Hepatocyte growth factor activator inhibitor type-1 in cancer: Advances and perspectives (Review)

QIAOLI ZHENG 1 , HAIJIAN WU 1,2 , JIANG CAO 1 and JINGJIA YE 1

¹Clinical Research Center and ²Department of Neurosurgery, The Second Affiliated Hospital, Zhejiang University School of Medicine, Hangzhou, Zhejiang 310009, P.R. China

Received December 1, 2013; Accepted June 5, 2014

DOI: 10.3892/mmr.2014.2628

Abstract. Cancer is one of the most common diseases, with high morbidity and mortality rates. Large-scale efforts have been made to understand the pathogenesis of the disease, particularly in the advanced stages, in order to develop effective therapeutic approaches. Hepatocyte growth factor activator inhibitor type-1 (HAI-1), also known as serine protease inhibitor Kunitz type 1, inhibits the activity of several trypsin-like serine proteases. In particular, HAI-1 suppresses hepatocyte growth factor (HGF) activator and matriptase, resulting in subsequent inhibition of HGF/scatter factor and macrophage-stimulating protein (MSP). HGF and MSP are involved in cancer development and progression, via the receptors Met receptor tyrosine kinase (RTK) and Ron RTK, respectively. Therefore, HAI-1-mediated downregulation of HGF and MSP signaling may suppress tumorigenesis and progression in certain types of cancers. Abnormal HAI-1 expression levels have been observed in various types of human cancer. The exact function of HAI-1 in cancer pathogenesis, however, has not been fully elucidated. In this review, the focus is on the potential impact of aberrant HAI-1 expression levels on tumorigenesis and progression, the underlying mechanisms, and areas that require further investigation to clarify the precise role of HAI-1 in cancer.

Contents

- 1. Introduction
- 2. HAI-1 functional domains and cognate proteases inhibited by HAI-1

Correspondence to: Dr Jingjia Ye, Clinical Research Center, The Second Affiliated Hospital, Zhejiang University School of Medicine, 88 Jiefang Road, Hangzhou, Zhejiang 310009, P.R. China E-mail: yej001@zju.edu.cn

Key words: hepatocyte growth factor activator inhibitor type-1, serine protease inhibitor Kunitz type 1, cancer, mechanism, hepatocyte growth factor/scatter factor, hepatocyte growth factor activator, matriptase

- 3. Aberrant HAI-1 expression levels in cancer correlate with malignant phenotypes and clinicopathological parameters
- 4. Molecular mechanisms of HAI-1 in cancer
- 5. Conclusion and future perspectives

1. Introduction

Hepatocyte growth factor activator inhibitor type-1 (HAI-1), encoded by the *serine protease inhibitor*, *Kunitz type 1* gene, is a membrane-bound Kunitz-type serine protease inhibitor (1). HAI-1 was firstly purified from the conditioned medium of the MKN45 human stomach carcinoma cell line and identified as an inhibitor of hepatocyte growth factor activator (HGFA) (2). HAI-1 has also been demonstrated to inhibit a number of type-II transmembrane serine proteases (TTSPs), including matriptase, hepsin, transmembrane protease serine 13 (TMPRSS13) and human airway trypsin-like protease (HAT) (3-6). As a protein predominantly expressed in epithelial cells, HAI-1 is vital for cell growth, survival and mobility (1).

Increasing evidence has demonstrated that HAI-1 suppresses tumorigenesis and progression via regulation of the activity of a range of serine proteases in the tumor microenvironment. HGFA, a target trypsin-like serine protease of HAI-1, is secreted as a single-chain zymogen precursor and is activated by thrombin during blood coagulation. The activated HGFA induces the activation of two known macromolecular substrates, namely hepatocyte growth factor (HGF) and macrophage-stimulating protein (MSP), which are critical proteins involved in cancer pathogenesis (7,8). Downregulation of the activity of these two substrates through HAI-1-mediated HGFA inhibition, therefore, suppresses tumorigenesis and progression. In addition, TTSPs, another subtype of target HAI-1 proteases, facilitate epithelial carcinogenesis and progression (9). Therefore, HAI-1 is an important and promising therapeutic target in tumor treatment. This review focuses on recent advances in the understanding of HAI-1 with regard to the development and progression of cancer, and future studies concerning HAI-1 are proposed.

2. HAI-1 functional domains and proteases inhibited by HAI-1

HAI-1 is composed of an N-terminal extracellular region with two Kunitz domains (KD1 and KD2) separated by a

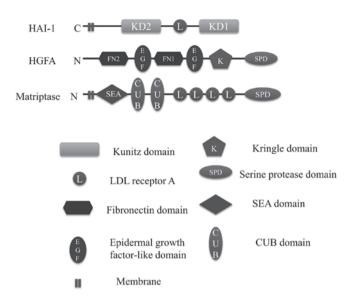


Figure 1. Domain structures of HAI-1, HGFA and matriptase. HAI-1, hepatocyte growth factor activator inhibitor type-1; HGFA, hepatocyte growth factor activator; LDL, low-density lipoprotein; SEA, sea urchin sperm protein/enteropeptidase/agrin; CUB, Cls/Clr, urchin embryonic growth factor, bone morphogenetic protein-1.

