

# *Zingiber mioga* (Thunb.) Roscoe attenuates allergic asthma induced by ovalbumin challenge

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**Abstract.** *Zingiber mioga* (Thunb.) Roscoe (ZM) is a traditional medicine, used to treat inflammatory diseases. The present study aimed to evaluate the inhibitory effects of ZM on the inflammatory response in lipopolysaccharide (LPS)-stimulated RAW264.7 murine macrophage cells and in a mouse model of ovalbumin (OVA)-induced allergic asthma. Mice received OVA sensitization on day 0 and 14, and were challenged with OVA between days 21 and 23. ZM was administered to the mice at a dose of 30 mg/kg, 1 h prior to OVA challenge. In LPS-stimulated RAW264.7 cells, ZM significantly decreased nitric oxide (NO) and tumor necrosis factor (TNF)- $\alpha$  production in a concentration-dependent manner, and mRNA expression of inducible NO synthase (iNOS), TNF- $\alpha$  and matrix metalloproteinase (MMP)-9 was reduced. In addition, treatment with ZM decreased the inflammatory cell count in bronchoalveolar lavage fluid from the mice, and reduced the expression of interleukin (IL)-4, IL-5, IL-13, eotaxin and immunoglobulin E. ZM also reduced airway hyperresponsiveness in OVA-challenged mice, and attenuated the infiltration of inflammatory cells and mucus production in the airways, with a decrease in the expression of iNOS and MMP-9 in lung tissue. In conclusion, the results of the present study indicate that ZM effectively inhibits inflammatory

responses. Therefore, it may be that ZM has potential as a therapeutic agent for use in inflammatory diseases.

## Introduction

Allergic asthma is a chronic respiratory disease, the prevalence of which is increasing rapidly worldwide (1). The characteristic pathophysiological features of asthma include chronic eosinophilic inflammation of the airways, airway hyperresponsiveness (AHR) and eventual progression to airway remodeling (1). Nitric oxide (NO) is derived from inducible NO synthase (iNOS), and has a crucial role in airway inflammation (2). The development of allergic asthma is associated with overexpression of iNOS in the airways, which results in an asthmatic response. Previous studies have demonstrated that iNOS expression is associated with the induced activation of T-helper (Th)2 lymphocytes, which results in the pathophysiological alterations that occur in asthma (2,3). Th2 lymphocytes produce various Th2 cytokines that have important roles in the development of allergic asthma (3). Th2 cytokines include interleukin (IL)-4, IL-5 and IL-13, and induce secretion of allergen-specific immunoglobulin (Ig) E, chemokines and eosinophils, as well as mucus production. Chemokines, such as eotaxin, are essential for the delivery of eosinophils to the airways (4). Matrix metalloproteinase 9 (MMP-9) is also associated with the development of allergic asthma. MMP-9 belongs to a family of extracellular proteases that are responsible for the degradation of the extracellular matrix during tissue remodeling (5). MMP-9 in particular appears to have an important role in chronic airway inflammation and remodeling in asthma, and was shown to be upregulated in a murine asthma model in association with an accumulation of inflammatory cells (6). The suppression of NO production and MMP-9 expression is an important therapeutic target for asthma.

*Zingiber mioga* (Thunb.) Roscoe (ZM) is a medicinal herb belonging to the Mioga family. ZM is globally used to treat inflammatory diseases, including stomatitis (7). In addition to their extensive use as spices, numerous types of ginger have been used in traditional Oriental medicine in order to

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ameliorate symptoms, such as inflammation, rheumatic disorders and gastrointestinal discomforts (7). Previous studies have demonstrated that ZM exhibits anti-inflammatory, anticancer and antioxidant effects *in vitro* (7,8). Kim *et al* (8) previously showed that ZM was able to inhibit the production of inflammatory mediators in lipopolysaccharide (LPS)-stimulated macrophage cells. However, to the best of our knowledge, no study has investigated the protective effects of ZM on airway inflammation in an OVA-induced allergic asthma model.

The present study aimed to investigate the inhibitory potential of ZM on airway inflammation in asthma, using a murine model of OVA-induced asthma and LPS-stimulated RAW264.7 cells. In order to investigate the possible mechanism underlying the effects of ZM, the abundance of pro-inflammatory proteins in lung tissue was examined, as well as the expression of mRNA in RAW264.7 cells.

