The combination of positive anti-WDR1 antibodies with negative anti-CFL1 antibodies in serum is a poor prognostic factor for patients with esophageal carcinoma

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Abstract. WD repeat-containing protein 1 (WDR1) regulates the cofilin 1 (CFL1) activity, promotes cytoskeleton remodeling, and thus, facilitates cell migration and invasion. A previous study reported that autoantibodies against CFL1 and β-actin were useful biomarkers for diagnosing and predicting the prognosis of patients with esophageal carcinoma. Therefore, the present study aimed to evaluate the serum levels of anti-WDR1 antibodies (s-WDR1-Abs) combined with serum levels of anti-CFL1 antibodies (s-CFL1-Abs) in patients with esophageal carcinoma. Serum samples obtained from 192 patients with esophageal carcinoma and other solid cancers. And s-WDR1-Ab and s-CFL1-Ab titers were analyzed using the amplified luminescent proximity homogeneous assay-linked immunosorbent assay. Compared with those of healthy donors, the s-WDR1-Ab levels were significantly higher in the 192 patients with esophageal, whereas these were not significantly higher in the samples from patients with gastric, colorectal, lung, or breast cancer. In 91 patients treated with surgery, sex, tumor depth, lymph node metastasis, stage and C-reactive protein levels were significantly associated with overall survival, as determined using the log-rank test, whereas the squamous cell carcinoma antigen, p53 antibody and s-WDR1-Ab levels tended to be associated with a worse prognosis. Although no significant difference was observed in the survival between the positive and negative groups of s-WDR1-Abs or s-CFL1-Abs alone in the Kaplan-Meier test, the patients in the s-WDR1-Ab-positive and s-CFL1-Ab-negative groups exhibited a significantly poorer prognosis in the overall survival analysis. On the whole, the present study demonstrates that the combination of positive anti-WDR1 antibodies with negative anti-CFL1 antibodies in serum may be a poor prognostic factor for patients with esophageal carcinoma.

Introduction

Esophageal carcinoma is a rapidly progressive disease and is life-threatening even at the early stages. The therapeutic efficacy of esophageal carcinoma is less satisfactory than that of several other types of cancer; therefore, the detection and diagnosis at the early stage of carcinoma are indispensable for improving the therapeutic outcomes of patients (1).

The serum levels of several antigens and antibodies have long been measured using the serological analysis of recombinant tumor cDNA expression libraries (SEREX) method, a useful screening method for tumor markers (2). SEREX uses patient serum to immunoscreen cDNA libraries prepared from cancer specimens. By sequencing the isolated cDNA clones, the SEREX method is suitable as for the large-scale screening for tumor antigens. Large-scale SEREX screening has already identified numerous antibodies or antigens, such as trophoblast cell surface antigen 2 (3), solute carrier family 2 member 1 (4), striatin 4 (5), LDL receptor related protein associated protein 1 (6), proprotein convertase subtilisin/kexin type 9 (7), cofilin 1 (CFL1) and β -actin (ACTB) (8). CFL1 can depolymerize F-actin in a pH-dependent manner and is also involved in lung, pancreatic, and gastric cancer invasion and metastasis (9-11).

WD repeat-containing protein 1 (WDR1; also known as actin-interacting protein 1) regulates cofilin activity, promotes cytoskeletal remodeling, and thus facilitates cell migration and invasion (12). The presence of autoantibodies against WDR1 has been found in the sera of patients who have suffered a transient ischemic attack using SEREX screening (13). The

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authors have previously reported that serum ACTB and CFL1 antibody titers were inversely associated with the overall survival of patients (8), suggesting the importance of actin polymerization and depolymerization in determining the prognosis. Thus, the present study aimed to evaluate the serum anti-WDR1 antibody (s-WDR1-Ab) titers in patients with various types of cancer compared with healthy donors. The overall survival was then compared among those with positive and negative s-WDR1-Ab and serum anti-CFL1 antibody (s-CFL1-Ab) titers.