low-density lipoprotein receptor (LDLR)-like domain, a transmembrane region and a short cytoplasmic region (Fig. 1) (10). The primary transmembrane form (66 kDa) of HAI-1 is released as several soluble proteins (58, 48, 40 and 39 kDa) into the extracellular milieu by proteolytic cleavage (11). The transmembrane and soluble forms of HAI-1 exhibit inhibitory activity against serine proteases in the pericellular microenvironment (11,12). Among these HAI-1 molecules, the 58 kDa and 40 kDa HAI-1 proteins are the predominant soluble forms in the conditioned medium from cancer cell cultures. The 40 kDa HAI-1, which lacks KD2, exhibits higher inhibitory activity against HGFA than the 58 kDa band (11). However, studies have demonstrated that KD1 is also responsible for protease inhibition via interaction with target proteins (12-15). Furthermore, KD1-protease complex formation is enhanced by the LDLR-like domain but attenuated by KD2 (12).

HAI-1 exerts marked inhibitory activity against a variety of serine proteases, including HGFA, matriptase, hepsin, plasmin, trypsin, prostasin, TMPRSS13 and HAT (1,3-6,16,17). Studies have demonstrated that the proteases inhibited by HAI-1 clearly promote carcinogenesis and progression. For instance, HGFA expression is upregulated in breast, colorectal and renal cell carcinomas accompanied by downregulation of HAI-1 (18-20). Matriptase, another protease inhibited by HAI-1, is overexpressed in a variety of malignant tumors, and possesses the ability to promote oncogenesis and progression (1). Hepsinencoding gene, Hpn, is among the most consistently and quantitatively overexpressed genes in human prostate cancer, as detected by cDNA microarray and tissue array assays, and hepsin is the most reliable single marker to distinguish prostatic neoplasia from benign prostate hyperplasia (21-23). Prostasin, one glycosylphosphatidylinositol-anchored serine protease, has been reported to be upregulated in ovarian cancer but downregulated in high-grade prostate cancer (17,24,25). Therefore, HAI-1 may contribute to the prevention of cancer growth and progression via inhibition of these serine protease activities.

3. Aberrant HAI-1 expression levels in cancer correlate with malignant phenotypes and clinicopathological parameters

Aberrant HAI-1 expression levels have been demonstrated in various types of cancer and have diagnostic and prognostic implications. The expression profiles and functions of HAI-1 have been investigated extensively in pre-clinical and clinical studies (Table I).

Abnormal HAI-1 gene expression levels have been detected in a wide variety of human cancer cell lines; certain cell lines with a highly invasive nature exhibited low HAI-1 mRNA expression levels (26). In addition, as determined by in vitro and in vivo models, HAI-1 exerts a potential inhibitory effect on cancer cell invasion and migration, important hallmarks of cancer (27). Forced expression of HAI-1 significantly inhibited the invasion and migration of cervical, endometrial and uterine cancer cells in vitro (28-30). Furthermore, breast, pancreatic, prostate and oral carcinoma cells exhibited enhanced invasive properties in vitro in response to HAI-1 knockdown (31-34), a result also validated in nude mice bearing xenografts (33,35,36). Such findings are important for the analysis of the pathogenic role of HAI-1 in cancer cells. However, whether HAI-1 suppresses or promotes proliferation of cancer cells remains elusive (28-34,37).

Immunohistochemical (IHC) staining has detected reduced HAI-1 expression levels in endometrial, cervical and colorectal carcinomas and uterine leiomyosarcoma, compared with adjacent normal tissues (19,28-30). In addition, reduced HAI-1 mRNA levels have been detected by polymerase chain reaction in breast, gastric, colorectal and renal cell carcinoma (RCC) tissues (18-20,38). Further detailed analysis revealed that reduced HAI-1 expression levels were associated with worse clinicopathological parameters (advanced stage, lymph node metastasis and distant metastasis) and/or poor prognosis (reduced disease-free survival and overall survival times) in ovarian, gastric, cervical, endometrial, renal cell

Table I. Expressional and functional studies of HAI-1 in cancer.

