

RNA silencing of integrin-linked kinase increases the sensitivity of the A549 lung cancer cell line to cisplatin and promotes its apoptosis

XIAOZHEN ZHAO¹, ZHENYE XU¹, ZHONGQI WANG¹, ZHONGHUA WU²,
YABIN GONG¹, LIJUAN ZHOU¹ and YI XIANG¹

¹Department of Oncology, Longhua Hospital, Shanghai University of Traditional Chinese Medicine, Shanghai 200032;

²Experiment Center for Science and Technology, Shanghai University of Traditional Chinese Medicine, Shanghai 201203, P.R. China

Received February 13, 2014; Accepted November 3, 2014

DOI: 10.3892/mmr.2015.3471

Abstract. The expression of integrin-linked kinase (ILK) has been reported to be involved in the regulation of integrin-mediated processes, including cancer cell proliferation, migration and invasion. Previous studies have demonstrated that inhibition of ILK may be an underlying approach for treating cancer. However, whether the knock down of ILK affects growth and apoptosis of lung cancer cells remains to be elucidated. Importantly, whether downregulation of ILK increases the sensitivity of lung cancer cells to cisplatin and amplifies cell apoptosis also remains to be elucidated. In the present study, ILK downregulation was mediated by lentivirus-mediated RNA interference. The expression levels of associated genes were determined by reverse-transcription quantitative polymerase chain reaction and western blotting. Cell proliferation was evaluated using a modified 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide assay and clone formation assay. The cell cycle and apoptosis were analyzed using flow cytometry. The current data revealed that lentivirus-mediated ILK gene silencing alone inhibited A549 cell proliferation and promotes cell cycle arrest, however, had no detectable effect on cell apoptosis. However, combined treatment with lentivirus-mediated ILK interference and cisplatin chemotherapy induced significantly more cell apoptosis than mono-chemotherapy or knockdown. The increased cell apoptosis and proliferation inhibition were attributed to abnormal downstream protein expression of ILK, including phospho-glycogen synthase kinase 3 β , p-AKT, activator protein-1, β -catenin, cyclin D1 and matrix metalloproteinase-9.

ILK inhibition may suppress the proliferation of A549 and increase A549 sensitivity to cisplatin. The combined treatment of ILK gene knockdown and chemotherapy has the potential to improve anticancer efficacy.

Introduction

Lung cancer is currently the leading cause of cancer-associated mortality worldwide, being responsible for an estimated 87,750 mortalities in males and 72,590 in females in 2012 (1). The majority of patients have advanced disease (stage III/IV) at initial diagnosis, which contributes to the poor prognosis of these patients. It is reported that the 5-year overall survival rate is <5% among patients with advanced-stage disease (2). Chemotherapy, including cisplatin (CDDP), carboplatin and oxaliplatin, remains a crucial treatment for advanced patients (3). Platinum drugs cross-link at the nucleophilic centers on the DNA of tumor cells, forming intra- (on the same strand) and/or inter-strand (between two opposite strands) adducts between guanines or guanine and adenine, which inhibit DNA replication and trigger cell cycle arrest and apoptosis (4,5). However, the major limitation in the clinical applications of platinum compounds is the development of tumor drug resistance (6-8). Therefore, it is essential to develop more effective treatment strategies to overcome this drug resistance.

Integrin-linked kinase (ILK), a highly conserved, 59 kDa serine/threonine kinase, has been implicated in the regulation of various biological processes that are crucial to the progression of malignant disease (9). ILK expression and activity have been revealed to be increased in association with tumor grade, T status, lymph node metastasis and survival in lung cancer patients (10-12). ILK promotes lung cancer cell migration and invasion via upregulation of matrix metalloproteinase-9 (MMP-9) (13) and epithelial-mesenchymal transition-associated genes, including vimentin, fibronectin, Snail and Slug (14). Further studies indicate that silencing ILK through targeting small interfering RNA (siRNA) inhibits cell proliferation and growth, induces cell cycle arrest and the apoptosis of bladder (15) and pancreatic cancer cells (16),

Correspondence to: Dr Xiaozhen Zhao, Department of Oncology, Longhua Hospital, Shanghai University of Traditional Chinese Medicine, 725 Wanping Road, Shanghai 200032, P.R. China
E-mail: zxiaozhen@hotmail.com

Key words: lentivirus, RNA interference, integrin-linked kinase, cisplatin, lung cancer

suggesting the inhibition of ILK may be a novel approach for treating lung cancer. Notably, Song *et al* (17) previously demonstrated that downregulation of ILK by siRNA arrests the growth and increases the CDDP sensitivity and apoptotic rate of human gastric cell line cells that are resistant to SGC7901/CDDP. Thus, it is hypothesized that there may be a synergistic interaction between downregulation of ILK and CDDP administration for treating lung cancer by creating cytotoxic DNA lesions and affecting apoptosis in lung cancer A549 cells. To the best of our knowledge, the present study is the first to examine this mechanism.

Materials and methods

Cell culture. The human lung adenocarcinoma cell line A549 and human embryo kidney (HEK) 293T cells (American Type Culture Collection, Manassas, VA, USA) were maintained in Dulbecco's modified Eagle's medium (Invitrogen Life Technologies, Carlsbad, CA, USA) containing 10% fetal bovine serum (Invitrogen Life Technologies) and cultured in a humidified atmosphere of 5% CO₂ at 37°C.

