAP-1, TAP-2 and HLA-I are affected in NPC and investigate whether these proteins may be prognostic factors for NPC.

### Materials and methods

Case specimens. A total of 78 paraffin specimens (biopsy specimens obtained during surgery) from 58 patients and 20 healthy controls were collected from the Pathology Department of The Third Affiliated Hospital of Kunmming Medical University, between 2000 and 2002. The characteristics of the 58 patients are summarized in Table I. The control group comprised 20 healthy individuals who were examined at the hospital and NPC was excluded by pathological examination. Immunohistochemistry confirmed that all the specimens were collected prior to medical treatment. Peripheral blood (2 ml) was collected from each patient in heparin tubes. Approval for this study was granted by the Ethics Committee of The Third Affiliated Hospital of Kunming Medical University. The patients provided their permission for the collection of the specimens. The NPC patients received medical treatment according to the NCCN Practice Guidelines for Head and Neck Cancer following pathological diagnosis. All the patients were followed up after treatment.

Immunohistochemistry. Rabbit polyclonal anti-TAP-1 antibody (1:100 dilution; Santa Cruz Biotechnology Inc., Santa Cruz, CA, USA), rabbit monoclonal anti-HLA-I antibody (1:250 dilution; Epitomics Inc., Burlingame, CA, USA) and rabbit polyclonal anti-TAP-2 antibody (1:100 dilution; Abcam, Hong Kong, China) were used in this study. Specimens used as the positive contrast were purchased from Beijing Zhongshan Golden Bridge Biotechnology Co., Ltd. (Beijing, China) and phosphate-buffered saline (PBS) was used for negative contrast. Tumor sections (4  $\mu$ m) were deparaffinised and heated in a microwave oven for 10 min for antigen repair. After cooling, the slides were submerged in a peroxidase quenching solution containing 1 part of 30% hydrogen peroxide to 9 parts of absolute methanol for 10 min and were then washed with PBS. Subsequently, 10% serum was added to each section, followed by incubation for 20 min and draining. The sections were incubated with the primary antibodies overnight in a 4°C chamber, then rinsed with PBS. After rinsing, the sections were treated with Table I. Characteristics of study population in NPC (n=58).

Case	No. (%)
Gender	
Male	38 (65.5)
Female	20 (34.5)
Age (years)	
Mean (range)	49.23 (14-84)
Clinical stage	
Ι	4 (6.9)
II	18 (31.0)
III	10 (17.3)
IV	26 (44.8)
Histological differentiation	
High or moderate	5 (8.6)
Poor	53 (91.4)
Lymph node metastasis	
N <sub>0</sub>	17 (29.3)
N <sub>1-3</sub>	41 (70.7)
Distant metastasis	
$M_0$	48 (82.8)
M <sub>1</sub>	10 (17.2)

NPC, nasopharyngeal carcinoma.

biotin-conjugated antibody for 10 min followed by washing with PBS. Horseradish peroxidase polymer conjugate was applied to each section and incubated for 10 min, followed by washing with PBS. Finally, 3,3'-diaminobenzidine was applied to each section and incubated for 10 min. The samples were rinsed thoroughly with distilled water. Subsequently, the slides were counterstained with hematoxylin, dehydrated and sealed with neutral gum.

The staining of the cores was scored based on signal intensity (0-3) and the percentage of positive cells (0, <5%; 1, 5-10%; 2, 11-50%; 3, 51%-80%; and  $4 \ge 50\%$ ) (12). The results defined as positive were scored based on the product of the two data as follows: 0, negative (-); 1-4, weak positive (+); 5-8, medium positive (++); and 9-12, strong positive (+++).

