

Differential changes in Neuregulin-1 signaling in major brain regions in a lipopolysaccharide-induced neuroinflammation mouse model

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Abstract. Neuregulin 1 (Nrg1) is involved in multiple biological processes in the nervous system. The present study investigated changes in Nrg1 signaling in the major brain regions of mice subjected to lipopolysaccharide (LPS)-induced neuroinflammation. At 24 h post-intraperitoneal injection of LPS, mouse brain tissues, including tissues from the cortex, striatum, hippocampus and hypothalamus, were collected. Reverse transcription-polymerase chain reaction was used to determine the expression of Nrg1 and its receptors, Neu and ErbB4, at the mRNA level. Western blotting was performed to determine the levels of these proteins and the protein levels of phosphorylated extracellular signal-regulated kinases (Erk)1/2 and Akt1. Immunohistochemical staining was utilized to detect the levels of pNeu and pErbB4 in these regions. LPS successfully induced sites of neuroinflammation in these regions, in which changes in Nrg1, Neu and ErbB4 at the mRNA and protein levels were identified compared with controls. LPS induced a reduction in pNeu and pErbB4 in the striatum and hypothalamus, although marginally increased pErbB4 levels were found in the hippocampus. LPS increased the overall phosphorylation of Src but this effect was reduced in the hypothalamus. Moreover, increased phosphorylation of Akt1 was found in the striatum and hippocampus. These data suggest diverse roles for Nrg1 signaling in these regions during the process of neuroinflammation.

Introduction

Neuregulin 1 (Nrg1) is a member of the epidermal growth factor family and functions through the activation of the tyrosine kinase domain of ErbB receptors (1). The bioactivity of Nrg1 is primarily mediated via homodimers consisting of its cognate receptor ErbB4 (2) or via heterodimeric complexes of ErbB3/Neu and ErbB4/Neu (3). These receptors, including Neu and ErbB4, have a high affinity for Nrg1 and contain an active tyrosine kinase domain (4). The interaction between ligands and receptors regulates a wide spectrum of biological processes via the initiation of a complicated cascade of intracellular signaling pathways, in which extracellular signal-regulated kinases (Erk), Akt, mitogen-activated protein kinase (MAPK), PI3 γ , protein kinase C (PKC) and Jak-STAT are involved (5-8). Moreover, the activation of these signaling pathways may lead to cell cycle arrest, cell proliferation, differentiation, tumorigenic development and anti-apoptotic processes (3,9,10). A previous study demonstrated that the Nrg1-ErbB signaling pathway is essential in organ development, cell differentiation and tumorigenesis (7).

Expression and functions of Nrg1 have also been systematically described in the nervous system, particularly in the spinal cord, cortex and hippocampus. Moreover, recent data suggest that Nrg1-ErbB4 signaling is key in the pathogenesis of a diverse range of neurodegenerative and neurological diseases, including Alzheimer's disease (11,12), multiple sclerosis (13), schizophrenia (14), neurospongioma (15) and brain damage (5). Perturbed Nrg1 signaling in these diseases extends our understanding of the biological functions of Nrg1.

Neuroinflammation is essential in diseases affecting the central nervous system (CNS). Neuronal apoptosis occurs in response to multiple cytokines, chemokines and toxic factors secreted by glial cells during this process (16). Inflammatory signals, such as prostaglandins, proinflammatory cytokines or lipopolysaccharide (LPS), can cross the intact or ruptured blood-brain barrier (BBB) of the CNS (17), and controlled and balanced inflammation contribute to the repair of damaged tissues following brain injury (18).

Although multiple factors have been determined to be associated with the neuroinflammatory process, the involvement

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of Nrg1-ErbB signaling in this condition remains largely unknown. Nrg1, like other specific members of the cytokine family, such as interleukin-1 and epidermal growth factor (EGF), was affected following brain infection and injury; thus, Nrg1 is hypothesized to partially contribute to the development of schizophrenia (19). Nrg1 mRNA and intracellular protein levels increased in response to the duration of LPS exposure in human umbilical venous endothelial cells, supporting its potential role as a systemic endogenous inhibitor of perinatal inflammatory brain damage during fetal development (20).

It was hypothesized that inflammation can affect the expression of Nrg1 and result in intracellular signaling changes, thus contributing to the pathology of neuroinflammation-based neurodegenerative diseases. LPS is a component of the outer cell wall of Gram-negative bacteria, and it has been widely used to induce inflammation in the brain (11,21). Therefore, the effects of LPS-induced neuroinflammation on the expression of Nrg1 and its related cell signaling molecules were analyzed in several major brain structures *in vivo* using a mouse model treated with intraperitoneal injection of LPS.

