

Wip1 regulates SKOV3 cell apoptosis through the p38 MAPK signaling pathway

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Received January 8, 2016; Accepted January 19, 2017

DOI: 10.3892/mmr.2017.6469

Abstract. The aim of the present study was to explore the effect of silencing wild-type p53-induced phosphatase 1 (Wip1) on apoptosis of human ovarian cancer SKOV3 cells. SKOV3 cells cultured *in vitro* were divided into three groups: untreated cells, cells transfected with control small interfering RNA (siRNA) and cells transfected with siRNA targeting Wip1. Flow cytometry analysis was used to detect cell apoptosis. Western blot analysis was performed to determine expression of tumor protein 53 (p53), cleaved caspase-3, caspase-3, BCL2 associated X (Bax), BCL2 apoptosis regulator (Bcl-2), p38 mitogen-activated protein kinase (p38 MAPK) and phosphorylated (p)-p38 MAPK. Reverse transcription-quantitative polymerase chain reaction was used to detect expression of p53, Bax, Bcl-2 and caspase-3 mRNAs. Compared with control, apoptosis of SKOV3 cell was significantly increased following Wip1 siRNA silencing. Wip1 silencing also resulted in a significant increase of p53 and p-p38 MAPK expression, as well as increased cleaved caspase-3/caspase-3 and Bax/Bcl-2 protein ratios. No significant differences were observed in apoptosis and apoptosis-related protein expression in the control siRNA transfected cells. The present study demonstrated that Wip1 silencing promotes apoptosis of human ovarian cancer SKOV3 cells by activation of the p38 MAPK signaling pathways and through subsequent upregulation of p53, and cleaved caspase-3/caspase-3 and Bax/Bcl-2 protein ratios. Overall, the findings of the present study suggest that targeting Wip1 may be a potential therapeutic avenue for the treatment of human ovarian cancer in the future.

Introduction

Ovarian cancer, one of the most common gynecologic malignancies, is an insidious cancer with a 5-year survival rate of merely 20-30% (1). Currently, the pathogenesis of ovarian cancer remains incompletely understood. Apoptosis is a highly regulated, crucial biological process leading to cell death: Regulatory imbalance can cause excessive cell proliferation or apoptosis, leading to diseases including ovarian cancer. Several studies have demonstrated that abnormal apoptosis regulation in ovarian cells contributes to the development of ovarian cancer (2,3). Wild-type p53-induced phosphatase 1 [Wip1; official name: protein phosphatase, Mg²⁺/Mn²⁺ dependent 1D (PPM1D)], a p53-dependent proto-oncogene first identified in 1997 (4), is widely involved in the regulation of multiple cell signaling pathways, and is important in the pathogenesis of cancers, including ovarian cancer (5-7).

Previous studies have demonstrated that Wip1 is closely associated with apoptosis. Ma *et al* (8) revealed that manganese (Mn) exposure led to neuronal necrosis in rats, accompanied by a significant increase in neuronal apoptosis and a notable reduction in Wip1 expression in nerve tissues and cells. Sun *et al* (9,10) reported that Wip1 expression was significantly higher in nasopharyngeal cancer and renal cancer tissues than in normal tissues. Wip1 silencing led to a markedly accelerated apoptosis in these types of cancer cells, indicating involvement of Wip1 in suppressing apoptosis. By contrast, elevated Wip1 expression exhibits an inhibitory effect on apoptosis (8-10). To the best of our knowledge, the mechanism by which Wip1 regulates apoptosis in ovarian cancer cells has not been reported to date.

The present study aimed to investigate *in vitro* the role of Wip1 in apoptosis of ovarian cancer SKOV3 cells and its potential mechanism of action.

Materials and methods

Cell culture. The human ovarian cancer cell lines SKOV3, CAOV3, AZ780, ES2 and the normal ovarian epithelial cell line were purchased from Cell Center, Peking Union Medical College (Beijing, China). They were cultured in Dulbecco's modified eagle's medium-1640 supplemented with 5% fetal bovine serum (FBS), 2 mM l-glutamine, 100 U/ml penicillin

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Key words: ovarian cancer, wild-type p53-induced phosphatase 1, human ovarian cancer SKOV3 cells, apoptosis

and 100 µg/ml streptomycin in an atmosphere containing 95% air, 5% CO₂. Cells were plated (1 × 10³ cells/well) in 96-well plates for 24 h and incubated at 37°C for 4 h in 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT), which was purchased from Sigma-Aldrich; Merck KGaA (Darmstadt, Germany). The medium was removed, 50 µl DMSO was added to each well and incubated at room temperature for 45 min while shaking. Absorbance was measured at a wavelength of 570 nm, using a SynergyMx microplate reader (Bio Tek Instruments, Inc., Winooski, VT, USA) to determine the viable cell fraction.

Cells at a 75-85% confluence were either left untreated, transfected with Wip1 siRNA plasmid or control siRNA plasmid which was performed with Lipofectamine[®] 2000 (Invitrogen; Thermo Fisher Scientific, Inc., Waltham, MA, USA) according to the manufacturer's protocol, and then collected for experimental assay 48 h following transfection.

