

microRNA-205 acts as a tumor suppressor and directly targets YAP1 in glioma

TAO JI¹, XIEJUN ZHANG^{1,2} and WEIPING LI^{1,2}

¹Department of Neurosurgery, Shenzhen Second People's Hospital, Clinical College of Anhui Medical University; ²Shenzhen Key Laboratory of Neurosurgery, Shenzhen, Guangdong 518035, P.R. China

Received June 4, 2016; Accepted April 10, 2017

DOI: 10.3892/mmr.2017.6748

Abstract. Glioma is the most common form of primary malignant tumor that occurs in the central nervous system. The underlying molecular mechanism of the carcinogenesis and progression of glioma remains to be elucidated. It is well-established that microRNAs (miRs) are associated with the regulation of glioma initiation and progression, and may represent a novel effective therapeutic strategy for the treatment of glioma. In the present study, the expression, roles and molecular mechanisms of miR-205 in glioma were investigated. The expression levels of miR-205 in glioma tissues, normal brain tissues, human glioma and normal HEB glial cell lines were determined using reverse transcription-quantitative polymerase chain reaction (RT-qPCR). To explore the functional roles of miR-205 in glioma cells, a Cell Counting kit 8 assay, and Transwell migration and invasion assays were employed. The molecular mechanisms underlying the roles of miR-205 in glioma cells were investigated using bioinformatics analysis, a luciferase reporter assay, RT-qPCR and western blot analysis. The results of the present study demonstrated that miR-205 expression was markedly low in glioma tissues and cell lines compared with normal brain tissue and a glial cell line. Upregulation of miR-205 *in vitro* decreased cell viability, migration and invasion in glioma. Further investigation of the potential molecular mechanism demonstrated that the tumor suppressive functions of miR-205 in regulating the proliferation, migration and invasion of glioma cells were mediated by a direct target gene, yes associated protein 1 (YAP1). The results of the present study suggested that miR-205 inhibited glioma growth and metastasis by directly targeting YAP1, and that miR-205 should be investigated as a novel therapeutic target for anti-cancer treatment.

Introduction

Glioma is the most common form of primary malignant tumor that occurs in the central nervous system (1). The underlying molecular mechanism of the carcinogenesis and progression of glioma remains unclear, and may be associated with a number of risk factors, including tumor origin, genetic factors, biochemical environment, ionizing radiation, nitroso compounds, air pollution, lifestyle factors and infection (2). Despite major advancements in the combination of surgery, radiotherapy and chemotherapy, the prognosis for patients with glioma remains poor (3). The average 5-year survival rate of glioma is 4-5%, and the mean survival time following diagnosis is 12-15 months (4,5). This poor prognosis is largely due to the rapid growth and metastasis of the glioma cells, frequently over long distances, through the narrow extracellular spaces in the brain (6,7). Therefore, further research is required to increase understanding of the molecular mechanism underlying the rapid growth and metastasis of glioma, and to investigate more effective therapeutic targets for the treatment of this disease.

The discovery of microRNAs (miRNAs) in the regulation of glioma initiation and progression has provided novel therapeutic strategies for the treatment of glioma (8). miRNAs represent a large family of non-protein-coding, endogenous, single stranded short RNA molecules of 20-23 nucleotides in length (9). miRNAs negatively modulate gene expression through the post-transcriptional silencing of their target mRNAs, which occurs due to complementary binding to the 3'untranslated region (UTR) of their direct target genes, resulting in gene degradation or translational inhibition (10). By negatively regulating the protein expression of their target genes, miRNAs exert adverse effects on a variety of biological processes, including cell proliferation, the cell cycle, apoptosis, migration, invasion, metastasis and differentiation (11).

It is well-established that ~50% of miRNAs are located at fragile sites and cancer susceptibility loci, demonstrating the potential roles of miRNAs in cancer (12). Previous studies have demonstrated that the abnormal expression of miRNAs may be associated with the development and progression of various types of human cancer, and that miRNAs may act as oncogenes or tumor suppressors, depending on the characteristics of their target genes (13-15).