## Materials and methods

**Cell culture and cell viability.** The RAW264.7 murine macrophage cells (American Type Tissue Collection, Manassas, VA, USA) were maintained in Dulbecco's modified Eagle's medium (DMEM; Gibco Life Technologies, Grand Island, NY, USA), supplemented with 10% heat-inactivated fetal bovine serum (FBS; Gibco Life Technologies) and antibiotics (100 U/ml penicillin, 100 µg/ml streptomycin, Gibco Life Technologies) at 37°C in an incubator containing 5% CO<sub>2</sub> and 95% air. The cell viability of cells treated with ZM was measured using a 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT; Amresco, LLC, Solon, OH, USA) assay. The RAW264.7 cells were cultured in 96-well plates at a density of 1×10<sup>4</sup> cells/well. The ZM extract was obtained from The Plant Extract Bank at the Korea Research Institute of Bioscience and Biotechnology (PB2176.2; Chungbuk, Korea). The ZM extract was added to each individual well at a concentration of 20, 40, 80 and 100 µg/ml, and then incubated for 24 h. The control cells were treated with an equivalent volume of phosphate-buffered saline (PBS; pH 7.4). MTT solution (10 µl) was added to each well, and the plates were incubated for 4 h at 37°C. After 4 h, 100 µl dimethyl sulfoxide (Samchun Pure Chemical Co., Ltd., Daejeon, Korea) was added to each well to solubilize the formazan crystals. The optical density was measured at 570 nm using a microplate reader (VersaMax; Molecular Devices, Sunnyvale, CA, USA).

**Determination of NO production.** The cells (5×10<sup>4</sup> cells/well) were seeded in 96-well plates in FBS-free DMEM, and were treated with various concentrations of ZM (10, 20 and 40 µg/ml) for 1 h. Subsequently, the cells were incubated in the presence of LPS (0.5 µg/ml) for 24 h. The nitrite accumulation in the culture medium was measured using Griess reagent (Promega Corporation, Madison, WI, USA). The absorbance was then measured at 540 nm using a microplate reader (iMark Microplate absorbance reader; Bio-Rad Laboratories, Inc. Hercules, CA, USA).

**Determination of pro-inflammatory cytokines.** Cells were treated as described above, and the concentration of tumor necrosis factor-α (TNF-α) was quantified in the culture

medium, using a competitive enzyme-linked immunosorbent assay (ELISA) kit (BD Biosciences, San Jose, CA, USA), according to the manufacturer's instructions. The absorbance was measured at 450 nm in a microplate reader (Bio-Rad Laboratories, Inc.). The absolute concentrations were obtained by running standard curves on the identical ELISA plates.

**Reverse transcriptase-polymerase chain reaction (RT-PCR).** Total RNA was extracted from the cells using TRIzol® reagent (Life Technologies, Carlsbad, CA, USA). For RT-PCR, single-strand cDNA (Qiagen, Hilden, Germany) was synthesized from 2 µg total RNA using a commercial PCR kit (166-2600EDU; Bio-Rad Laboratories Inc.) and a T100 thermal cycler (186-1096; Bio-Rad Laboratories Inc.). The primer sequences used in the PCR were as follows: Forward: 5'-CAAGAGTTTGACCAGAGGACC-3', and reverse: 5'-TGGAACCACTCGTACTTGGGA-3' for iNOS, forward: 5'-AAGCACATGCAGAATGAGTACCG-3' and reverse: 5'-GTGGGACAGCTTCTGGTCGAT-3' for MMP-9 and forward: 5'-CGCTCATGCGGATAGTGAT-3' and reverse: 5'-TGTTTGAGACCTTCAACACC-3' for β-actin (Bioneer, Daejeon, Korea). The following cycle conditions were set for each PCR reaction: iNOS, forward, 95°C for 5 min, then 94°C for 30 sec, 58°C for 30 sec and 72°C for 45 sec for 32 cycles, and a final extension step at 72°C for 5 min; MMP-9 and β-actin, forward, 94°C for 5min, then 94°C for 30 sec, 50°C for 30 sec and 72°C for 45 sec for 30 cycles, and a final extension step at 72°C for 10 min. The PCR products were then subjected to 1.5% agarose gel electrophoresis and were stained with 5 µg/ml ethidium bromide (Bio-Rad Laboratories Inc.). Images were captured using an Olympus C4000 zoom camera system (Olympus America Inc., Center Valley, PA, USA).

**Mice.** Specific pathogen-free female BALB/c mice (19-21 g, 6-8 weeks-old) were obtained from Koatech Co. (Pyeongtaek, Korea). The 28 mice were housed under standard conditions (temperature, 22±2°C; humidity, 55±5%; 12 h-light/dark cycle) with food and water. All of the experimental procedures were approved by the Institutional Animal Care and Use Committee of the Korea Research Institute of Bioscience and Biotechnology.

