

Therapeutic effect of Zeng Ye decoction on primary Sjögren's syndrome via upregulation of aquaporin-1 and aquaporin-5 expression levels

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Abstract. The aim of this study was to investigate the curative effect of Zeng Ye decoction on primary Sjögren's syndrome (pSS) and further explore its underlying mechanism involving aquaporin (AQP)-1 and AQP-5. The pSS model was established based on the immune induction method, and the saliva flow, submandibular gland index, morphological structures of salivary glands, and AQP-1 and AQP-5 protein expression levels in the salivary glands were determined. The saliva flow and the submandibular gland index were significantly reduced in the model group ($P < 0.01$, compared with those in the control group), and significantly increased following interferon (IFN), Zeng Ye decoction extraction (ZYE) and Zeng Ye decoction (ZY) treatment ($P < 0.01$, compared with those of the model group). Submandibular gland atrophy, fibrous tissue hyperplasia and multiple focal lymphocytic infiltration were observed in the model group and were attenuated when subjected to IFN, ZYE and ZY treatment. The AQP-1 and AQP-5 expression levels increased following IFN, ZYE and ZY treatment ($P < 0.01$, compared with those of the model group), particularly in the ZYE-35 group. This result indicated that ZYE had a significant protective effect on pSS via upregulation of the expression levels of AQP-1 and/or AQP-5. However, the AQP-1 expression levels increased and the AQP-5 expression levels decreased in the model groups compared with those in the control group, which indicated different regulatory

pathways of the salivary gland damage on the basis of AQP-1 and AQP-5. This study provided a significant reference for the prevention and treatment of pSS.

Introduction

Sjögren's syndrome (SS) is a multifaceted, chronic and systemic autoimmune disease, which predominantly occurs in females (1-4). The disease includes two processes: Primary SS (pSS) and secondary SS (sSS). pSS occurs in individuals with no other types of rheumatic disease (5,6) and sSS occurs in individuals who have another type of rheumatic disease, which is most often systemic lupus erythematosus or rheumatoid arthritis (7-9). SS predominantly affects the salivary and lacrimal glands and further results in dry eyes and, in particular, dry mouth (10-12). The damage to the salivary glands in SS cannot be reversed, but the symptoms are controllable. The curative effect of type-I interferon (IFN) treatment in SS has been well-documented (13-15) and thus, it is used as a positive control in the present study.

Zeng Ye decoction is extracted from figwort, *Ophiopogon japonicus* and *Rehmannia glutinosa* Libosch. In the field of traditional Chinese medicine (TCM), Zeng Ye decoction, as an important Chinese medicinal agent, has been widely used for relieving constipation due to body fluid deficiency, the condition which is figuratively described as 'boat stranding with water depletion' (16). The theory of 'increasing body fluid for curing constipation' was proposed by the famous medical scholar JuTong Wu with regard to epidemic febrile diseases (17). However, the protective effect of Zeng Ye decoction in pSS and its underlying mechanism has not previously been reported to the best of our knowledge. Therefore, the present study aimed to investigate the therapeutic effect of Zeng Ye decoction on pSS and further explore its underlying mechanism.

Materials and methods

Experimental animals and groups. A total of 48 female specific pathogen-free mice (age, 8-10 weeks; weight,

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18-20 g; Vital River Laboratory Animal Technology Co. Ltd., Beijing, China) were randomly divided into eight groups ($n=6$), including the control, adjuvant, model, IFN, Zeng Ye decoction extraction treatment after 50 days pSS (ZYE-50), Zeng Ye decoction treatment after 50 days pSS (ZY-50), Zeng Ye decoction extraction treatment after 35 days pSS (ZYE-35), and Zeng Ye decoction treatment after 35 days pSS (ZY-35) groups. This experiment was approved by the Ethics Committee of Beijing University of Chinese Medicine (Beijing, China). The Zeng Ye decoction consisted of 30 g figwort (*Xuan Shen*), 24 g *Ophiopogon japonicus* (Mai Dong), and 24 g *Rehmannia glutinosa* Libosch (Sheng Di) (Beijing Tongrentang Co., Ltd., Beijing, China), decocted for ~30 min to produce the solution of 1 g raw herbs per 1 ml decoction. To obtain the extracting solution of Zeng Ye decoction, the central composite design-response surface methodology and high-performance liquid chromatography were performed to extract and determine the active ingredients from three herbs (Ruscogenin from Mai Dong, Catalpol from Sheng Di, Harpagide and Harpagoside from Xuan Shen). These ingredients were then mixed and adjusted to produce a solution of 1 g raw herbs per 1 ml.