The present study aimed to investigate the expression and roles of miRNA-205 (miR-205) in glioma. It was observed that

Correspondence to: Dr Weiping Li, Department of Neurosurgery, Shenzhen Second People's Hospital, Clinical College of Anhui Medical University, 3002 Sungang West Road, Shenzhen, Guangdong 518035, P.R. China
E-mail: liweiping0755@163.com

Key words: microRNA-205, tumor suppressor, glioma, yes associated protein 1

miR-205 was significantly downregulated in glioma tissues and cell lines compared with normal controls. Overexpression of miR-205 suppressed glioma cell proliferation, migration and invasion. The mechanistic investigation demonstrated that miR-205 regulated glioma tumorigenesis and progression by directly targeting yes associated protein 1 (YAP1). The results of the present study demonstrated the expression pattern and roles of miR-205 in regulating the growth and metastasis of glioma cells, and exhibited a potential therapeutic target for patients with glioma.

Materials and methods

Human samples. The present study was approved by the Ethical Committee of Shenzhen Second People's Hospital (Shenzhen, China) and written informed consent was obtained from all subjects prior to enrollment in the present study. A total of 19 glioma tissue samples were obtained from patients (age, range 31-69 years, median 45 years; 12 male and 7 female subjects) undergoing tumor resection surgery at the Department of Neurosurgery, Shenzhen Second People's Hospital between February 2013 and May 2015. A total of 8 healthy brain tissue samples were obtained from patients with cerebral trauma (age, range 24-57 years, median 36 years; 6 male and 2 female subjects) that underwent brain surgery between February 2013 and May 2015 at the Department of Neurosurgery, Shenzhen Second People's Hospital. None of the patients had received prior treatments, including radiation or chemotherapy. The tissues were immediately frozen and stored at -80°C until use.

Cell lines, culture condition and oligonucleotide transfection. U87, U251, LN229, LN18 human glioma cell lines and HEB normal human glial cell line were purchased from the American Type Culture Collection (ATCC, Manassas, VA, USA). All cell lines were maintained in Dulbecco's modified Eagle's medium (Gibco; Thermo Fisher Scientific, Inc., Waltham, MA, USA) containing 10% fetal bovine serum (FBS; Gibco; Thermo Fisher Scientific, Inc.), in a humidified chamber supplemented with 5% CO₂ at 37°C.

miR-205 mimics (5'-UCCUUC AUU CCA CCG GAG UCU G-3') and negative control (NC) miRNA mimics (5'-UUC UCC GAA CGU GUC ACG UTT-3') were obtained from Shanghai GenePharma Co., Ltd. (Shanghai, China). YAP1 small interfering RNA (siRNA; 5'-GGUGAUACUAUCAACCAATT-3') and NC siRNA (5'-UUCUCCGAACGUGUCACGUTT-3') were synthesized and purified by Guangzhou RiboBio Co., Ltd. (Guangzhou, China). According to the manufacturer's protocol, cells were transfected with miRNA mimics or siRNA using Lipofectamine 2000 (Invitrogen; Thermo Fisher Scientific, Inc.). Following transfection for 24 h, reverse transcription-quantitative polymerase chain reaction (RT-qPCR) and a Cell Counting kit 8 (CCK8) assay were performed. Transwell migration and invasion assays, and western blot analysis were performed at 48 and 72 h post-transfection, respectively.