Construction of lentiviral vectors expressing siRNA targeting ILK and transfection. The oligonucleotides encoding a negative control (NC) siRNA with no homology to the human genome (5'-AAT GTA CTG CGC GTG GAG A-3') and ILK siRNA (5'-CCT TCA ACT TTG TGC TCA T-3') were designed and synthesized by Shanghai Jikai Gene Chemical Co., Ltd, (Shanghai, China) and cloned into the *AgeI/EcoRI* linearized pGCSIL-GFP viral vector (GeneChem, Shanghai, China) to generate the lentiviral vectors. Expression of the lentiviral shRNA was confirmed by DNA sequencing. Plasmids along with 20 µg of pGCSIL-shILK or -shNC lentiviral vectors, 15 µg of packaging vectors pHelper 1.0 and 10 µg of packaging vectors pHelper 2.0 were mixed with 200 µl of Opti-MEM and 15 µl of Lipofectamine 2000 and then transfected into HEK293T cells. After 48 h transfection, lentiviruses were harvested in serum-free medium, filtered and concentrated in primed Centricon Plus-20 filter devices (Millipore, Billerica, MA, USA). Subsequently, A549 cells were infected with ILK-shRNA lentivirus or control lentivirus at a multiplicity of infection of 20. The number of green fluorescent protein (GFP)-positive cells was determined microscopically (MicroPublisher 3.3RTV; Olympus, Tokyo, Japan) three days post-transduction.

RNA extraction and reverse transcription quantitative polymerase chain reaction (RT-qPCR). Total RNA was extracted from cells with TRIzol reagent (Invitrogen Life Technologies, Carlsbad, CA, USA) according to the manufacturer's instructions. Gene expression was detected by RT-qPCR using the standard SYBR Green RT-PCR kit (Takara Bio, Inc., Shiga, Japan). Briefly, the cDNA was synthesized using the RevertAid First-Strand cDNA Synthesis kit (Fermentas, Vilnius, Lithuania) according to the manufacturer's instructions. The specific primer pairs and the amplified products were as follows: *ILK* (212 bp), sense 5'-TCCACCTGCTCCTCATCC-3' and antisense 5'-CCTCATCAATCATTACACTACGG-3' and *GAPDH* (121 bp), sense 5'-TGACTTCAACAGCGACACCCA-3' and antisense 5'-CACCTGTTGCTGTAGCCAAA-3'. The relative levels of gene mRNA transcripts were normalized to the internal

control *GAPDH*. Relative gene expression was quantified using the GraphPad Prism 4.0 software (GraphPad Software, San Diego, CA, USA).

Western blotting. Cells were lysed in 0.1 ml lysis buffer (0.1% SDS, 1% NP-40, 50 mM HEPES, pH 7.4, 2 mM EDTA, 100 mM NaCl, 5 mM sodium orthovanadate and 1% protease inhibitor mixture set I; Calbiochem, San Diego, CA, USA) on ice for 30 min. Following centrifugation at 13,400 × g (Eppendorf centrifuge 5415 D; Eppendorf AG, Hamburg, Germany) for 15 min, the supernatants were removed and total protein concentration was determined using a bicinchoninic acid protein assay kit (Pierce Biotechnology, Inc., Rockford, IL, USA). Proteins (20 µg) were separated in 10% sodium dodecyl sulfate polyacrylamide gel electrophoresis and transferred onto a polyvinylidene difluoride (PVDF) membrane at 400 mA for 2 h. Following blocking at room temperature in 5% bovine serum albumin for 1.5 h, PVDF membranes were incubated with the following antibodies: Rabbit anti-ILK polyclonal antibody (Santa Cruz Biotechnology, Inc., Santa Cruz, CA, USA), rabbit anti-phospho-glycogen synthase kinase (GSK)-3β-S9 monoclonal antibody (1:1,000), rabbit anti-p-Akt-S473 polyclonal antibody (1:1,000), rabbit anti-MMP-9 (1:200), rabbit anti-activator protein (AP-1; 1:800), rabbit anti-β-catenin polyclonal antibody (1:1,000), rabbit anti-cyclin D1 monoclonal antibody (1:500), rabbit anti-vascular endothelial growth factor polyclonal antibody (VEGF; 1:500), all purchased from Cell Signaling Technology, Inc. (Danvers, MA, USA) and mouse anti-GAPDH monoclonal antibody (Santa Cruz Biotechnology, Inc.; 1:4,000), followed by incubation with the correspondent peroxidase-conjugated secondary antibodies (goat anti-rabbit IgG; 1:4,000; goat anti-mouse IgG; Santa Cruz Biotechnology, Inc.; 1:4,000). Chemiluminescent detection was performed with the enhanced chemiluminescence kit (Pierce Biotechnology, Inc.).

3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay for assessment of proliferation and drug sensitivity. The experimental cells in the exponential phase of growth were plated at a final concentration of 2×10³ cells/well in 96-well culture plates for different culture time periods. Varied concentrations of CDDP (5, 10, 15, 20 and 25%) were added to each well and the cells were incubated for 72 h. MTT (10 µl, 5 mg/ml) was then added. Following an additional 4 h of incubation, the reaction was terminated by removal of the supernatant and addition of 100 µl dimethyl sulfoxide for 30 min. The optical density of each well was measured at 490 nm using an ELISA reader (ELx808; Bio-Tek Instruments, Winooski, VT, USA).