Cancer type	Model	HAI-1 expression	Consequence/cancer association	Reference
Breast	MCF-7 cell line	High expression levels	Low invasion	Parr and Jiang(26)
	MDA MB-231 cell line	Low expression levels	High invasion	Parr and Jiang (26)
	MDA-MB-231 cell line	Knockdown	Enhanced migration, proliferation, invasion	Parr and Jiang (31)
	Breast cancer specimens	Lower levels in grade 3	Decreased in poorly differentiated tumors	Parr <i>et al</i> (18)
Colorectal	Primary colorectal carcinoma specimens	Lower levels in carcinoma tissue	Associated with disease progression	Kataoka et al (19)
Pancreatic	SUIT-2 cell line	Knockdown	Reduced cell growth, but enhance invasion	Cheng et al (34)
	SUIT-2 cell line, nude mice	Knockdown	Enhanced pulmonary metastasis	Fukushima et al (35)
Ovarian	Ovarian cancer specimen	Lower levels in stage III/IV	Loss of expression associated with advanced stage	Oberst et al (38)
Gastric	Gastric cancer specimens	Low expression levels	Associated with invasion and lymph node metastasis	Zeng et al (39)
Cervical	SiHa and HeLa cell lines	Overexpression	Inhibited growth, invasion, lead to apoptosis	Nakamura et al (28)
	Cervical cancer specimens	Low expression levels	Poor prognosis	Nakamura et al (28)
Endometrial	KLE and HEC-251 cell lines	Overexpression	Inhibited growth, invasion and migration	Nakamura et al (30)
	Endometrial cancer specimens	Low expression levels	Poor prognosis	Nakamura et al (30)
Uterine	SK-LMS-1 and SKN cell lines	Overexpression	Inhibited growth, invasion and migration	Nakamura et al (29)
	Uterine leiomyosarcoma specimens	Low expression levels	Poor prognosis	Nakamura et al (29)
Prostate	PC-3 and DU-145 cell lines	Knockdown	Inhibited growth, enhance invasion and migration	Sanders et al (32)
	Prostate cancer specimens	Low expression levels	Associated with increasing aggressiveness	Saleem et al (40)
	Prostate cancer samples	High mean serum level	Distant metastasis and hormone resistance	Nagakawa et al (47)
Kidney	Renal cell carcinoma specimen	Low expression levels	Involved in cancer progression	Yamauchi et al (20)
Oral cavity	HSC-3 and SAS cell lines	Knockdown	Reduced growth, but enhanced migration	Baba <i>et al</i> (33)
	SAS cell line, nude mice	Knockdown	Enhanced tumorigenicity	Baba <i>et al</i> (33)
	Oral squamous cell carcinoma specimens	Reduced expression levels at the invasion front	Associated with invasion, lymph node metastasis	Baba <i>et al</i> (33)
Liver	Hep3B cell line	Knockdown	Inhibited growth	Nagata et al (37)
	Hepatocellular carcinoma specimens	Positive in 35% cancer tissues	Involved in cancer progression	Nagata et al (37)
	Hepatocellular carcinoma specimens	Positive in 31% cancer tissues	Associated with poor prognosis	Funagayama et al (41)

HAI-1, hepatocyte growth factor activator inhibitor type-1.

and oral squamous cell carcinomas, and uterine leiomyosarcoma (19,20,28-30,33,39-40), but not in hepatocellular carcinoma (HCC) (37,41). Notably, HAI-1 was only marginally detectable in normal hepatocytes (42), while >30% HCC tissues were identified as HAI-1-positive by IHC (41), thus increased HAI-1 expression levels appear to be associated with advanced tumor stage and poor prognosis in HCC (37,41).

However, the exact role of HAI-1 in several types of cancer, including breast, colorectal and prostate cancer, remains controversial. Although reduced HAI-1 expression levels were associated with poorly differentiated breast cancer (18), high-level expression of HAI-I was found to be associated with poor patient outcome in a breast cancer tissue microarray analysis (43). HAI-1 downregulation in colorectal

cancer has been observed in a number of studies, but enhanced immunoreactivity of HAI-1 was detected in colorectal cancer cells at the invasion front, which may be involved in distant metastasis, although this trend was not statistically significant (44). In human prostate cancer tissues, the HAI-1 protein levels were elevated compared with those of benign prostate tissues (45,46). The mean serum levels of HAI-1 in 118 patients with prostate cancer were reported to be significantly higher than those in 27 patients with benign prostatic hyperplasia. Furthermore, increased HAI-1 levels in serum were associated with distant metastasis and the development of hormone-resistance in prostate cancer (47). However, another study observed using immunohistochemistry indicated that HAI-1 expression levels were reduced in all grades of prostate cancer specimens (40).

According to current research, HAI-1 may exhibit different functions in different types of cancer or even at different stages/sites in the same type of cancer (26,32,37,40,47). However, the differences in measuring HAI-1 expression levels, the lack of standardized methods (including antibody) among studies create difficulties in reaching a conclusion regarding HAI-1 expression in cancer and its association with clinicopathological parameters. Further studies with large samples and standardized criteria are warranted to elucidate the role of HAI-1 in tumor pathology, and to determine the diagnostic and prognostic value of HAI-1 expression.

4. Molecular mechanisms of HAI-1 in cancer

As described above, HAI-1 exerts a suppressive effect on cancer invasion and metastasis, processes which result in a poor prognosis for cancer patients (48); however the molecular basis of HAI-1-mediated cancer inhibition remains poorly understood. In the present review, advances in the understanding of the diverse molecular mechanisms regulating HAI-1-mediated effects via target serine proteases, particularly HGFA and matriptase, are summarized. Studies have shown that increased expression levels of HGFA and/or matriptase were accompanied by significantly downregulated HAI-1 expression. Thus, the net balance between HGFA/matriptase and HAI-1 was shifted in favor of HGFA/matriptase in various types of carcinoma, including breast, ovarian, renal, prostate and colorectal carcinoma (18-20,38,40,43,49,50). In addition, in vitro studies have validated the finding that HAI-1 knockdown-induced enhanced migration is partially reversed by silencing of matriptase or other serine protease expression (33,34,36).

