

Inhibitory effect of metformin combined with gemcitabine on pancreatic cancer cells *in vitro* and *in vivo*

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Abstract. Pancreatic cancer is a malignant digestive system tumor with a particularly poor prognosis, and is the fourth leading cause of cancer-associated mortality in the USA. The anti-diabetic therapeutic agent, metformin (MET) has been demonstrated to exert anti-tumor effects. The present study assessed the ability of MET, alone or in combination with gemcitabine (GEM), to inhibit the growth of the human CFPAC-1 pancreatic cancer cell line *in vitro* and *in vivo*. Cell counting kit-8 assays were performed to measure CFPAC-1 cell viability and apoptosis was detected with annexin V/propidium iodide. Cell cycle analysis was conducted by flow cytometry. The mRNA and protein levels of B-cell lymphoma-extra large (Bcl-xL), Bcl2 associated X protein (Bax), caspase-3, survivin and cyclin D1 in CFPAC-1 cells and tumor tissues were detected by reverse transcription-polymerase chain reaction and western blotting, respectively. Furthermore, the expression levels of caspase-3 and proliferating cell nuclear antigen in tumor tissues were detected by immunohistochemistry. The results demonstrated that following MET treatment, the growth of CFPAC-1 cells and xenografts in nude mice was inhibited, the expression levels of Bcl-xL, survivin and cyclin D1 were downregulated, while the expression levels of Bax and caspase-3 were upregulated. These effects were enhanced when MET was administered in combination with GEM. The mechanism underlying the anti-tumor effect of MET may be associated with the induction of cell apoptosis and the inhibition of proliferation.

Introduction

Pancreatic cancer is the fourth leading cause of cancer-associated mortality for men and women in the USA, with a 5-year survival rate of <5% (1). To date, the sole potential cure for pancreatic cancer is surgery (2). However, ~80% patients present when the cancer is too advanced for surgery (3). For patients with locally advanced or metastatic inoperable pancreatic cancer, chemotherapy and radiotherapy are the standard treatments. Gemcitabine (GEM) is currently the first-line therapeutic agent prescribed for advanced pancreatic cancer; however, its efficacy is limited, with a median survival extension of just six weeks (4). Therefore, adjuvant strategies have been increasingly investigated to improve these outcomes.

Metformin (MET) is widely used in the treatment of type 2 diabetes mellitus and has been investigated due to its anti-tumor effects (5). Previous studies have suggested that MET has an effect on various cancer types (6), including pancreatic cancer. Certain *in vitro* and *in vivo* studies using mouse xenograft models have indicated that MET may exert direct anti-tumor activity via the inhibition of cancer cell proliferation, migration and invasion, promotion of cancer cell apoptosis, and suppression of tumor growth (7-9). However, the underlying mechanism of these effects and the combination of MET with conventional chemotherapy remains to be clarified.

The aim of the present study was to investigate the underlying anti-tumor mechanism of MET on the growth of the CFPAC-1 human pancreatic cell line, and to evaluate the effects of combination therapy with MET and GEM *in vitro* and *in vivo*.

Materials and methods

Cell lines and animals. The human CFPAC-1 pancreatic ductal adenocarcinoma cell line was donated by the Institute of Hematology of Soochow University (Suzhou, China). The cells were incubated in 75-cm² cell culture flasks and maintained in RPMI-1640 (Gibco; Thermo Fisher Scientific, Inc., Waltham, MA, USA), supplemented with 10% fetal bovine serum (Gibco; Thermo Fisher Scientific, Inc.), 100 U/ml penicillin and 100 µg/ml streptomycin in a humidified atmosphere composed of 5% CO₂ at 37°C. A total of 20 specific-pathogen-free female athymic BALB/c nu/nu mice (age, 4-5 weeks; weight, 15-16 g) were purchased from the Shanghai Laboratory Animal Center (Shanghai, China). Mice were allowed free access to

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sterilized water and food, and were housed in individual ventilated cages at 23±5°C under a 12-h light/dark cycle. The experimental protocol was approved by the Animal Care and Use Committee of Soochow University.

Cell counting kit (CCK)-8 assay. The CFPAC-1 cells were trypsinized with 0.05% trypsin and then plated in 96-well plates with 100 µl medium per well. Following an overnight incubation in 5% CO₂ at 37°C, cells were treated with normal saline (NS) or MET (0, 1, 2.5, 5, 10, 20, 40 or 60 mmol/l; Sigma-Aldrich, St. Louis, MO, USA), with six replicates per concentration. Following incubations of 24, 48 or 72 h at 37°C, cells were incubated for 1-4 h with 10 µl CCK-8 reagent per well. Cell viability was measured with CCK-8 according to the manufacturer's instructions (Peptide Institute, Inc., Osaka, Japan), and the survival and growth inhibition rates were calculated as follows: Survival rate = absorbance of MET (1, 2.5, 5, 10, 20, 40 or 60 mmol/l) / absorbance of MET (0 mmol/l) × 100; inhibition rate = [absorbance of MET (0 mmol/l) - absorbance of MET (1, 2.5, 5, 10, 20, 40 or 60 mmol/l)] / absorbance of MET (0 mmol/l) × 100. In the combination therapy experiment, cells were treated with MET (20 mmol/l) and GEM (5 µmol/l) alone or in combination and cell viability was measured as above.