pSS model construction. The aforementioned mice were multi-point injected with pertussis vaccine (batch number: 20111258-2; Wuhan Institute of Biological Products Co., Ltd., Wuhan, China), complete Freund's adjuvant (CFA) and submandibular gland antigen (200 μ g/ml; volume: 0.5 ml) in the footpad and subcutaneously, and intraperitoneally injected with 0.05 ml pertussis vaccine. The control group did not receive the treatment and the adjuvant group was only injected with CFA (0.1 ml/footpad). One and seven days after the first immunization, the immunization was boosted with intraperitoneal injection of 2.9×10^{10} *Bordetella pertussis* to each group with the exception of the control group. After 21 days from the first immunization, further booster immunizations were performed every 10 days in the model group and treatment group, which involved intraperitoneal injection of 2.9×10^{10} *Bordetella pertussis* and dorsal subcutaneous multipoint injection of emulsified antigen. The adjuvant group were subjected to a further booster immunization with dorsal subcutaneous injection of CFA. Thirty-five and 50 days later, the ZYE-35, ZY-35, ZYE-50 and ZY-50 groups were lavaged with 0.8 ml/10 g Zeng Ye decoction, the control group were lavaged with an equal volume double-distilled H_2O and the IFN group was intraperitoneally injected with IFN α -2b (batch number: 20110571; Beijing Kawin Technology Share-Holding Co., Ltd., Beijing, China). Subsequently, the mice were sacrificed with cervical dislocation, the salivary gland was removed for sectioning, and the total protein was extracted to determine the aquaporin (AQP)-1 and AQP-5 expression levels.

Hematoxylin and eosin staining. The slides were deparaffinized and rehydrated, and the sections were marginally over-stained with hematoxylin (5 min) and fixed, and then excessive stain was removed with tap water. The slides were differentiated and destained for a few seconds in acidic alcohol until they appeared red (4-5 dips). The sections were briefly rinsed in tap water to remove the acid and stained blue in bicarbonate until the nuclei stood out sharply (~2 min).

The sections were rinsed under running tap water for 8 min, dehydrated and cleaned, or stained with eosin. Hematoxylin stained slides from the last tap water rinse were placed in 70% ethanol for 3 min. The slides were placed in eosin for 2 min, taken through three changes of 95% ethanol for 5 min and then transferred to the first absolute ethanol of the clearing series. The images were captured and analyzed by ImageJ software, version 1.47 (<http://rsb.info.nih.gov/ij/download.html>).

Immunohistochemical assay. The slides were deparaffinized, rehydrated and washed three times with phosphate-buffered saline (PBS; 5 min/time). Endogenous peroxidase was inactivated by incubating the sections with 3% H_2O_2 for 30 min. The sections underwent sequential incubations in 10% normal goat serum in PBS for 30 min at room temperature. Subsequently, the sections were incubated in mouse monoclonal anti-AQP-1 and AQP-5 (1:100; Abcam, Cambridge, UK) in PBS containing 0.3% Triton X-100 at 4°C overnight. After washing three times for 5 min with PBS, the sections were incubated in peroxidase-conjugated goat anti-mouse IgG (1:200; Zymed Laboratories, Carlsbad, CA, USA) for 1 h at room temperature. The sections were developed with diaminobenzidine (Sigma-Aldrich, St. Louis, MO, USA) in Tris-buffered saline containing 0.001% H_2O_2 for 30-50 min. The number of positive cells was measured by Image-Pro Plus software, version 7.0 (http://www.mediacy.com/index.aspx?page=IP_Premier) and analyzed by Origin software, version 9.0 (<http://www.originlab.com/index.aspx?go=Products/OriginPro>).