It was observed that LPS induced differential changes in Nrg1 and its receptors Neu and ErbB4 in the cortex, striatum, hippocampus and hypothalamus. Notably, a profound reduction of Nrg1 expression was found in the hypothalamus and striatum, suggesting that the Nrg1 signaling in these locations is more prone to be influenced by LPS-induced neuroinflammation. These findings may provide greater insight into the function of Nrg1 in inflammation-related neuronal diseases, including AD, PD and SCI, and may further aid in the development of novel clinical therapies.

Materials and methods

Animals. Eleven female C57BL/6 mice (age, 3 months; weight, ~25 g) were purchased from the Guangdong Medical Laboratory Animal Center (Guangzhou, China) and were maintained at 25°C under a 12-h dark/light cycle, with access to food and water *ad libitum*. All experimental procedures conducted on animals were approved by the Animal Ethics Committees of Shantou University Medical College (Shantou, China) and the authorities of Guangdong Province.

LPS treatment. A total of 1.0 mg LPS (Sigma-Aldrich Israel, Ltd., Rehovot, Israel) was dissolved in 10 ml normal saline (pH 7.4), such that the final concentration of LPS solution was 0.01%. Six mice were intraperitoneally injected with LPS at a dose of 1.0 mg/kg (10 ml/kg), and 5 mice in the control group were injected with normal saline at 10 ml/kg.

Tissue preparation. Mice were then sacrificed by decapitation 24 h after LPS or saline injection. For morphological studies, 2 mice from each group were subjected to intracardiac perfusion of phosphate-buffered saline-buffered paraformaldehyde for fixation. For molecular studies, seven brains, three from the normal saline-treated mice and four from LPS-treated mice, were collected. The right and left hemisphere from each brain was separated. Moreover, the frontoparietal cortex, hippocampus, striatum and hypothalamus from each side were dissected and stored at -80°C for either reverse transcription-polymerase chain reaction (RT-PCR) or western blot analysis (22).

RT-PCR analysis. Total RNA from brain tissue was extracted using the DNAiso Reagent (Tiangen Biotech (Beijing) Co., Ltd., Beijing, China) according to the manufacturer's protocol. Reverse transcription was performed using StarScript II First-strand cDNA Synthesis mix (Gene-star Biosolutions Co., Ltd., Beijing, China), and RT-PCR was performed using 2X Taq PCR StarMix (Genstar Solutions). Sequences of primers are listed in Table I. PCR amplification was conducted with an initial denaturing step at 94°C for 2 min, then 35 cycles at 94°C for 30 sec, at 60°C for 30 sec, and at 72°C for 30 sec, and a further extension at 72°C for 10 min. For glyceraldehyde 3-phosphate dehydrogenase (GAPDH) cDNA amplification, 28 reaction cycles were employed. Then, 5 µl of the PCR products were used to perform gel electrophoresis using a 2.0% agarose gel (GeneChoice, Inc., Frederick, MD, USA) containing Gelred (1:10,000; Biotium, Inc., Hayward, CA, USA). The bands were identified under ultraviolet light.