Patients and methods

Collection of serum samples. The present study was approved by the Ethics Committee of Toho University Graduate School of Medicine (nos. A18103 A17052 A16035 A16001_26095_25024_24038_22047,M21038_20197_19213), Chiba University Graduate School of Medicine (no. 2018-320) (Japan) and Kashiwado Memorial Foundation (no. 2012-001). Serum samples were collected from patients who provided written formal informed consent. A total of 672 serum samples were obtained from cancer patients. These included 192 samples from patients with esophageal carcinoma, 96 samples from patients with gastric cancer, 192 samples from patients with colorectal cancer, 96 samples from patients with lung cancer and 96 samples from patients with breast cancer; all samples were obtained at the Toho University Omori Hospital (Tokyo, Japan) from June, 2010 to February, 2016. Serum samples from patients with esophageal carcinoma were collected prior to any treatments at Toho University, Omori Medical Center. Among these, 91 patients underwent radical surgery, comprising 70 males and 21 females. The median age of the operated esophageal carcinoma patients was 67 years. A total of 63 patients were candidates for neoadjuvant chemotherapy. Each patient was followed-up until June, 2018 or until death, whichever occurred first. All data regarding clinicopathological characteristics and prognoses were retrospectively obtained. Serum samples from 96 healthy donors (48 males, 48 females) were obtained by excluding residual samples at the Port Square Kashiwado Clinic, China, Japan.

Preparation and purification of antigenic WDR1 and CFL1 proteins. cDNA clones for CFL1 and WDR1 were separated using SEREX screening from the λ ZAP II phage cDNA library for the human esophageal carcinoma cell line, T.Tn (3,4,8), and human aortic endothelial cells (14,15), respectively. Full-length cDNAs of WDR1 were recombined into pGEX-4T-1 (Cytiva). ECOS[™] competent Escherichia coli BL-21 cells (Nippon Gene, Co., Ltd.) were transformed with prokaryotic expression plasmids, pGEX-4T-1, pGEX-4T-1-WDR1 and pGEX-4T-1-CFL1 (Cytiva), and then cultured for 3 h in a 200 ml Luria broth containing 0.1 mM isopropyl β-D-thiogalactopyranoside (IPTG; FUJIFILM Wako Pure Chemical Corporation). The cells were lysed by sonication in BugBuster Protein Extraction Reagent (Merck), and GST, GST-WDR1 and GST-CFL1 proteins were purified by affinity chromatography using glutathione-Sepharose columns (Cytiva), as previously described (16-18).

Measurement of s-WDR1-Ab and s-CFL1-Ab levels and conventional serum markers. Serum samples were obtained prior to any treatment, and centrifuged at 3,000 x g for 10 min at 25°C, and stored at -80°C. The s-WDR1-Ab and s-CFL1-Ab levels were measured using the amplified luminescence proximity homogeneous assay-linked immunosorbent assay (AlphaLISA) with WDR1 and CFL1. AlphaLISA was conducted using 384-well microtiter plates (white opaque OptiPlate[™], PerkinElmer, Inc.) containing 2.5 µl 1/100-diluted sera and 2.5 µl GST, GST-WDR1, or GST-CFL1 (10 µg/ml) in AlphaLISA buffer [25 mM HEPES (pH 7.4), 0.1% casein, 0.5% Triton X-100, 1 mg/ml dextran-500 and 0.05% Proclin-300] following the manufacturer's instructions (PerkinElmer, Inc.) and as previously described (19). The reaction mixture was incubated at room temperature for 6-8 h. Subsequently, anti-human IgG-conjugated acceptor beads (2.5 µl of 40 µg/ml) and glutathione-conjugated donor beads (2.5 μ l of 40 μ g/ml) (PerkinElmer, Inc.) were added followed by incubation for a further 7-28 days at room temperature in the dark. Chemical emission was read on an EnSpire Alpha microplate reader (PerkinElmer, Inc.; http://www.perkinelmer.com/lab-solutions/resources/docs/GDE_ELISA-to-AlphaLISA.pdf) as previously described (5,15,20,21). By subtracting the alpha photon counts of the GST control from those of the GST fusion proteins, specific reactions were calculated.

The serum levels anti-p53 antibodies (s-p53-Abs) (22,23) and squamous cell carcinoma antigen (SCC-Ag) (24), which were often measured worldwide (22-24), were also measured using standard procedures. The cut-off values for s-p53-Abs was set at 1.3 IU/ml and those for SCC-Ag was set at and 1.5 ng/ml.

Statistical analyses. Differences between the two variables were analyzed using Fisher's exact test. Corresponding differences between the three variables were determined using the Kruskal-Wallis test with Bonferroni's correction and the Mann-Whitney U-test. Receiver operating characteristic (ROC) curve analysis was used to determine the predictive qualities of putative disease markers, and cut-off values were determined to maximize the total sensitivity and specificity and the Youden index. Optimal cut-off values for serum antibody levels that affect overall survival were determined using X-tile 3.6.1 software (Yale University, New Haven, CT, USA), as previously described (25). The Kaplan-Meier method was used to analyze survival and survival curves were drawn. In addition, the survival distributions of two groups were compared using the log-rank test. Statistical analyses were performed using EZR software (https://www.jichi.ac.jp/saitama-sct/SaitamaHP. files/statmed.html) (version 1.55) (26). P<0.05 was considered to indicate a statistically significant difference.