Detection of apoptosis by flow cytometry. Cells were stained with fluorescein isothiocyanate (FITC)-labeled annexin-V and simultaneously with propidium iodide (PI), to discriminate intact cells (annexin-/PI-) from apoptotic cells (annexin+/PI-) and necrotic cells (annexin+/PI+). A total of 1.0×10⁶ cells were washed twice with ice-cold phosphate-buffered saline (PBS) and incubated for 30 min in a binding buffer (1 µg/ml PI and 1 µg/ml FITC-labeled annexin-V), respectively. Fluorescence-activated cell sorting analysis for annexin-V and PI staining was performed using a flow cytometer (Beckman Coulter, Inc., Fullerton, CA, USA). All experiments were performed in triplicate.

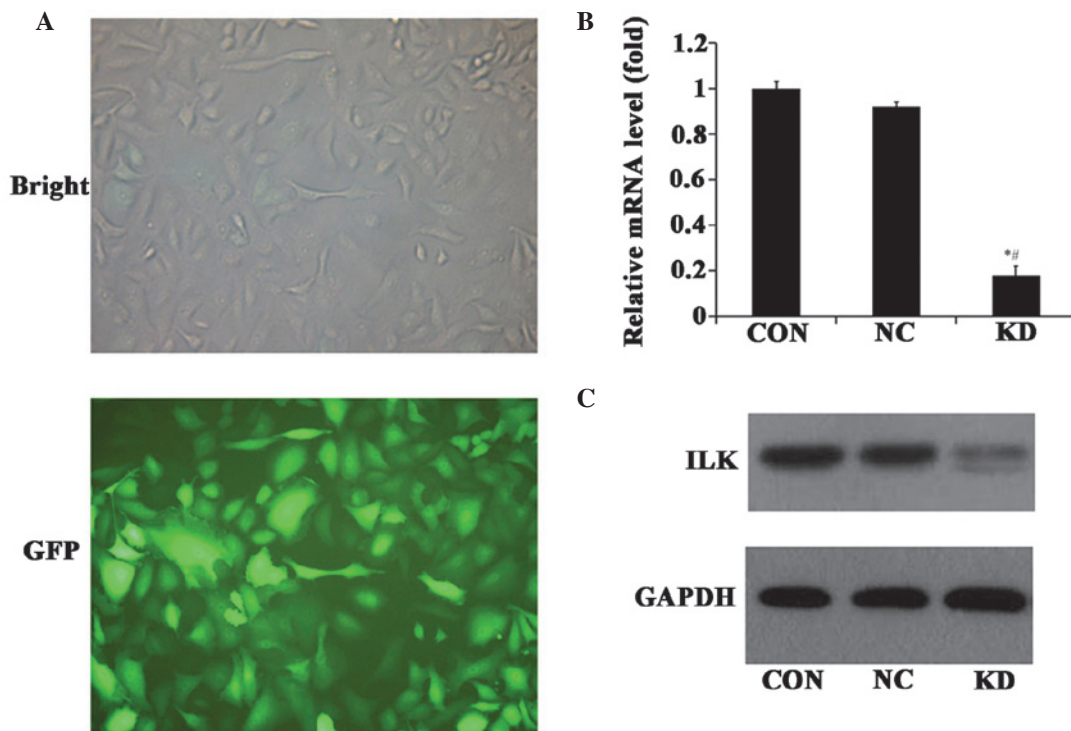


Figure 1. Downregulation of ILK expression by lentivirus-mediated RNAi. (A) Expression of lentivirus-mediated RNAi in A549 cells. At 3 days after infection with lentivirus, A549 cells were visualized using a fluorescence microscope. The phase contrast (bright) and fluorescence (GFP; green) images were captured with the same exposure times at magnification, x200. (B) Relative levels of *ILK* mRNA transcripts were analyzed by RT-qPCR. Data are presented as the mean \pm standard deviation of a representative experiment performed in triplicate (n=3; *P<0.05, compared with the Con group; #P<0.05, compared with the NC group). (C) Silencing effects of ILK protein were measured by western blot analysis. CON, parent cells; NC, cells transfected with negative control RNAi; KD, cells transfected with ILK specific RNAi; ILK, integrin-linked kinase; GFP, green fluorescent protein.

Cell cycle assay. Cells were seeded into a 6-well plate, harvested by centrifugation at 13,40 x g for 5 min (Eppendorf centrifuge 5415 D; Eppendorf AG). Following being washed twice in pre-cooled PBS (pH 7.4), cells were fixed in 70% alcohol. The percentage of cells in each stage of the cell cycle was determined by staining with PI. The analysis of cell cycle distribution was performed by FACSscan flow cytometer (Becton-Dickinson, Franklin Lakes, NJ, USA) in accordance with the manufacturer's instructions.

Colony-forming assay. Exponentially growing cells transfected with ILK-RNAi-lentivirus and negative control lentivirus were suspended in complete growth medium and seeded in 6-well plates at 200 cells per well. The plates were maintained at 37°C in a humidified incubator with 5% CO₂ for 2 weeks. The visible colonies were subsequently recorded under an inverted fluorescence microscope (MicroPublisher 3.3RTV; Olympus). Following fixation in paraformaldehyde, the colonies were subjected to Giemsa (Karyomax; Gibco, Grand Island, NY, USA) staining for 10 min followed by acquisition of images with an Olympus C5050 digital camera attached to an Olympus CKX1 inverted microscope (Olympus).