Western blot analysis. For western blot analysis, tissue was dissolved in 200 µl radioimmunoprecipitation buffer containing PMSF (1%) (Solarbio Biotech Corp, Beijing, China) and homogenized using a motor-driven micro tissue grinder (Kimble Chase, Vineland, NJ, USA). The supernatants were collected following centrifugation at 14,000 x g for 15 min at 4°C. Protein concentrations were determined by the bicinchoninic acid assay (Solarbio Biotech Corp). Equivalent quantities (20 µg) of tissue lysates were heated at 95°C in 20% sample loading buffer consisting of 0.125 mol/l Tris-HCl, pH 1.8; 20% glycerol, 10% sodium dodecyl sulfate, 0.1% bromophenol blue and 5% β-mercaptoethanol, and were then resolved using 10% sodium dodecyl sulfate-polyacrylamide gel electrophoresis and electroblotted onto a polyvinylidene difluoride membrane (Millipore, Billerica, MA, USA). Non-specific protein-binding sites were blocked with 5% bovine serum albumin (Amresco, LLC, Solon, OH, USA) or non-fat milk diluted in Tris-buffered saline buffer containing 0.1% Tween-20 (TBST, pH 7.4). The membranes were incubated with the following anti-mouse primary antibodies: Polyclonal rabbit p-ErbB-4 (Tyr 1056; 1:500; #sc-33040); polyclonal rabbit ErbB-4 (C-18; 1:500; #sc-283); polyclonal mouse p-ErbB-2/p-Neu (7F8; 1:500; #sc-81508); monoclonal mouse Neu (3B5; 1:500; #sc-33684); polyclonal rabbit p-c-Src (Tyr 419; 1:500; #sc-101802); monoclonal mouse Src-1 (1135/H4; 1:500; #sc-32789); monoclonal mouse anti-p-Akt1/2/3 (11E6; 1:1,000; #sc-81433); monoclonal mouse Akt1 (G-5; 1:1,000; #sc-55523); monoclonal mouse p-Erk (E-4; 1:1,000; #sc-7383); monoclonal mouse Erk1/2 (MK1; 1:1,000; #sc-135900); monoclonal mouse GAPDH (G-9; 1:1,000; #sc-3650620) (all 1:1,000 Santa Cruz Biotechnology, Inc., Dallas, TX, USA) and monoclonal mouse Nrg1 Ab-1 (7D5; 1:500; MS-272-P1; LabVision; Thermo Fisher Scientific, Inc., Waltham, MA, USA) overnight at 4°C. After 3 washes for 5 min each, horseradish peroxidase-conjugated goat anti-mouse (#BA1051) and rabbit anti-goat (#BA1055) secondary antibodies (1:1,000; Wuhan Boster Biological Technology, Ltd., Wuhan, China) diluted in 3% non-fat milk in TBST were applied, followed by 3 washes with TBST for 5 min each at room temperature. The antigens were visualized using an enhanced chemiluminescence (ECL) solution (Beyotime Institute of Biotechnology, Haimen, China). The signal intensity was quantified using Image J software,

Table I. Sequences of primers for reverse transcription-polymerase chain reaction.

Gene	Domain	Primers	Accession no.	Annealing temperature (°C)	Expected length (bp)
ErbB4	5' terminus	5'-GCCAAAAATGAAGCTGGCGA-3'	NM_010154.1	60	219
	3' terminus	5'-TTGTGCTCGATGCTGGTGAT-3'			
Neu	5' terminus	5'-TAGCTGGGCATTACCCCTAC-3'	BC053078.1	55	358
	3' terminus	5'-AGGGCATAAGCTGTGTCACC-3'			
Nrg1	5' terminus	5'-GTCTCAGAGGGTGCCTCAAC-3'	NM_178591.2	60	332/230
	3' terminus	5'-GTTCTTCCGGGTGGGTACTG-3'			
GAPDH	5' terminus	5'-GTGGAGTCATACTGGAACATGTAG-3'		60	150
	3' terminus	5'-AATGGTGAAGGTTCGGTGTG-3'			

Nrg1, neuregulin 1; GAPDH, glyceraldehyde 3-phosphate dehydrogenase.