*Flow cytometry (FCM)*. The collected peripheral blood cells were immunostained with fluorochrome-conjugated anti-human antibodies (FITC-conjugated anti-human CD4, PE-conjugated anti-human CD4 and PC5-conjugated anti-human CD45; Beckman Coulter Inc., Brea, CA, USA) for 25 min at 4°C. The peripheral blood was dissolved using Q-prep autohemolysis equipment. The samples were analyzed with a flow cytometer (EPICS-XL; Beckman Coulter Inc.). The frequency of CD4<sup>+</sup> T cells (CD4-FITC/CD45-PC5) and CD8<sup>+</sup> T cells (CD8-PE/CD45-PC5) among the lymphocytes of different subsets was calculated, with IgG-FITC/CD45-PC5 and IgG-PE/CD45-PC5 as the negative controls. Data was analyzed with Expo32 software.

*Measurement of IL-10 by enzyme-linked immunosorbent assay* (*ELISA*). The blood samples were centrifuged at 3,000 rpm

	TAP-1				TAP-2				HLA-I	
Group	No.	_	+	P-value	_	+	P-value	_	+	P-value
Chronic	20	2	18	<0.001	1	10	<0.001	1	10	<0.001
NPC	20 58	33	18 25	<0.001	31	19 27	<0.001	29	19 29	<0.001

Table II. Expression of TAP-1, TAP-2 and HLA-I in the NPC and the control group.

TAP, transporters associated with antigen processing; HLA, human leukocyte antigen; NPC, nasopharyngeal carcinoma. The Chi-square values were 13.22, 14.43 and 12.72, respectively.

Table III. Association of TAP-1, TAP-2 and HLA-I expression with clinical factors in NPC.

	TAP-1			TA	TAP-2		HI	LA-I	
	Positive no. (%)	Negative no. (%)	P-value	Positive no. (%)	Negative no. (%)	P-value	Positive no. (%)	Negative no. (%)	P-value
Age (years)									
≤43	12 (48)	16 (48.5)	1.00	15 (55.5)	13 (41.9)	0.43	10 (34.5)	18 (62.1)	0.065
>43	13 (52)	17 (51.5)		12 (44.4)	18 (58.1)		19 (65.5)	11 (37.9)	
Gender									
Male	17 (68)	21 (63.6)	0.786	16 (59.3)	22 (71)	0.413	20 (69)	18 (62.1)	0.783
Female	8 (32)	12 (36.4)		11 (40.7)	9 (29)		9 (31)	11 (37.9)	
Clinical stage									
I+II	13 (52)	7 (21.2)	0.015	15 (55.6)	5 (16.1)	0.002	15 (51.7)	5 (17.2)	0.012
III+IV	12 (48)	26 (78.8)		12 (44.4)	26 (83.9)		14 (48.3)	24 (82.8)	
Lymph node metastasis									
Yes	11 (44)	30 (90.9)	0.00	12 (44.4)	29 (93.5)	0.00	13 (44.8)	28 (96.6)	0.00
No	14 (56)	3 (9.1)		15 (55.6)	2 (6.5)		16 (55.2)	1 (3.4)	
Distant metastasis									
Yes	1 (4)	9 (27.3)	0.033	1 (3.7)	9 (29)	0.014	0 (0)	10 (35.7)	0.00
No	24 (96)	24 (72.7)		26 (96.3)	22 (71)		29 (100)	19 (64.3)	

TAP, transporters associated with antigen processing; HLA, human leukocyte antigen; NPC, nasopharyngeal carcinoma.

(825 x g) for 25 min. The supernatant was collected, aliquoted and stored at -80°C until use. The concentrations of IL-10 were measured using commercial ELISA kits (Biosource International Inc., Boshide Company, Wuhan, China), according to the manufacturer's instructions. the multivariate Cox proportional hazards model. P<0.05 was considered to indicate a statistically significant difference.