The two best-characterized HAI-1-inhibited proteases (HGFA and matriptase) activate pro-HGF and pro-MSP, and are responsible for the subsequent activation of Met receptor tyrosine kinase (RTK) and Ron RTK, respectively (1,7,51). Dysregulation of the HGF-Met signaling pathway has been implicated in the development and metastasis of human cancer (52,53). Tumor xenografts with overexpressed HGF or Met exhibit high metastatic ability in mouse models (54-58). In addition, angiogenesis and lymphangiogenesis are promoted in tumors due to the induction of endothelial cell growth by HGF-Met cascade, as revealed by *in vitro* and *in vivo* studies (59-61). The downstream effectors of Met RTK activate several distinct signaling cascades, among

which the RAS-mitogen activated protein kinase (MAPK) and phosphatidylinositol 3 kinase (PI3K)-AKT signaling pathways are predominant. The RAS-MAPK cascade eventually activates the extracellular signal-regulated kinases (ERKs), which transmit signals downstream, and results in the transcription of genes controlling cell proliferation, differentiation, adhesion, migration and apoptosis (62). Activation of the PI3K-AKT-mammalian target of rapamycin signaling pathway results in cancer cell proliferation and invasion (63). Met activation may also enhance the function of Rap1 and modulate the adhesion molecules cadherin and integrin, and therefore promote cell migration (64,65). As with the signaling activation pattern activated by HGF-Met, MSP-Ron signaling is also mediated by the RAS-MAPK and PI3K-AKT signaling pathways (Fig. 2) (7). Therefore, HAI-1 inhibits tumor development and progression via suppression of protease-mediated downstream signaling pathways.

In addition, the activation of the RAS-MAPK and PI3K-AKT signaling pathways is crucial for RTK-mediated epithelial-to-mesenchymal transition (EMT) in cancer cells (7). EMT is recognized as a potential mechanism for carcinoma metastasis and the loss of E-cadherin is a hallmark of EMT (66). The predominant transcriptional repressors of E-cadherin are zinc finger transcription factors, including Snail (Snail), Slug, smad-interacting protein 1 (SIP1) and a basic helix-loop-helix transcription factor, Twist (67). An increasing amount of evidence has demonstrated that the interactions among HAI-1 and target serine proteases contribute to EMT in certain carcinoma cells. Support for this concept includes the finding that human pancreatic cancer cells with stable knockdown of HAI-1 exhibited an elongated spindle-like morphology and an enhanced migratory ability. Vimentin, SIP1 and matrix metalloproteinase (MMP)-9 expression was upregulated in these cells but E-cadherin expression was downregulated. The subsequent silencing of matriptase in these HAI-1 knockdown cells resulted in reversal in the expression levels of MMP-9 accompanied by a recovery of E-cadherin expression levels (34). In another study, HAI-1 overexpression resulted in a significant increase in E-cadherin expression levels but a reduction in Vimentin, SIP1, Snail and Twist expression levels in human endometrial cancer cell lines (30). The involvement of HAI-1 in EMT was further confirmed by other studies: Reduced E-cadherin expression levels in HAI-1-knockdown pancreatic cancer cells was reversed by recombinant KD1 (35) and HAI-1 knockdown oral squamous cell carcinoma cell lines exhibited more elongated morphology and reduced E-cadherin expression levels (33). All evidence reveals that HAI-1 inhibits tumor metastasis, partly by inhibiting EMT.

HAI-1 may also suppress the invasion and metastasis of tumor cells by inhibiting the activity of certain cognate serine proteases that activate fibrinolytic enzymes, MMPs and single-chain urokinase-type plasminogen activator (4,68-70). These enzymes are responsible for the degradation of extracellular matrix components and further potentiate local tumor invasion and metastasis (71).

Recently, a transgenic mouse model revealed that HAI-1 suppressed intestinal tumorigenesis. Enhanced tumor formation was observed in mice with deficient intestinal HAI-1 expression. Notably, a total of 22 genes (including those

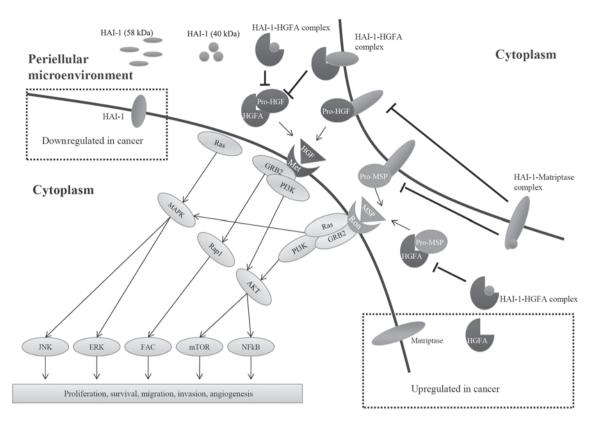


Figure 2. Signaling pathways mediated by HAI-1. HAI-1 is downregulated in cancer cells, whereas the HAI-inhibited proteases (HGFA and matriptase) are upregulated. Thus, the balance between HGFA (or matriptase) and HAI-1 is shifted in favor of HGFA (or matriptase) in cancer cells. HGFA and matriptase convert pro-HGF and pro-MSP into active HGF and MSP, which further bind to Met RTK and Ron RTK, respectively. The RAS-MAPK and PI3K-AKT signaling pathways are activated downstream of the HGF-Met and MSP-Ron complexes. These signaling pathways promote cell proliferation, survival, migration, invasion and angiogenesis. HAI-1, hepatocyte growth factor activator inhibitor type-1; HGF, hepatocyte growth factor; HGFA, HGF activator; MSP, macrophage-stimulating protein; RTK, receptor tyrosine kinase; MAPK, mitogen-activated protein kinase; PI3K, phosphatidylinositol 3 kinase; AKT, protein kinase B; GRB2, growth factor receptor-bound protein 2; JNK, c-Jun N-terminal kinase; ERK, extracellular signal-regulated kinase; mTOR, mammalian target of rapamycin; NFκB, nuclear factor kappa-light-chain-enhancer of activated B cells.