**Induction of asthma in BALB/c mice and drug administration.** The mice were divided into four groups (n=7): Normal control, PBS sensitization/challenge; OVA, OVA sensitization/challenge + PBS; Mon, OVA sensitization/challenge + montelukast (30 mg/kg; Sigma-Aldrich, St. Louis, MO, USA); ZM, OVA sensitization/challenge + ZM (30 mg/kg). Airway inflammation was induced by an OVA (Sigma-Aldrich) challenge. Each mouse was immunized by an intraperitoneal injection of 20 µg OVA, emulsified with 2 mg aluminum hydroxide (Thermo Fisher Scientific, Waltham, MA, USA) in 200 µl PBS (pH 7.4), on days 0 and 14. On days 21, 22 and 23 the mice were exposed, by inhalation, to 1% (w/v) OVA aerosolized solution using an ultrasonic nebulizer (NE-U12; Omron Corp., Tokyo, Japan) for 1 h. The ZM groups of asthma-induced mice were treated orally with 30 mg/kg ZM, or with an oral injection of 30 mg/kg montelukast 1 h prior to the OVA challenge. Montelukast is a cysteinyl leukotriene (cys-LT)-1

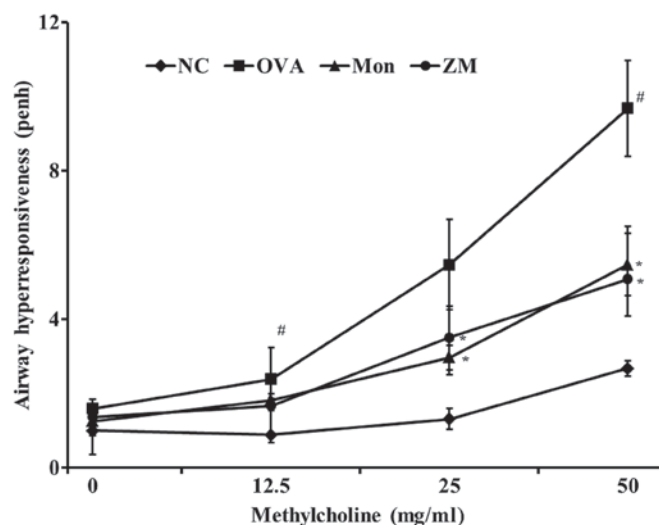


Figure 1. ZM reduced airway hyperresponsiveness in OVA-challenged mice. Airway hyperresponsiveness was examined 24 h following the final challenge of mice with various concentrations of methylcholine (12.5, 25 or 50 mg/ml), using single-chamber whole body plethysmography. Data are expressed as the mean  $\pm$  standard deviation (n=7/group). <sup>#</sup>P<0.05, compared with NC and <sup>\*</sup>P<0.05, compared with OVA. NC, normal control mice; OVA, ovalbumin-challenged mice; Mon, montelukast (30 mg/kg) + OVA-challenged mice; ZM, *Zingiber mioga* (Thunb.) Roscoe (30 mg/kg) + OVA-challenged mice. NC, normal control.

receptor agonist, and was used as a positive control in the present study. The control mice were treated orally with an equivalent volume of PBS.

**Measurement of AHR.** A total of 24 hours after the final aerosol challenge, AHR was measured indirectly using single-chamber whole body plethysmography (Allmedicus, Seoul, Korea) following exposure to aerosolized PBS, followed by aerosols containing increasing concentrations of methacholine (12.5, 25 and 50 mg/ml; Sigma-Aldrich) in PBS, for 3 min at each exposure level.

**Collection of bronchoalveolar lavage fluid (BALF) and serum.** For collection of BALF, the mice were sacrificed 24 h after AHR measurement with an intraperitoneal injection of phenobarbital (50 mg/kg; Hanlim Pharm., Co., Ltd., Seoul, Korea), and tracheotomy was performed as described previously (9). The lung was lavaged three times, and each time, fluid was withdrawn via the trachea with 0.5 ml PBS (total volume, 1.5 ml). The number of inflammatory cells was determined by counting the cells in  $\geq 5$  squares of a hemocytometer (Marienfeld, Lauda-Königshofen, Germany), following exclusion of the dead cells by Trypan blue (Sigma-Aldrich) staining. The differential cell count of the BALF was performed using Diff-Quik<sup>®</sup> staining reagent (B4132-1A; IMEB Inc., San Marcos, CA, USA), according to the manufacturer's instructions. The blood was collected from the inferior vena cava and centrifuged at 200  $\times$  g for 10 min at 4°C. The supernatant was then stored at -70°C until further use.

**Measurement of immunoglobulin (Ig)E in serum and cytokines in BALF.** The Th2 cytokines (IL-4, IL-5, IL-13) and eotaxin in BALF were measured using Mouse IL-4, IL-5 and IL-13 Quantikine ELISA kits (R&D Systems, Minneapolis, MN, USA), according to the manufacturer's instructions. The total IgE and OVA-specific IgE serum levels were measured by ELISA. Briefly, microtiter plates were coated with

anti-IgE antibodies (anti-mouse IgE; 10 g/ml; AbD Serotec, Oxford, UK) in PBS-Tween 20, and incubated with BALF or plasma samples. The plates were then washed four times with washing solution (contained in ELISA kits), and 200  $\mu$ l o-phenylenediamine dihydrochloride (Sigma-Aldrich) was added to each well. The plates were incubated for 10 min in the dark and the absorbance was measured at 450 nm using the iMark microplate absorbance reader.