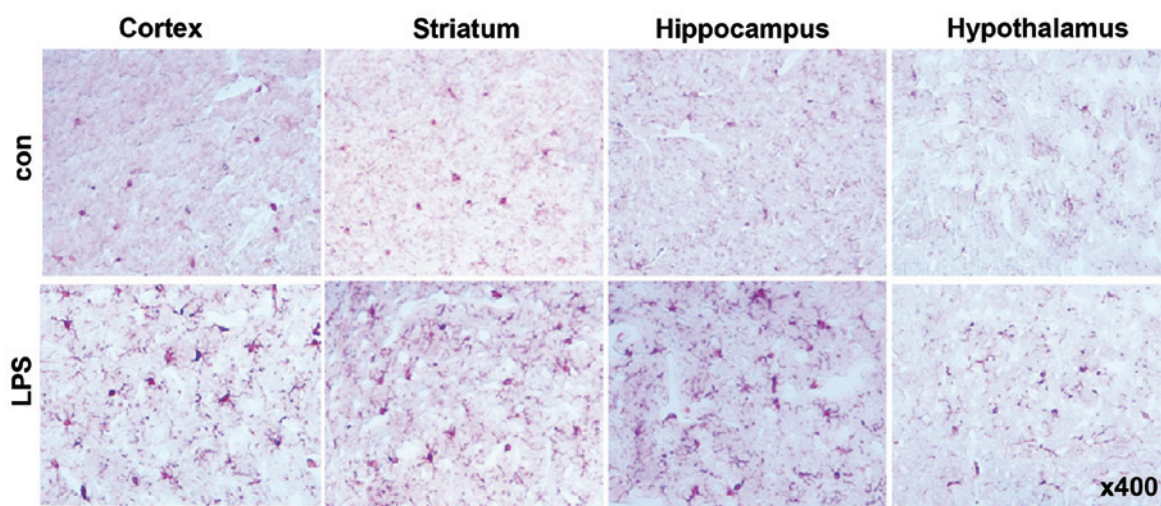


Figure 1. Systemic LPS injection induced neuroinflammation in major brain regions. Immunohistochemical staining demonstrated a wide distribution of Iba-1-positive activated microglia in the cortex, striatum and hippocampus. By contrast, the number of activated microglia was limited in the hypothalamus. LPS, lipopolysaccharide.

version 1.48 (rsb.info.nih.gov/ij/) as the average densitometry multiplied by the area (measured as the number of pixels).

Immunohistochemical staining. Mice were perfused with a 4% paraformaldehyde solution followed by decapitation and then cryoprotected in 20% sucrose. Brain sections (8 μ m thick) were prepared using a cryostat microtome (Leica CM1860, Leica Biosystems, Germany). Antigen retrieval was performed using 10-mM citrate buffer (pH 6.0) for 40 min at 99°C. Then, the tissue sections were incubated in 3% H₂O₂ to clear endogenous peroxidase and were then saturated with 10% normal goat serum for 10 min. Next, sections were incubated with the following primary antibodies: Rabbit polyclonal anti-pNeu and pErbB4 antibodies (1:100; Santa Cruz Biotechnology, Inc.). After being washed in PBS 3 times for 5 min each, the biotinylated secondary antibody and the streptavidin-peroxidase conjugate were applied. The enzyme activity was developed using an AEC kit (ZLI-9036; ZSGB Biotechnology Co., Ltd., Beijing, China). the antigen-antibody complexes were visualized using the AEC method, according to the manufacturer's protocols.

Statistical analysis. Statistical analyses were performed with SPSS17.0 (SPSS software Inc., Chicago, IL, USA). Data were analyzed using Student's t-test for independent samples. $P < 0.05$ was considered to indicate a statistically significant difference.

Results

Neuroinflammatory response to LPS administration in major mouse brain regions. Intraperitoneal administration of LPS induced an apparent increase in the number of activated microglial cells positive for Iba-1 in the cortex, striatum, hippocampus and hypothalamus (Fig. 1), indicating the successful induction of neuroinflammation in our model.

Determination of Nrg1, Neu and ErbB4 expression in different brain regions in response to LPS. RT-PCR was used to investigate the expression changes of Nrg1, Neu and ErbB4 at the mRNA level in major brain regions in response to LPS administration for 24 h. It was demonstrated that LPS induced an apparent increase in the Neu mRNA level in the cortex, striatum and hippocampus, but a profound reduction