Results

Identification of WDR1 and CFL1 using SEREX screening. WDR1 was identified using large-scale SEREX screening with the sera of patients with a transient ischemic attack as an antigen recognized by serum IgG antibodies (13). The 172-1233 region of WDR1 cDNA (accession no. NM_017491) was isolated along with the coding sequence from 235-735. The cDNA was then recombined into pGEX-4T-1 and the



Figure 1. (A) Comparison of s-WDR1-Ab levels between healthy donors and patients with esophageal, gastric, colorectal, lung, and breast cancers examined using an amplified luminescence proximity homogeneous assay-linked immunosorbent assay (AlphaLISA). The s-WDR1-Ab levels are shown in a scatter dot plot. ***P<0.001; NS, not significant. (B) s-WDR1-Ab and s-CFL1-Ab levels using the ROC curve analysis between the alive and deceased cases in 192 patients with esophageal carcinoma. ROC curves for WDR1-Ab and CFL1-Ab are presented. The area under the ROC curve, 95% confidence interval, cut-off levels, specificity and sensitivity, and P-values are presented in Table SI. Closed circles indicate the positions with the highest sum of sensitivity plus specificity, i.e., the Youden index. The ROC curve analysis was used to calculate the P-values. WDR1, WD repeat-containing protein 1; CFL1, cofilin 1; s-WDR1-Ab, serum anti-WDR1 antibody; s-CFL1-Ab, serum anti-CFL1 antibody; ROC, receiver operating characteristic.

Table I. Comparison of serum levels according to clinicopathological characters of the patients with esophageal carcinoma using univariate analysis.

Variable	WDR1-Ab >1865	WDR1-Ab ≤1865	P-value
Sex			
Male	24	46	0.012
Female	14	7	
Age, years			
>65	22	31	>0.999
≤65	16	22	
Tumor location			
Upper	5	9	0.771
Lower	33	44	
Tumor depth			
T1	15	14	0.254
T2-T4	23	39	
Lymph node metastasis			
N0	17	24	>0.999
N1	21	29	
Tumor stage			
0	6	3	
Ι	6	8	
II	8	17	
III	16	18	
IV	2	7	
SCC-Ag (ng/ml)			
>1.5	10	21	0.260
≤1.5	27	31	
p53-Ab (U/ml)			
>1.30	7	10	>0.999
≤1.30	29	43	
WBC $(/\mu l)$			
>8.000	4	7	0.757
≤8.000	34	46	
Neutrophils (%)			
>70	8	11	>0.999
≤70	30	42	
Lymphocytes (%)			
>35	6	11	0.597
≤35	32	42	
Hemoglobin (g/dl)			
>12	25	36	>0.999
≤12	13	17	
Platelets $(/\mu]$)			
>150.000	37	36	0.232
≤150.000	- 1	6	
CRP (mg/dl)	-	ž	
>0.3	10	19	0.370
≤0.3	27	33	

Table I. Continued.

Variable	WDR1-Ab >1865	WDR1-Ab ≤1865	P-value
Albumin (g/dl)			
>3.5	29	39	0.811
≤3.5	9	14	

Fisher's exact probability test was used for statistical analysis. Stage was determined using the Classification by the Japan Esophageal Society (36). As regards tumor location, the oral side from the tracheal bifurcation was defined as the 'upper' location and the anal side from the tracheal bifurcation was defined as the 'lower' location. WDR1, WD repeat-containing protein 1; WDR1-Ab, serum anti-WDR1 antibody; N0, no lymph node metastasis; N1, lymph node metastasis exists; SCC-Ag, squamous cell carcinoma antigen; WBC, white blood cell; CRP, C-reactive protein. There were some cases for which SCC-Ag, p53-Ab and CRP were not measured.

recombinant GST-fused WDR1 protein was purified. As WDR1 regulates actin polymerization, GST-CFL1 protein was also purified as previously described (8).

Comparison of s-WDR1-Ab levels between patients with solid cancers and healthy donors. The s-WDR1-Ab level was measured with AlphaLISA using GST-WDR1 for antigens. The results revealed that the s-WDR1-Ab levels in patients with esophageal carcinoma were significantly higher than those from healthy donors (Fig. 1A; P<0.001). However, the s-WDR1-Ab levels in serum samples from patients gastric, colorectal, lung and breast cancers did not differ significantly from those in the serum of healthy donors, as determined using Bonferroni's correction.