**Immunoblotting.** The mouse lung tissue was homogenized (1/10 w/v) in Tissue Lysis/Extraction reagent (Sigma-Aldrich) that contained a protease inhibitor cocktail (Sigma-Aldrich) using a homogenizer (IKA, Guangzhou, China). In order to extract the nuclear protein, the lung tissue was homogenized using an extraction kit (Thermo Fisher Scientific). The protein concentrations of the samples were determined using Bradford reagent (Bio-Rad Laboratories, Inc.). Equal quantities of total protein (30  $\mu$ g) were separated by 10% SDS-polyacrylamide gel electrophoresis and transferred onto a polyvinylidene fluoride membrane (IPVH00010; Merck Millipore, Darmstadt, Germany). The membranes were then incubated with blocking solution (5% non-fat milk), followed by an overnight incubation at 4°C with the appropriate primary antibodies. The following primary antibodies and dilutions were used: Rabbit monoclonal anti- $\beta$ -actin (1:2,000 dilution; cat. no. P60709; Cell Signaling Technologies, Inc., Danvers, MA, USA); rabbit polyclonal anti-iNOS (1:1,000 dilution; cat. no. ADI-KAS-NO001-F; Enzo Life Sciences Inc., Farmingdale, NY, USA) and rabbit polyclonal anti-MMP-9 (1:1,000 dilution; cat. no. ab38898; Cell Signaling Technologies, Inc.). The blots were washed three times with tris-buffered saline containing Tween-20 (TBST; Biosesang, Seongnam, Korea) and then incubated with a 1:10,000 dilution of horseradish peroxidase (HRP)-conjugated secondary antibody (cat. no. 211-542-171; Jackson ImmunoResearch Laboratories, Inc., West Grove, PA, USA) for 30 min at room temperature. The blots were washed a further three

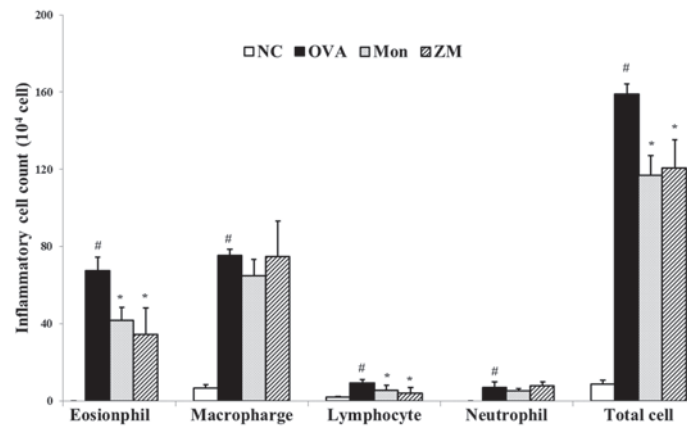


Figure 2. ZM inhibited the number of inflammatory cells in the bronchoalveolar lavage fluid of the mice. The cells were isolated by centrifugation and stained with Diff-Quik stain reagent. Data are expressed as the mean  $\pm$  standard deviation ( $n=7$ /group). <sup>#</sup> $P<0.05$ , compared with NC and <sup>\*</sup> $P<0.05$ , compared with OVA. NC, normal control mice; OVA, ovalbumin-challenged mice; Mon, montelukast (30 mg/kg) + OVA-challenged mice; ZM, *Zingiber mioga* (Thunb.) Roscoe (30 mg/kg) + OVA-challenged mice.