Cell cycle analysis and annexin V/propidium iodide (PI) assay. The CFPAC-1 cells were placed in 6-well plates at 5×10⁴ cells/well and treated with NS or MET (10, 20 or 40 mmol/l). Separate cell cultures were collected and trypsinized 48 h later, then washed twice with cold phosphate-buffered saline (PBS). The cells were subsequently incubated with an annexin V/PI double staining solution (Sigma-Aldrich) at room temperature. Fifteen min later, the stained cells were analyzed by flow cytometry and the percentage of apoptotic cells (those in the lower right quadrant) was calculated with ModFit LT software version 4.0 (Verity Software House, Inc., Topsham, ME, USA). Cell cycle analysis was performed by flow cytometry using CellQuest Pro software version 5.1 (BD Biosciences, Franklin Lakes, NJ, USA). In combination therapy experiments, cells were treated with MET (20 mmol/l) and GEM (5 µmol/l), alone or in combination for 48 h. The percentage of apoptotic cells was calculated as above.

Mice xenograft model. The CFPAC-1 cells were resuspended in serum-free RPMI-1640 at a concentration of 1×10⁷ cells/ml. A total volume of 0.2 ml cell suspension (2×10⁶ cells) was then injected subcutaneously into the right anterior armpit of nude mice, to establish a xenograft model. The 20 injected mice were randomly divided into four groups: NS-treated control, MET-treated, GEM-treated and combination therapy (n=5 per group). MET (200 mg/kg, daily), GEM (50 mg/kg, twice/week) or combination therapy [MET (200 mg/kg, daily)+GEM (50 mg/kg, twice/week)] were administered intraperitoneally for four weeks. The tumor volume was measured with calipers every second day and calculated as follows: Volume (mm³) = 4π / 3 × (width / 2)² × (length / 2). The mice were weighed weekly then sacrificed by cervical dislocation following four weeks of treatment and the tumors were collected.

Reverse transcription-polymerase chain reaction (RT-PCR) assay. Total RNA was extracted from CFPAC-1 cells and tumor tissues using TRIzol® reagent (Invitrogen; Thermo Fisher Scientific, Inc.). The first strand cDNA was synthesized with the Omniscript RT kit (Qiagen GmbH, Hilden, Germany), using 2,000 ng RNA per 20 µl reaction and an oligo (dT) primer. PCR reactions were performed for 35 cycles. Each cycle was performed as follows: Denaturation for 30 sec at 94°C, annealing for 30 sec at 60°C and polymerization for 30 sec at 72°C. Primers used in the PCR reaction were as follows: Forward, 5'-CGGGCATTTCAGTGACCTGAC-3' and reverse, 5'-TCAGGAACCAGCGGTTGAAG-3' for B-cell lymphoma-extra large (Bcl-xL); forward, 5'-GCGTCCACC AAGAAGCTGA-3' and reverse, 5'-ACCACCCTGGTCTTG GATCC-3' for Bcl2 associated X protein (Bax); forward, 5'-GTCTCAATGCCACAGTCCAGT-3' and reverse, 5'-AGC AAACCTCAGGGAAACATT-3' for caspase-3; forward, 5'-TACGCCTGTAATACCAGCAC-3' and reverse, 5'-TCT CCGCAGTTTCCTCAA-3' for survivin; forward, 5'-TGT TCGTGGCCTCTAAGATG-3' and reverse, 5'-ACTCCAGAA GGGCTTCAATC-3' for cyclin D1; and forward, 5'-AGCGGG AAATCGTGCGTG-3' and reverse, 5'-CAGGGTACATGG TGGTGCC-3' for β-actin. The PCR products were detected using Molecular Analyst software version 2.10e (BioRad Laboratories, Inc., Hercules, CA, USA). β-actin served as an internal standard.

Western blot assay. Cells or tumor tissue were lysed with radioimmunoprecipitation assay buffer on ice for 30 min and centrifuged at 12,000 × g, 4°C for 10 min. The protein samples (20 µg) were separated on a 10% SDS-PAGE gel, at 80 V (stacking gel) and 100 V (separating gel), and transferred onto nitrocellulose membranes. The membranes were blocked with PBS-Tween (PBS-T) containing 5% skimmed milk powder for 1 h at room temperature. The membranes were then incubated at 4°C overnight with primary mouse monoclonal antibodies, diluted 1:1,000, against Bcl-xL (catalog no. sc-8392), Bax (catalog no. sc-7480), caspase-3 (catalog no. sc-65497), survivin (catalog no. sc-374616) and cyclin D1 (catalog no. sc-8396), purchased from Santa Cruz Biotechnology, Inc. (Dallas, TX, USA). Subsequently, the membranes were washed with PBS-T three times and incubated for 1 h with a goat anti-mouse IgG antibody conjugated to horseradish peroxidase (HRP) (1:1,000; catalog no. sc-2031; Santa Cruz Biotechnology, Inc.). Membranes were then washed three times with PBS-T, and protein bands were visualized using an enhanced chemiluminescence (ECL) -detection system (Beyotime Institute of Biotechnology, Haimen, China).