ROC analysis resulted in an area under the ROC curve (AUC) for s-WDR1-Ab of 0.648 for esophageal carcinoma (Fig. 1B and Table SI). The sensitivity and specificity were 59.9 and 67.7%, respectively, with the determined cut-off value (1865) of the Youden index to maximize the sum of sensitivity and specificity. Setting the cut-off level of s-CFL1-Ab to 50718, the AUC value for s-CFL1-Ab was 0.694 with the sensitivity and specificity of 53.3 and 68.8%, respectively (Fig. 1B). Notably, the ROC curves of s-WDR-Ab and s-CFL-Ab were relatively similar.

Association between the esophageal carcinoma patient clinicopathological parameters and overall survival. The associations between the overall survival and patient characteristics, including sex, age, tumor location, tumor depth, lymph node metastasis, stage, SCC-Ag, p53-Ab, s-WDR1-Ab, white blood cell count, as well as neutrophil, lymphocyte, hemo-globin, platelet, C-reactive protein (CRP) and albumin levels were evaluated using univariate analysis. The s-WDR1-Ab levels were divided into the high- and low-antibody groups (>1865/≤1865) based on the cut-off value calculated using X-tile analysis. Fisher's exact probability test revealed that the s-WDR1-Ab levels were significantly higher in male than those in female patients (Table I). However, the other parameters were not found to be significantly associated with the s-WDR1-Ab

levels. As regards the stage, as the tumor stage progressed, the ratio of s-WDR1-Ab-positive cases vs. negative cases increased. The parameters of sex, tumor depth, lymph node metastasis and CRP levels were significantly associated with overall survival, as determined using the log-rank test (Table II), whereas the SCC-Ag, p53-Ab, and s-WDR1-Ab levels were only partly, but not significantly associated with the overall survival (P<0.066, 0.063 and 0.078, respectively). Since there was a significant difference in prognosis between males and females (Table II), the present study compared the prognosis of male and female patients in the high s-WDR1-Ab and low s-WDR1-Ab groups. The results did not reveal any significant difference, either in males or females (Fig. S1). Therefore, this result suggested that the difference in prognosis by sex appeared to have minimal relevance to the s-WDR1-Ab titer.

Combined analysis of the s-WDR1-Ab and s-CFL1-Ab levels in association with patient survival. To evaluate the prognostic characteristics of s-WDR1-Ab, survival curves were drawn using the Kaplan-Meier method. According to the cut-off level determined using the X-tile analysis, the s-WDR1-Ab and c-CFL1-Ab levels were divided into the positive and negative groups. Although no significant difference was observed in the survival of patients between the positive and negative groups of s-WDR1-Ab or s-CFL1-Ab levels alone (Fig. 2A and B), the combined group consisting of positive s-WDR1-Ab and negative s-CFL1-Ab exhibited a significantly poor prognosis in the overall survival analysis (Fig. 2C). Similarly, the group consisting of negative s-WDR1-Ab and positive s-CFL1-Ab exhibited a relatively favorable prognosis (Fig. 2C). After 50 months, approximately half of the patients in the latter group still survived, whereas those in the former group did not survive.

Discussion

The present study found that the s-WDR1-Ab levels were significantly higher in patients with esophageal carcinoma than in healthy subjects (Fig. 1A). Moreover, the s-WDR1-Ab-positive group tended to have a poorer prognosis than the negative group (Fig. 2A). The combination of s-WDR1-Ab positivity and s-CFL1-Ab negativity revealed a significant difference vs. that of s-WDR1-Ab negativity and s-CFL1-Ab positivity (Fig. 2C). WDR1 expression has been shown to be upregulated in the highly metastatic gallbladder cancer cell line (GBC-SD18H) (27) and to be involved in the metastasis and poor prognosis of patients with hepatocellular carcinoma (28). Recently, the analysis of the role of WDR1 in tumors has focused on the aberrant expression of WDR1 in several tumors, including breast cancer, thyroid neoplasia, ovarian carcinoma and glioblastoma (29-32). In particular, Izawa et al (30) reported that anti-WDR1 antibody levels were significantly high in thyroid carcinoma and could be a novel serological biomarker for papillary thyroid carcinoma. The results of the present study also revealed significantly high levels of WDR1 antibody in esophageal carcinoma. The high expression levels of WDR1 protein may be one of the causes of the development of s-WDR1-Abs.

It is well known that the polymerization and depolymerization of actin microfilaments play indispensable roles in cell division, migration, invasion and metastasis. Thus, regulated Table II. Univariate analysis of the risk factors associated with the overall survival of 91 patients with surgically treated for esophageal carcinoma.