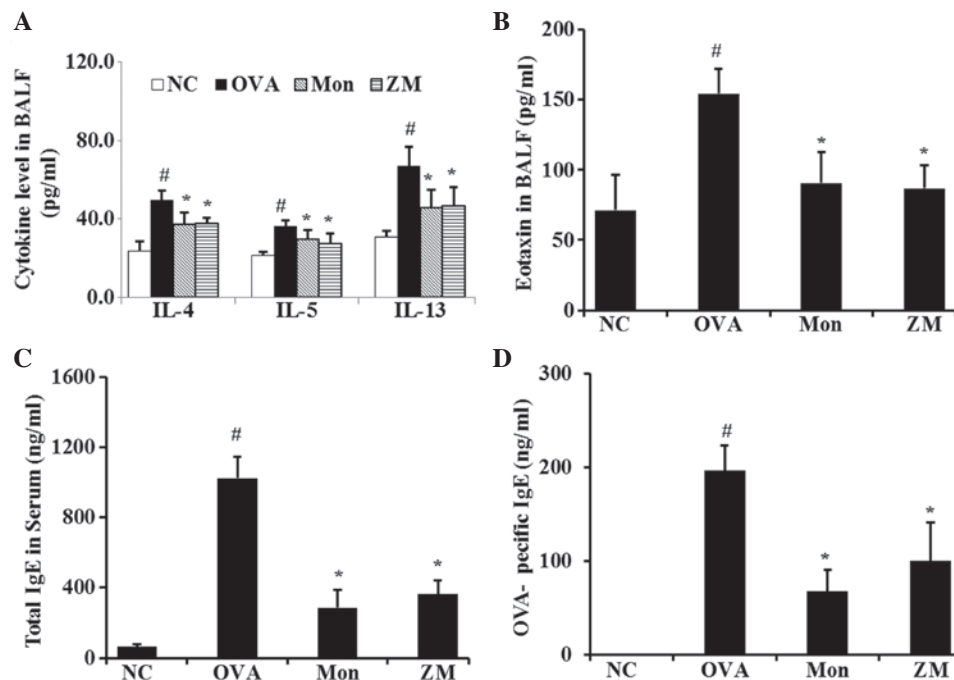


Figure 3. ZM inhibited the levels of pro-inflammatory cytokines, eotaxin and IgE. Levels of (A) pro-inflammatory cytokines in BALF, (B) chemokine in BALF, (C) total IgE in serum and (D) OVA-specific IgE in serum, as determined by ELISA. Data are expressed as the mean  $\pm$  standard deviation ( $n=7$ /group). <sup>#</sup> $P<0.05$ , compared with NC and <sup>\*</sup> $P<0.05$ , compared with OVA. NC, normal control mice; OVA, ovalbumin-challenged mice; Mon, montelukast (30 mg/kg) + OVA-challenged mice; ZM, *Zingiber mioga* (Thunb.) Roscoe (30 mg/kg) + OVA-challenged mice; BALF, bronchoalveolar lavage fluid; IL, interleukin; IgE, immunoglobulin E; NC, normal control.

times with TBST, and then developed using an Enhanced Chemiluminescence kit (Thermo Fisher Scientific).

**Lung tissue histopathology.** The lung tissue was fixed in 4% (v/v) paraformaldehyde, embedded in paraffin, and cut into 4- $\mu$ m sections, prior to staining with hematoxylin and eosin solution (Sigma-Aldrich) and periodic acid-Schiff (IMEB Inc.), in order to detect the inflammation and mucus production of the lung sections, respectively. Tissue sections were mounted and coverslipped. Pathological changes to the lung tissue were examined using a microscope (Leica DM1000; Leica Microsystems, Wetzlar, Germany).

**Statistical analysis.** Data are expressed as the mean  $\pm$  standard deviation. The statistical significance of the results were determined by analysis of variance, followed by a multiple comparison test with Dunnett's adjustment. Statistical analyses were performed using GraphPad InStat v 3.0 (GraphPad, La Jolla, CA, USA).  $P<0.05$  was considered to indicate a statistically significant difference.

## Results

**ZM reduces OVA-induced AHR.** The OVA-induced mice exhibited a significant increase in AHR with increasing



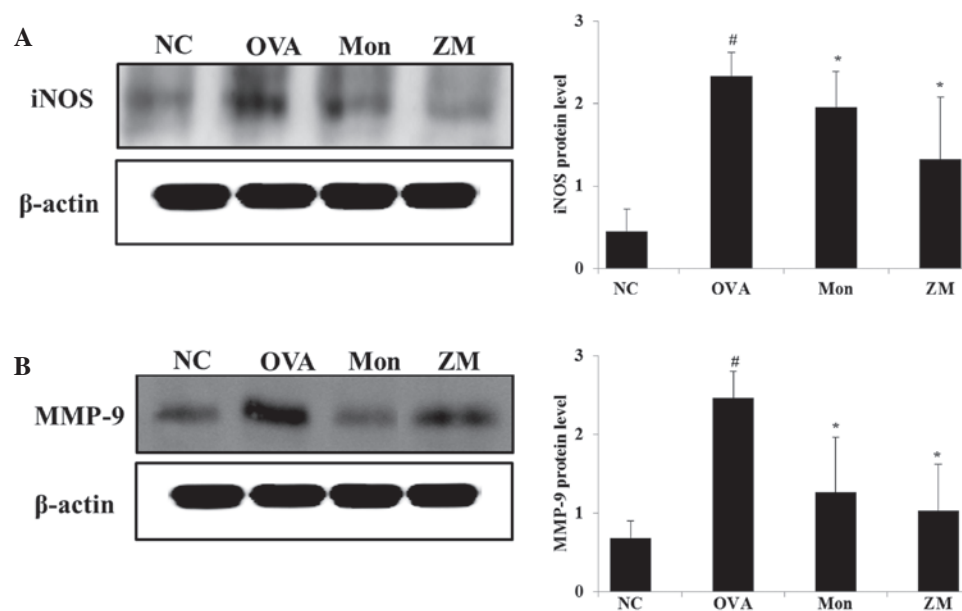


Figure 4. ZM reduced the protein expression levels of iNOS and MMP-9 in the lung tissue. The expression levels of iNOS and MMP-9 were detected by western blot analysis. Detection of  $\beta$ -actin was conducted in order to confirm the equal loading of proteins. (A) Protein expression levels of iNOS. (B) Protein expression levels of MMP-9. Data are expressed as the mean  $\pm$  standard deviation ( $n=7$ /group). <sup>\*</sup> $P<0.05$ , compared with NC and <sup>#</sup> $P<0.05$ , compared with OVA. NC, normal control mice; OVA, ovalbumin-challenged mice; Mon, montelukast (30 mg/kg) + OVA-challenged mice; ZM, *Zingiber mioga* (Thunb.) Roscoe (30 mg/kg) + OVA-challenged mice; iNOS, inducible nitric oxide synthase; MMP-9, matrix metalloproteinase 9.