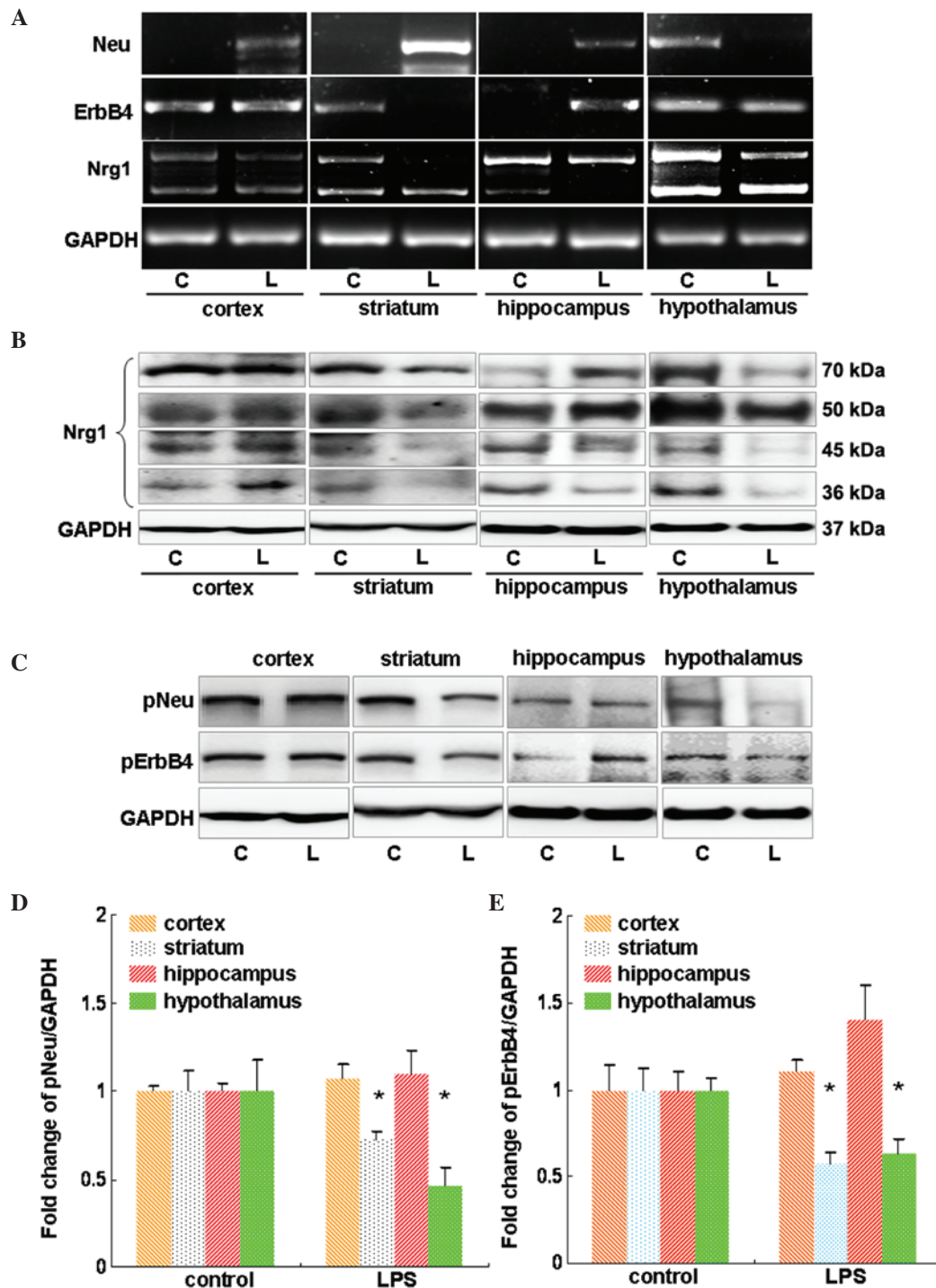


Figure 2. Evaluation of the effects of LPS on Nrg1 expression and Neu and ErbB4 expression and activation in mouse cortex, striatum, hippocampus and hypothalamus. (A) Reverse transcription-polymerase chain reaction analysis of the influence of LPS on the mRNA levels of Nrg1, Neu and ErbB4. (B) Western blot analysis of the effect of LPS on the protein levels of multiple isoforms of Nrg1. (C) Western blot analysis of the effect of LPS on the phosphorylation-induced activation of Neu and ErbB4. Quantification of the (D) Neu and (E) ErbB4 western blot results. Independent Student's t-test was applied. * $P < 0.05$ vs. the control group (control group, $n = 3$; LPS group, $n = 4$). LPS, lipopolysaccharide; Nrg1, neuregulin-1; C, control group; L, LPS group; GAPDH, glyceraldehyde 3-phosphate dehydrogenase; p-, phosphorylated.

in the Neu mRNA level was observed in the hypothalamus (Fig. 2A). LPS showed no effect on the mRNA levels of ErbB4 in the cortex and hypothalamus (Fig. 2A). However, the mRNA levels of ErbB4 were reduced in the striatum but were increased in the hippocampus in the LPS-treated mice compared with that of the controls (Fig. 2A). In addition, LPS reduced the mRNA levels of type I Nrg1 in the striatum and

hypothalamus (Fig. 2A). In accordance with these findings, in these 2 regions decreased protein levels of Nrg1 were observed, the molecular weights of which were 70, 50, 45 and 36 kDa. By contrast, the mRNA level of type II Nrg1 was reduced in the hippocampus, which also showed reduced 45 and 36 kDa Nrg1 protein levels (Fig. 2B). Notably, the LPS-treated mice showed no apparent changes in the mRNA levels of both types of Nrg1

