

Effects of maternal acrolein exposure during pregnancy on testicular testosterone production in fetal rats

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Abstract. Acrolein has been reported to have diverse toxic effects on various organs, including the reproductive system. However, little is known regarding the effects of maternal acrolein exposure on testicular steroidogenesis in male offspring. The present study investigated the effects of acrolein on fetal testosterone production and associated genes. Pregnant Sprague-Dawley rats were intraperitoneally injected with vehicle (normal saline) or 1, 2 or 5 mg/kg acrolein from gestational day (GD) 14-20, and fetal testes were examined on GD 21. Fetal body and testicular weights were markedly reduced in pups following exposure to high doses of acrolein (5 mg/kg) in late pregnancy. Notably, *in utero* exposure of 5 mg/kg acrolein significantly decreased the testicular testosterone level and downregulated the expression levels of steroidogenic acute regulatory protein (StAR) and 3 β -hydroxysteroid dehydrogenase (3 β -HSD), whereas the levels of other steroidogenic enzymes, including scavenger receptor class B, cholesterol side-chain cleavage enzyme and steroid 17 α -hydroxylase/17,20 lyase, were unaffected. Furthermore, the 3 β -HSD immunoreactive area in the interstitial region of the fetal testes was reduced at a 5 mg/kg dose, whereas the protein expression levels of 4-hydroxynonenal were dose-dependently increased following maternal exposure to acrolein. mRNA expression levels of insulin-like factor 3, a critical gene involved in testicular descent, were unaltered following maternal acrolein exposure. Taken together, the results of the present study suggested that maternal exposure to high doses of acrolein inhibited fetal testosterone synthesis, and abnormal expression of StAR and 3 β -HSD may be associated with impairment of the steroidogenic capacity.

Introduction

Cryptorchidism and hypospadias in newborn males, and infertility and testicular germ cell cancer in adult males, are common male reproductive system disorders worldwide (1,2). These disorders have been hypothesized to be associated with testicular dysgenesis syndrome (TDS), which originates in male fetal life (3). An important factor contributing to TDS is androgen dysfunction during the masculinization programming window. Masculinization is a pivotal event during reproductive tract development, driven by androgen produced by the fetal testes (4). Specific factors influencing this process, including exposure to certain environmental pollutants, may cause inadequate production of androgen, and ultimately lead to abnormal reproductive development (5-7).

Acrolein, a cyclophosphamide metabolite, is a common environmental and dietary pollutant arising from the combustion of fuels, plastic and fried food, and is additionally a primary component of tobacco (8). Acrolein may be generated endogenously during cellular metabolism by lipid peroxidation and degradation of threonine and polyamines. It has been identified as one of the most harmful non-biological air pollutants in residences in the United States (9,10). Increased acrolein exposure has been reported to be associated with various diseases, including diabetes mellitus, hepatotoxicity, cardiovascular disease and Alzheimer's disease (11-13).

Acrolein has been demonstrated to induce embryo lethality and teratogenicity in cultured rat embryos, and causes reproductive toxicity in a yeast gametogenesis model (14,15). Cyclophosphamide (CP), the precursor of acrolein, is used as a therapeutic agent for the treatment of childhood cancers; however, it may be associated with a high risk of infertility and long-term gonadal toxicity in male survivors (16). Additionally, cytochrome P450 3A 4 and 5, the two key enzymes that metabolize CP into acrolein, are highly expressed in mammalian testes, which may lead to an increased concentration of acrolein in the testes of patients treated with CP (17,18). As acrolein efficiently reaches the testes, it may interfere with steroidogenesis during fetal development, when testosterone production by Leydig cells is critical for normal sexual development. However, little is known regarding the effects of acrolein exposure in maternal rats on the steroidogenic function and sexual development of male offspring. The present study aimed to investigate the dose-dependent effects of

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acrolein on prenatal testosterone production, and the expression levels of the factors involved in testosterone biosynthesis in the fetal rat testes.

Materials and methods

Ethical approval. The present study was performed in compliance with the regulations of the Medical Ethics Committee of Peking University Third Hospital (Beijing, China; ethical approval no. LA2015205).

Chemicals and reagents. Acrolein (10 mg/ml; CAS107-02-8) was purchased from AccuStandard, Inc. (New Haven, CT, USA). Anti-3 β -hydroxysteroid dehydrogenase (3 β -HSD; catalog no. ab150384), anti-steroidogenic acute regulatory protein (StAR; catalog no. ab203193), anti-4-hydroxynonenal (4-HNE; catalog no. ab46545) and anti-glyceraldehyde-3-phosphate dehydrogenase (GAPDH; catalog no. ab181602) rabbit primary antibodies were obtained from Abcam (Cambridge, MA, USA). All other reagents were purchased from Sigma-Aldrich; Merck Millipore (Darmstadt, Germany) or as otherwise specified.