# Results

Statistical analysis. Data were analyzed using SPSS software, version 12.0 (SPSS Inc., Chicago, IL, USA). The correlation of TAP-1, TAP-2 and HLA-I expression with the clinicopathological variables was performed with the Chi-square test. The expression of CD4<sup>+</sup> T cells, CD8<sup>+</sup> T cells and IL-10 in the different groups was assessed with the independent-samples t-test. Survival was assessed with the Kaplan-Meier analysis and the log-rank score was used for determining statistical significance. The relative risk was assessed with

Low TAP-1, TAP-2 and HLA-I expression in NPC tissue. In NPC, the TAP-1 and TAP-2 immunoreactivity displayed weak diffuse cytoplasmic staining, whereas HLA-I displayed weak diffuse cytoplasmic and cytomembranous staining (Fig. 1A, C and E). By contrast, normal nasopharyngeal tissues exhibited intense staining for these proteins (Fig. 1B, D and F). A total of 25, 27 and 29 of the 58 NPC samples exhibited low expression of TAP-1, TAP-2 and HLA-I, respectively, whereas 18, 19 and 19 of the 20 normal nasopharyngeal tissue samples exhibited



Figure 1. Expression of TAP-1, TAP-2 and HLA-I in NPC and normal nasopharyngeal tissue samples. Staining for the three proteins was performed as described in Materials and methods. NPC tissues exhibited weak positive or absent staining for (A) TAP-1, (C) TAP-2 and (E) HLA-I (magnification, x400). Normal nasopharyngeal tissues exhibited strong staining for (B) TAP-1, (D) TAP-2 and (F) HLA-I (magnification, x400). TAP, transporters associated with antigen processing; HLA, human leukocyte antigen; NPC, nasopharyngeal carcinoma.



Figure 2. Percentage of  $CD4^+$  and  $CD8^+$  T cells to peripheral blood (PB) lymphocytes in NPC and control samples. The percentage of  $CD4^+$  and  $CD8^+$  T cells was analyzed by flow cytometry. The percentage of  $CD4^+$  T cells in NPC (A) was 33.0%, whereas in the control group (B) it was 42.4%. The percentage of  $CD8^+$  T cells in NPC (C) was 25.2% and in the control group (D) it was 22.4%. A representative experiment is shown. (E) The percentages of  $CD4^+$  T cells,  $CD8^+$  T cells and the expression of IL-10 were assessed with independent-samples t-test in the NPC and control groups. \*P<0.05 was considered to indicate a statistically significant difference. NPC, nasopharyngeal carcinoma; NO NPC, normal nasopharyngeal tissues.

Table IV. Analysis of CD4<sup>+</sup> T cells, CD8<sup>+</sup> T cells and IL-10 expression in different TAP-1, -2 and HLA-I expression groups in NPC.

	TAP-1			TAP-2			HL	HLA-I	
	Positive	Negative	P-value	Positive	Negative	P-value	Positive	Negative	P-value
CD4 <sup>+</sup> T cell (%)	33.19±9.20	33.35±8.05	0.970	33.58±6.40	32.52±5.20	0.815	34.34±6.58	30.22±3.49	0.379
CD8 <sup>+</sup> T cell (%)	27.93±4.04	18.43±2.37	0.013	28.47±3.62	19.18±2.11	0.018	31.35±4.72	16.65±2.07	0.005
IL-10 (ng/ml)	9.87±1.24	16.20±1.48	0.021	8.93±0.56	16.04±1.75	0.015	6.28±0.86	17.99±2.01	0.008

TAP, transporters associated with antigen processing; HLA, human leukocyte antigen; NPC, nasopharyngeal carcinoma; IL, interleukin.



Figure 3. Kaplan-Meier curves for survival by different clinicopathological factors. (A) NPC patients with stage I+II disease exhibited significantly higher survival rates compared to those with stage III+IV disease. (B) and (C) NPC patients with lymph node or distant organ metastases exhibited lower survival rates compared to those without lymph node or distant organ metastasis. (D) Poor histological differentiation was associated with higher survival rates compared to high or moderate differentiation. (E), (F) and (G) NPC patients with positive expression exhibited higher survival rates compared to those with negative expression of TAP-1, TAP-2 and HLA-I. NPC, nasopharyngeal carcinoma; TAP, transporters associated with antigen processing; HLA, human leukocyte antigen; Cum, cumulative.

high expression of TAP-1, TAP-2 and HLA-I, respectively. Therefore, the expression of TAP-1, TAP-2 and HLA-I in the NPC samples was distinctly lower compared to that in the normal nasopharyngeal tissue samples (P<0.05, Table II).