Antibodies and siRNAs. Antibodies towards Wip1 (cat. no. D4F7), p38 mitogen-activated protein kinase (p38 MAPK; cat. no. 9212), phosphorylated (p-) p38 MAPK (Thr180/Tyr182; catalog no. 3D7), tumor protein 53 (p53; cat. no. 7F5), mitogen-activated protein kinase 1 (ERK; cat. no. 137F5), phosphorylated (p-) ERK (Thr202/Tyr204; cat. no. D13.14.4E), mitogen-activated protein kinase 8 (JNK; cat. no. 56G8), phosphorylated (p-) JNK (Thr183/Tyr185; cat. no. G9) and cleaved caspase-3 (cat. no. 9661) and the MAPK inhibitor SB203580 were purchased from Cell Signaling Technology, Inc. (1:1,000; Danvers, MA, USA). Mouse anti-BCL2 (cat. no. ab7923) associated X (Bax; cat. no. ab77566) monoclonal antibody, rabbit anti-BCL2 apoptosis regulator (Bcl-2; cat. no. ab7973), caspase-3 (cat. no. ab32499) antibody were diluted at 1:1,000 and purchased from Abcam, Cambridge, UK. Pro-Light horseradish peroxidase chemiluminescence detection reagents were purchased from Tiangen Biotech Co., Ltd. (Beijing, China). siRNAs were purchased from Sigma-Aldrich; Merck KGaA. siRNA sequences were as follows: Wip1 siRNA-1, 5'-UUGUGAGUGAGUCGAGGUCGUUCC-3'; Wip1 siRNA-2, 5'-UAUCCUAAAAGUCAGGGCUUAGCG-3'; Wip1 siRNA-3, 5'-CCTCACAGCGAAAGAACTCTGTAA-3'; and control non-targeting N-siRNA, 5'-GAGUGGGUCUGGGUCUCCCCGUAGA-3'.

Apoptosis analysis by Annexin V-fluorescein isothiocyanate (FITC)/propidium iodide (PI) staining. Apoptotic cells in different groups were determined using an Annexin V/PI apoptosis detection kit according to the manufacturer's protocol (Multi Sciences Biotech Co., Ltd., Hangzhou, China). Briefly, the cell pellet was resuspended in 1x binding buffer followed by incubation with 5 ml of Annexin V (conjugated with FITC) and 10 ml of PI, in the dark for 5 min. Cell fluorescence was then analyzed using a flow cytometer (Epics-XLII, Becton Coulter, Inc., Brea, CA, USA). This test discriminates intact cells (Annexin V-/PI-), early apoptotic cells (Annexin V+/PI-) and late apoptotic cells (Annexin V+/PI+).

Western blot. Cells were lysed in lysis buffer for 2 h on ice. Confluent cell layers were washed with PBS and lysed for 30 min at 4°C with 1% Nonidet P-40, 0.1% Triton X-100, 30 mM sodium phosphate (pH 7.4) containing 1 mM sodium

orthovanadate, 2.5 mM Tris-HCl (pH 7.5), 100 mM NaCl, and 10 µg/ml of leupeptin, aprotinin. Following centrifugation at 13,000 × g for 20 min at 4°C, the supernatant was collected. Protein concentration was measured using a Lowry protein assay. Total cell lysates (40 µg protein) were separated by polyacrylamide 10% SDS-PAGE gels, and electrotransferred to a polyvinylidene fluoride membrane. The membrane was blocked by incubation with 0.05% nonfat milk powder in TBST for 2 h at 37°C, and then primary antibodies to cleaved caspase-3, caspase-3, p53, p38 MAPK, p-p38 MAPK, Bax and Bcl-2 were added for incubation overnight at 4°C. Following washing, the membrane was incubated with horseradish peroxidase-labeled goat anti-mouse or rabbit secondary antibody (1:5,000 dilution in TBST containing 1% bovine serum albumin; Santa Cruz Biotechnology, Inc., Dallas, TX, USA) for 120 min at room temperature and then washed three times with TBST at 37°C. Finally, blots were visualized by chemiluminescence and autoradiography. LabWorks 4.5 analysis software (UVP, Inc., Upland, CA, USA) was used to perform quantification of bands in the western blots.