RT-qPCR. Total RNA was extracted from cells using TRIzol® reagent (Invitrogen; Thermo Fisher Scientific, Inc.), according to the manufacturer's protocol. Total RNA was reverse-transcribed into cDNA using a First-Strand cDNA Synthesis kit (Invitrogen; Thermo Fisher Scientific, Inc.).

qPCR was performed using the SYBR Green PCR mixture (Applied Biosystems; Thermo Fisher Scientific, Inc.) with an ABI Prism 7500 Sequence Detection System (Applied Biosystems; Thermo Fisher Scientific, Inc.). Thermocycling conditions were as follows: Initial 1 step at 95°C for 10 min, followed by 40 cycles at 95°C for 15 sec and at 60°C for 1 min. U6 and GAPDH were used as internal standards to normalize the expression of miR-205 and YAP1 mRNA, respectively. The primer sequences are presented in Table I. Relative gene expression was calculated using the 2^{-ΔΔCq} method (16).

CCK8 assay. Cell proliferation was evaluated by performing a CCK-8 assay (Dojindo Molecular Technologies, Inc., Kumamoto, Japan). Transfected cells were harvested, counted and seeded into a 96-well plate at a density of 2,000 cells/well. The CCK-8 assay was performed at 24, 48, 72, and 96 h following incubation. At each time point, 10 μl CCK-8 solution was added to each well and incubated at 37°C for a further 2 h. The absorbance was detected at 450 nm using a microplate reader (Bio-Rad Laboratories, Inc., Hercules, CA, USA). Each assay was performed in triplicate.

Transwell migration and invasion assays. Cellular migration and invasion was examined using a Transwell migration and invasion assay with Transwell chambers (8 mm pores; Costar; Corning Incorporated, Corning, NY, USA). For the Transwell invasion assay, the Transwell chambers were coated with Matrigel (BD Biosciences, San Jose, CA, USA). For the migration and invasion assays, transfected cells were harvested, counted and re-suspended. A total of 5×10⁴ cells in 200 μl FBS-free culture medium was added to the upper chamber, while 500 μl culture medium supplemented with 20% FBS was added to the lower chamber. Following incubation for 48 h at 37°C, cells that had migrated or invaded through the Transwell chamber were fixed with 100% methanol at room temperature for 10 min, stained with 0.1% crystal violet at room temperature for 10 min, and counted in five random areas of each Transwell chamber using an inverted microscope (Olympus Corporation, Tokyo, Japan).

Bioinformatics analysis and luciferase reporter assay. The direct target genes of miR-205 were analyzed using TargetScan (targetscan.org) and miRanda (microrna.org).

Luciferase reporter plasmids, psiCHECK2-YAP1-3'UTR wild type (Wt) and psiCHECK2-YAP1-3'UTR mutant (Mut), were synthesized by Shanghai GenePharma Co., Ltd. HEK293T cells (ATCC) were seeded in 24-well plates at 50-60% confluence. Following incubation overnight, cells were transfected with psiCHECK2-YAP1-3'UTR Wt or psiCHECK2-YAP1-3'UTR Mut, and miR-205 mimics or NC, using Lipofectamine 2000. Following incubation for 48 h at 37°C, cells were collected, and firefly and *Renilla* luciferase activities were detected using a Dual-Luciferase Reporter Assay System (Promega Corporation, Madison, WI, USA), according to the manufacturer's protocol.

Western blot analysis. Transfected cells were lysed with radioimmunoprecipitation assay lysis buffer (Beyotime Institute of Biotechnology, Haimen, China), supplemented with a protease inhibitor cocktail (Roche Applied Science,

Table I. Reverse transcription-quantitative polymerase chain reaction primers.

Gene	Forward, 5'-3'	Reverse, 5'-3'
miR-205	GCTCCTTCATTCCACCGG	CAGTGCAGGGTCCGAGGT
U6	CTCGCTTCGGCAGCACATATACT	ACGCTTCACGAATTTGCGTGTC
YAP1	TATCAATCCCAGCACAG	GGAATGGCTTCAAGGTAG
GADPH	CATCACCATCTTCCAGGAGCG	TGACCTTGCCCACAGCCTTG

miR, microRNA; YAP1, yes associated protein 1.