encoding ligands, receptors, transcription factors and downstream genes) associated with the Wnt signaling pathway were identified by microarray analysis to be augmented in the tumors. Furthermore, the expression of several other genes involved in mucosal permeability and angiogenesis, including *cldn2*, *lt1*, *cdh13*, *cdh5* and *tnfrsf12a*, was also upregulated (72). As barrier dysfunction may contribute to epithelial malignancy (73,74), HAI-1 may suppress tumorigenesis resulting from inhibition of the expression of these associated genes.

HAI-1 may exhibit different functions depending on the presence of cognate serine proteases in the intra- or extra-cellular milieu. The membrane-form HAI-1 acts not only as an inhibitor to HGFA, but also as an acceptor molecule, generating a reservoir of active HGFA on the cell surface; the HAI-1-HGFA complex on cell membrane may be dissociated and release the active HGFA into the surrounding microenvironment (75). Another study demonstrated that HAI-1 acted as an essential cofactor in the activation of pro-matriptase (76). Therefore, abnormal HAI-1 function may potentially contribute to tumor development and progression under specific conditions.

In conclusion, these findings established that HAI-1 is key in the development and progression of cancer; however, identification of the acute mechanism remains incomplete and requires further investigation.

5. Conclusion and future perspectives

HAI-1 is a vital protein involved in a number of biological and pathological processes due to its ability to inhibit cognate serine proteases in the extracellular milieu. The majority of these serine proteases are involved in the development and progression of cancer; therefore, HAI-1 exerts a suppressive function in cancer through regulation of these proteases.

Thus far, considerable achievements have been gained in the understanding of the pathological role of HAI-1 in tumors, particularly in the impact of aberrant HAI-1 expression levels on tumor growth, invasion, angiogenesis and metastasis. Existing studies have identified several of the molecular mechanisms mediated by HAI-1 and the target serine proteases. As determined by these findings, the prognostic and pharmaceutical properties of HAI-1 render the molecule a promising factor in cancer diagnosis and treatment.

However, paradoxical results have been obtained regarding HAI-1 expression patterns in certain types of cancer. The regulatory mechanisms that result in aberrant HAI-1 expression levels under different circumstances remain elusive. Further investigation into HAI-1 is important not only for providing greater insight into the molecular aspects of HAI-1 in cancer, but also for the possible development of novel diagnostic and therapeutic approaches. Even at the early stages of HAI-1 clinical investigation, understanding the acute roles of HAI-1

in cancer no doubt contributes, at least partly, to the eventual control of human cancer.

Acknowledgements

This study was supported by the National Natural Science Foundation of China (nos. 30271450 and 30672365) and the Natural Science Foundation of Zhejiang Province (no. 300466).