Immunohistochemistry. Tumor xenografts were fixed in formalin and embedded in paraffin blocks, which were sliced into 4-µm thick sections for immunohistochemical staining.

Following deparaffinization, the sections were incubated at 4°C overnight with primary mouse monoclonal antibodies, diluted 1:200, against caspase-3 (catalog no. 65497) or proliferating cell nuclear antigen (PCNA; catalog no. sc-25280), purchased from Santa Cruz Biotechnology, Inc. The sections were washed three times with PBS-T, and incubated for 1 h at room temperature with a goat anti-mouse IgG antibody conjugated to HRP (1:200; catalog no. sc-2031; Santa Cruz

Biotechnology, Inc.). The slices were washed with PBS-T three times, and incubated in diaminobenzidine (Sangon Biotech Co., Ltd., Shanghai, China) and counterstained with hematoxylin (Sangon Biotech Co., Ltd.) for 1 min. Images were captured using a light microscope (magnification, x400; model CKX41-A32RC; Olympus Corporation, Tokyo, Japan).

Statistical analysis. Data were expressed as the mean \pm standard error and analyzed using SPSS version 18.0 (SPSS, Inc., Chicago, IL, USA). Statistical analysis was performed using one-way analysis of variance. The Student-Newman-Keuls (SNK) method was used as a post-hoc test. The Kruskal-Wallis test was used to evaluate the differences of categorical values, followed by the Mann-Whitney U test as a post-hoc test. $P < 0.05$ was considered to indicate a statistically significant difference.

Results

Effect of MET on growth of CFPAC-1 cells. The human CFPAC-1 pancreatic ductal adenocarcinoma cells were treated with various concentrations of MET for 24, 48 or 72 h, and survival and growth inhibition rates were evaluated with the CCK-8 assay. The proliferation and survival of CFPAC-1 cells was inhibited in a dose-and time-dependent manner (Fig. 1A and B). Cell apoptosis was determined by annexin V/PI assay. The percentage of apoptotic cells increased in a dose-dependent manner following a 48 h incubation with MET (Fig. 1C; Table I). Cell cycle analysis revealed that cells in the G₀/G₁ phase increased significantly following MET treatment in a dose-dependent manner (Fig. 1D and E; Table II). Furthermore, RT-PCR and western blot analysis demonstrated that the expression levels of Bcl-xL, survivin and cyclin D1 were reduced following 48 h of MET treatment (Fig. 1F and G). By contrast, the expression levels of Bax and caspase-3 increased following MET treatment.

Effect of MET combined with GEM on growth of CFPAC-1 cells. The CFPAC-1 cells were treated with MET, GEM or a combination of the two agents. CCK-8 assay revealed that the proliferation and survival of CFPAC-1 cells were inhibited significantly following treatment with MET in combination with GEM, compared with MET or GEM alone (combined treatment vs. MET treatment at 24, 48 and 72 h, $P < 0.001$; combined treatment vs. GEM treatment at 24, 48 and 72 h, $P < 0.001$; Fig. 2A and B). In addition, the percentage of apoptotic cells increased following combination therapy compared with MET or GEM treatment alone (Fig. 2C; Table III). The mRNA and protein expression levels of Bcl-xL, survivin and cyclin D1 were decreased in MET and GEM alone treatment groups, and a greater decrease was observed in the combination therapy group (Fig. 2D and E). Similarly, a greater increase in Bax and caspase-3 expression levels was observed in the combination therapy group compared with the single treatment groups.

Growth inhibitory effect of MET *in vivo*. All mice survived for the duration of the experiment with no observable toxic effect, such as weight loss. Mice were sacrificed at 28 days and the weights of the xenografts were measured. In comparison to the NS-treated control group, xenografts from mice treated with

Table I. Impact of MET on viability of CFPAC-1 cells *in vitro*.

Parameter	MET concentration (mmol/l)			
	0	10	20	40
Apoptosis, %	3.01 \pm 0.49	13.77 \pm 1.31 ^a	22.63 \pm 1.45 ^a	32.97 \pm 3.19 ^a

As measured by an annexin V/propidium iodide assay. Data are expressed as the mean \pm standard error. ^a $P < 0.05$ vs. normal saline treated control cells. MET, metformin.