Western blot assay. The total protein of each group was extracted and quantified, and ~35 mg protein was separated by 12.5% SDS-PAGE. The separated protein was transferred to polyvinylidene difluoride membranes, and then incubated overnight with rabbit anti-mouse AQP-1 antibody (1:500; Abcam) and mouse anti-mouse AQP-5 antibody (1:500; Abcam). The blotted membranes were incubated for 2.5 h with horseradish peroxidase-labeled goat anti-rabbit secondary antibody (1:1,000; Santa Cruz Biotechnology, Inc., Santa Cruz, CA, USA). Subsequently, the protein bands were read with an electronic scanner and analyzed with the Image-Pro Plus software, version 7.0.

Statistical analysis. All data are expressed as the mean \pm standard deviation. Statistical analysis of the morphometrical quantification of the AQP-1 and AQP-5 positive cells was performed by means of the one-way analysis of variance test. Scheffe's test for group mean comparisons was used when two means were compared. $P<0.05$ and $P<0.01$ were considered to indicate a statistically significant difference.

Results

Construction of the pSS model. The saliva flow and the submandibular gland index were reduced in the model group compared with those in the control group and increased in the IFN, ZYE-50, ZY-50, ZYE-35 and ZY-35 groups compared with those in the model group ($P<0.01$; Fig. 1). This result indicated that the pSS model was correctly constructed, and

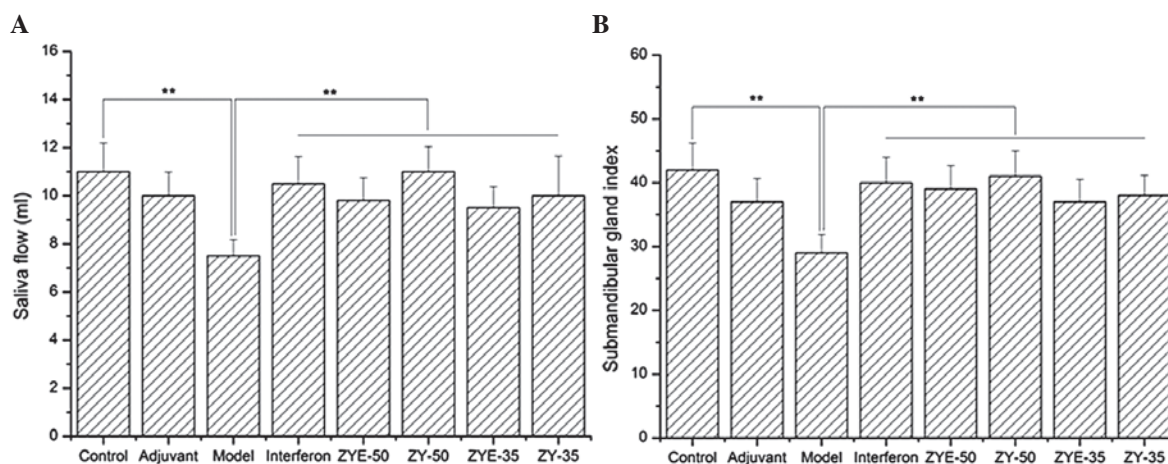


Figure 1. Histograms displaying the salivary gland flow and submandibular gland index. A histogram of (A) the salivary gland flow analysis and (B) the submandibular gland index analysis. The graphs indicated that Zeng Ye decoction increases the saliva flow and submandibular gland index. ** $P < 0.01$. ZYE, Zeng Ye decoction extraction; ZY, Zeng Ye decoction.