References

- Parr C, Sanders AJ and Jiang WG: Hepatocyte growth factor activation inhibitors - therapeutic potential in cancer. Anticancer Agents Med Chem 10: 47-57, 2010.
- 2. Shimomura T, Denda K, Kitamura A, *et al*: Hepatocyte growth factor activator inhibitor, a novel Kunitz-type serine protease inhibitor. J Biol Chem 272: 6370-6376, 1997.
- Lin CY, Anders J, Johnson M and Dickson RB: Purification and characterization of a complex containing matriptase and a Kunitz-type serine protease inhibitor from human milk. J Biol Chem 274: 18237-18242, 1999.
- 4. Kirchhofer D, Peek M, Lipari MT, *et al*: Hepsin activates pro-hepatocyte growth factor and is inhibited by hepatocyte growth factor activator inhibitor-1B (HAI-1B) and HAI-2. FEBS Lett 579: 1945-1950, 2005.
- Hashimoto T, Kato M, Shimomura T and Kitamura N: TMPRSS13, a type II transmembrane serine protease, is inhibited by hepatocyte growth factor activator inhibitor type 1 and activates pro-hepatocyte growth factor. FEBS J 277: 4888-4900, 2010.
- 6. Kato M, Hashimoto T, Shimomura T, *et al*: Hepatocyte growth factor activator inhibitor type 1 inhibits protease activity and proteolytic activation of human airway trypsin-like protease. J Biochem 151: 179-187, 2012.
- 7. Yao HP, Zhou YQ, Zhang R and Wang MH: MSP-RON signalling in cancer: pathogenesis and therapeutic potential. Nat Rev Cancer 13: 466-481, 2013.
- 8. Eigenbrot C, Ganesan R and Kirchhofer D: Hepatocyte growth factor activator (HGFA): molecular structure and interactions with HGFA inhibitor-1 (HAI-1). FEBS J 277: 2215-2222, 2010.
- 9. Bugge TH, Antalis TM and Wu Q: Type II transmembrane serine proteases. J Biol Chem 284: 23177-23181, 2009.
- Kataoka H, Miyata S, Uchinokura S and Itoh H: Roles of hepatocyte growth factor (HGF) activator and HGF activator inhibitor in the pericellular activation of HGF/scatter factor. Cancer Metastasis Rev 22: 223-236, 2003.
- 11. Shimomura T, Denda K, Kawaguchi T, *et al*: Multiple sites of proteolytic cleavage to release soluble forms of hepatocyte growth factor activator inhibitor type 1 from a transmembrane form. J Biochem 126: 821-828, 1999.
- 12. Kojima K, Tsuzuki S, Fushiki T and Inouye K: Roles of functional and structural domains of hepatocyte growth factor activator inhibitor type 1 in the inhibition of matriptase. J Biol Chem 283: 2478-2487, 2008.
- 13. Denda K, Shimomura T, Kawaguchi T, Miyazawa K and Kitamura N: Functional characterization of Kunitz domains in hepatocyte growth factor activator inhibitor type 1. J Biol Chem 277: 14053-14059, 2002.
- 14. Kirchhofer D, Peek M, Li W, et al: Tissue expression, protease specificity, and Kunitz domain functions of hepatocyte growth factor activator inhibitor-1B (HAI-1B), a new splice variant of HAI-1. J Biol Chem 278: 36341-36349, 2003.
- 15. Miyata S, Fukushima T, Kohama K, *et al*: Roles of Kunitz domains in the anti-invasive effect of hepatocyte growth factor activator inhibitor type 1 in human glioblastoma cells. Hum Cell 20: 100-106, 2007.
- 16. Shia S, Stamos J, Kirchhofer D, et al: Conformational lability in serine protease active sites: structures of hepatocyte growth factor activator (HGFA) alone and with the inhibitory domain from HGFA inhibitor-1B. J Mol Bio 346: 1335-1349, 2005.
- 17. Fan B, Wu TD, Li W and Kirchhofer D: Identification of hepatocyte growth factor activator inhibitor-1B as a potential physiological inhibitor of prostasin. J Biol Chem 280: 34513-34520, 2005.
- Parr C, Watkins G, Mansel RE and Jiang WG: The hepatocyte growth factor regulatory factors in human breast cancer. Clin Cancer Res 10: 202-211, 2004.

- Kataoka H, Hamasuna R, Itoh H, Kitamura N and Koono M: Activation of hepatocyte growth factor/scatter factor in colorectal carcinoma. Cancer Res 60: 6148-6159, 2000.
- 20. Yamauchi M, Kataoka H, Itoh H, *et al:* Hepatocyte growth factor activator inhibitor types 1 and 2 are expressed by tubular epithelium in kidney and down-regulated in renal cell carcinoma. J Urol 171: 890-896, 2004.
- 21. Dhanasekaran SM, Barrette TR, Ghosh D, *et al*: Delineation of prognostic biomarkers in prostate cancer. Nature 412: 822-826, 2001.
- 22. Wu QY and Parry G: Hepsin and prostate cancer. Front Biosci 12: 5052-5059, 2007.
- 23. Klezovitch O, Chevillet J, Mirosevich J, *et al*: Hepsin promotes prostate cancer progression and metastasis. Cancer Cell 6: 185-195, 2004.
- 24. Mok SC, Chao J, Skates S, *et al*: Prostasin, a potential serum marker for ovarian cancer: identification through microarray technology. J Natl Cancer Inst 93: 1458-1464, 2001.
- 25. Takahashi S, Suzuki S, Inaguma S, *et al*: Down-regulated expression of prostasin in high-grade or hormone-refractory human prostate cancers. Prostate 54: 187-193, 2003.
- 26. Parr C and Jiang WG: Expression of hepatocyte growth factor/scatter factor, its activator, inhibitors and the c-Met receptor in human cancer cells. Int J Oncol 19: 857-863, 2001.
- 27. Hanahan D and Weinberg RA: Hallmarks of cancer: the next generation. Cell 144: 646-674, 2011.
- 28. Nakamura K, Abarzua F, Hongo A, *et al*: The role of hepatocyte growth factor activator inhibitor-1 (HAI-1) as a prognostic indicator in cervical cancer. Int J Oncol 35: 239-248, 2009.
- Nakamura K, Abarzua F, Hongo A, et al: Hepatocyte growth factor activator inhibitors (HAI-1 and HAI-2) are potential targets in uterine leiomyosarcoma. Int J Oncol 37: 605-614, 2010.
- 30. Nakamura K, Hongo A, Kodama J and Hiramatsu Y: The role of hepatocyte growth factor activator inhibitor (HAI)-1 and HAI-2 in endometrial cancer. Int J Cancer 128: 2613-2624, 2011.
- 31. Parr C and Jiang WG: Hepatocyte growth factor activation inhibitors (HAI-1 and HAI-2) regulate HGF-induced invasion of human breast cancer cells. Int J Cancer 119: 1176-1183, 2006.
- 32. Sanders AJ, Parr C, Mason MD and Jiang WG: Suppression of hepatocyte growth factor activator inhibitor-1 leads to a more aggressive phenotype of prostate cancer cells *in vitro*. Int J Mol Med 20: 613-619, 2007.
- 33. Baba T, Kawaguchi M, Fukushima T, *et al*: Loss of membrane-bound serine protease inhibitor HAI-1 induces oral squamous cell carcinoma cells' invasiveness. J Pathol 228: 181-192, 2012.
- 34. Cheng H, Fukushima T, Takahashi N, Tanaka H and Kataoka H: Hepatocyte growth factor activator inhibitor type 1 regulates epithelial to mesenchymal transition through membrane-bound serine proteinases. Cancer Res 69: 1828-1835, 2009.
- 35. Fukushima T, Kawaguchi M, Yamasaki M, *et al*: Hepatocyte growth factor activator inhibitor type 1 suppresses metastatic pulmonary colonization of pancreatic carcinoma cells. Cancer Sci 102: 407-413, 2011.
- 36. Ye J, Kawaguchi M, Haruyama Y, *et al*: Loss of hepatocyte growth factor activator inhibitor type 1 participates in metastatic spreading of human pancreatic cancer cells in a mouse orthotopic transplantation model. Cancer Sci 105: 44-51, 2014.
- 37. Nagata K, Hirono S, Ido A, *et al*: Expression of hepatocyte growth factor activator and hepatocyte growth factor activator inhibitor type 1 in human hepatocellular carcinoma. Biochem Biophys Res Commun 289: 205-211, 2001.
- 38. Zeng L, Cao J and Zhang X: Expression of serine protease SNC19/matriptase and its inhibitor hepatocyte growth factor activator inhibitor type 1 in normal and malignant tissues of gastrointestinal tract. World J Gastroenterol 11: 6202-6207, 2005.
- 39. Oberst MD, Johnson MD, Dickson RB, *et al*: Expression of the serine protease matriptase and its inhibitor HAI-1 in epithelial ovarian cancer: correlation with clinical outcome and tumor clinicopathological parameters. Clin Cancer Res 8: 1101-1107, 2002.
- 40. Saleem M, Adhami VM, Zhong W, et al: A novel biomarker for staging human prostate adenocarcinoma: overexpression of matriptase with concomitant loss of its inhibitor, hepatocyte growth factor activator inhibitor-1. Cancer Epidemiol Biomarkers Prev 15: 217-227, 2006.
- 41. Funagayama M, Kondo K, Chijiiwa K and Kataoka H: Expression of hepatocyte growth factor activator inhibitor type 1 in human hepatocellular carcinoma and postoperative outcomes. World J Surg 34: 1563-1571, 2010.