Reverse transcription-quantitative polymerase chain reaction (RT-qPCR). Total RNA was extracted using TRIzol (Thermo Fisher Scientific, Inc., Waltham, MA, USA) and reverse transcribed. Total RNA (2 µg) was reverse transcribed using random primers and M-MLV at 42°C for 1 h and then heated to 94°C for 5 min in a total reaction volume of 20 µl. Equal amounts of the product of the reverse transcription reaction were subjected to PCR amplification. qPCR was performed at a final volume of 20 µl, with 20 µl SYBR Premix Ex Taq[™] II (2X; Takara Bio, Inc., Otsu, Japan), 0.4 µl ROX Reference Dye (50X; Takara Bio, Inc.), 2 µl reverse transcription cDNA, 0.8 µl of mixed forward and reverse primers (at a final concentration of 1 ng/ml) and 6 µl double-distilled water. The reaction was performed for 40 cycles as follows: 95°C for 30 sec, 95°C for 5 sec, 55°C for 30 sec and 72°C for 30 sec. Relative fold changes in mRNA expression were calculated using the 2^{-ΔΔC_q} method (11). Primer sequences were as follows: Bax, forward 5'-GCTTGCTTCAGGGTTTCATCCA-3' and reverse 5'-TGTCCACGGCGGCAATCATC-3'; Bcl-2, forward 5'-GGCGGA GAACAGGGTACGATAAC-3' and reverse 5'-CGGGAT GCGGCTGGATGGGG-3'; p53, forward 5'-CTAACCGCG GTCCCTTCCCAGAAAACCTAC-3' and reverse 5'-TACAGT CAGAGCCAACCTCAGGCG-3'; caspase-3, forward 5'-AAT GTCATCTCGCTCTTGGT-3' and reverse 5'-GCTTAGAAT CACACACAC-3'; 18S ribosomal RNA, forward 5'-ACACGG ACAGGATTGACAGA-3' and reverse 5'-GGACATCTAAGG GCATCACAG-3'.

Statistical analysis. Experimental data were expressed as the mean + standard deviation. Statistical analysis was performed with the SPSS 13.0 statistical software (SPSS, Inc. Chicago, IL, USA). One-way analysis of variance was followed by Dunnett's test for multiple comparisons. P<0.05 was considered to indicate a statistically significant difference.

Results

Screening of ovarian cancer cell lines. Wip1 expression was analyzed in various ovarian cancer cell lines by RT-qPCR

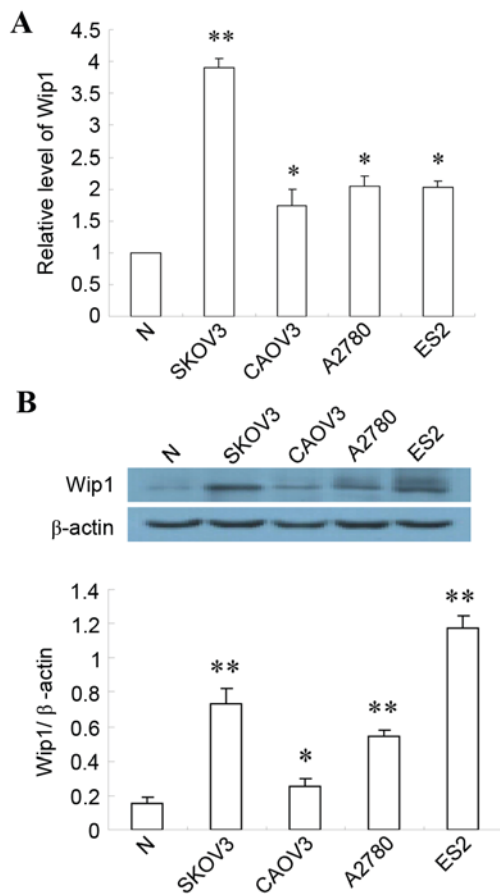


Figure 1. Expression of Wip1 in ovarian cancer cell lines SKOV3, CAOV3, A2780, ES2 and ovarian epithelial cell line. (A) Wip1 mRNA expression levels were assessed by reverse transcription-quantitative polymerase chain reaction. Results are plotted relative to normal control (n=6). (B) Western blot analysis of Wip1 protein expression levels with quantification relative to β -actin control (n=6). * $P<0.05$ and ** $P<0.01$ vs. N. Wip1, wild-type p53-induced phosphatase 1; N, ovarian epithelial cell line.

and western blotting. Both mRNA and protein expression levels of Wip1 were significantly elevated in SKOV3, CAOV3, A2780 and ES2 cells ovarian cancer cell lines compared with normal (ovarian epithelial cell line), indicating that overexpression of Wip1 is a feature of ovarian cancer cells (Fig. 1). Expression of Wip1 mRNA and protein was significantly higher in SKOV3 cells than in the other three cell lines (Fig. 1). Consequently, SKOV3 cells were selected to further explore the relationship between Wip1 and ovarian cancer.

Wip1 silencing by siRNA on SKOV3 cells. In order to efficiently silence expression of Wip1 in SKOV3 cells, three different siRNA sequences were tested, as well as various conditions for the transfection. As demonstrated in Fig. 2A, reduced Wip1 protein expression was observed in SKOV3 cells following siRNA transfection, compared with untransfected cells, with siRNA-2 being the most efficient. Wip1 protein expression was decreased in Wip1 siRNA-2 transfected cells compared with control, reaching the lowest level at 80 nmol/l siRNA (Fig. 2B) and at 48 h following transfection (Fig. 2C). Assessment of cell viability by MTT assay demonstrated that SKOV3 cells were still viable following Wip1 siRNA transfection, proliferation

was significantly decreased compared to control ($P<0.01$, Fig. 2D).