	Univariate analysis P-value ^a
Sex Male/Female	0.046
Age, years >65/≤65	0.425
Tumor location Upper/lower	0.284
Tumor depth T1/T2-4	<0.001
Lymph node metastasis N⁻/N⁺	<0.001
Stage 0, I, II/III, IV	<0.001
SCC-Ag (ng/ml) >1.5/≤1.5	0.066
p53-Ab (U/ml) >1.30/≤1.30	0.063
WDR1-Ab >1865/≤1865	0.078
WBC (/µl) >8,000/≤8,000	0.994
Neutrophils (%) >70/≤70	0.127
Lymphocytes (%) >35/≤35	0.498
Hemoglobin (g/dl) ≤12/>12	0.095
Platelets (/µ1) ≤150,000/>150,000	0.194
CRP (mg/dl) >0.3/≤0.3	<0.001
Albumin (g/dl) ≤3.5/>3.5	0.172

^aP-values were obtained using the log-rank test. P-values in bold font indicate statistically significant differences (P<0.05). As regards tumor location, the oral side from the tracheal bifurcation was defined as the 'upper' location and the anal side from the tracheal bifurcation was defined as the 'lower' location. WDR1, WD repeat-containing protein 1; WDR1-Ab, serum anti-WDR1 antibody; N0, no lymph node metastasis; N1, lymph node metastasis exists; SCC-Ag, squamous cell carcinoma antigen; WBC, white blood cell; CRP, C-reactive protein.

polymerization and depolymerization may be required for cancer progression. As WDR1 enhances CFL1-mediated actin disassembly (33), the high expression of both WDR1 and CFL1 may synergistically disrupt cancer progression. Similar to the CFL1 activity, actin polymerization inhibitors, such as



Figure 2. Comparison and combination of the overall survival of patients with esophageal carcinoma in the s-WDR1-Ab and s-CFL1-Ab groups. The levels of s-WDR1-Ab and s-CFL1-Ab were divided into two groups (positive, s-WDR1-Ab⁺ and s-XFL1-Ab⁺; negative, s-WDR1-Ab- and s-CFL1-Ab⁻) using the cut-off values calculated using the X-tile analysis. Overall survival was examined using the Kaplan-Meier method. (A) s-WDR1-Ab⁺ and s-WDR1-Ab⁺, (B) s-CFL1-Ab⁺ and s-CFL1-Ab⁺, and (C) s-WDR1-Ab⁺/s-CFL1-Ab⁻ and s-WDR1-Ab⁺/s-CFL1-Ab⁺. The log-rank test was used to evaluate any significant differences between each group. WDR1, WD repeat-containing protein 1; CFL1, cofilin 1; s-WDR1-Ab, serum anti-WDR1 antibody; s-CFL1-Ab, serum anti-CFL1 antibody.

cytochalasin D and aplyronine A may be used as anticancer drugs (34,35). The low expression of CFL1, which cannot depolymerize an actin filament, may be compensated by the high expression of WDR1. This may account for the poor patient prognosis observed herein in the combined s-CFL1-Ab-negative and s-WDR1-Ab-positive groups (Fig. 2C).

Examining the expression levels of intracellular proteins in cancer tissues may be difficult. If the serum autoantibody levels reflect the intracellular antigenic protein expression levels, examining serum antibody levels is much easier and non-invasive. Some antigenic proteins can be leaked out from necrotic or apoptotic cancer cells but are hardly detectable because of their rapid degradation in the serum. Conversely, IgG antibodies are highly stable. Thus, antibody biomarkers are much more sensitive and stable to enable early detection and diagnosis. Although antibody biomarkers are not yet widely known, p53-Ab has been commonly used in clinical practice (22,23).

In conclusion, the present study demonstrated that serum anti-WDR1 antibody titers were significantly higher in patients with esophageal carcinoma than in healthy subjects. The combination of positive anti-WDR1 antibodies with negative anti-CFL1 antibodies may be a poor prognostic factor for patients with esophageal carcinoma.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

MI, YI, TH and HS conceived and designed the study. SY, TS, YO, TN, MS, FS and HT collected the blood samples and the clinicopathological data. SYL, LH and HW performed the experiments and acquired the data. HW, LH and HT contributed the reagents, materials, analysis tools or patient data. MI and SYL analyzed and interpreted the data. MI and SYL performed the statistical analyses. MI, TH and HS drafted the manuscript. MI, TH and HS confirm the authenticity of all the raw data. All authors have read and approved the final manuscript.

Ethics approval and consent to participate

The present study was conducted according to the guidelines of the Declaration of Helsinki. The study was approved by the Ethics Committee of Toho University, Graduate School of Medicine, Chiba University Graduate School of Medicine and the Kashiwado Memorial Foundation. Serum samples were collected from patients who had provided written informed consent.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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