Changes in  $CD4^+$  T cells,  $CD8^+$  T cells and IL-10 in the peripheral blood of NPC patients. FCM was applied to detect  $CD4^+$  T cells,  $CD8^+$  T cells and IL-10 was analyzed by ELISA. The percentage of  $CD4^+$  T cells in the peripheral

Factor	No.	3-year survival rate (%)	5-year survival rate (%)	Log-rank value	P-value
Age (years)					
≤43	38	68.42	47.37	1.63	0.202
>43	20	60.00	40.00		
Gender					
Male	38	79.50	65.00	0.26	0.610
Female	20	71.05	44.74		
Clinical stage					
I+II	22	77.27	56.25	10.14	0.001
III+IV	36	55.56	38.89		
N stage					
N <sub>0</sub>	17	76.47	47.06	5.36	0.021
N <sub>1-3</sub>	41	58.54	43.90		
M stage					
$M_0$	48	69.57	54.35	6.77	0.009
$M_1$	10	41.67	8.33		
Histological differentia	tion				
High/moderate	5	36.00	0	6.02	0.014
Poor	53	67.92	49.06		
TAP-1 expression					
Positive	25	80.00	60.00	11.77	0.001
Negative	33	51.52	33.33		
TAP-2 expression					
Positive	27	81.48	62.96	19.38	0.000
Negative	31	48.39	29.03		
HLA-I expression					
Positive	29	82.76	65.52	18.90	0.000
Negative	29	44.83	24.14		

Table V. Univariate survival a	nalysis in	patients	with NPC.
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NPC, nasopharyngeal carcinoma; TAP, transporters associated with antigen processing; HLA, human leukocyte antigen.

blood of NPC patients was  $33.41\pm10.04\%$  (Fig. 2A), which was lower compared to that of the normal subjects, which was  $40.15\pm3.56\%$  (Fig. 2B) (P<0.05, Fig. 2E). There was no significant difference between the NPC and control groups regarding CD8<sup>+</sup> T cells ( $25.32\pm8.29$  vs.  $22.89\pm2.24\%$ , P>0.05, Fig. 2C, D and E). The ELISA results indicated that the expression of IL-10 in NPC was higher compared to that in the control group ( $13.12\pm1.23$  vs.  $3.69\pm1.03$  ng/ml, respectively; P<0.05, Fig. 2E).

Association between TAP-1, TAP-2 and HLA-I status and clinical variables. As shown in Table III, the low expression of TAP-1, TAP-2 and HLA-1 was significantly associated with TNM stage, lymph node metastasis and distant metastasis (P<0.05). No association was observed between their expression with age and gender (P>0.05).

Correlations of TAP-1, TAP-2 and HLA-I expression with CD4<sup>+</sup> T cells, CD8<sup>+</sup> T cells and IL-10. In order to elucidate whether TAP-1, TAP-2 and HLA-I expression affected