Statistical analysis. Data are expressed as the mean \pm standard deviation. Statistical analysis was performed using SPSS 11.0 software (SPSS, Inc., Chicago, IL, USA). The difference between two groups was analyzed using Student's t-test. P<0.05 was considered to indicate a statistically significant difference.

Results

ILK expression in A549 cells following treatment with lentivirus-mediated RNAi. At 3 days after transfection, A549 cells were visualized using a fluorescence microscope and the mRNA and protein levels of ILK were also analyzed. The results revealed that >80% of cells had green fluorescent signals compared with the bright field (Fig. 1A). The level of ILK mRNA in ILK specific-RNAi infected cells was significantly decreased by ~70% (P<0.05), compared with the control RNAi-infected cells (Fig. 1B). In accordance with the silencing of mRNA expression, ILK protein was also down-regulated in ILK RNAi cells (Fig. 1C). These findings suggest that ILK-specific RNAi may downregulate ILK expression efficiently.

Effect of ILK knockdown on cell proliferation, clone formation, the cell cycle and cell apoptosis in vitro. To assess the effects of ILK knockdown on cell proliferation of the cell line, A549, an MTT assay was performed. As expected, cells with ILK RNAi exhibited an inhibited cell proliferation ability compared with the control cells (Fig. 2A). In line with the MTT assay, the quantity and size of the colony in the ILK RNAi-infected cells were also significantly decreased compared with the control cells (Fig. 2B). In order to elucidate whether lentivirus-mediated ILK RNAi had any effects on the cell cycle of A549 cells, all three groups of A549 cells were subjected to a flow cytometry assay after 3 days of infection. The results revealed that ILK-shRNA-lentivirus infected cells