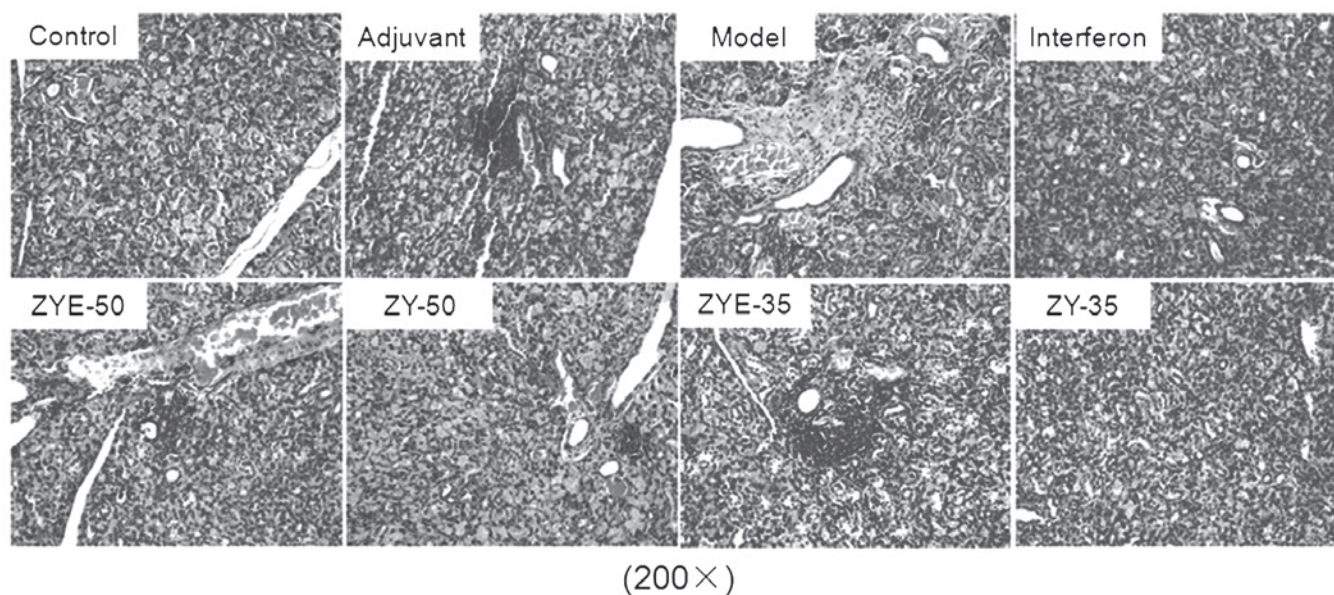


Figure 2. Morphological structure assay of the salivary glands by hematoxylin and eosin staining. The assay indicates that interferon and Zeng Ye decoction treatment effectively alleviate pSS damage (** $P < 0.01$, compared with that of the model group). ZYE, Zeng Ye decoction extraction; ZY, Zeng Ye decoction.

that Zeng Ye decoction facilitated the secretion of saliva and recovered the submandibular gland index.

Zeng Ye decoction alleviates the salivary gland damage after pSS, particularly in the ZYE-35 and ZYE-50 groups. The number of stained cells in the salivary gland was increased after the induction of pSS and the damage was effectively alleviated following IFN, ZYE and ZY treatment (Fig. 2). Submandibular gland atrophy, fibrous tissue hyperplasia and multiple focal lymphocytic infiltration were observed in the model group and were recovered when the mice were subjected to IFN, ZYE and ZY treatment.

Zeng Ye decoction increases AQP-1 and AQP-5 expression levels after pSS induction, particularly in the ZYE-35 group. The AQP-1 expression levels were elevated in the model

group compared with those in the control group ($P < 0.01$), and the AQP-1 expression levels were further increased when subjected to ZYE and ZY treatment. Furthermore, the number of cells positive for AQP-1 was significantly increased in the ZYE-35 group compared with that in the ZY-35 (Fig. 3; $P < 0.01$). The AQP-5 expression levels were reduced in the model group compared with those in the control group ($P < 0.01$), and the AQP-5 expression levels were elevated following the Zeng Ye decoction treatment. Furthermore, the number of cells positive for AQP-5 was significantly increased in the ZYE-35 group compared with that in the ZY-35 group (Fig. 4; $P < 0.01$).