CD4<sup>+</sup> T cells, CD8<sup>+</sup> T cells and IL-10, we divided the 58 NPC patients into TAP-1-positive and -negative groups, TAP-2-positive and -negative groups and HLA-I-positive and -negative groups and then assessed the proportion of CD4+ T and CD8<sup>+</sup> T cells and the expression of IL-10 in the subgroups. As shown in Table IV, there was no significant difference between the positive and negative subgroups regarding the expression of CD4+ T cells (33.19±9.2 vs. 33.35±8.05%; 33.58±6.4 vs. 32.52±5.2%; and 34.34±6.58 vs. 30.22±3.49%, respectively; P>0.05). The percentage of CD8<sup>+</sup> T cells was higher in the positive compared to the negative TAP-1, TAP-2 and HLA-I groups (27.93±4.04 vs. 18.43±2.37%; 28.47±3.62 vs. 19.18±2.11%; and 31.35±4.72 vs. 16.65±2.07%, respectively; P<0.05). The expression of IL-10 was lower in the positive compared to the negative TAP-1, TAP-2 and HLA-I groups (9.87±1.24 vs. 16.20±1.48 ng/ ml; 8.93±0.56 vs. 16.04±1.75 ng/ml; and 6.28±0.86 vs. 17.99±2.01 ng/ml, respectively; P<0.05).

Follow-up and univariate survival analysis. A total of 58 patients received standard treatment and were regularly

Variable	Wald	OR	95.0% CI	P-value
Age	0.595	1.290	0.675-2.465	0.440
Gender	0.014	1.037	0.570-1.886	0.904
TNM stage	0.004	0.952	0.220-4.128	0.948
Lymph node metastasis	0.112	1.155	0.497-2.687	0.737
Distant metastasis	4.598	2.609	0.971-5.974	0.041
Histological differentiation	0.016	0.922	0.262-3.242	0.899
TAP-1 expression	0.005	1.071	0.168-6.819	0.942
TAP-2 expression	0.412	1.925	0.260-14.250	0.521
HLA-I expression	4.557	2.586	0.389-8.186	0.042

Table VI. Multivariate Cox regression analysis.

followed up. The Kaplan-Meier analysis was used to analyze single factors considered to affect disease progression including age, gender, TNM stage, lymph node metastasis or lack thereof, distant organ metastasis or lack thereof, pathological type, TAP-1 expression, TAP-2 expression and HLA-I expression. We observed that clinical stage, lymph node metastasis, distant organ metastasis, histological differentiation, TAP-1 expression, TAP-2 expression and HLA-I expression, TAP-2 expression and HLA-I expression exerted a significant effect on the overall survival of NPC patients (Fig. 3 and Table V, P<0.05).

*Multivariate survival analysis.* Since the factors mentioned above were shown to exert a significant effect on the survival of NPC patients, a multivariate analysis was performed to assess the independent predictive value of each of these factors for overall survival. Of note, distant metastasis and HLA-I expression status were shown to be potential independent prognostic factors for NPC patients (P=0.041 and P=0.042, respectively, Table VI).

# Discussion

During the tumor immune response process, the MHC class I molecules (mainly HLA-I) play a crucial role in the elimination of virally infected and transformed cells by cytotoxic T cells (CTLs). The CTLs recognize virally infected or malignant T cells by foreign peptides presented on the cell surface in association with class I antigens. The HLA-I antigen processing and presentation pathway starts with proteins being degraded by the proteasome, which consists of multiple catalytic subunits (mainly LMP-2 and -7). The peptides are then translocated across the endoplasmic reticulum (ER) membrane via the antigen-processing subunits TAP-1 and -2. The HLA-I heavy chain is synthesized in the ER, where it forms a complex with  $\beta_2 M$ . The HLA-I/ $\beta_2 M$  complex then interacts with TAP-associated peptides and facilitates peptide loading into HLA-I/\beta\_2M/peptide complexes, which are transported to the surface.

TAP plays a pivotal role in the peptide loading of the HLA-I molecules and is therefore essential for their expression on the cell surface. Thus, the transport of the HLA-I complexes to the cell surface may be prevented if TAP is malfunctioning,

leading to antigen not being recognized by CTLs and escaping the immune supervision (13-15). The absence of TAP-1, LMP-2, LMP-7 and HLA-I have been reported to occur in small-cell lung carcinoma (16), colorectal cancer, breast cancer (17), malignant melanoma, malignant tumors of the head and neck (18) and malignant brain tumors (19) and may represent a mechanism of tumor escape from the control of the immune system. An increase in the expression of LMP-2, LMP-7 and HLA-I may be induced if TAP transfection is performed.