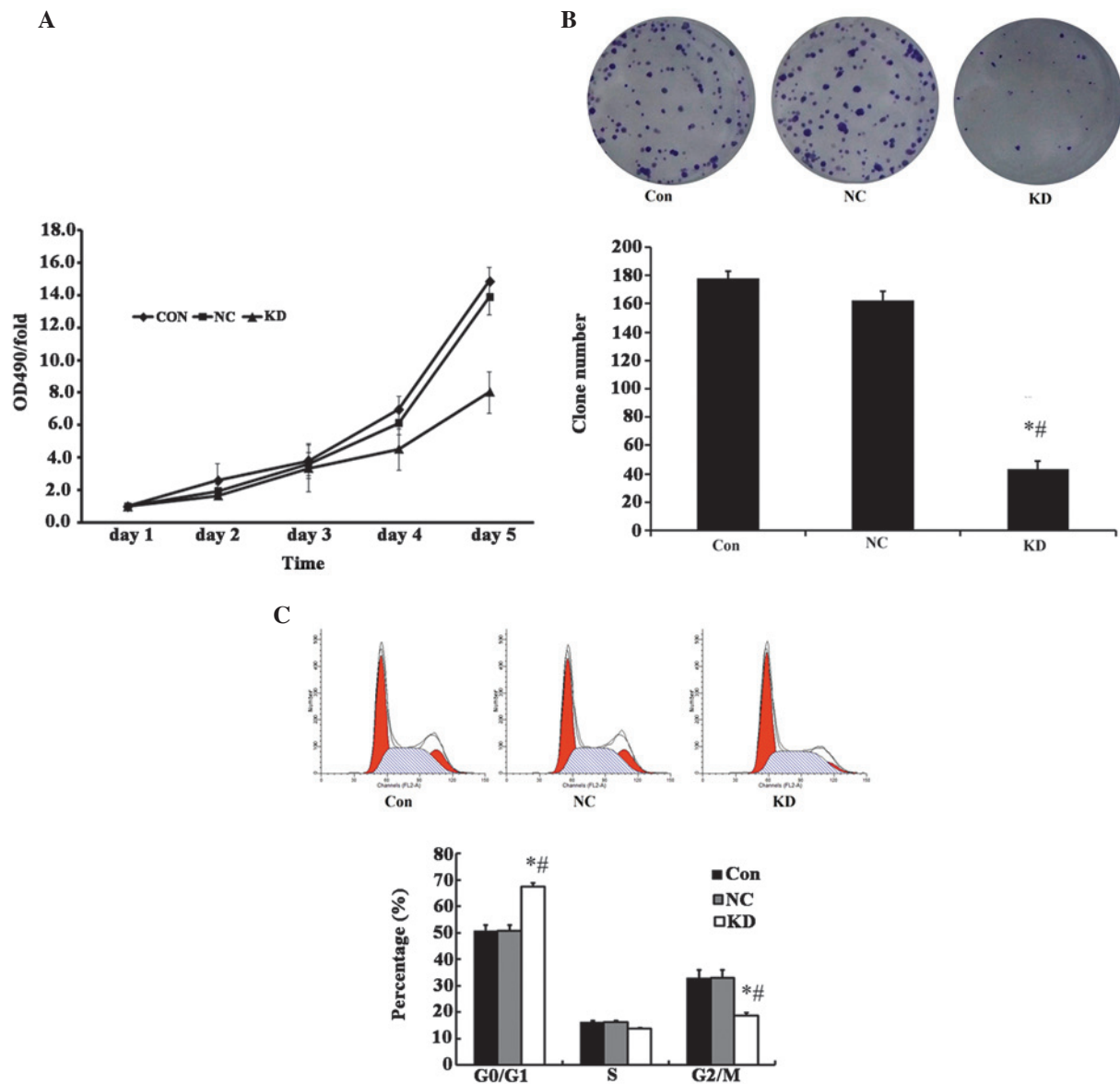


Figure 2. Functional effect of ILK knockdown. (A) Knockdown of ILK inhibited the cell proliferation of A549 cells [measured using a 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide assay], (B) reduced the colony formation ability (measured by colony formation assay), (C) augmented the proportion of G0/G1 phase (the proportion of different cell cycle phases was quantitated by propidium iodide staining followed by flow cytometric analysis). * $P < 0.05$, compared with the Con group; # $P < 0.05$, compared with the NC group. CON, parent cells; NC, cells transfected with negative control RNAi; KD, cells transfected with ILK specific RNAi; ILK, integrin-linked kinase; OD, optical density.

exhibited an increased proportion in G0/G1 phase compared with the control-shRNA-lentivirus infected cells (Fig. 2C). As shown in Fig. 3B and C, although levels of cell apoptosis were higher in the ILK knockdown groups than that in the normal and control groups, no significant differences were observed, implying that downregulation of ILK alone is not an optimal approach for the treatment of lung cancer.

Synergistic effects between CDDP and ILK RNAi on proliferation and apoptosis in A549 cells. Using drug sensitivity analysis, it was found that 10% CDDP treatment produced the maximal decrease in cell viability (5% CDDP, 0.876 ± 0.015 vs 10% CDDP, 0.921 ± 0.009 , $P < 0.05$; no difference between 10, 15, 20 and 25% treatment was observed, 0.921 ± 0.009 vs 0.934 ± 0.005 , 0.934 ± 0.004 , 0.941 ± 0.003 ; $P > 0.05$). Therefore, proliferation and apoptosis in A549 cells undergoing ILK

RNAi infection and 10% CDDP treatment was evaluated. Cells infected with ILK specific-RNAi and addition of CDDP exhibited significantly increased apoptotic levels and inhibited cell viability compared with the control cells (Fig. 3A, B and C), indicating a synergistic effect between CDDP and ILK RNAi.

Effect of downstream gene expression of ILK on the regulation of cell survival and apoptosis. In order to investigate how cell survival and apoptosis were affected by ILK RNAi and CDDP, whether ILK RNAi regulated downstream gene expression was investigated. It was found that MMP-9, p-GSK3 β , p-AKT, AP-1, β -catenin and cyclin D1 protein levels were downregulated in the ILK specific-RNAi transfected cells. VEGF exhibited no change following ILK knockdown (Fig. 4).

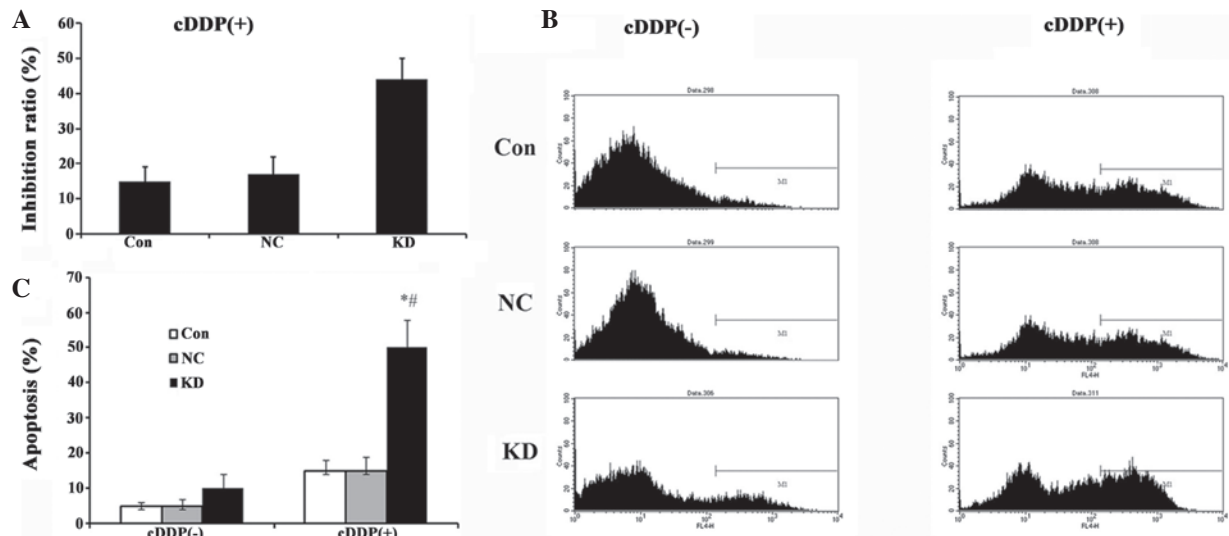


Figure 3. A549 cell proliferation and apoptosis in cells by transfection with lentivirus expressing KD RNAi, NC-RNAi or the parent A549 cells as CON combined with CDDP. (A) Cytotoxicity measured using a 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide assay. Proliferation in A549 cells was significantly inhibited by ILK RNAi and CDDP. (B) Apoptosis measured by flow cytometry. (C) Statistical comparisons. The results demonstrated that transfection with KD and CDDP treatment significantly enhanced cell apoptosis. Data are presented as the mean \pm standard deviation of a representative experiment performed in triplicate (n=3). ^{*}P<0.05, compared with the Con group; ^{##}P<0.05, compared with the NC group. NC, cells transfected with negative control RNAi; CON, control; KD, cells transfected with ILK specific RNAi; ILK, integrin-linked kinase; CDDP, cisplatin.