Animals and treatments. Pregnant Sprague-Dawley rats (n=32, with n=8 used for preliminary studies and 24 for formal studies; Vital River Laboratories, Co., Ltd., Beijing, China) were individually housed and maintained under a 12-h light/dark cycle at a controlled temperature (20–25°C) and humidity (50 \pm 5%) for one week prior to the experiments. The study was performed according to the Guide for the Care and Use of Laboratory Animals published by the National Institutes of Health (Bethesda, MD, USA). Acrolein was administered intraperitoneally (i.p.), in accordance with previous studies of maternal exposure (19). Our preliminary study revealed that rats died 4 days following injection of 10 mg/kg acrolein, and as testosterone levels in the fetal testes decreased significantly in the 5 mg/kg group, a dose of 5 mg/kg was used in the present study. A total of 24 pregnant rats at gestational days (GD) 14–20 were divided into four groups (n=6) and injected i.p. with 1, 2 or 5 mg/kg acrolein, with an equal volume of saline serving as the control according to previous studies (20,21). Pregnant rats were anesthetized with an i.p. injection of 50 mg/kg sodium pentobarbital (Sinopharm Chemical Reagent, Co., Ltd., Beijing, China) at GD 21. The fetal rats were harvested by cesarean section, weighed and dissected under a stereomicroscope. Gender was determined by morphology and the gonads. All male fetuses were sacrificed by decapitation and whole blood was collected in a tube with heparin for testosterone analysis. Fetal testes were aseptically removed and stored at -80°C for analysis of testosterone levels, reverse transcription-quantitative polymerase chain reaction (RT-qPCR) and histology.

RT-qPCR. Total RNA was extracted using TRIzol[®] reagent (Invitrogen; Thermo Fisher Scientific, Inc., Waltham, MA, USA). The quantity and quality of the purified RNA was evaluated by spectroscopy. cDNA was synthesized using a RevertAid First Strand cDNA Synthesis Kit (catalog no. K1621; Thermo Fisher Scientific, Inc.), according to the manufacturer's protocol. qPCR was performed using the SYBR[®] Green Master mix (Fermentas; Thermo Fisher Scientific, Inc.) and

the Applied Biosystems 7500 Real-Time PCR system (Applied Biosystems; Thermo Fisher Scientific, Inc.). The cycling conditions were as follows: 1 cycle at 95°C for 5 min, and 40 cycles of amplification at 95°C for 15 sec and 60°C for 1 min. All the samples were run in duplicate using the threshold suggested by the software for the instrument to calculate the quantitation cycle (Cq). Following RT-qPCR, a melting curve analysis was performed to demonstrate the specificity of the PCR products, which revealed that the melting curve for the PCR product of each gene transcript had a single peak (data not shown). To normalize the readings, Cq values from GAPDH served as internal controls for each run, obtaining a Δ Cq value for each gene. Relative alterations in the gene expression data were analyzed using the $2^{-\Delta\Delta Cq}$ method (22). Primer sequences are presented in Table I.

Western blot analysis. Fetal testes were washed twice with ice-cold phosphate buffered saline (PBS) and homogenized with RIPA lysis buffer containing protease inhibitors (Beijing Solarbio Science & Technology Co., Ltd., Beijing, China). The protein concentration was quantified using the Bicinchoninic Acid Protein assay kit (Beijing CoWin Biotech, Co., Ltd., Beijing, China). Proteins (30 μ g) in lysates were separated by 10% SDS-PAGE, transferred to nitrocellulose membranes (Applygen Technologies, Inc., Beijing, China), and blocked with 5% bovine serum albumin (BSA; Sigma-Aldrich; Merck Millipore) at room temperature for 1 h. Membranes were subsequently incubated at 4°C overnight with the following rabbit primary antibodies, all at a 1:1,000 dilution: Anti-StAR, anti-4-HNE or anti-GAPDH, following which corresponding IRDye-conjugated goat anti-rabbit IgG secondary fluorescent antibodies (cat. no. 925-32211; 1:10,000 dilution; LI-COR Biosciences, Lincoln, NE, USA) were added and incubated for 1 h at room temperature in a dark place. Membranes were scanned using the Odyssey[®] CLx Imaging system (LI-COR Biosciences) and the protein expression was quantified using Image Studio[™] Software (LI-COR Biosciences).