Zeng Ye decoction increases AQP-1 and AQP-5 protein expression levels in pSS. The western blot analysis results demonstrated that the AQP-1 expression levels were increased

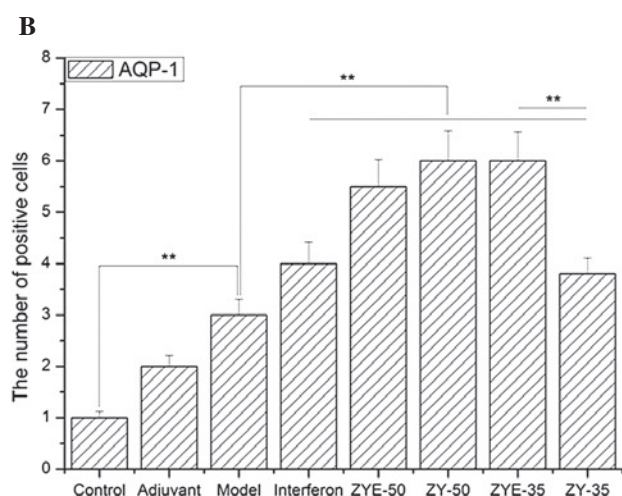
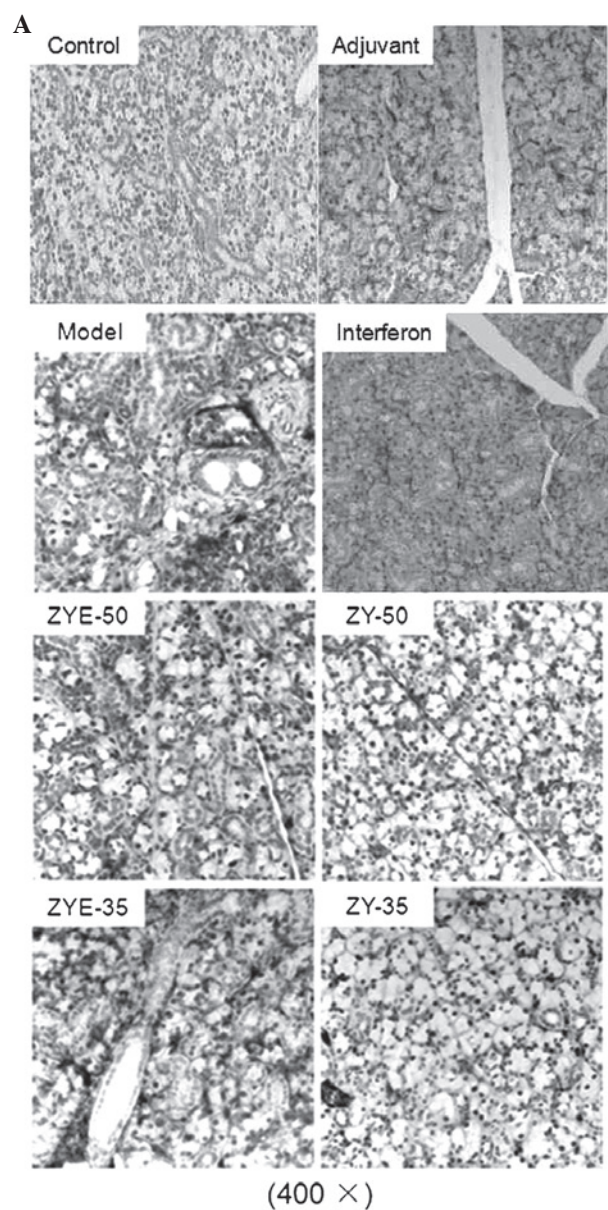


Figure 3. AQP-1 expression level assay in the salivary glands by immunohistochemistry and histogram analysis. (A) Immunohistochemical assay and (B) histogram analysis of the AQP-1 expression levels in the salivary glands. The results indicated that the AQP-1 expression levels were elevated when subjected to ZYE and ZY, particularly in the ZYE-35 group (** $P < 0.01$). ZYE, Zeng Ye docuction extraction; ZY, Zeng Ye docuction; AQP, aquaporin.