Penzberg, Germany). The bicinchoninic acid protein assay kit (Pierce; Thermo Fisher Scientific, Inc.) was used to detect the concentration of protein. Equal amounts of protein (30 μ g) were subjected to 10% SDS-PAGE, transferred to polyvinylidene difluoride membranes (EMD Millipore, Billerica, MA, USA), and blocked in 5% skimmed milk in TBS with 0.1% Tween-20 (TBST) at room temperature for 1 h. Subsequently, the membranes were incubated with the primary antibodies mouse anti-human monoclonal YAP1 (1:1,000; cat no. ab124474; Abcam, Cambridge, UK), and mouse anti-human monoclonal GADPH antibody (1:1,000; cat no. ab125247; Abcam), at 4°C overnight. Following washing three times with TBST, the membranes were incubated with the corresponding horseradish peroxidase-conjugated secondary antibody (1:5,000; cat no. ab6789; Abcam) for 1 h at room temperature. Protein bands were developed using enhanced chemiluminescence reagents (EMD Millipore). GADPH was used as an internal control for YAP1.

Statistical analysis. Data are expressed as the mean \pm standard deviation of 3 independent experiments. The statistical significance of the differences between groups was assessed using a two-tailed Student's *t*-test for pair-wise comparisons, or a one-way analysis of variance followed by a post hoc Student-Newman-Keuls test for multiple comparisons. Statistical analysis was performed using SPSS software version 16.0 (SPSS, Inc., Chicago, IL, USA). $P < 0.05$ was considered to indicate a statistically significant difference.

Results

Expression of miR-205 in glioma tissues and cell lines. In order to determine whether miR-205 is involved in the tumorigenesis and progression of glioma, the expression of miR-205 in glioma tissues was analyzed using RT-qPCR. It was observed that miR-205 expression was significantly reduced in glioma tissues compared with healthy brain tissues (Fig. 1A; $P < 0.05$). The expression levels of miR-205 were additionally measured in glioma cell lines (U87, U251, LN229, LN18) and a normal human glial cell line (HEB). Compared with HEB, the expression of miR-205 was significantly reduced in glioma cell lines (Fig. 1B; $P < 0.05$).

Restoration of miR-205 expression inhibits cell proliferation, migration and invasion in glioma. In order to further investigate the biological roles of miR-205 in glioma cells, U87 and LN229 cells, that express a relatively decreased level of

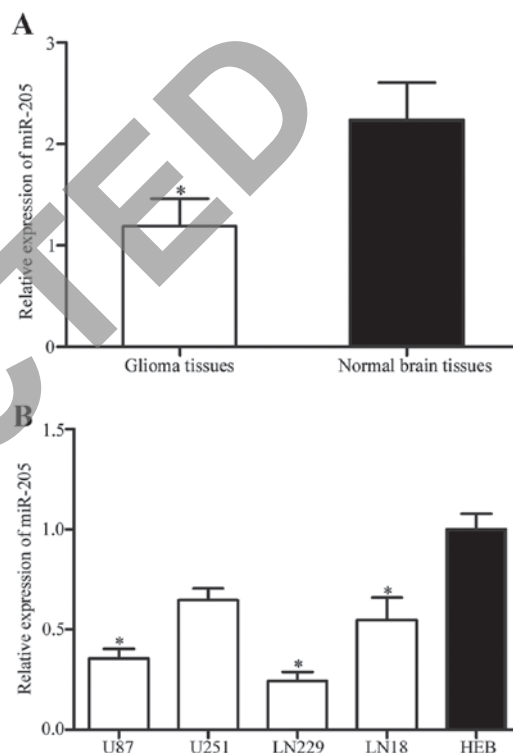


Figure 1. miR-205 is downregulated in glioma tissues and cell lines. (A) miR-205 relative expression levels in glioma tissues and normal brain tissues were measured using RT-qPCR. Data are expressed as the mean \pm standard deviation. * $P < 0.05$ vs. normal brain tissues. (B) miR-205 expression levels in glioma cell lines (U87, U251, LN229, LN18) and the normal human glial cell line (HEB) were assessed using RT-qPCR. Data are expressed as the mean \pm standard deviation. * $P < 0.05$ vs. HEB cells. miR, microRNA; RT-qPCR, reverse transcription-quantitative polymerase chain reaction.

miR-205 compared with normal glial cells, were transfected with miR-205 or NC mimics. Following transfection, RT-qPCR analysis was performed to measure miR-205 expression. As presented in Fig. 2A, miR-205 was significantly upregulated in U87 and LN229 cells transfected with miR-205 mimics compared with the NC groups ($P < 0.05$).