Radioimmunoassay (RIA) for testosterone analysis. Serum was separated from blood collected from male offspring by centrifugation at 3000 \times g for 10 min at room temperature and stored at -80°C until required for the testosterone assay. Fetal testes were rinsed with 0.01 M PBS and homogenized in 150 μ l 0.01 M PBS. The homogenate was centrifuged for 10 min at 5000 \times g at 4°C, and the supernatant was collected and stored at -80°C until required. ¹²⁵I-based RIA kits were purchased from the Beijing North Institute of Biological Technology (Beijing, China). Testosterone levels were measured according to the manufacturer's protocol, and expressed as ng/ml.

Histopathology and immunohistochemistry. The testes were immersed in 4% paraformaldehyde for fixation, dehydrated via a graded series of ethanol washes followed by xylene, and embedded in paraffin. Paraffin-embedded tissues were serially sectioned (5- μ m thick), mounted onto glass slides coated with poly-L-lysine, deparaffinized with xylene and rehydrated with graded ethanol. At least two non-serial sections were stained with hematoxylin and eosin (H&E) using standard procedures for morphological analyses. For histological evaluation of apoptosis, DNA fragmentation was examined using

Table I. Sequences of the specific oligonucleotide primers used for polymerase chain reaction amplification.

Target gene	Genbank no.	Product length (bp)	Primer sequence (5'-3')
Scavenger receptor class B	NM_031541	134	F:ctcctgactttctccgtcttcc R:caggatctggaactgcttgt
Steroidogenic acute regulatory protein	NM_031558	125	F:tcaactggaagcaacactctac R:cctgctggcttctctt
Cytochrome P-450 side chain cleavage	NM_017286	156	F:ctggtgacaatggtggataaac R:ccttagggtccaggatgtaaac
3 β -hydroxysteroid dehydrogenase	M38178	141	F:tgttggtgcaggagaaagaa R:ggtactgggcatccagaatc
Cytochrome P-450, family 17	NM_012753	170	F:gcctttgcagatgctgta R:ggcgtggacaggctctat
17 β -hydroxysteroid dehydrogenase	NM_012851	180	F:aggctttaccagggtcttcc R:cagtgtcctctcaatctcttc
Insulin-like factor 3	NM_053680	130	F:gacccagcaagaccttt R:tagggatcctccaaggcaat
GAPDH	NM_017008	155	F:actccattcttccaccttg R:gtccagggtttcttactccttg

F, forward; R, reverse.

the Terminal Deoxynucleotidyl Transferase dUTP Nick-End Labeling assay (TUNEL) Detection kit (cat. no. 11684817910; Roche Diagnostics, Basel, Switzerland) according to the manufacturer's protocol. Slides were incubated with 20 μ g/ml proteinase K for 15 min at room temperature and washed with PBS three times. The slides were subsequently incubated with TUNEL reaction buffer for 60 min at 37°C in a humidified atmosphere in the dark. Following a further wash with PBS, the slides were incubated with an anti-converter-peroxidase secondary antibody, which was part of the TUNEL kit (Roche Diagnostics) for 30 min at 37°C, and the signal was visualized with diaminobenzidine (DAB; OriGene Technologies, Inc., Beijing, China). The number of positive cells was calculated for analysis under a light microscope.

Testicular Leydig cells were identified in 5- μ m thick paraffin sections by immunohistochemistry for 3 β -HSD. Antigen retrieval was performed by microwave oven heating for 5 min in 0.01 M citrate buffer (pH 6.0). The slides were incubated for 10 min in 3% (v/v) hydrogen peroxide in PBS to block endogenous peroxidase activity and subsequently washed with PBS. Following blocking with normal goat serum (Beijing Zhongshan Golden Bridge Biotechnology; OriGene Technologies, Inc., Rockville, MD, USA) diluted 1:5 in PBS containing 5% BSA, the slides were incubated overnight at 4°C with a rabbit polyclonal anti-3 β -HSD antibody (diluted 1:100 in antibody dilutions liquid (cat. no. ZLI-9028; Beijing Zhongshan Golden Bridge Biotechnology; OriGene Technologies, Inc.). Following this, the slides were washed with PBS and incubated with the horseradish peroxidase-conjugated goat anti-rabbit IgG secondary antibody (provided at working dilution; cat. no. PV-6001; Beijing Zhongshan Golden Bridge Biotechnology; OriGene Technologies, Inc.) at 37°C for 30 min. The slides were subsequently stained using a DAB kit, washed with water,

then stained with hematoxylin, dehydrated using sequential concentrations ethanol, being washed for 2 min in each starting with 80%, followed by 95% and finishing with 100% ethanol twice, and placed under cover slips. The density of 3 β -HSD-immunoreactivity was detected as described previously (23). Briefly, photomicrographic digital images were obtained from 3 β -HSD-immunostained sections of the fetal testes, and regions of these photomicrographs were analyzed to measure the density of 3 β -HSD immunoreactivity in the interstitial region of the testes. The surface area of the total interstitial region and the 3 β -HSD-immunoreactive area were subsequently measured using ImageJ software version 1.46 (National Institute of Health). Areas darker than 100 of 256 pixels were determined to be 3 β -HSD-immunoreactive. The positive 3 β -HSD-immunoreactive area was normalized by dividing by the total area of the interstitial region of interest, and was expressed as the 3 β -HSD-immunoreactive area/1-mm² area of the interstitial region of the fetal testes.