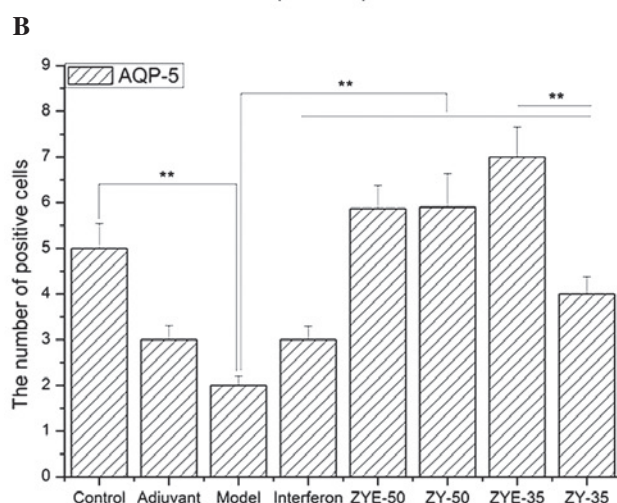
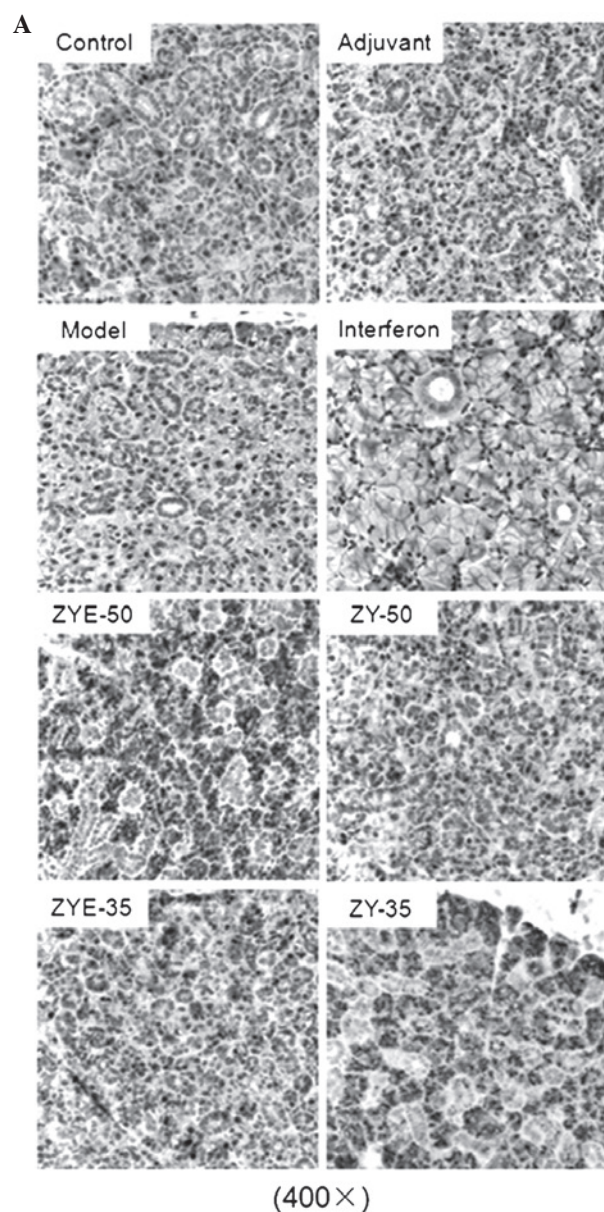


Figure 4. AQP-5 expression level assay in the salivary glands by immunohistochemistry and histogram analysis. (A) Immunohistochemical assay and (B) histogram analysis of the AQP-5 expression levels in the salivary glands. The results indicated that the AQP-5 expression levels were elevated when subjected to ZYE and ZY, particularly in the ZYE-35 group (** $P < 0.01$). ZYE, Zeng Ye docuction extraction; ZY, Zeng Ye docuction; AQP, aquaporin.