MET, GEM or combination therapy were smaller in size and weight (MET vs. normal saline treatment, $P = 0.007$; GEM vs. normal saline treatment, $P = 0.001$; combined vs. normal saline treatment, $P < 0.001$; Fig. 3A-C). Furthermore, the tumor volume and weight were decreased significantly following combined treatment (combined vs. MET treatment, $P < 0.001$; combined vs. GEM treatment, $P = 0.001$). Bcl-xL, survivin, Bax, caspase-3 and cyclin D1 mRNA and protein expression levels followed a similar pattern to that of the *in vitro* experiment (Fig. 3D and E). The protein expression level of caspase-3 was markedly upregulated in the combination therapy group, and this was confirmed by immunohistochemistry (Fig. 4A). However, expression of PCNA protein was decreased following treatment (Fig. 4B).

Discussion

The glucose-lowering effect of MET is associated with an increase in insulin sensitivity *in vivo*, resulting in increased glucose uptake, decreased gluconeogenesis and reduced plasma glucose concentrations. Emerging evidence from *in vitro*, *in vivo* and epidemiological studies suggests an anti-tumor role of MET (10-12). The underlying mechanisms are complex and require further investigation. It has been demonstrated that MET activates the 5' adenosine monophosphate-activated protein kinase/mammalian target of rapamycin signaling pathway through inhibition of complex 1 of the mitochondrial respiratory chain (13,14). In addition, MET has indirect anti-tumor effects via the reduction of circulating insulin and insulin growth factor 1 levels (15). *In vitro*, MET induces cell cycle arrest, thus inhibiting proliferation and inducing cell apoptosis. Furthermore, MET affects the proliferation of cancer stem cells, reduces DNA damage and inhibits the inflammatory response (16-18).

In the present study, the influence of MET on the apoptosis and proliferation of the human CFPAC-1 pancreatic ductal adenocarcinoma cell line was investigated *in vitro* and *in vivo*. The mechanism underlying induction of apoptosis in cancer cells by MET may be associated with altered expression of pro-apoptotic and anti-apoptotic molecules. In addition, MET arrested cells in the G₀/G₁ phase and downregulated the expression of cyclin D1 and PCNA, demonstrating anti-proliferative activity. Furthermore, the pro-apoptotic and anti-proliferative activities of MET were enhanced when it was administered in combination with GEM.

Appropriate apoptotic signaling is crucial for preserving tissue homeostasis by maintaining a healthy balance between