Statistical analysis. Statistical analysis was performed using SPSS software version 12.0 (SPSS, Inc., Chicago, IL, USA). The data were examined for normal distribution and homogeneity of variance. Normally distributed and variance homogeneous data were analyzed by one-way analysis of variance. Dunnett's post hoc test was used to compare the values from acrolein-treated animals with the control group. Data are presented as the mean \pm standard deviation. $P < 0.05$ was considered to indicate a statistically significant difference.

Results

Effects of maternal acrolein exposure on pregnant rats and fetus development. To investigate the effects of maternal acrolein exposure during pregnancy on maternal weight gain and

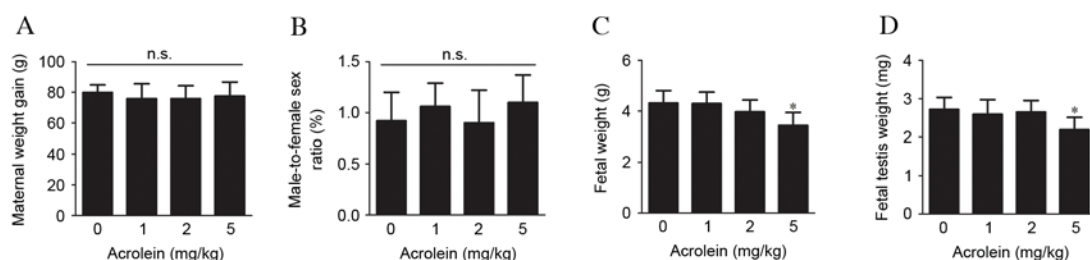


Figure 1. Effects of maternal acrolein exposure on offspring sex and male fetuses. (A) Maternal weight gain, (B) male to female ratio of offspring, and the (C) body and (D) testicular weight of male fetuses. Data are presented as the mean \pm standard deviation (n=32-39). *P<0.01 vs. 0 mg/kg acrolein. n.s., non-significant.

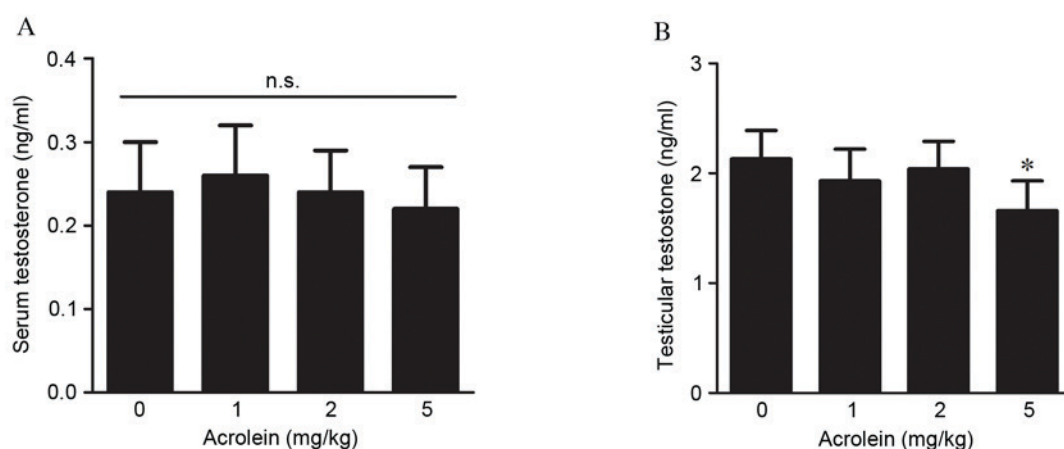


Figure 2. Effects of maternal acrolein exposure on testosterone production. (A) Serum and (B) testicular testosterone concentration. Data are presented as the mean \pm standard deviation (n=12-15). *P<0.01 vs. 0 mg/kg acrolein. n.s., non-significant.

fetus development, pregnant rats were injected i.p. with 0, 1, 2 or 5 mg/kg acrolein daily from GD 14-20. No significant differences in body weight gain in the pregnant rats were observed between the acrolein-treated and control groups (Fig. 1A). Although no significant differences were observed in the male-to-female sex ratio (Fig. 1B), the weight of the pups was markedly reduced when pregnant rats received acrolein at 5 mg/kg compared with the control (P=0.003, Fig. 1C). However, no significant effect on fetal weight was observed in groups administered with 1 or 2 mg/kg acrolein. In addition, the weight of the fetal testes in the group treated with 5 mg/kg acrolein was reduced compared with the control group (P=0.001), while no significant alterations were observed in the other groups (Fig. 1D).