In our study, the expression of TAP-1, TAP-2 and HLA-I in NPC were distinctly reduced. In the peripheral blood of NPC patients, the expression of IL-10, which has immunosuppressive function, was increased and the percentage of CD4<sup>+</sup> T cells, which help B lymphocytes secrete antibodies, was decreased. The percentage of the CD8<sup>+</sup> T cells, which are crucial in eliminating the infected or malignant T cells had no detectable change. However, we observed that the percentage of CD8<sup>+</sup> T cells exhibited a consistently positive correlation with the expression of TAP-1, TAP-2 and HLA-I, whereas the expression of IL-10 had a negative correlation with the expression of TAP-1, TAP-2 and HLA-I in NPC. These results suggest that the reduction in the expression of TAP-1, TAP-2 and HLA-I may contribute to the immunosuppression associated with in NPC, which may help tumor cells escape immune surveillance (20).

An association between the downregulation of TAP-1, TAP-2 and HLA-I expression and cancer prognosis has been reported in a wide range of malignancies (21-24). The correlation between TAP-1, TAP-2 and HLA-I and prognosis in NPC has been investigated (25). Our data demonstrated that the downregulation of the expression of TAP-1, TAP-2 and HLA-I became more prominent in more advanced clinical stages. The Cox regression model indicated that HLA-I expression and distant metastasis were independent prognostic factors. Distant metastasis was the major cause of mortality in patients with NPC, even following successful locoregional control with radiotherapy or/ and chemotherapy. According to a previous study, ~11-36% of NPC patients with controlled locoregional disease will develop distant metastasis (26). Distant metastasis was shown to be a negative prognostic factor in the present study, although the difference was not considered statistically significant (P=0.041).

Several factors may have affected the results: i) distant metastasis may not have been diagnosed by pathological examination. It has been suggested that <sup>18</sup>F-fluorodeoxyglucose-positron emission tomography/computed tomography is the most sensitive, specific and accurate modality for distant metastasis staging of NPC; ii) gender, pretreatment quality of life variables, treatment, metastatic spread to different organs and short-term treatment response may affect the prognosis of NPC with distant metastasis (27,28); iii) an increase in the follow-up of NPC cases is required. Of note, the expression of HLA-I, but not that of TAP-1 and/or TAP-2, was an independent prognostic factor in NPC. The HLA-I molecules play a central role in antigen submission, during which TAP molecules are involved in antigen transport and the antigen/HLA assembly. Thus, the downregulation of HLA-I expression correlates in part with the decrease in the TAPs (12). This study suggested that the downregulation of HLA-I expression was particularly associated with a poor prognosis in NPC patients. Due to the limited patient sample, it could not be determined whether this constitutes a biological property or is due to the limited data availability.

In conclusion, we demonstrated that the expression of TAP-1, TAP-2 and HLA-I were downregulated in NPC and this downregulation may contribute to immunosuppression in NPC patients. Of note, distant metastasis and HLA-I expression may be considered as independent prognostic factors in NPC. Further studies are required to elucidate the molecular mechanisms through which TAP-1, TAP-2 and HLA-I expression affect immunity and investigate the possibility of designing a biological treatment that will enhance HLA-I molecule expression in NPC patients.

# Acknowledgements

We would like to thank the patients and healthy donors for participating in this study. This study was supported in part by grants from the National Natural Science Foundation of China (no. 81260312), the Research Project Foundation of Health Science and Technology of Yunnan Province (no. 2011WS0068), the Technological Plan of Society Development of Yunnan Province (Fundamental Research Program) (no. 2009ZC119M) and the Technological Plan of Society Development of Yunnan Province (Key Fundamental Research Program) (no. 2009CC026).

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