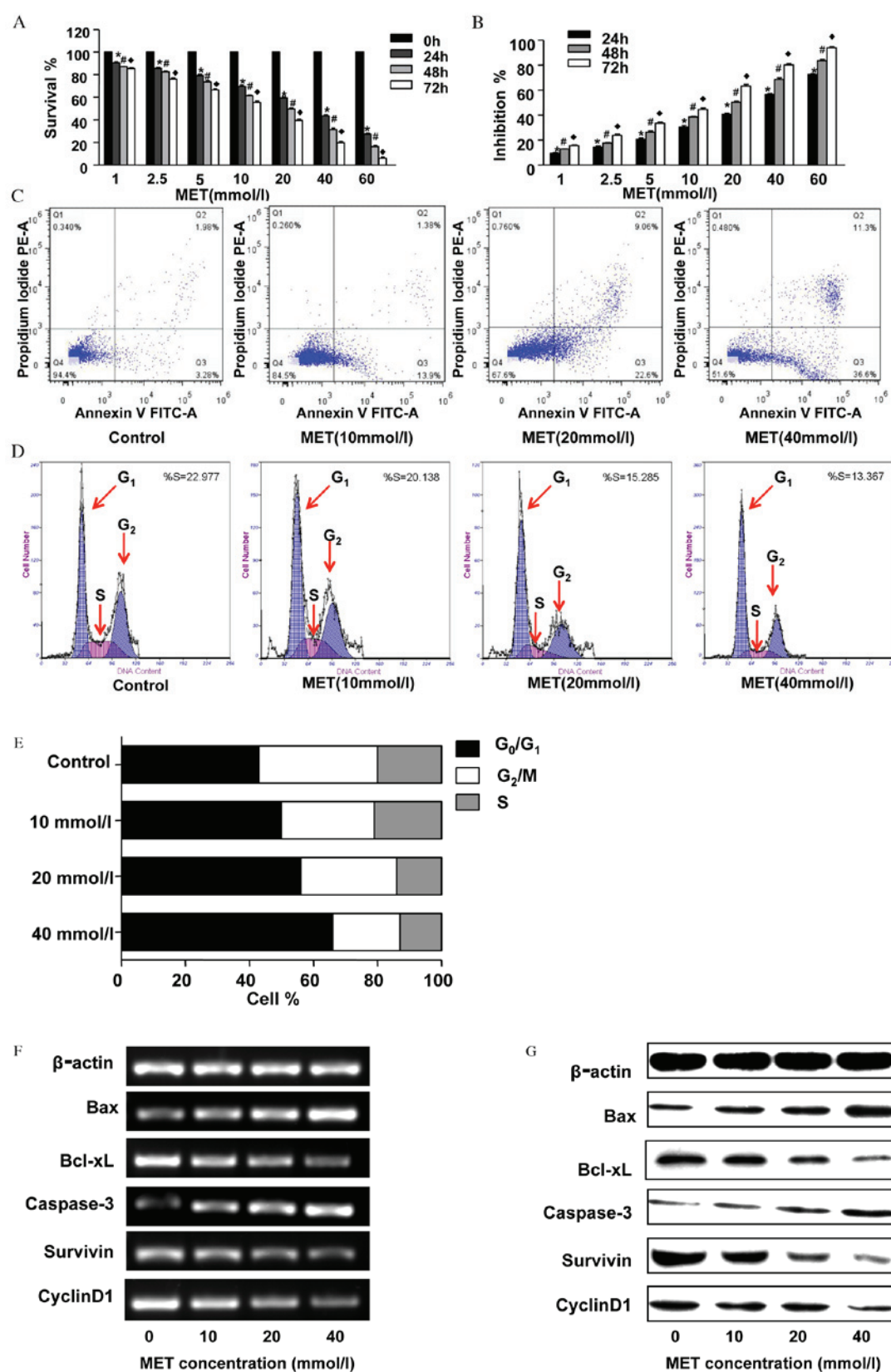


Figure 1. Inhibitory effect of MET on the growth of the human CFPAC-1 pancreatic cancer cell line *in vitro*. CFPAC-1 cells were incubated with various concentrations of MET, from 0-60 mmol/l, for 0, 24, 48 or 72 h. The cell counting kit-8 assay was then conducted to analyze (A) cell viability and (B) growth inhibition rates. MET treatment decreased survival and inhibited growth of CFPAC-1 cells. Data are presented as the mean \pm standard error. (C) The percentage of apoptotic CFPAC-1 cells (the lower right quadrant, annexin V⁺ PI⁺) was analyzed using an annexin V/propidium iodide assay. (D) Cell cycle analysis was conducted by flow cytometry. (E) Percentage of cells in G₀/G₁ phases increased following MET treatment. Subsequent to treatment with MET, CFPAC-1 cells were harvested and the (F) mRNA and (G) protein expression levels of Bax, Bcl-xL, caspase-3, survivin and cyclin D1 were analyzed by reverse transcription-polymerase chain reaction and western blotting, respectively. β -actin served as an internal standard. The expression levels of Bax and caspase-3 increased following MET treatment, while Bcl-xL, survivin and cyclin D1 were reduced. * $P < 0.05$ vs. 0 h; # $P < 0.05$ vs. 24 h; * $P < 0.05$ vs. 48 h. MET, metformin; Bcl-xL, B-cell lymphoma-extra large; Bax, Bcl2 associated X protein.

Table II. Impact of MET on cell cycle of CFPAC-1 cells *in vitro*.

Cell cycle phase	MET concentration (mmol/l)			
	0	10	20	40
G ₀ /G ₁ , %	42.89±1.02	49.59±3.15 ^a	56.03±1.49 ^a	65.93±0.27 ^a
G ₂ /M, %	38.28±4.93	28.72±1.32 ^a	30.70±2.75 ^a	22.01±2.95 ^a
S, %	20.12±3.38	21.68±4.18	13.27±1.78 ^a	13.58±0.43 ^a

Data are expressed as the mean ± standard error. ^aP<0.05 vs. normal saline treated control cells. MET, metformin.

Table III. Impact of MET combined with GEM on the viability of CFPAC-1 cells *in vitro*.

Parameter	Treatment group			
	Control	20 mmol/l MET	5 μmol/l GEM	20 mmol/l MET+5 μmol/l GEM
Apoptosis, %	3.01±0.49	24.53±2.18 ^a	22.37±1.61 ^a	52.07±2.81 ^{a,b}

As measured by an annexin V/propidium iodide assay. Data are expressed as the mean ± standard error. ^aP<0.05 vs. normal saline treated control cells; ^bP<0.05 vs. MET- and GEM-treatment MET, metformin; GEM, gemcitabine.