Effects of maternal acrolein exposure on testosterone production in fetal rats. The effects of maternal acrolein exposure during pregnancy on testosterone production in the serum and testes were analyzed. As presented in Fig. 2A, no significant effects of acrolein on serum testosterone were observed in fetal rats. However, compared with the control, the intratesticular testosterone concentration was markedly reduced following treatment with 5 mg/kg acrolein (P<0.001, Fig. 2B). Although testosterone in the groups treated with 1 and 2 mg/kg acrolein decreased compared with the control group, no statistically significant differences were observed.

Effects of maternal acrolein exposure on the expression levels of steroidogenic genes and insulin-like factor 3 (InsI3). A panel of genetic markers associated with testosterone production was used to assess the steroidogenic function in the fetal testes following prenatal exposure to acrolein. No significant differences were observed in the mRNA expression levels of the scavenger receptor class B member 1 (SR-B1; Fig. 3A). However, there was a slight increase in 17 β -HSD mRNA expression levels in the 1 mg/kg group (P=0.032, Fig. 3B). No significant differences in mRNA expression levels were observed in the other steroidogenic enzymes assessed, cholesterol side-chain cleavage enzyme (P450scc; Fig. 3C) and steroid 17 α -hydroxylase/17,20 lyase (P450c17; Fig. 3D). The mRNA expression levels of 3 β -HSD were only reduced in the 5 mg/kg group (P=0.020, Fig. 3E). These alterations were consistent with reduced intratesticular testosterone levels. No significant alterations in the mRNA expression levels of InsI-3, a critical gene involved in testicular descent, were observed in any of the groups treated with acrolein, compared with the control group (Fig. 3F). The mRNA expression levels of the cholesterol transporter StAR in fetal testes were reduced following exposure to 2 and 5 mg/kg acrolein (P=0.009 and P=0.038, respectively), compared with the control group, while no significant differences were observed in the 1 mg/kg group (Fig. 3G). Alterations in protein expression levels of StAR were in accordance with mRNA expression levels (Fig. 3H).