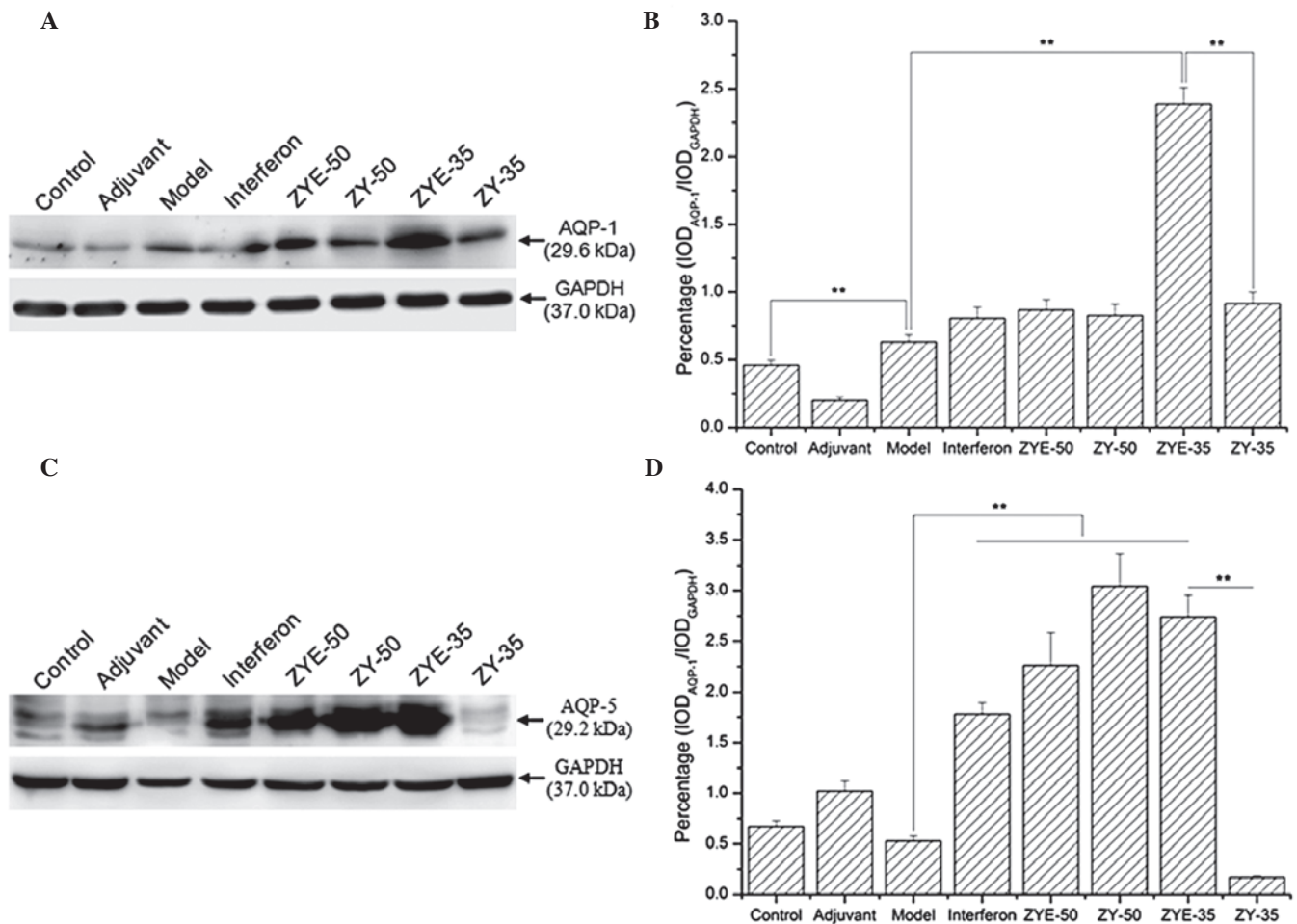


Figure 5. AQP-1 and AQP-5 expression levels assay in the salivary glands by western blot and histogram analysis. The AQP-1 expression levels assay in the salivary glands by (A) western blot and (B) histogram analysis. The AQP-5 expression levels assay in the salivary glands by (C) western blot and (D) histogram analysis. The results indicate that the AQP-1 and AQP-5 expression levels were increased when subjected to ZYE and ZY, particularly the ZYE-35 (** $P < 0.01$, compared with those of the model, ZY-35, ZYE-50 and ZY-50 groups). ZYE, Zeng Ye decoction extraction; ZY, Zeng Ye decoction; AQP, aquaporin.