Effect of Wip1 silencing on apoptosis of SKOV3 cells. Assessment of apoptosis by Annexin V/PI staining revealed a significant increase in SKOV3 apoptosis following Wip1 silencing by siRNA, compared with control untransfected cells (Control; $P<0.01$, Fig. 3), or with non-targeting siRNA transfected cells (N-siRNA; $P<0.01$, Fig. 3). The apoptosis rate of SKOV3 cells was similar in both the control untransfected cells and the non-targeting siRNA transfected cells.

Effect of Wip1 silencing on mRNA expression of apoptosis-related proteins. To gain insight into the mechanism of SKOV3 apoptosis caused by Wip1 silencing, the expression of apoptosis-related proteins Bax, caspase-3, p53 and Bcl-2 was examined. Upon Wip1 silencing, relative expression of Bax (2.5 ± 0.034), caspase-3 (2.32 ± 0.032) and p53 (1.87 ± 0.068) mRNA was significantly higher in SKOV3 cells transfected with Wip1 siRNA, compared with control ($P<0.01$; Fig. 4A). The relative expression of Bcl-2 mRNA (0.47 ± 0.035) was significantly reduced compared with control ($P<0.05$; Fig. 4A), resulting in an increased Bax/Bcl-2 ratio. The mRNA expression of apoptosis-related proteins did not differ significantly in the control untransfected cells and the cells transfected with the non-targeting siRNA (data not shown). The above findings suggested that silencing Wip1 promoted expression of pro-apoptotic markers in SKOV3 cells.

Effect of Wip1 silencing on SKOV3 apoptosis-related protein expression. Protein expression of apoptosis-related markers Bax, cleaved caspase-3, caspase-3, p53 and Bcl-2 was detected by western blotting (Fig. 4B). In SKOV3 cells, expression of Bax/Bcl-2 protein ratio (0.86 ± 0.041), cleaved caspase-3/caspase-3 protein ratio (0.78 ± 0.031) and p53 protein (1.02 ± 0.051) was significantly higher in Wip1 siRNA cells than control (0.38 ± 0.059 ; $P<0.01$; Fig. 4B). There were no significant differences in expression of apoptosis-related proteins in the control untransfected cells (Control) compared with the cells transfected with the non-targeting siRNA (N-siRNA; Fig. 4B). These results indicated that silencing Wip1 promoted expression of pro-apoptotic proteins in SKOV3 cells.

Wip1 acts through the p38 MAPK signaling pathway. Following Wip1 silencing, expression of the major signaling molecules ERK, p-ERK, JNK, p-JNK, p38 MAPK and p-p38 MAPK was analyzed by western blot. The results demonstrated a significant increase in p-p38 MAPK protein ($P<0.05$, Fig. 5A), suggesting involvement of p38 MAPK signaling in the Wip1-mediated apoptosis effect in SKOV3 cells. To confirm this, the p38 MAPK inhibitor SB203580 was utilized to block p38 MAPK signaling: Treatment with SB203580 significantly reduced p38 MAPK phosphorylation ($P<0.01$, Fig. 5B). Assessment of cell proliferation by MTT assay revealed that SB203580 treatment completely reversed the proliferation-inhibition effect of Wip1 silencing in SKOV3 cells, suggesting that p38 MAPK is the main mediator of this effect ($P<0.01$, Fig. 5C). Based on these findings, Wip1 silencing promoted SKOV3 apoptosis by regulating the p38 MAPK pathway.