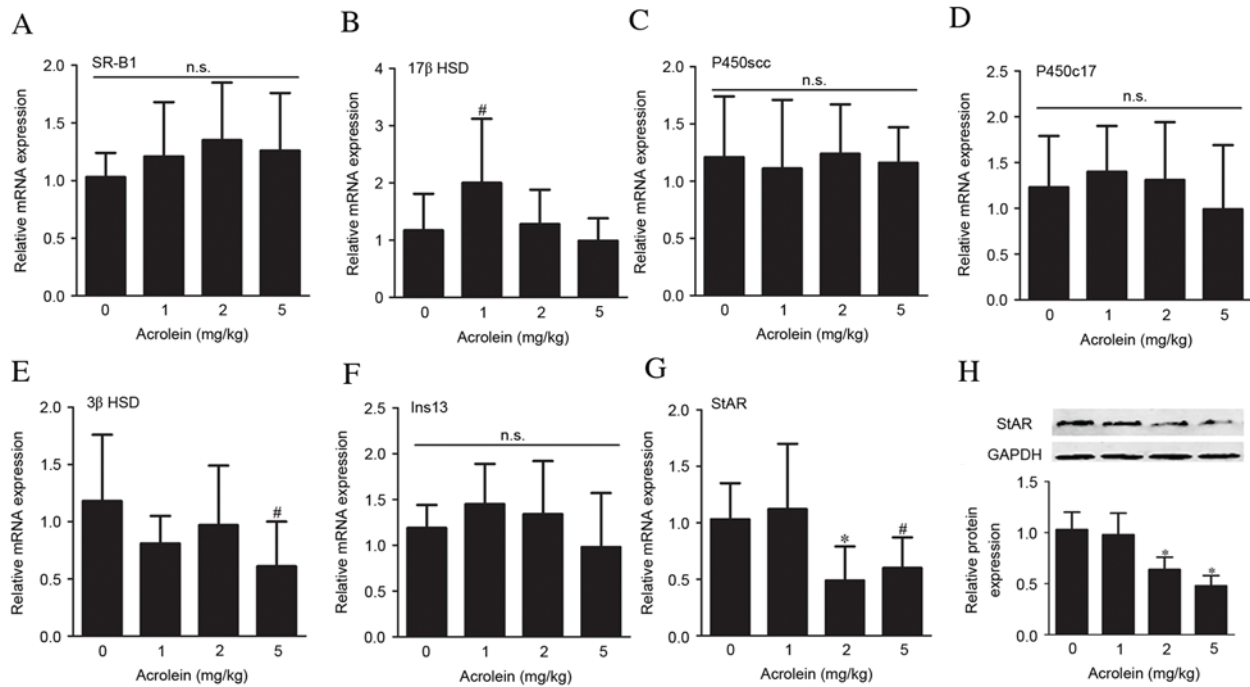


Figure 3. Gene expression analysis. Reverse transcription-quantitative polymerase chain reaction analysis of the expression of (A) SR-B1, (B) 17β HSD, (C) P450scc, (D) P450c17, (E) 3β HSD, (F) Ins13 and (G) StAR. (H) Representative western blot images and analysis of StAR protein expression levels in fetal testes following maternal acrolein exposure. Data are presented as the mean ± standard deviation (n=6-8). *P<0.05 and #P<0.01 vs. 0 mg/kg acrolein. SR-B1, scavenger receptor class B; StAR, steroidogenic acute regulatory protein; P450scc, cholesterol side-chain cleavage enzyme; P450c17, steroid 17 alpha-hydroxylase/17,20 lyase; HSD, hydroxysteroid dehydrogenase; Ins13, insulin-like factor 13; n.s., non-significant.

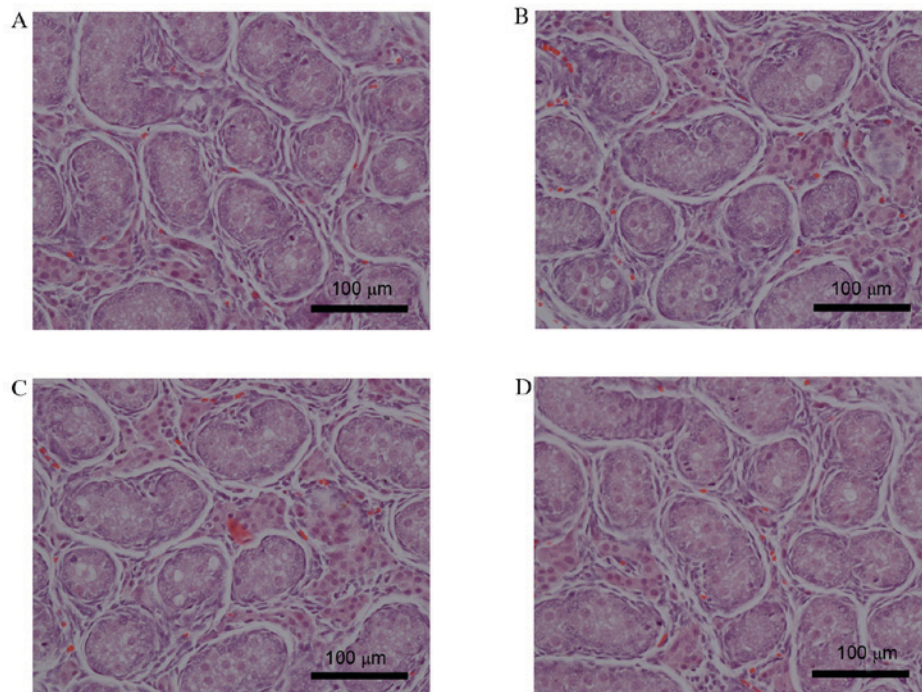


Figure 4. Effects of maternal acrolein exposure on testicular histology. Representative hematoxylin and eosin stained sections from fetal testes following exposure to (A) 0, (B) 1, (C) 2 and (D) 5 mg/kg acrolein. Scale bar=100 μm.

Effects of maternal acrolein exposure on fetal testes histopathology. The effects of maternal acrolein exposure during pregnancy on fetal testes histology were determined by

H&E staining. As presented in Fig. 4, no abnormal morphology was observed in the fetal testes of the acrolein-treated rats compared with the control group. The distribution of Leydig