- 42. Kataoka H, Suganuma T, Shimomura T, et al: Distribution of hepatocyte growth factor activator inhibitor type 1 (HAI-1) in human tissues. Cellular surface localization of HAI-1 in simple columnar epithelium and its modulated expression in injured and regenerative tissues. J Histochem Cytochem 47: 673-682,
- 43. Kang JY, Dolled-Filhart M, Ocal IT, et al: Tissue microarray analysis of hepatocyte growth factor/Met pathway components reveals a role for Met, matriptase, and hepatocyte growth factor activator inhibitor 1 in the progression of node-negative breast cancer. Cancer Res 63: 1101-1105, 2003.
- 44. Nagaike K, Kohama K, Uchiyama S, et al: Paradoxically enhanced immunoreactivity of hepatocyte growth factor activator inhibitor type 1 (HAI-1) in cancer cells at the invasion front. Cancer Sci 95: 728-735, 2004.
- 45. Warren M, Twohig M, Pier T, et al: Protein expression of matriptase and its cognate inhibitor HAI-1 in human prostate cancer: a tissue microarray and automated quantitative analysis. Appl Immunohistochem Mol Morphol 17: 23-30, 2009.
- 46. Yasuda K, Komiya A, Watanabe A, et al: Expression of Hepatocyte growth factor activator inhibitor type-Î (HAI-1) in prostate cancer. Anticancer Res 33: 575-581, 2013.
- 47. Nagakawa O, Yamagishi T, Akashi T, Nagaike K and Fuse H: Serum hepatocyte growth factor activator inhibitor type I (HAI-I) and type 2 (HAI-2) in prostate cancer. Prostate 66: 447-452, 2006.
- 48. Hudson BD, Kulp KS and Loots GG: Prostate cancer invasion and metastasis: insights from mining genomic data. Brief Funct Genomics 12: 397-410, 2013.
- 49. Vogel LK, Saebø M, Skjelbred CF, et al: The ratio of Matriptase/HAI-l mRNA is higher in colorectal cancer adenomas and carcinomas than corresponding tissue from control individuals. BMC Cancer 6: 176, 2006.
- 50. Tsai WC, Sheu LF, Chao YC, et al: Decreased Matriptase/HAI-1 ratio in advanced colorectal adenocarcinoma of Chinese patients. Chin J Physiol 50: 225-231, 2007.
- 51. Naldini L, Weidner KM, Vigna E, et al: Scatter factor and hepatocyte growth factor are indistinguishable ligands for the MET receptor. EMBO J 10: 2867-2878, 1991.
- 52. Graveel C, Su Y, Koeman J, et al: Activating Met mutations produce unique tumor profiles in mice with selective duplication of the mutant allele. Proc Natl Acad Sci USA 101: 17198-17203, 2004
- 53. Ponzo MG, Lesurf R, Petkiewicz S, et al: Met induces mammary tumors with diverse histologies and is associated with poor outcome and human basal breast cancer. Proc Natl Acad Sci USA 106: 12903-12908, 2009.
- 54. Rong S, Segal S, Anver M, Resau JH and Vande Woude GF: Invasiveness and metastasis of NIH 3T3 cells induced by Met-hepatocyte growth factor/scatter factor autocrine stimulation. Proc Natl Acad Sci USA 91: 4731-4735, 1994.
- 55. Meiners S, Brinkmann V, Naundorf H and Birchmeier W: Role of morphogenetic factors in metastasis of mammary carcinoma cells. Ôncogene 16: 9-20, 1998.
- 56. Gallego MI, Bierie B and Hennighausen L: Targeted expression of HGF/SF in mouse mammary epithelium leads to metastatic adenosquamous carcinomas through the activation of multiple signal transduction pathways. Oncogene 22: 8498-8508, 2003.
- 57. Jeffers M, Fiscella M, Webb CP, et al: The mutationally activated Met receptor mediates motility and metastasis. Proc Natl Acad Sci USA 95: 14417-14422, 1998.