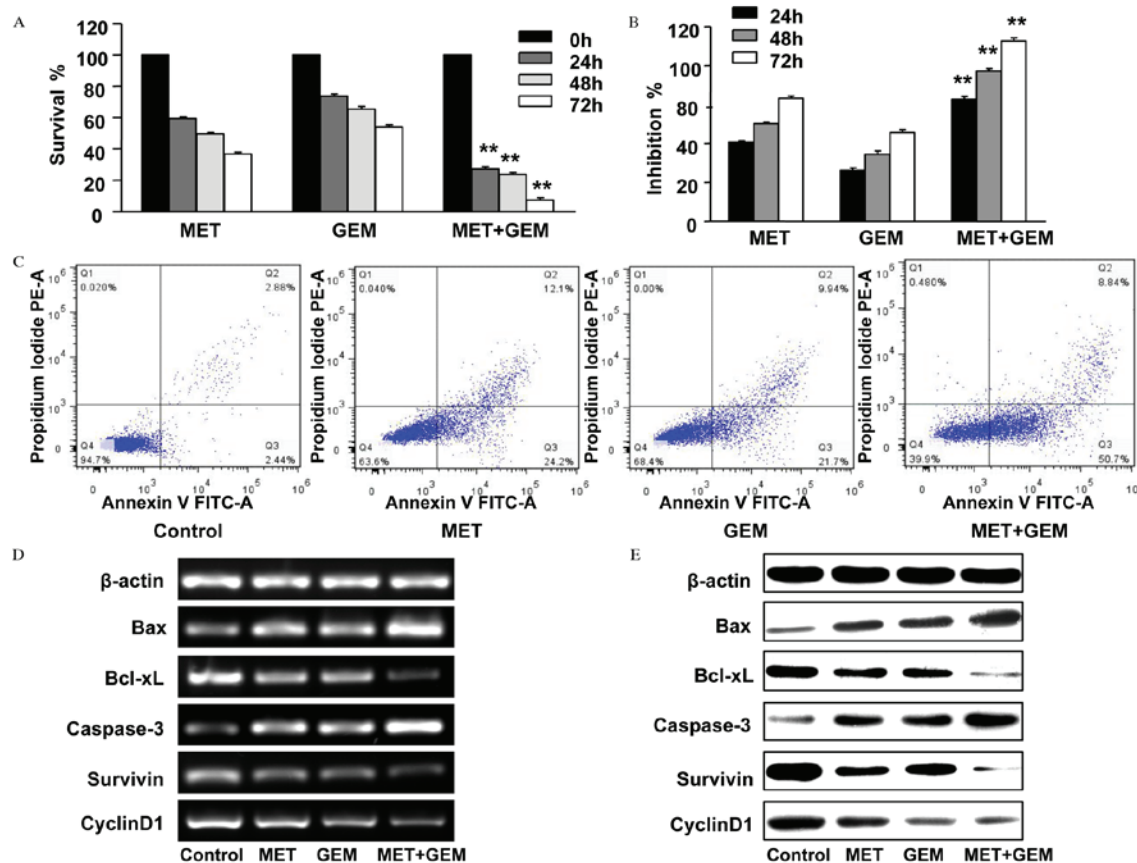


Figure 2. Inhibitory effect of MET combined with GEM on the growth of the human CFPAC-1 pancreatic cancer cell line *in vitro*. CFPAC-1 cells were incubated with 20 mmol/l MET, 5 μmol/l GEM or MET and GEM in combination for 0, 24, 48 or 72 h. The cell counting kit-8 assay was conducted to analyze (A) cell viability and (B) growth inhibition rates. Combination therapy significantly increased the effectiveness of MET and GEM. The data are presented as the mean ± standard error. (C) The percentage of apoptotic CFPAC-1 cells (the lower right quadrant, annexin V⁺ PI⁺) was analyzed using an annexin V/propidium iodide assay. Subsequent to treatment with 20 mmol/l MET, 5 μmol/l GEM or MET and GEM combined, CFPAC-1 cells were harvested and the (D) mRNA and (E) protein expression levels of Bax, Bcl-xL, caspase-3, survivin and cyclin D1 were analyzed by reverse transcription-polymerase chain reaction and western blotting, respectively. ^{**}P<0.05 vs. MET or GEM treatment alone. MET, metformin; GEM, gemcitabine; Bcl-xL, B-cell lymphoma-extra large; Bax, Bcl2 associated X protein.