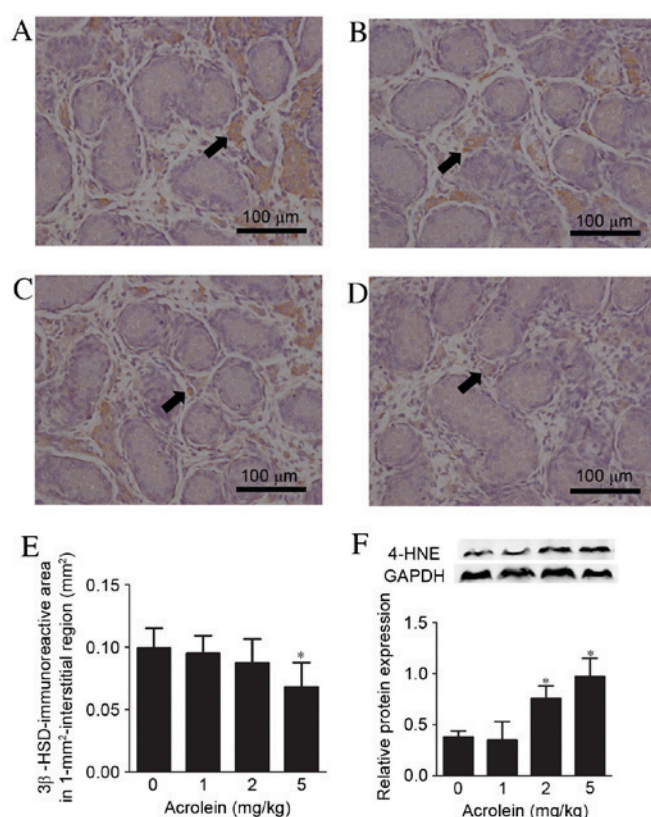


Figure 5. Effects of maternal acrolein exposure on 3β-HSD immunoreactivity in the interstitial region and the protein expression levels of 4-HNE in fetal testes. Representative images of 3β-HSD stained sections from fetal testes following exposure to (A) 0, (B) 1, (C) 2 and (D) 5 mg/kg acrolein. Scale bar=100 μm. Arrows indicate 3β-HSD-positive staining. (E) Quantification of the 3β-HSD immunoreactive area in each group. (F) Representative western blot images and analysis of the protein expression levels of 4-HNE in fetal testes following maternal exposure to acrolein. Data are presented as the mean ± standard deviation (n=6-8). *P<0.01 vs. 0 mg/kg acrolein. 3β-HSD, 3β-hydroxysteroid dehydrogenase; 4-HNE, 4-hydroxynonenal.

cells was identified by immunohistochemical staining for the Leydig cell type-specific marker, 3β-HSD (Fig. 5A-D). The 3β-HSD immunoreactive area in the interstitial region of the fetal testes was reduced following exposure to 5 mg/kg acrolein during pregnancy ($P=0.004$), whereas the immunoreactive areas in the 1 and 2 mg/kg groups were unaltered compared with controls (Fig. 5E). ATUNEL assay revealed that there were no significant differences in cell death in fetal testes between the acrolein exposure and control groups (data not shown). The protein expression levels of 4-HNE, an indicator of oxidative stress, were subsequently examined in fetal testes. As presented in Fig. 5F, the protein expression levels of 4-HNE increased following fetal exposure to acrolein in a dose-dependent manner ($P<0.001$).

Discussion

Acrolein has been reported to have diverse toxic effects in numerous organs, including the reproductive system (11). Various studies have demonstrated that acrolein exposure impairs male germ cells, Sertoli cells and spermatogenesis (14,24,25). In the present study, the dose-associated effects of acrolein exposure during pregnancy on fetal testosterone production and

gene expression were investigated. The results demonstrated that the weight of pups and fetal testes were significantly decreased when prenatally exposed to high doses of acrolein (5 mg/kg). Furthermore, maternal acrolein exposure during pregnancy resulted in a reduction of intratesticular testosterone production and the expression of steroidogenic genes, including StAR and 3β-HSD, and impaired function of Leydig cells. These results indicated that high doses of acrolein exposure *in utero* impair steroidogenic capacity during the fetal period.

Testosterone is primarily synthesized by testicular Leydig cells. Fetal Leydig cells are a distinct population that exhibit a specific origin, structure and capacity for testosterone production, and are regulated by hormones and growth factors (26,27). Leydig cells begin to appear in the interstitial tissue of the developing testes following the formation of testicular cords, reach peak numbers around birth and gradually disappear following postnatal day 7 (28). Previous studies have demonstrated that various environmental toxicants, including phthalate and perfluorooctane sulfonate, may cause impairment of rat fetal Leydig cells and reduction of testosterone production (29,30). Although no direct evidence has indicated that acrolein impairs steroidogenic capacity, males treated with high doses of CP, the precursor of acrolein, for sarcoma during childhood had abnormally elevated gonadotrophin-releasing hormone-stimulated luteinizing hormone levels, suggesting a degree of Leydig cell insufficiency (16). In the present study, a high dose of acrolein (5 mg/kg) significantly decreased fetal testicular testosterone concentration following prenatal exposure, whereas fetal testosterone was unaffected following treatment with low doses of acrolein (1 and 2 mg/kg). Exposure to pollutants, including toluene, during pregnancy is associated with an abnormal 3β-HSD immunoreactive area in fetal testes (23); the present study additionally demonstrated that prenatal acrolein treatment had similar effects. Furthermore, high doses of acrolein exposure (5 mg/kg) during pregnancy were revealed to lead to weight reduction in the male fetus and fetal testes.