- 58. Moshitch-Moshkovitz S, Tsarfaty G, Kaufman DW, et al: In vivo direct molecular imaging of early tumorigenesis and malignant progression induced by transgenic expression of GFP-Met. Neoplasia 8: 353-363, 2006.
- 59. Abounader R and Laterra J: Scatter factor/hepatocyte growth factor in brain tumor growth and angiogenesis. Neuro Oncol 7: 436-451, 2005
- 60. Bussolino F, Di Renzo MF, Ziche M, et al: Hepatocyte growth factor is a potent angiogenic factor which stimulates endothelial cell motility and growth. J Cell Biol 119: 629-641, 1992.
- 61. Grant DS, Kleinman HK, Goldberg ID, et al: Scatter factor induces blood vessel formation in vivo. Proc Natl Acad Sci USA 90: 1937-1941, 1993
- 62. Santarpia L, Lippman SM and El-Naggar AK: Targeting the MAPK-RAS-RAF signaling pathway in cancer therapy. Expert Opin Ther Targets 16: 103-119, 2012.
- 63. Sami A and Karsy M: Targeting the PI3K/AKT/mTOR signaling pathway in glioblastoma: novel therapeutic agents and advances in understanding. Tumour Biol 34: 1991-2002, 2013.
- 64. Gherardi E, Birchmeier W, Birchmeier C and Vande Woude G: Targeting MET in cancer: rationale and progress. Nat Rev Cancer 12: 89-103, 2012.
- 65. Boettner B and Van Aelst L: Control of cell adhesion dynamics by Rap1 signaling. Curr Opin Cell Biol 21: 684-693, 2009.
- 66. Thiery JP: Epithelial-mesenchymal transitions in tumour
- progression. Nat Rev Cancer 2: 442-454, 2002. 67. Berx G, Raspé E, Christofori G, Thiery JP and Sleeman JP: Pre-EMTing metastasis? Recapitulation of morphogenetic processes in cancer. Clin Exp Metastasis 24: 587-597, 2007.
- 68. Herter S, Piper DE, Aaron W, et al: Hepatocyte growth factor is a preferred in vitro substrate for human hepsin, a membrane-anchored serine protease implicated in prostate and ovarian cancers. Biochem J 390: 125-136, 2005.
- 69. Lee SL, Dickson RB and Lin CY: Activation of hepatocyte growth factor and urokinase/plasminogen activator by matriptase, an epithelial membrane serine protease. J Biol Chem 275: 36720-36725, 2000.
- 70. Takeuchi T, Harris JL, Huang W, et al: Cellular localization of membrane-type serine protease 1 and identification of protease-activated receptor-2 and single-chain urokinase-type plasminogen activator as substrates. J Biol Chem 275: 26333-26342, 2000.
- 71. Liotta LA: Tumor invasion and metastases role of the extracellular matrix: Rhoads Memorial Award lecture. Cancer Res 46: 1-7, 1986.
- 72. Hoshiko S, Kawaguchi M, Fukushima T, et al: Hepatocyte growth factor activator inhibitor type 1 is a suppressor of intestinal tumorigenesis. Cancer Res 73: 2659-2670, 2013.
- 73. Turner JR: Intestinal mucosal barrier function in health and disease. Nat Rev Immunol 9: 799-809, 2009.
- 74. Turksen K and Troy TC: Junctions gone bad: Claudins and loss of the barrier in cancer. Biochim Biophys Acta 1816: 73-79, 2011.
- 75. Kataoka H, Shimomura T, Kawaguchi T, et al: Hepatocyte growth factor activator inhibitor type 1 is a specific cell surface binding protein of hepatocyte growth factor activator (HGFA) and regulates HGFA activity in the pericellular microenvironment. J Biol Chem 275: 40453-40462, 2000.

 76. Oberst MD, Williams CA, Dickson RB, Johnson MD and
- Lin CY: The activation of matriptase requires its noncatalytic domains, serine protease domain, and its cognate inhibitor. J Biol Chem 278: 26773-26779, 2003.