The results of the CCK-8 assay demonstrated that treatment with miR-205 mimics reduced the viability of U87 and LN229 cells (Fig. 2B; $P < 0.05$). Transwell migration and invasion assays were used to investigate the effect of miR-205 on the migratory and invasive behavior of glioma cells. The results demonstrated that upregulation of miR-205 suppressed the migratory and invasive capabilities of U87 and LN229 cells

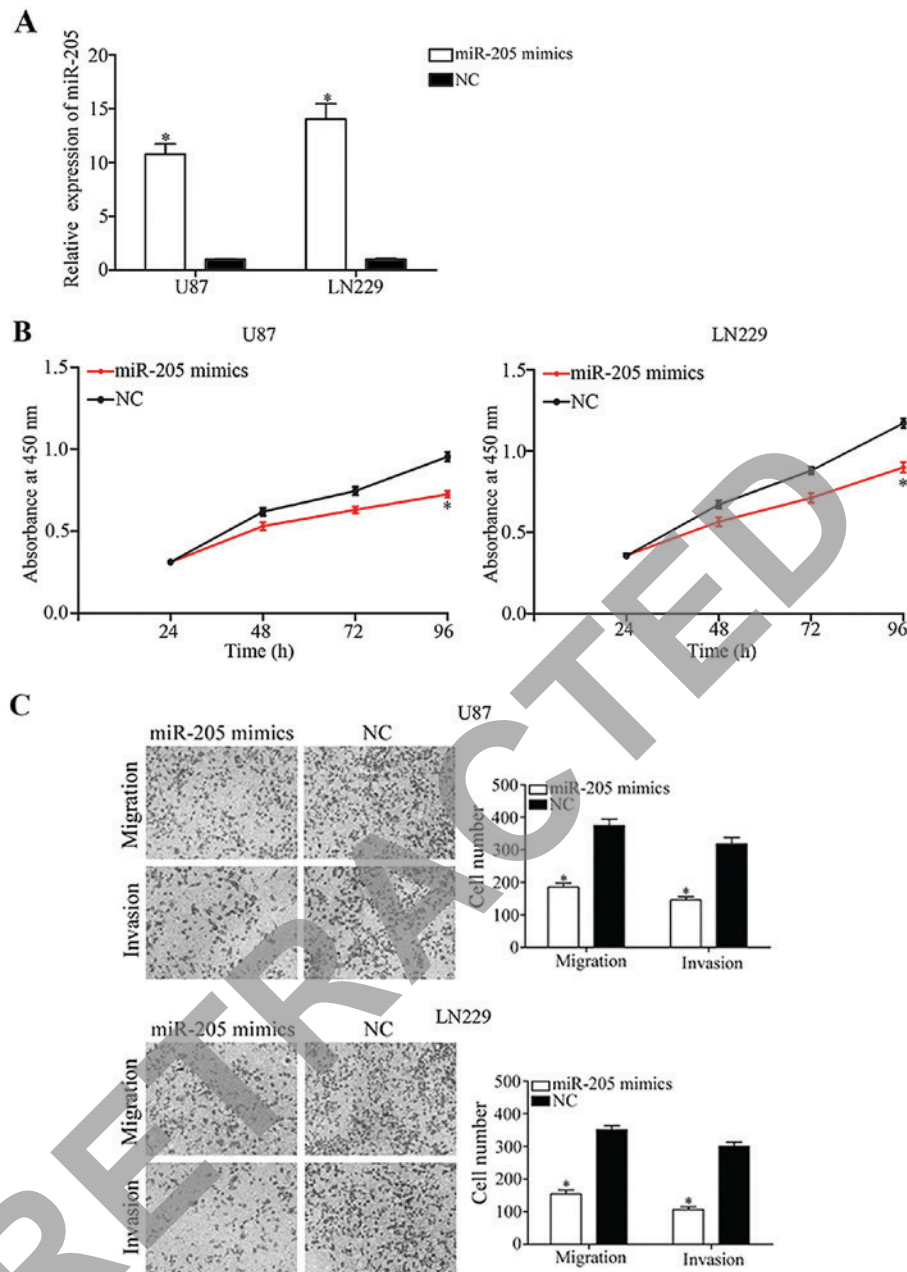


Figure 2. Effects of miR-205 overexpression on glioma cells. (A) The relative expression levels of miR-205 in U87 and LN229 cell lines were detected by reverse transcription-quantitative polymerase chain reaction following transfection with miR-205 mimics or NC. (B) Cell proliferation was determined using a Cell Counting kit 8 assay in U87 and LN229 cells transfected with miR-205 mimics or NC. (C) The migratory and invasive capabilities of U87 and LN229 cells transfected with miR-205 mimics or NC were determined using Transwell migration and invasion assays. Data are expressed as the mean \pm standard deviation. * $P < 0.05$ vs. NC. miR, microRNA; NC, negative control.

(Fig. 2C; $P < 0.05$). The results of the present study suggested that miR-205 may function as a negative regulator of glioma progression.

YAP1 is a direct target of miR-205. In order to examine the molecular mechanism underlying the tumor suppressive roles of miR-205 in glioma, bioinformatics analysis was performed using TargetScan and miRanda. The analysis demonstrated that YAP1 was a putative target gene of miR-205 (Fig. 3A). Therefore, a luciferase reporter assay was performed to investigate whether miR-205 directly interacted with the 3'UTR of YAP1. The results of the present study demonstrated that

miR-205 significantly decreased the luciferase activity of psiCHECK2-YAP1-3'UTR Wt (Fig. 3B; $P < 0.05$). However, upregulation of miR-205 failed to affect the luciferase activities of psiCHECK2-YAP1-3'UTR Mut. In order to determine if miR-205 decreased YAP1 expression, RT-qPCR analysis and western blotting were performed, and the results demonstrated that restoring miR-205 expression significantly decreased YAP1 expression at the mRNA (Fig. 3C; $P < 0.05$) and protein levels (Fig. 3D; $P < 0.05$), in U87 and LN229 cells. The results of the present study demonstrated that miR-205 directly targeted the 3'UTR of YAP1 and negatively regulated its expression.