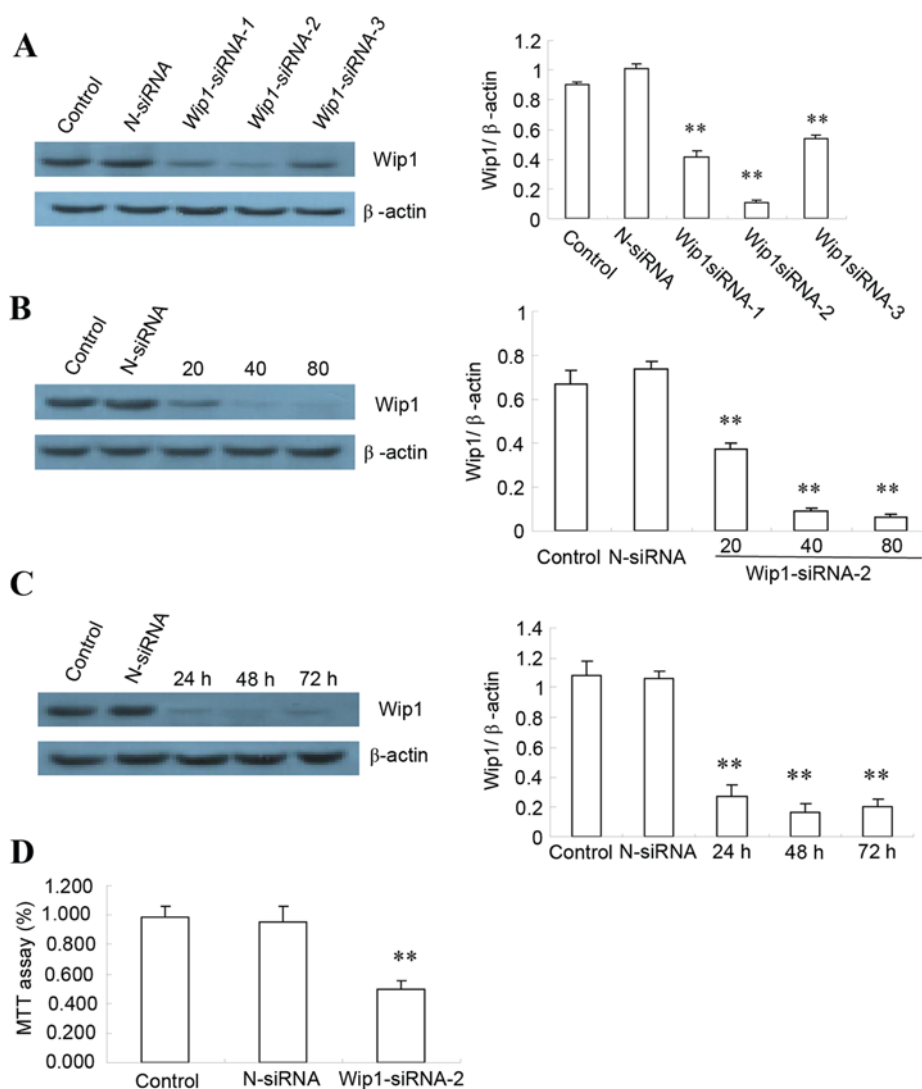


Figure 2. Wip1 silencing by siRNA in SKOV3 ovarian cancer cells. (A) Wip1 protein expression levels were assessed by western blotting in untransfected cells (Control) and cells transfected with either N-siRNA or one of the three different siRNA sequences (siRNA-1, -2 or -3), with quantification relative to β -actin. (B) SKOV3 cells were either left untransfected (Control) or transfected with either N-siRNA or with 20, 40, or 80 nmol/l Wip1-siRNA-2. The effect on Wip1 expression was analyzed by western blot. Representative images and quantification is reported here as fold ratio to β -actin. (C) SKOV3 cells were either left untransfected (Control) or transfected with either N-siRNA or Wip1-siRNA-2 (80 nmol/l Wip1-siRNA-2), for 24, 48 or 72 h. The effect on Wip1 expression was analyzed by western blot. Representative images and quantification is reported here as fold ratio to β -actin. (D) SKOV3 cells were either left untransfected (Control) or transfected with either N-siRNA or Wip1-siRNA-2 (80 nmol/l Wip1-siRNA-2 for 48 h), and the proliferation rate was analyzed by MTT assay (n=4 repeats). **P<0.01 vs. Control. Wip1, wild-type p53-induced phosphatase 1; siRNA, small interfering RNA; N-siRNA, non-targeting siRNA.

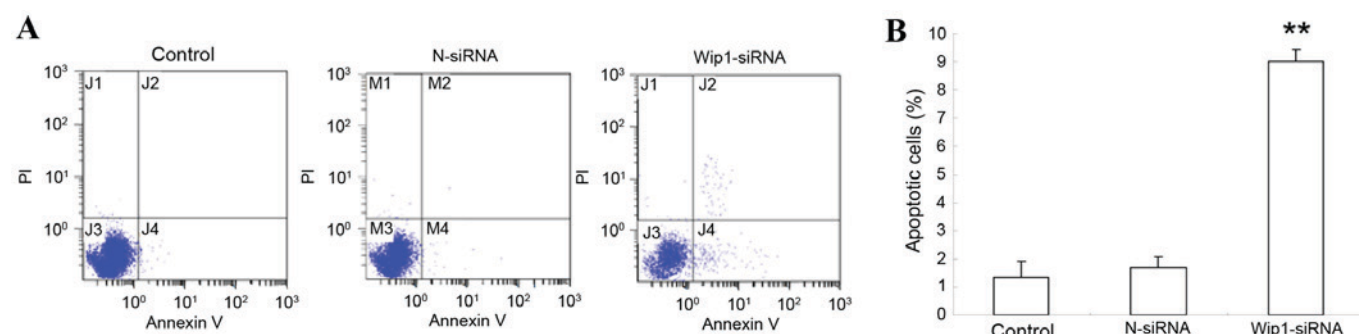


Figure 3. Effect of Wip1 siRNA on SKOV3 cell apoptosis. SKOV3 cells were either left untransfected (Control) or were transfected with either N-siRNA or Wip1-siRNA, following which they were analyzed for apoptosis by Annexin V-FITC/PI staining and flow cytometry. (A) Representative flow cytometry plots. Apoptotic cells were divided into two stages: early apoptotic (Annexin V⁺/PI⁻, J4 quadrant) and late apoptotic (Annexin V⁺/PI⁺, J2 quadrant) cells. (B) Quantification of flow cytometry analysis, reporting results for the early and late apoptotic cells as % of total population analyzed (n=4 repeats). **P<0.01 vs. Control. Wip1, wild-type p53-induced phosphatase 1; siRNA, small interfering RNA; N-siRNA, non-targeting siRNA; FITC, fluorescein isothiocyanate; PI, propidium iodide.