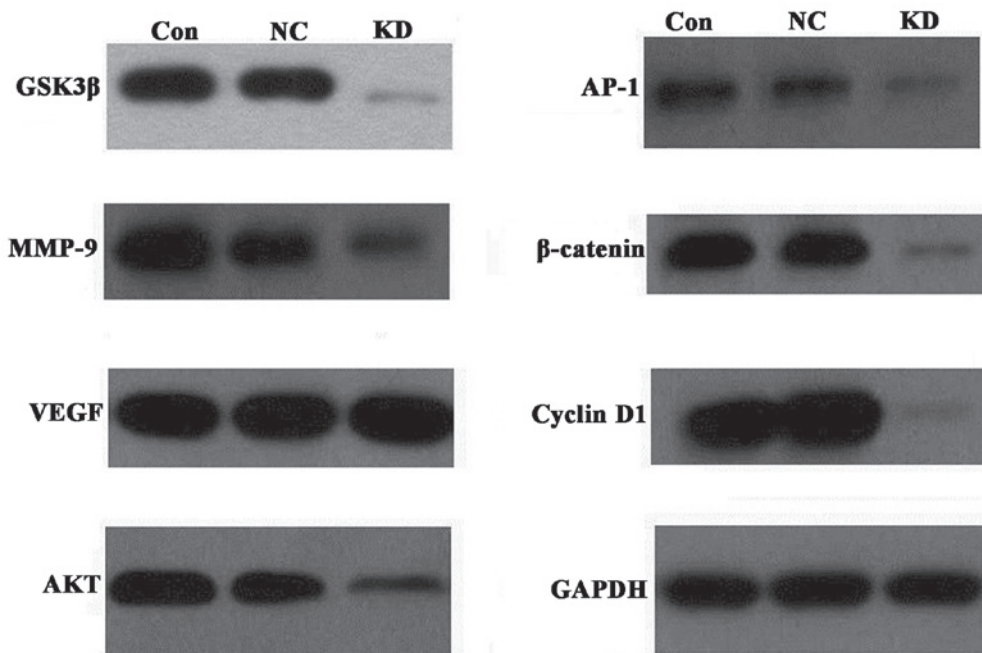


Figure 4. Effect of ILK knockdown on downstream protein expression by western blotting. The protein expression of GSK3 β , AKT, AP-1, β -catenin, cyclin D1 and MMP-9 were found to be downregulated in the ILK specific-RNAi transfected cells. VEGF demonstrated no change following ILK knockdown. GAPDH was used as an internal control. CON, parent cells; NC, cells transfected with negative control RNAi; KD, cells transfected with ILK specific RNAi; GSK, glycogen-synthase kinase; VEGF, vascular endothelial growth factor; ILK, integrin-linked kinase; AP, activator protein; MMP, matrix metalloproteinase.

Discussion

A significant number of studies have identified that ILK is a potential oncogene and inhibition of ILK by siRNA (15,16) or small molecules (18) may be a potentially useful therapeutic approach for treating cancer. However, whether the knockdown of ILK affects growth and apoptosis of lung cancer cells remains to be elucidated. In the present study, data revealed

that lentivirus-mediated ILK gene silencing may significantly inhibit A549 cell proliferation and alter cell cycle progression. However, treatment with ILK RNAi alone had significant effects on cell apoptosis. CDDP-based combination chemotherapy is currently one of the most active treatments for advanced lung cancer. The mechanism of cisplatin-based chemotherapy is generally accepted as its ability to form adducts with DNA and cause DNA strand breaks in the nucleus, which interferes with

normal transcription and/or DNA replication, leading to either repair of the DNA damage and cell survival or activation of the irreversible cell death program as a consequence (4,5). However, in the present study no significant differences in apoptosis in parent A549 cells and negative control-RNAi transfection cells with or without CDDP treatment were identified. This may be attributed to cisplatin insensitivity of A549 cell lines (19). In consideration of the mechanism of the above two approaches, lentivirus-mediated ILK siRNA and cisplatin-based chemotherapy were combined to investigate whether there is a synergistic interaction between them to promote cell apoptosis and inhibit cell growth. As expected, the present results demonstrated that apoptosis was significantly increased in cells infected with ILK specific-RNAi and addition of CDDP compared with other groups (Fig. 3). These findings suggest combined treatment modalities with lentivirus-mediated ILK interference and cisplatin chemotherapy may be more effective for cell apoptosis induction than mono-chemotherapy or knockdown. This contrasted with the conclusion drawn from the study by Kalra *et al* (20), who demonstrated that combination of CDDP and QLT0267, an ILK inhibitor, produced antagonistic interactions in a breast cancer model. This may result from the different pharmacological effects of these two compounds.