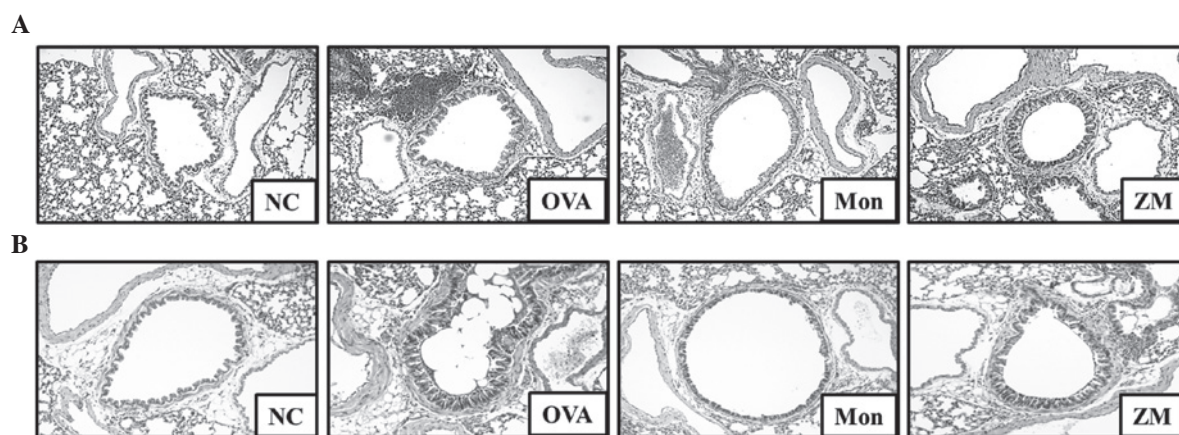


Figure 5. ZM inhibited airway inflammation and mucus production. (A) Histological examination of airway inflammation in lung tissue by hematoxylin and eosin staining. (B) Histological examination of mucus production in the lung tissue by periodic acid-Schiff staining. Magnification,  $\times 200$ . NC, normal control mice; OVA, ovalbumin-challenged mice; Mon, montelukast (30 mg/kg) + OVA-challenged mice; ZM, *Zingiber mioga* (Thunb.) Roscoe (30 mg/kg) + OVA-challenged mice.

concentrations of methylcholine exposure, compared with the normal control mice. The montelukast-treated mice exhibited significantly reduced AHR, compared with the OVA-induced mice. The ZM-treated mice also exhibited decreased AHR at methylcholine concentrations of 25 and 50 mg/ml, compared with the OVA-challenged mice. These reductions were similar to the results observed in the montelukast-treated mice (Fig. 1).

**ZM decreases the number of inflammatory cells in BALF.** Significantly increased numbers of inflammatory cells, particularly eosinophils, were observed in the BALF samples from OVA-induced mice compared with samples from the normal controls. BALF samples from the ZM-treated mice exhibited a markedly decreased number of inflammatory

cells, including eosinophils, macrophages, lymphocytes and neutrophils, compared with samples from the OVA-challenged mice (Fig. 2).

**ZM reduces the levels of proinflammatory cytokines in BALF, and levels of IgE in serum.** The levels of IL-4, IL-5, IL-13 and eotaxin were significantly increased in the BALF from OVA-challenged mice, compared with that from the normal controls. However, montelukast-treated mice had significantly reduced levels of these cytokines, as compared with the OVA-challenged mice. The levels of these cytokines were also significantly decreased in the BALF of the ZM-treated mice, compared with that of the OVA-challenged mice (Fig. 3A and B). The total IgE and OVA-specific IgE