The synthesis of testosterone in Leydig cells requires a series of steroidogenic enzymes, including SR-B1, StAR, P450_{scc}, 3β-HSD, P450_{c17} and 17β-HSD (31). The mechanisms underlying reduced fetal testosterone levels following exposure to acrolein were investigated by assessing the mRNA expression levels of these steroidogenic enzymes. SR-B1 is responsible for the transport of high-density lipoprotein (HDL) cholesteryl esters into the cell; the present study demonstrated that the expression levels of SR-B1 were unaffected following exposure to acrolein. StAR is important for the delivery of cholesterol from the outer to the inner mitochondrial membrane in fetal testes (32). In the present study, the mRNA and protein expression levels of this cholesterol transport molecule were markedly decreased following treatment with 2 and 5 mg/kg acrolein. Additionally, previous studies have demonstrated that acrolein impairs cholesterol transport by modification of apolipoprotein A-I and HDL, and that cholesterol is essential for testosterone biosynthesis (33). P450_{scc} is involved in catalyzing the conversion of cholesterol to pregnenolone, which is subsequently transported to the smooth endoplasmic reticulum in the cytoplasm, where 3β-HSD converts it to progesterone, following conversion to

testosterone by P450c17 and 17 β -HSD (34). In the present study, the mRNA expression levels of 3 β -HSD were reduced by exposure to 5 mg/kg acrolein, while the mRNA expression levels of P450scc, P450c17 and 17 β -HSD were not significantly altered, with the exception of a slight increase in 17 β -HSD in the 2 mg/kg group. These results suggested that high doses of acrolein exposure impairs the steroidogenic capacity in testosterone synthesis by reducing StAR and 3 β -HSD expression levels in fetal testes, and that the inhibitory effect on steroidogenesis appears to result from selective alterations of gene expression.

Ins13 is another important factor secreted by fetal Leydig cells, and specifically binds to the leucine-rich repeat-containing G protein-coupled receptor 8 in the gubernaculum to induce scrotal descent of the testes (35). Therefore, interference with the expression of Ins13 may lead to cryptorchidism. The present data suggested that prenatal acrolein exposure does not affect the mRNA expression levels of Ins13. In addition, fetal testes were not histologically damaged following maternal acrolein exposure. Previous studies have demonstrated that acrolein impairs the sertoli cytoskeleton and induces germ cell apoptosis by induction of oxidative stress *in vitro* (24,25), and that it negatively regulates meiosis in a yeast gametogenesis model by inhibiting premeiotic DNA synthesis (14). The present results revealed that prenatal acrolein *in vivo* does not lead to an evident morphological alteration, which is consistent with a study that demonstrated that the mating pattern and fertility of rats were unaffected following acrolein treatment (36). Additionally, Kuwada *et al* (37) revealed that neonatal endocrine disruptors exposure decreases weight and steroidogenesis of juvenile testes, whereas spermatogenesis was restored during puberty. However, the effects of prenatal acrolein exposure on the function of sertoli and germ cells remain to be fully elucidated. Taken together, these studies indicated that maternal acrolein exposure during pregnancy does not cause a distinct pathological impairment in fetal testes, with the exception of reduced steroidogenic capacity at a dose of 5 mg/kg acrolein.

Exposure to environmental pollutants often generates excessive levels of reactive oxygen species (ROS) and induces oxidative stress responses in cells (38). ROS, including oxygen radicals, have been demonstrated to disrupt the balance of the endocrine system and inhibit testicular steroidogenesis (39,40). Acrolein, an unsaturated α , β -aldehyde, reacts with and depletes cell antioxidants, including glutathione, and affects the cellular redox balance (25). 4-HNE is an aldehydic product of lipid peroxidation and has been considered an indicator of oxidative stress-induced cell death (41). Thus, the protein expression levels of 4-HNE in fetal testes were examined following maternal acrolein exposure. It was demonstrated that the protein expression levels of 4-HNE were increased in testes following fetal exposure to acrolein in a dose-dependent manner, indicating that acrolein-induced oxidative stress damage may be implicated in the etiology of fetal testicular pathological alterations.

In conclusion, the results of the present study indicated that prenatal exposure to high doses of acrolein significantly reduced fetal and testes weight and most notably, testicular testosterone production capacity. The abnormal expression

of StAR and 3 β -HSD, and oxidative stress damage, may contribute to the impairment of steroidogenesis. Consequently, these alterations may result in the maldevelopment of the testes and affect masculinization.

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