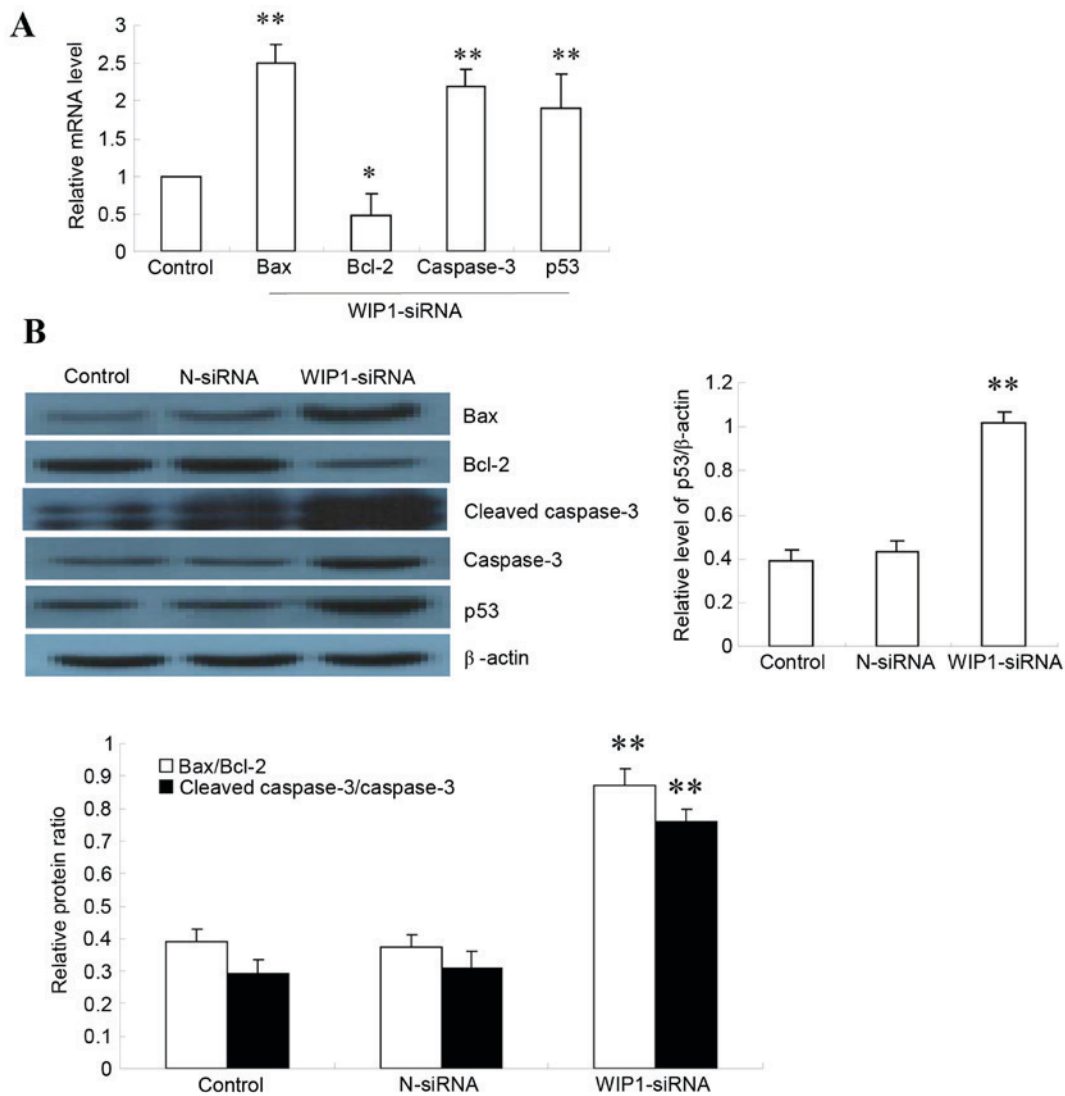


Figure 4. Effect of Wip1 siRNA on expression of apoptotic markers in SKOV3 cells. (A) The mRNA expression levels of Bax, Bcl-2, caspase-3 and p53 were analyzed by reverse transcription-quantitative polymerase chain reaction in SKOV3 cells transfected with Wip1 siRNA and control untransfected cells (n=6). Results are plotted relative to the control. (B) The protein levels of Bax, Bcl-2, cleaved caspase-3, caspase-3 and p53 were analyzed by western blot (n=6 repeats). Results for p53 are plotted relative to β-actin, and for the other proteins as ratios of Bax/Bcl-2 expression and of cleaved caspase-3/caspase-3 expression. *P<0.05 and **P<0.01 vs. Control. Wip1, wild-type p53-induced phosphatase 1; siRNA, small interfering RNA; Bax, BCL2 associated X; Bcl-2, BCL2 apoptosis regulator; p53, tumour protein 53; N-siRNA, non-targeting siRNA.