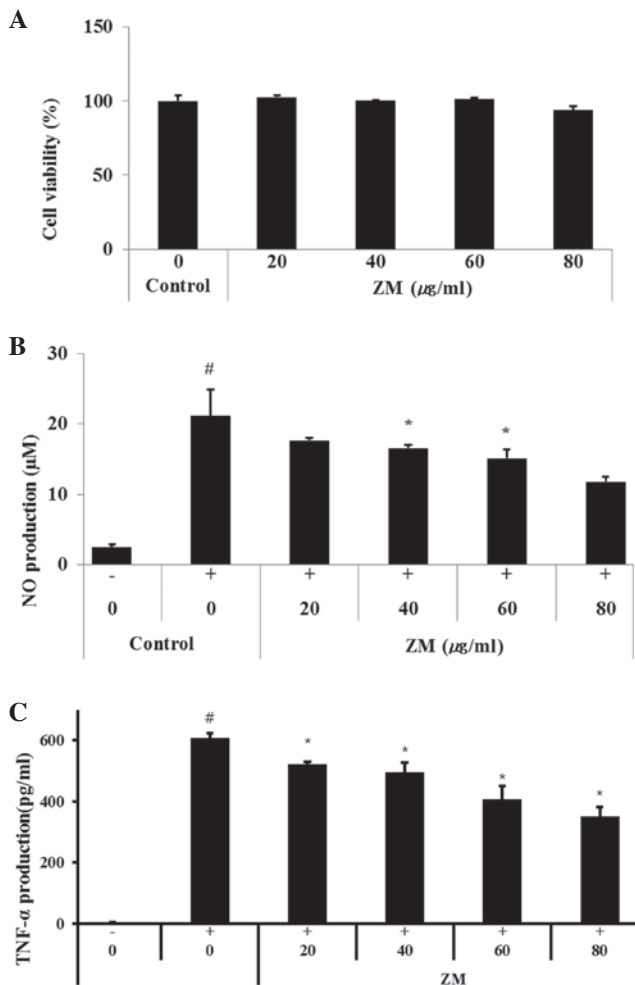


Figure 6. RAW264.7 murine macrophage cells were treated with 20, 40, 60 or 80 µg/ml of ZM for 24 h. (A) Cell survival rates were determined using an MTT assay. ZM inhibited the production of (B) NO and (C) TNF-α in the LPS stimulated RAW264.7 cells. Cells were treated with ZM (20, 40, 60 or 80 µg/ml) for 1 h and with LPS (0.5 µg/ml) for 24 h. Data are expressed as the mean ± standard deviation. <sup>#</sup>P<0.05, compared with control cells and <sup>\*</sup>P<0.05, compared with LPS-treated cells. ZM, *Zingiber mioga* (Thunb.) Roscoe; LPS, lipopolysaccharide; NO, nitric oxide; TNF-α, tumor necrosis factor-α.

serum levels in the OVA-challenged mice were significantly elevated, compared with levels in the normal controls. Conversely, montelukast-treated mice had markedly decreased levels of total and OVA-specific IgE, as compared with the OVA-challenged mice. The ZM-treated mice also exhibited a significant reduction in the total IgE and OVA-specific IgE serum levels, compared with those in the OVA-challenged mice (Fig. 3C and D).

*ZM inhibits protein expression levels of iNOS and MMP-9 in the lung tissue of OVA-challenged mice.* OVA-challenged mice exhibited significantly increased protein expression levels of iNOS in the lung tissue, compared with the normal controls. However, montelukast-treated mice exhibited significantly reduced iNOS expression in lung tissue, as compared with the OVA-challenged mice. In addition, in the ZM-treated mice, a marked reduction in the expression levels of iNOS were detected, as compared with the OVA-challenged mice (Fig. 4A). In the OVA-challenged mice, MMP-9 expression was increased, compared with the normal controls,

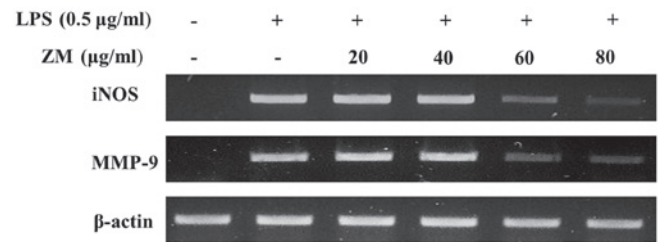


Figure 7. Treatment with ZM reduced the mRNA expression levels of iNOS and MMP-9 in the LPS-stimulated RAW264.7 murine macrophage cells. Detection of β-actin was conducted in order to confirm the equal loading of mRNA. ZM, *Zingiber mioga* (Thunb.) Roscoe; LPS, lipopolysaccharide; iNOS, inducible nitric oxide synthase; MMP-9, matrix metalloproteinase 9.

whereas they were significantly decreased in the montelukast- and ZM-treated mice, compared with the OVA-challenged mice (Fig. 4B).

*ZM decreases inflammatory cell infiltration and mucus production.* Lung tissue from the OVA-challenged mice exhibited infiltration of inflammatory cells into the peribronchial and perivascular lesions, and mucus hypersecretion in airway epithelial cells (Fig. 5A and B). However, lung tissue from montelukast-treated mice exhibited a reduction in inflammatory cell infiltration and mucus secretion. In addition, lung tissue from ZM-treated mice exhibited significantly lower inflammatory cell infiltration and mucus secretion, compared with the OVA-challenged mice.

*ZM reduces NO production and TNF-α in LPS-stimulated cells.* In the present study, nontoxic concentrations of ZM (20, 40, 60 and 80 µg/ml; Fig. 6A) were tested for their ability to inhibit LPS-stimulated NO production. The LPS-stimulated cells exhibited increased cellular NO levels, compared with non-stimulated cells. By contrast, the production of NO was inhibited in the cells pre-treated with ZM, in a concentration-dependent manner (Fig. 6B).