in the model group compared with those in the control and further increased in the IFN, ZYE-50, ZY-50, ZYE-35 and ZY-35 groups, particularly in the ZYE-35 group (Fig. 5A; $P < 0.01$, compared with those of the model group). The AQP-5 expression levels were reduced in the model group and increased in the ZYE-50, ZYE-35 and ZY-50 groups compared with those in the control group. However, the AQP-5 expression levels were significantly reduced in the ZY-35 group, and indicated that ZYE facilitated the recovery of salivary gland damage via upregulation of the expression levels of AQP proteins and further alleviated the inflammatory response (Fig. 5A; $P < 0.01$, compared with those in the ZY group).

Discussion

The present study demonstrated that Zeng Ye decoction, a TCM, had a significant curative effect on pSS via upregulation of the levels of AQP-1 and AQP-5. The high-yield of these water channel proteins in the salivary glands facilitated the secretion of fluid, and was beneficial to the recovery from pSS. These results indicated that AQP-1 and AQP-5 may be novel targets in pSS and may provide a significant reference for the prevention and treatment of pSS.

SS is a chronic, multifaceted, high incidence, female-prone autoimmune disease, which specifically targets the salivary and lacrimal glands. The disease results in dry mouth and dry eyes, and further leads to multiple organ and tissue damage (9,18-20). The damage to the salivary glands in SS is severe, although several approaches are used to alleviate dry mouth in the clinic, including drinking water (21-24), chewing gum (25), using saliva substitutes (26) and using prescription medications (27,28). However, there are only methods to relieve the symptoms of the disease, none of which are curative, particularly in pSS. Therefore, the aim of the present study was to investigate the curative effect of Zeng Ye decoction on pSS and further study its underlying mechanism.

Zeng Ye decoction and its extraction had a significant curative effect on pSS in the present study via upregulation of AQP-1 and AQP-5 expression levels. The AQP-1 expression levels were upregulated and the AQP-5 expression levels were downregulated in the model group compared with those in the control group. This result indicated that SS had different regulatory patterns of AQP-1 and AQP-5. The different patterns may be associated with their different regulatory signaling pathways. Sabapathy (29) demonstrated that AQP-1 only regulated the JNK signaling pathway, and that AQP-5 is mainly involved in

the nuclear factor- κ B and/or p-c-Jun/c-Fos crosstalk signaling pathway in C3H/HeN mice following treatment with lipopolysaccharide and its inhibitor. Therefore, our future studies will further explore the regulatory signaling pathway of AQP1 and AQP5 in the pSS mouse model with Zeng Ye decoction.

In the present study, the different preparation techniques of Zeng Ye decoction, including extraction and water decoction, were selected to separate its effective constituent. The protective effect of the Zeng Ye decoction via different preparation techniques was consistent at 50 days of pSS. At 35 days of pSS, the AQP-1 and AQP-5 expression levels were significantly different between the extraction and water decoction groups. This result may be associated with the different immune mechanisms of the different ingredients of Zeng Ye decoction in the early development of pSS. Our future studies will also explore the detailed protective mechanisms of Zeng Ye decoction on the basis of AQP-1 and AQP-5.

Therefore, the present study demonstrated that Zeng Ye decoction had a significant curative effect in pSS. Zeng Ye decoction significantly upregulated the AQP-1 and AQP-5 expression levels via different mechanisms. This study provided a novel insight into a potential curative treatment for pSS and explored the implied possible mechanism of Zeng Ye decoction.

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