Furthermore, the present results also revealed that ILK siRNA may affect cell growth and apoptosis by regulating its downstream genes, including p-GSK3 β , p-AKT, AP-1, β -catenin, cyclin D1 and MMP-9. Indirectly, it was also demonstrated that these downstream genes may mediate cisplatin resistance in lung cancer cells. These conclusions appeared to be in accordance with previous studies: ILK kinase activity is rapidly stimulated by the engagement of integrins to the extracellular matrix components. These stimuli result in activation of protein kinase B/Akt, suppression of apoptosis and promotion of cell survival. Thus, targeting inhibition of ILK led to low expression of p-Akt and promoted cell apoptosis (21,22). Additionally, Akt activity is reported to be a determinant of CDDP resistance (23-25). Therefore, reduced expression of p-Akt may reduce this resistance, further inducing cell apoptosis. In addition to regulating the activity of PKB/Akt, ILK also inhibits the activity of GSK-3 by phosphorylation at Ser9 (26). Downregulation of ILK led to a decrease in p-GSK3 β and an increase in GSK-3 activity, which has been demonstrated to facilitate the cell apoptosis pathway (27-29). Further studies indicate that GSK-3 may be involved in cancer cell cycle arrest and apoptosis by regulating cyclin D1 expression, nuclear translocation of β -catenin and activation of the transcription factor AP-1 (26,30). Cyclin D1 is frequently overexpressed in lung cancer patients (31) and associated with poor survival of patients with lung cancer (32). Non-small cell lung cancer cells transfected with cyclin D1-targeted siRNA exhibited a marked decrease in cell growth rate and invasive capacity (33). AP-1 is a major transcription factor that regulates MMP-9 expression, which may contribute to the lower invasiveness and growth potential of cancer cells (34,35).

In conclusion, the results from the present study have identified for the first time, to the best of our knowledge, that downregulating ILK expression inhibits proliferation and cell cycle arrest in lung cancer cells. Downregulation of ILK

expression and CDDP treatment, in combination, is a more effective approach for the treatment of lung cancer through affecting downstream gene expression, including p-GSK3 β , p-AKT, AP-1, β -catenin, cyclin D1 and MMP-9.

Acknowledgements

This study was supported by the National Science Foundation (grant no. 81072739) and the Xinglin Scholar Program of Shanghai University of Traditional Chinese Medicine (grant no. 2209-13-03).

References

1. Siegel R, Naishadham D and Jemal A: Cancer statistics, 2012. *CA Cancer J Clin* 62: 10-29, 2012.
2. Jemal A, Siegel R, Ward E, *et al*: Cancer statistics, 2008. *CA Cancer J Clin* 58: 71-96, 2008.
3. Metro G, Chiari R, Mare M, *et al*: Carboplatin plus pemetrexed for platinum-pretreated, advanced non-small cell lung cancer: a retrospective study with pharmacogenetic evaluation. *Cancer Chemother Pharmacol* 68: 1405-1412, 2011.
4. Siddik ZH (ed): Mechanisms of action of cancer chemotherapeutic agents: DNA-interactive alkylating agents and antitumour platinum-based drugs. In: *The Cancer Handbook*. 1st Edition. Nature Publishing Group, London, pp1295-1313, 2002.
5. Rosell R, Lord RV, Taron M and Reguart N: DNA repair and cisplatin resistance in non-small-cell lung cancer. *Lung Cancer* 38: 217-227, 2002.
6. Bordin DL, Lima M, Lenz G, *et al*: DNA alkylation damage and autophagy induction. *Mutat Res* 753: 91-99, 2013.
7. Galluzzi L, Senovilla L, Vitale I, *et al*: Molecular mechanisms of cisplatin resistance. *Oncogene* 31: 1869-1883, 2011.
8. Cetintas VB, Kucukaslan AS, Kosova B, *et al*: Cisplatin resistance induced by decreased apoptotic activity in non-small-cell lung cancer cell lines. *Cell Biol Int* 36: 261-265, 2012.
9. Hannigan G, McDonald P, Walsh M and Dedhar S: Integrin-linked kinase: not so 'pseudo' after all. *Oncogene* 30: 4375-4385, 2011.
10. Takanami I: Increased expression of integrin-linked kinase is associated with shorter survival in non-small cell lung cancer. *BMC Cancer* 5: 1, 2005.
11. Watzka SB, Rauscher-Pötsch I, Stubenberger E, *et al*: Immunoreactivity of integrin-linked kinase in primary non-small-cell lung cancer and survival after curative resection. *Eur J Cardiothorac Surg* 38: 254-259, 2010.
12. Posch F, Setinek U, Flores RM, *et al*: Serum integrin-linked kinase (sILK) concentration and survival in non-small cell lung cancer: a pilot study. *Clin Transl Oncol* 16: 455-462, 2013.
13. Zhao M, Gao Y, Wang L, *et al*: Overexpression of integrin-linked kinase promotes lung cancer cell migration and invasion via NF- κ B-mediated upregulation of matrix metalloproteinase-9. *Int J Med Sci* 10: 995-1002, 2013.
14. Chen D, Zhang Y, Zhang X, *et al*: Overexpression of integrin-linked kinase correlates with malignant phenotype in non-small cell lung cancer and promotes lung cancer cell invasion and migration via regulating epithelial-mesenchymal transition (EMT)-related genes. *Acta Histochem* 115: 128-136, 2013.
15. Gao J, Zhu J, Li HY, Pan XY, Jiang R and Chen JX: Small interfering RNA targeting integrin-linked kinase inhibited the growth and induced apoptosis in human bladder cancer cells. *Int J Biochem Cell Biol* 43: 1294-1304, 2011.
16. Zhu XY, Liu N, Liu W, Song SW and Guo KJ: Silencing of the integrin-linked kinase gene suppresses the proliferation, migration and invasion of pancreatic cancer cells (Panc-1). *Genet Mol Biol* 35: 538-544, 2012.
17. Song W, Jiang R and Zhao CM: Role of integrin-linked kinase in multi-drug resistance of human gastric carcinoma SGC7901/DDP cells. *Asian Pac J Cancer Prev* 13: 5619-5625, 2012.
18. Younes MN, Yigitbasi OG, Yazici YD, *et al*: Effects of the integrin-linked kinase inhibitor QLT0267 on squamous cell carcinoma of the head and neck. *Arch Otolaryngol Head Neck Surg* 133: 15, 2007.