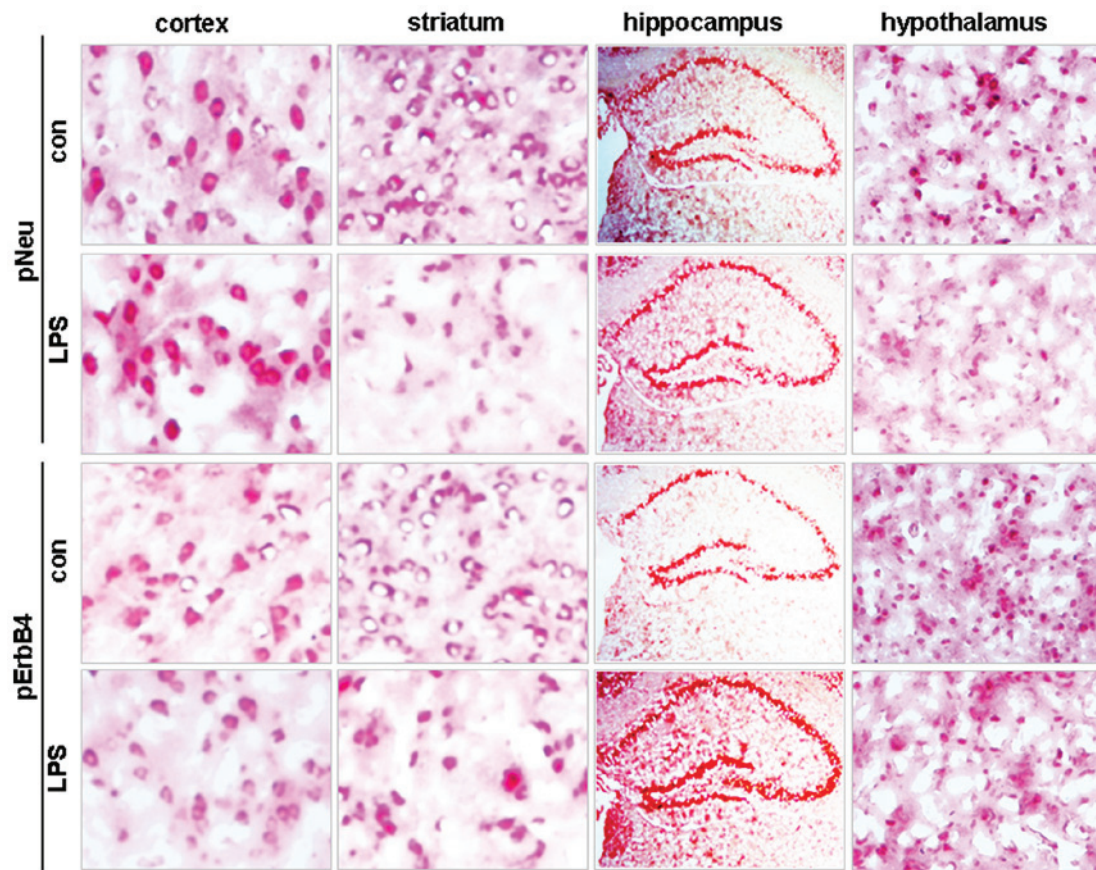


Figure 3. Immunohistochemical staining of pNeu and pErbB4 in the cortex, striatum, hippocampus and hypothalamus in mice treated with either saline (con) or LPS. Original magnification, x400 for cortex, striatum and hypothalamus; x100 for hippocampus. LPS, lipopolysaccharide; p-, phosphorylated.

in the cortex compared with the vehicle control mice (Fig. 2B). This was further confirmed by western blot analysis, which revealed no profound changes in the different Nrg1 isoforms at the protein level (Fig. 2B).

The effect of LPS on Neu and ErbB4 phosphorylation was then investigated in major brain regions. As shown in Fig. 2C and D, administration of LPS showed no apparent effect on the phosphorylation of Neu and ErbB4 in the cortex ($P > 0.05$ vs. the vehicle control); however, it induced a significant reduction in the phosphorylation of molecules in the striatum and hypothalamus ($P < 0.05$ vs. the vehicle control; Fig. 2C and D). In the hippocampus, LPS showed no effect on pNeu level, but it increased the pErbB4 level (Fig. 2C-E).

Effect of LPS on the phosphorylation-induced activation of Neu and ErbB4 in major brain regions in mice. In accordance with the western blot data, immunohistochemical staining demonstrated no changes in either pNeu or pErbB4 levels in cortical brain tissue, but it reduced the stained signals for the two molecules in the striatum and hypothalamus (Fig. 3). None of the hippocampal areas, including the CA1, CA2 and CA3 zones and the dentate gyrus, showed changes in pNeu levels, whereas increased pErbB4 levels were observed in these areas in LPS-treated mice compared with control (Fig. 3).