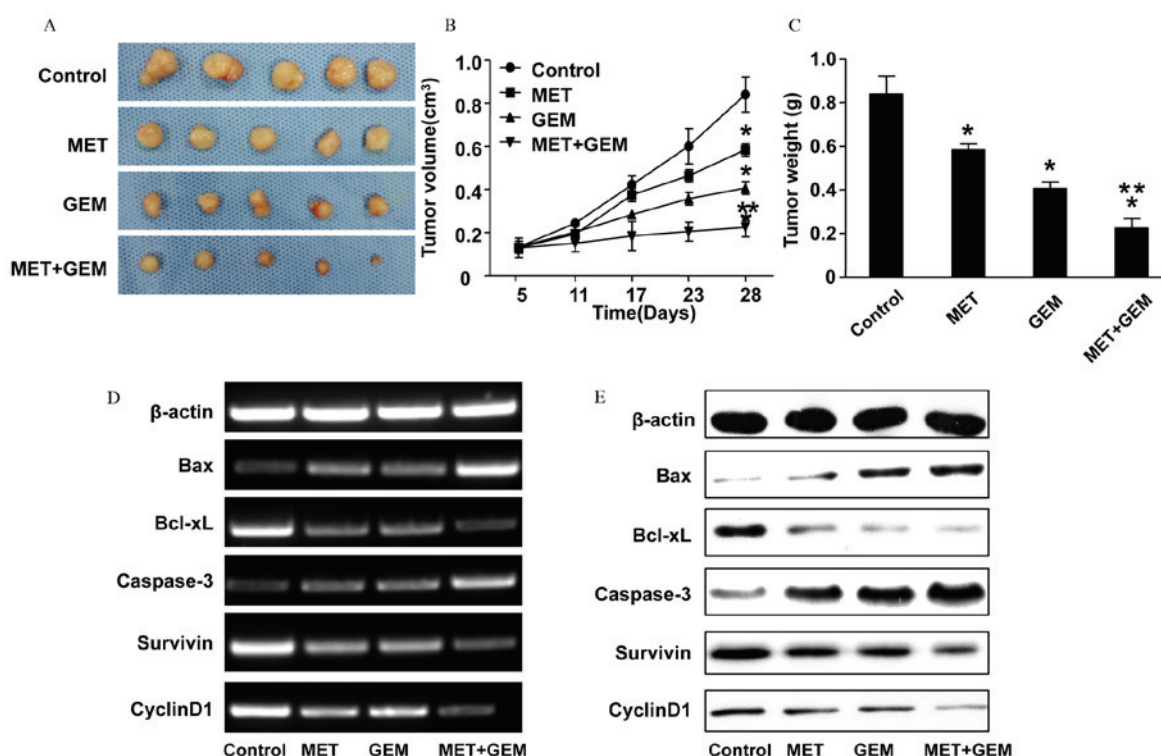


Figure 3. Inhibitory effect of combined treatment with MET and GEM on the growth of the human CFPAC-1 pancreatic cancer cell line *in vivo*. CFPAC-1 cells were injected into nude mice, which were then treated for 28 days with 200 mg/kg MET, 50 mg/kg GEM or MET and GEM in combination. (A) Gross morphology of tumors at sacrifice. (B) Volume changes in the xenografts. Treatment with MET, GEM or a combination of the two significantly inhibited tumor growth, compared with the control group. The data are presented as the mean \pm SE. (C) Following 28 days of treatment, the tumor weights were significantly decreased in all three treatment groups compared with the control group. The data are presented as the mean \pm SE. (D) mRNA and (E) protein expression levels of Bax, Bcl-xL, caspase-3, survivin and cyclin D1 in nude mice xenografts were analyzed by reverse transcription-polymerase chain reaction and western blotting, respectively. * $P < 0.05$ vs. normal saline-treated control group; ** $P < 0.05$ vs. MET or GEM treatment alone. MET, metformin; GEM, gemcitabine; Bcl-xL, B-cell lymphoma-extra large; Bax, Bcl2 associated X protein; SE, standard error.

cell death and cell survival. The ratio of pro- and anti-apoptotic molecules regulates cell death. Carcinogenesis may occur if the balance is disturbed and apoptosis in malignant cells is reduced. Bcl-xL and Bax are members of the Bcl-2 protein family and, as such, are important in apoptosis. Bax mediates the permeabilization of the outer mitochondrial membrane and the release of cytochrome *c* into the cytoplasm, which activates caspase-3 to induce chromosome cleavage and apoptosis (19). However, the effect triggered by Bax is blocked by Bcl-xL (20). It has been reported that >50% of cancers are associated with excessive expression of Bcl-xL (21). Survivin belongs to the inhibitor of apoptosis proteins (IAPs) family and is considered a node protein, inhibiting apoptosis and regulating cell mitosis (22). Promising cancer treatment strategies that target apoptotic inhibitors, including Bcl-2 family proteins and IAPs are currently under investigation (23).

The cell cycle is divided into three phases: G_0 , the interphase (G_1 , S and G_2) and M. Cells are quiescent in the G_0 phase. The G_1 checkpoint control mechanism ensures that the cell is prepared for DNA synthesis. Cyclin D1 is an important cell cycle regulatory protein, which performs a positive role during the crucial restriction point of the G_1 /S transition. Cyclin D1 expression and accumulation are induced by growth factors and occur at multiple levels, including increased transcription, translation and protein stability (24).

PCNA is a DNA clamp that acts as a processivity factor for DNA polymerase δ in eukaryotic cells, and the presence of PCNA is a specific marker of cell proliferation (25).

Pancreatic cancer remains one of the most fatal types of cancer despite the oncological advances achieved over the past two decades. Patients suffering from pancreatic cancer have a median survival time of 4-6 months (26). GEM, the first-line chemotherapeutic agent prescribed in unresectable cases, only marginally improves the outcome. GEM-based combination therapies, including cisplatin, capecitabine and exatecan have failed to make any significant improvements (27-29). Furthermore, combination therapies may be more toxic and, therefore, less well tolerated (27-29). The lack of effective and less toxic treatment options for pancreatic cancer has prompted investigations into novel combined treatment strategies.