Discussion

Previous studies have suggested that Wip1 may be important in regulation of treatment response in malignant tumors. Lee *et al* (12) revealed a significant increase in UV-induced apoptosis by Wip1 knockout in mice or human keratinocytes. A Wip1 inhibitor, CCT007093, has also been reported to increase apoptosis in breast cancer cells and skin keratinocytes (12). A previous study by Yang *et al* (13) assessed tumor specimens derived from breast cancer patients for Wip1 mRNA and protein expression using semi-quantitative RT-PCR, immunohistochemistry and western blotting, and demonstrated that expression of Wip1 was significantly higher in breast cancer tissues than in adjacent, normal breast tissues. High expression of Wip1 mRNA and protein may therefore promote breast cancer growth, suggesting that may be a novel target for breast cancer therapy. Thus, regulation of Wip1 levels may also be a potential treatment of ovarian cancer.

Wip1, a serine/threonine protein phosphatase located in human chromosome region 17q23/q24, is encoded by the gene protein phosphatase Mg²⁺/Mn²⁺ dependent 1D (PPM1D). The Wip1 gene is commonly overexpressed or amplified in human cancers, including primary breast cancers (14-16), gastric cancers (17), medulloblastomas (18-20), neuroblastomas (21), and ovarian clear cell adenocarcinomas (5,22). Previous studies have demonstrated a close association between Wip1 genes and apoptosis (23,24). Apoptosis is a process of programmed cell death, regulated by a balance of anti-apoptotic factors (such as Bcl-2) and pro-apoptotic factors (such as p53, Bax and caspase-3). The interaction between pro-apoptotic and anti-apoptotic factors is closely associated with the pathogenesis and development of tumors. Song *et al* (25) reported a significant increase in Wip1 levels following γ-irradiation in prostate cancer cells. In these cells, Wip1 interacted with and dephosphorylated Bax, resulting in Bax translocating to the mitochondria and inhibiting apoptosis. The study also