As shown in Fig. 6C, the LPS-stimulated cells exhibited markedly increased levels of TNF-α, compared with the normal controls, whereas the ZM-treated cells exhibited a significant concentration-dependent decrease in the levels of TNF-α, compared with the LPS-stimulated cells.

*ZM inhibits iNOS and MMP-9 mRNA expression in LPS-stimulated cells.* The LPS-stimulated cells exhibited a significant increase in mRNA expression levels of iNOS, TNF-α and MMP-9, compared with the control group. The LPS-induced mRNA expression levels of iNOS and MMP-9 were significantly reduced in the ZM-treated cells (Fig. 7).

## Discussion

The present study aimed to evaluate the anti-inflammatory effects of ZM in an OVA-induced allergic asthma murine model, and in LPS-stimulated RAW264.7 cells. ZM decreased the number of inflammatory cells, the Th2 cytokine levels and the levels of eotaxin in BALF from mice treated with OVA. In addition, mice treated with ZM produced significantly reduced serum levels of total and OVA-specific IgE, compared with the

OVA-challenged mice. ZM also decreased the protein expression levels of iNOS and MMP-9 in the lung tissue, as well as reducing AHR, airway inflammation and mucus production. Furthermore, ZM inhibited the production of NO and TNF- $\alpha$  in the LPS-stimulated RAW264.7 cells, and reduced the mRNA expression levels of iNOS, TNF- $\alpha$  and MMP-9.

Allergic asthma is a chronic inflammatory airway disease that is characterized by persistent AHR, Th2 lymphocyte infiltration and plasma exudation. Allergic asthma is associated with an elevated serum IgE level, eosinophilia and mucus production in the airways (10,11). NO is a particularly important inflammatory mediator of allergic asthma. NO is a gas molecule synthesized by NOS, which is associated with inflammatory responses. It has previously been suggested that iNOS is induced by inflammatory cytokines, such as TNF- $\alpha$ , and results in the production of much larger amounts of NO (12). The overproduction of NO underlies pathophysiology of airway inflammation. NO contributes to the infiltration of inflammatory cells, such as neutrophils, monocytes and eosinophils, which produce IL-4, IL-5 and IL-13 Th2 cytokines (6,13). IL-4 promotes B-cell isotype switching and the upregulation of IgE, whereas IL-5 induces the infiltration of eosinophils into the airways (14). IL-13 is also a potent stimulator of airway remodeling, since it induces lung and alveolar enlargement, as well as mucus metaplasia and AHR (15,16). In the present study, Th2 cytokines, including IL-4, IL-5, and IL-13, were markedly decreased in the ZM-treated mice. The ZM-treated mice also exhibited a significant reduction in the number of inflammatory cells, which was accompanied by decreased levels of total and OVA-specific IgE, and eotaxin. Treatment with ZM reduced airway inflammation and mucus production alongside inhibition of iNOS expression in the lung tissue. These findings indicated that ZM may effectively inhibit the development of allergic asthma induced by an OVA challenge. It was hypothesized that this may be associated with the suppression of iNOS expression. In addition, ZM reduced NO and TNF- $\alpha$  production, and iNOS levels in the LPS-stimulated RAW264.7 cells in a concentration-dependent manner.

MMPs are a family of extracellular proteases that are responsible for the degradation of the extracellular matrix during tissue remodeling. Within the MMP family, the MMP-9 gelatinases are specific to the denatured collagens and collagen-IV of the basement membrane (17). MMP-9 is controlled by the inflammatory mediator NO, as well as cytokines and chemokines, and is produced by various types of cells, including inflammatory, endothelial and epithelial cells (18). MMP-9 is associated with airway remodeling and inflammatory cell infiltration into the airways. Previous studies have demonstrated that increased expression levels of MMP-9 in patients with asthma are associated with an increase in the number of inflammatory cells (19,20). Inhibition of MMP-9 has been shown to significantly reduce the production of Th2 cytokines and IgE, in addition to the expression levels of adhesion molecules, which results in the alleviation of asthmatic responses, such as airway inflammation, mucus overproduction, and AHR (21). Numerous studies have explored novel therapeutic materials focusing on the suppression of MMP-9 expression in asthma (6,14,17). The present study evaluated the protein expression levels of

MMP-9 in lung tissue. ZM-treated mice exhibited reduced MMP-9 expression in lung tissue, compared with the OVA-challenged mice. These findings indicated that ZM may reduce asthmatic responses by downregulating the expression of MMP-9 in lung tissue.

In conclusion, ZM effectively suppressed inflammatory responses in the OVA-challenged mice and LPS-stimulated RAW264.7 cells. Based on these results, the anti-inflammatory effects of ZM were hypothesized to be associated with the suppression of iNOS and MMP-9 expression. The findings suggested that ZM may be a potent therapeutic agent in inflammatory diseases, such as allergic asthma.

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