There are complex associations between diabetes mellitus and pancreatic cancer, as diabetes may be a risk factor for, or a result of, pancreatic cancer (30). Previous studies have demonstrated that MET is beneficial for pancreatic cancer prognosis (31,32). A borderline significant relative survival benefit was observed in MET-treated patients compared with non-MET-treated patients [hazard ratio (HR), 0.80; 95% confidence interval (CI), 0.62-1.03] in a pooled analysis of four publications containing 1,429 patients (33). However, few clinical trials of MET in pancreatic cancer treatment have been reported.

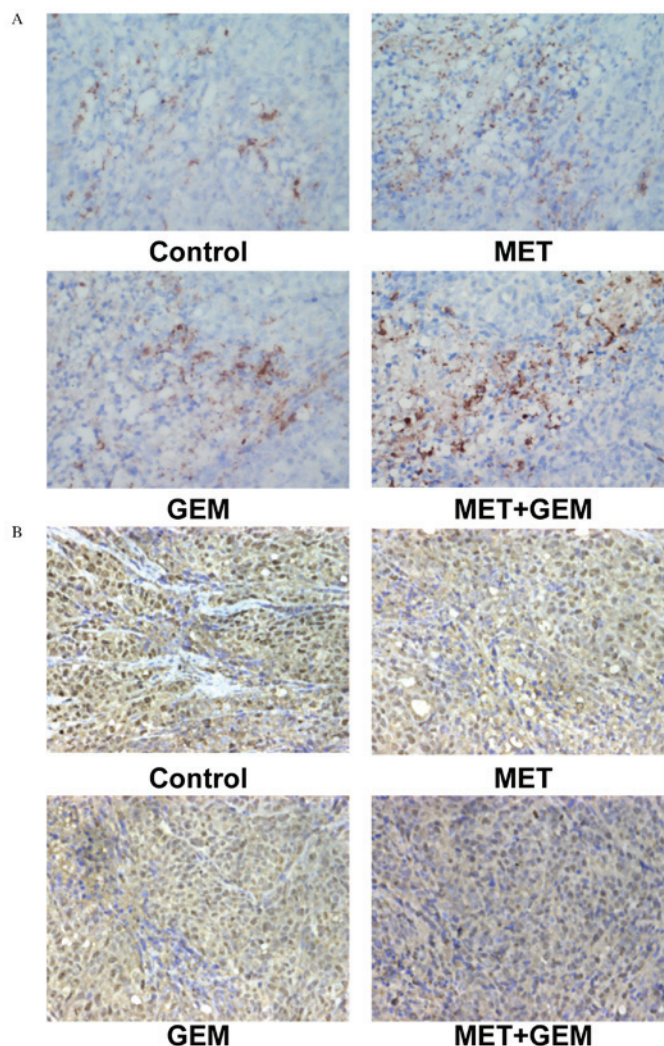


Figure 4. Effect of combination therapy with 200 mg/kg MET and 50 mg/kg GEM in the nude mouse xenograft model. Immunohistochemical detection of (A) caspase-3 and (B) PCNA. Staining of caspase-3 increased following combination therapy, while PCNA staining decreased. Magnification, x400. MET, metformin; GEM, gemcitabine; PCNA, proliferating cell nuclear antigen.

Between 2010 and 2014, Kordes *et al* (34) presented a randomized, placebo-controlled trial of MET in the treatment of patients with advanced pancreatic cancer. Despite the laboratory evidence of anti-tumor activity, MET addition did not improve the clinical outcome for these patients (median survival of 7.6 months in the placebo group vs. 6.8 months in the MET group; HR, 1.056; 95% CI, 0.72-1.55). The authors hypothesized that the conventional anti-diabetic dose of MET may fail to accumulate to a sufficient concentration to cause energetic stress (34). Blood MET concentrations were in the micromolar range, significantly lower than that of *in vitro* studies, which demonstrated the direct anti-tumor effects of MET. In addition, only patients with advanced pancreatic cancer were included in the trial (34). Future studies focusing on patients with hyperinsulinaemia or patients with tumors expressing markers of sensitivity to energetic stress are required.

In conclusion, MET inhibited the growth of pancreatic cancer cells via the induction of apoptosis and the reduction of proliferation. In addition, the efficacy of MET was significantly improved when administered in combination with GEM.

The findings of the present study may provide an alternative strategy for the treatment of pancreatic cancer, but requires validation in clinical trials. In addition, the mechanisms underlying the effects of apoptosis induction and proliferation reduction require additional investigation. Furthermore, future studies are required to identify inhibitors of oxidative phosphorylation, which are more potent than MET.

Acknowledgements

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