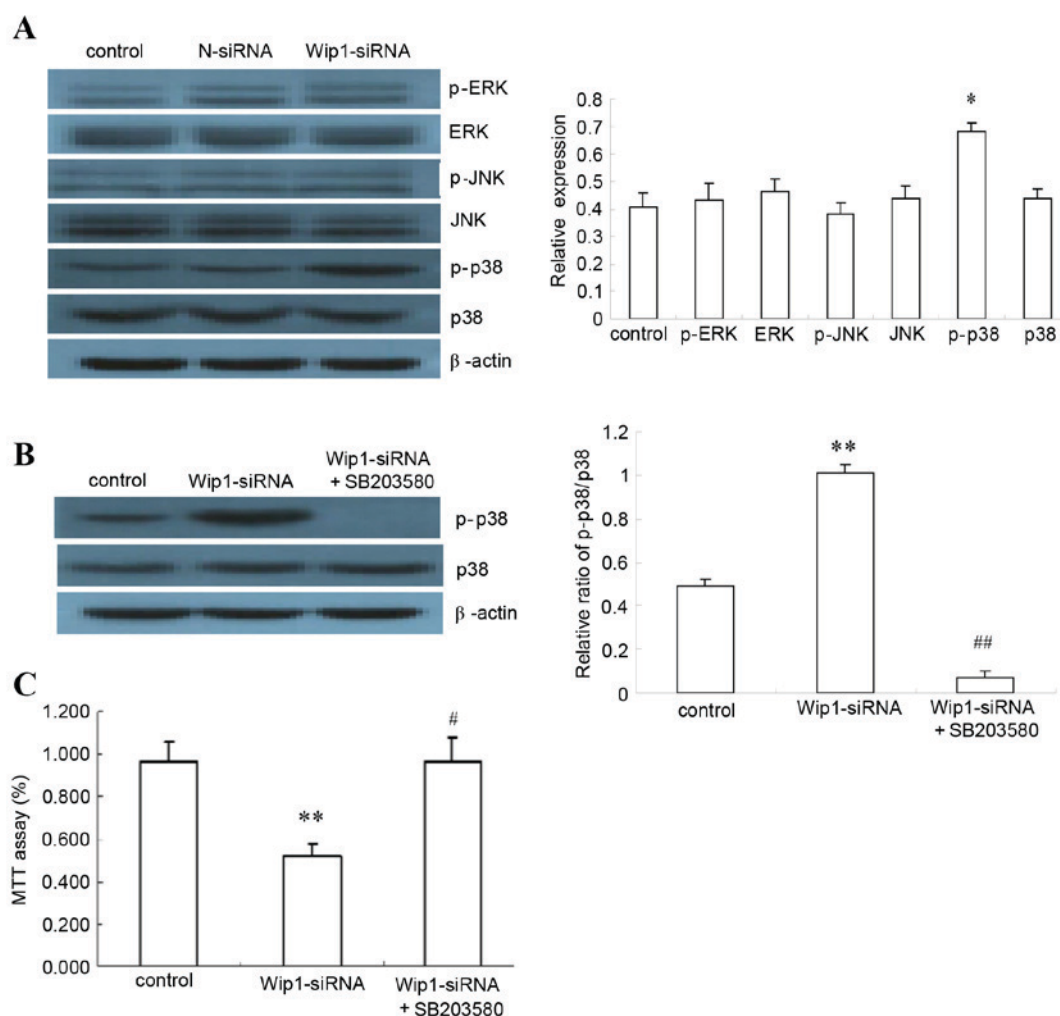


Figure 5. Effects of p38 MAPK inhibitor on Wip1-silenced SKOV3 cells. (A) SKOV3 cells were either left untransfected (Control, SKOV3 cells), or were transfected with N-siRNA or Wip1-siRNA. The protein expression of p-p38 MAPK, p38 MAPK, p-ERK, ERK, p-JNK and JNK were analyzed by western blot (n=6). Representative images and quantification are reported here, as relative level to β -actin. (B) Effects of p38 MAPK and p-p38 MAPK were analyzed by western blot, following transfection with Wip1 siRNA and treatment with p38 MAPK inhibitor SB203580 for 48 h (10 μ M) (n=6). Representative images and quantification are reported here, as the ratio of p-p38/p38. (C) The proliferation rate of SKOV3 cells was analyzed by MTT assay, following transfection with Wip1 siRNA and treatment with p38 MAPK inhibitor SB203580 for 48 h (10 μ M) (n=4). Results are plotted as % ratio relative to control (SKOV3 cells). * P <0.05 and ** P <0.01 vs. Control (SKOV3 cells). # P <0.01 and ## P <0.01 vs. Wip1-siRNA. Wip1, wild-type p53-induced phosphatase 1; siRNA, small interfering RNA; N-siRNA, non-targeting siRNA; p-, phosphorylated; p38 MAPK, p38 mitogen-activated protein kinase; ERK, mitogen-activated protein kinase 1; JNK, mitogen-activated protein kinase 8.

indicated that the apoptosis inhibiting effect of Wip1 could be reverted by its inhibitors. In a Mn neurotoxicity study by Ma *et al* (8), Wip1 expression in the neurons of Mn-exposed rats was progressively decreased, whereas p53 level and Bax/Bcl-XL ratio were elevated progressively, suggesting that Wip1 downregulation might induce apoptotic cell death. Another study also demonstrated a close association between Wip1 expression and apoptosis: In HeLa cells, knockdown of Wip1 significantly decreased growth and colony formation ability and increased apoptosis, as evident also by a significant increase in the protein expression levels of p-p38 MAPK, p53, and p-p53 (26). By contrast, Goloudina *et al* (27) demonstrated that overexpression of Wip1 in p53-negative tumor cells activated the apoptotic pathway through increased Bax/Bcl-XL ratio, thereby leading to cancer cell apoptosis. The present study demonstrated that Wip1 silencing promoted apoptosis in SKOV3 ovarian cancer cells and upregulated caspase-3 expression and Bax/Bcl-2 ratio.

The MAPK signaling pathway is an important intracellular signaling system, and is involved in cell growth, development and apoptosis. The p38 MAPK signaling pathway directly phosphorylates the key regulator p53, which activates cell cycle arrest. In addition, activation of the p38 MAPK pathway is essential for drug-induced tumor cell death. Holmes *et al* (28) have studied the mechanism of vitamin A as a drug for ovarian cancer, and demonstrated that CD437 (a synthetic analog for vitamin A) induces CAOV3 cell apoptosis through the p38 MAPK pathway, and p38 MAPK antagonists could block this effect. Arsenic trioxide (ATO), a potential cancer chemotherapeutic drug, is known to have a curative effect on acute promyelocytic leukemia (29,30). Yoda *et al* (31) confirmed that ATO induces phosphorylation of checkpoint kinase 2 (Chk2) and p38 MAPK and activates the p38 MAPK/p53 pathway, thus leading to apoptosis. Wip1 was demonstrated to suppress DNA damage-induced phosphorylation of Chk2 and p38 MAPK *in vitro*, an effect that was restored by ATO treatment.

Following suppression of Wip1 expression, ATO-induced activation of the p38 MAPK apoptotic pathway was significantly enhanced (31). Wip1 was therefore identified as a direct target of ATO in repression of tumor cell apoptosis. In the present study, silencing Wip1 significantly enhanced p38 MAPK phosphorylation, suggesting that Wip1 acts through the p38 MAPK pathway.

In conclusion, silencing Wip1 promoted apoptosis in SKOV3 ovarian cancer cells, most likely by promoting activation of the p38 MAPK signaling pathway. Therefore, Wip1 may be a potential therapeutic target in ovarian cancer.

Acknowledgements

This study was supported by the Key Project of Hebei Provincial Health and Family Planning Commission Fund (grant no. 20150204).

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