Western blot analysis of the effects of LPS on signaling pathways in different parts of the brain. To gain additional insight into the influence of LPS on the downstream signaling pathways

of Nrg1, the phosphorylation levels of Src, Akt1 and Erk were measured using western blot analysis (Fig. 4). The phosphorylation of c-Src was significantly upregulated in the cortical tissue ($P < 0.05$ vs. the vehicle control). By contrast, it was down-regulated in the hypothalamus ($P < 0.01$ vs. the vehicle control) (Fig. 4A and B). In addition, the Akt1 phosphorylation level, as indicated by the pAkt1/Akt1 ratio, was significantly increased in the striatum ($P < 0.05$ vs. the vehicle control), with no significant changes identified in the cortex and hypothalamus. The hippocampus also showed an increased average pAkt1/Akt1 ratio, although no significant difference was found when compared with the control mice (Fig. 4A and C). As indexed by pErk/Erk ratio, LPS resulted in marginally reduced Erk phosphorylation in the cortex but increased it in the striatum, although no significance was found when compared with the vehicle controls. No obvious changes in Erk phosphorylation were observed in either the hippocampus or hypothalamus (Fig. 4A and D).

Discussion

Traditionally, transforming growth factor α -transactivated ErbB1/epidermal growth factor receptor is required for the LPS response, and neuregulins are required to transactivate ErbB4 during the acid-stress response (23). Stability enhancement of neuregulin receptor degradation protein-1 inhibits the production of proinflammatory cytokines in macrophages triggered by toll-like receptors. This process may also reduce the production of LPS-induced proinflammatory mediators, such as inducible