19. Dolfen D, Schottler K, Valiahdi SM, *et al*: Synthesis, structures and in vitro cytotoxicity of some platinum (II) complexes containing thiocarbamate esters. *J Inorg Biochem* 102: 2067-2071, 2008.
20. Kalra J, Warburton C, Fang K, *et al*: QLT0267, a small molecule inhibitor targeting integrin-linked kinase (ILK) and docetaxel can combine to produce synergistic interactions linked to enhanced cytotoxicity, reductions in P-AKT levels, altered F-actin architecture and improved treatment outcomes in an orthotopic breast cancer model. *Breast Cancer Res* 11: R25, 2009.
21. Persad S, Attwell S, Gray V, *et al*: Inhibition of integrin-linked kinase (ILK) suppresses activation of protein kinase B/Akt and induces cell cycle arrest and apoptosis of PTEN-mutant prostate cancer cells. *Proc Natl Acad Sci USA* 97: 3207-3212, 2000.
22. Troussard AA, McDonald PC, Wederell ED, Mawji NM, *et al*: Preferential dependence of breast cancer cells versus normal cells on integrin-linked kinase for protein kinase B/Akt activation and cell survival. *Cancer Res* 66: 393-403, 2006.
23. Lin Y, Wang Z, Liu L and Chen L: Akt is the downstream target of GRP78 in mediating cisplatin resistance in ER stress-tolerant human lung cancer cells. *Lung Cancer* 71: 291-297, 2011.
24. Yang X, Fraser M, Moll UM, Basak A and Tsang BK: Akt-mediated cisplatin resistance in ovarian cancer: modulation of p53 action on caspase-dependent mitochondrial death pathway. *Cancer Res* 66: 3126-3136, 2006.
25. Peng DJ, Wang J, Zhou JY and Wu GS: Role of the Akt/mTOR survival pathway in cisplatin resistance in ovarian cancer cells. *Biochem Biophys Res Commun* 394: 600-605, 2010.
26. Maydan M, McDonald PC, Sanghera J, *et al*: Integrin-linked kinase is a functional Mn²⁺-dependent protein kinase that regulates glycogen synthase kinase-3 β (GSK-3 β) phosphorylation. *PLoS One* 5: e12356, 2010.
27. Beurel E and Jope RS: The paradoxical pro- and anti-apoptotic actions of GSK3 in the intrinsic and extrinsic apoptosis signaling pathways. *Prog Neurobiol* 79: 173-189, 2006.
28. Watcharasit P, Thiantanawat A and Satayavivad J: GSK3 promotes arsenite-induced apoptosis via facilitation of mitochondria disruption. *J Appl Toxicol* 28: 466-474, 2008.
29. Ngok-Ngam P, Watcharasit P, Thiantanawat A and Satayavivad J: Pharmacological inhibition of GSK3 attenuates DNA damage-induced apoptosis via reduction of p53 mitochondrial translocation and Bax oligomerization in neuroblastoma SH-SY5Y cells. *Cell Mol Biol Lett* 18: 58-74, 2013.
30. Ban JO, Kwak DH, Oh JH, *et al*: Suppression of NF- κ B and GSK-3 β is involved in colon cancer cell growth inhibition by the PPAR agonist troglitazone. *Chem Biol Interact* 188: 75-85, 2010.
31. Kosacka M, Piesiak P, Kowal A, Gołdecki M and Jankowska R: Galectin-3 and cyclin D1 expression in non-small cell lung cancer. *J Exp Clin Cancer Res* 30: 101-107, 2011.
32. Li R, An SJ, Chen ZH, *et al*: Expression of cyclin D1 splice variants is differentially associated with outcome in non-small cell lung cancer patients. *Hum Pathol* 39: 1792-1801, 2008.
33. Huang H, Hu YD, Li N and Zhu Y: Inhibition of tumor growth and metastasis by non-small cell lung cancer cells transfected with cyclin D1-targeted siRNA. *Oligonucleotides* 19: 151-162, 2009.
34. Troussard AA, Costello P, Yoganathan TN, Kumagai S, Roskelley CD and Dedhar S: The integrin linked kinase (ILK) induces an invasive phenotype via AP-1 transcription factor-dependent upregulation of matrix metalloproteinase 9 (MMP-9). *Oncogene* 19: 5444-5452, 2000.
35. Lakka SS, Gondi CS, Yanamandra N, *et al*: Inhibition of cathepsin B and MMP-9 gene expression in glioblastoma cell line via RNA interference reduces tumor cell invasion, tumor growth and angiogenesis. *Oncogene* 23: 4681-4689, 2004.