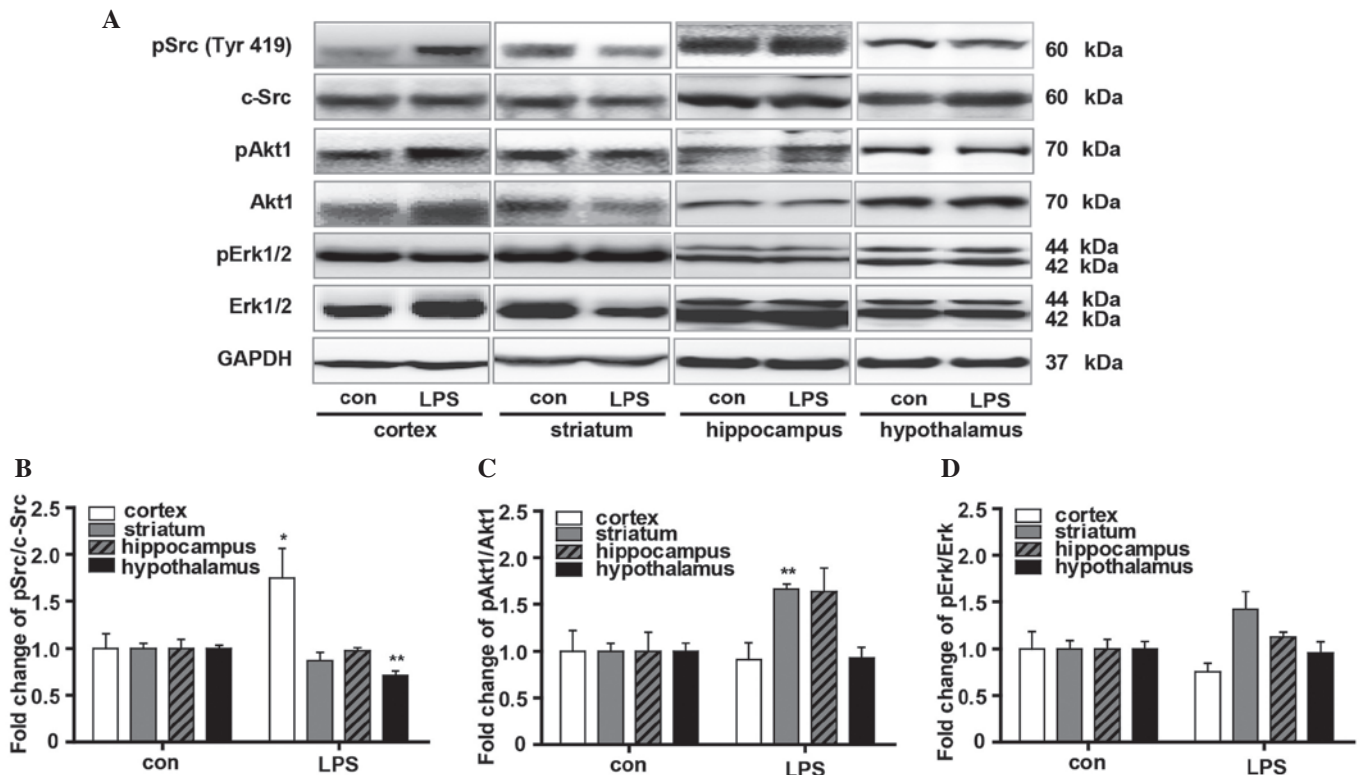


Figure 4. Evaluation of the effects of LPS on the signaling pathway downstream of Nrg1. (A) Western blot analysis of the effect of LPS on the phosphorylation of Src, Akt1 and Erk in the cortex, striatum, hippocampus and hypothalamus in mice. Graphs presenting quantification of (B) Src, (C) Akt1 and (D) Erk western blot results. Independent Student's t-test was applied. * $P < 0.05$ and ** $P < 0.01$ vs. the control group (control group, $n = 3$; LPS group, $n = 4$). LPS, lipopolysaccharide; Nrg1, neuregulin 1; Erk, extracellular signal-regulated kinase; p-, phosphorylated; GAPDH, glyceraldehyde 3-phosphate dehydrogenase.

nitric oxide synthase, nitric oxide, cyclooxygenase-2, and prostaglandin E_2 (24). A loss of ADAM17-mediated growth factor shedding was reported to abrogate cytokine release of primary SMC stimulated by LPS or the supernatant of acid-exposed epithelial cells (23). Previous studies have revealed that Nrg1 functions by blocking the delayed neuronal death and proinflammatory responses in rats subjected to focal ischemic stroke (25). Neuregulins can modulate a diverse array of biological effects based on the expression of the receptors and cell types that are associated with the survival and function of neuronal cells (26-30). To the best of our knowledge, the present study observed for the first time the differential changes in Nrg1 expression and the phosphorylation of its receptors Neu and ErbB4, as well as of its downstream signaling molecules, in different brain regions of the mouse following neuroinflammation induced by LPS. Either type I or type II Nrg1 mRNA levels were reduced by LPS in the hippocampus, striatum and hypothalamus, whereas the mRNA level of its receptor Neu was reduced only in the hypothalamus. Using western blotting, it was demonstrated that the expression of several isoforms of Nrg1 was upregulated in the cortex and hippocampus in response to LPS. However, their expression was downregulated in the LPS-affected striatum and hypothalamus, in which the phosphorylation levels of Neu and ErbB4 were notably downregulated. These combined results suggest that LPS can differentially modulate the expression of Nrg1 signaling, and that Nrg1 may differentially function in different brain regions upon neuroinflammation.

Inflammation is considered to activate transcription factors, including nuclear factor (NF)- κ B and Jak-STAT, which can

regulate pro-inflammatory gene expression. Nrg1 and its receptors ErbB3 and ErbB4 are highly expressed not only in the developing nervous system but also in the mature nervous system (31-33). Following brain lesion development, the levels of Nrg and the ErbB4 receptor are increased (34,35), suggesting that Nrg/ErbB4 signaling may function in synaptic plasticity and neuroprotection. Nrg1 is crucial in the development of the brain (36), laying the foundation for its roles in inflammation-related processes in the disease setting. It is considered that Nrg1 functions upstream of NF- κ B-mediated cell signaling in response to certain inflammatory conditions (37).

The mechanisms underlying the role of Nrg1 in neuroinflammation are unknown. It has been reported that Nrg1 β can exert neuroprotective effects on cell survival via the activation of the phosphatidylinositol-3-kinase (PI3K)/Akt signaling pathway (38,39). It was suggested that the potential neuroprotective effects of Nrg1 are due to its ability to modulate multiple biological processes during neuroinflammation. To test this, Nrg1 and the phosphorylation levels of certain signaling pathway proteins, including Src, Akt1 and Erk were examined. Using western blotting techniques, it was demonstrated that the phosphorylation level of Src was increased in the cortex but decreased in the hypothalamus. However, the phosphorylation levels of Akt1 and Erk were upregulated in the striatum and hippocampus.

In conclusion, the present results show differential changes in Nrg1 signaling in major mouse brain regions in response to LPS, suggesting a differential role for Nrg1 in these brain regions during the development of neuroinflammation. Modulating

Nrg1 signaling may assist in the treatment of neurotraumatic and neurodegenerative diseases that involve neuroinflammation.

Acknowledgements

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