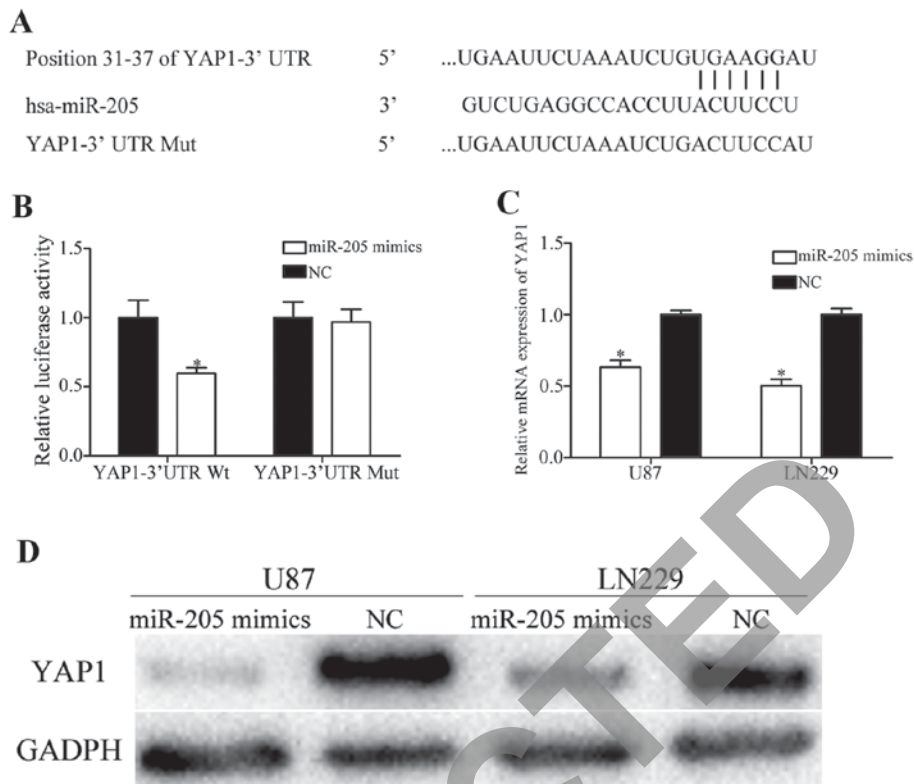


Figure 3. YAP1 is a direct target of miR-205. (A) Bioinformatics analysis demonstrated that YAP1 was a potential target for miR-205. (B) A luciferase reporter assay demonstrated that miR-205 suppressed the luciferase activities of psiCHECK2-YAP1-3'UTR Wt, and not the psiCHECK2-YAP1-3'UTR Mut luciferase reporter plasmid. (C) YAP1 mRNA expression was detected in U87 and LN229 cells transfected with miR-205 mimics or NC using reverse transcription-quantitative polymerase chain reaction. Data are expressed as the mean \pm standard deviation. * $P < 0.05$ vs. NC. (D) YAP1 protein levels were determined in U87 and LN229 cells transfected with miR-205 mimics or NC using western blotting. YAP1, yes associated protein 1; miR, microRNA; UTR, untranslated region; Mut, mutated; Wt, wild type; NC, negative control.

YAP1 knockdown mimics the tumor suppressive roles of miR-205 in glioma. In the present study, YAP1 was identified to be a direct target of miR-205 in glioma. Therefore, it was subsequently investigated whether YAP1 acts as a downstream effector of miR-205, mediating its tumor suppressive functions. U87 and LN229 cells were transfected with YAP1 or NC siRNA. Following transfection, RT-qPCR analysis and western blotting was performed to measure YAP1 mRNA and protein expression, respectively. The results demonstrated that YAP1 was significantly downregulated at the mRNA (Fig. 4A; $P < 0.05$) and protein levels (Fig. 4B; $P < 0.05$), in YAP1 siRNA-transfected U87 and LN229 cells.

Functional assays demonstrated that YAP1 knockdown decreased U87 and LN229 cellular viability (Fig. 4C; $P < 0.05$), migration and invasion (Fig. 4D; $P < 0.05$) *in vitro*, which was consistent with the effects induced by miR-205 overexpression. The results of the present study demonstrated that YAP1 may be a downstream effector of miR-205 in glioma.

Discussion

In the present study, it was observed that miR-205 was downregulated in glioma. In addition, YAP1 was identified to be a direct target of miR-205, and it was demonstrated that the overexpression of miR-205 inhibited glioma cell proliferation, migration and invasion by directly targeting YAP1. The results of the present study suggested that low miR-205

expression may promote glioma initiation and progression, and that miR-205 may be investigated as a therapeutic target for patients with glioma. To the best of our knowledge, the present study was the first to demonstrate the role of miR-205 in glioma tumorigenesis.

Dysregulation of miR-205 has been observed in various types of human cancer. Niu *et al* (17) reported that the expression levels of miR-205 were decreased in renal cell carcinoma tissues and cell lines, compared with matched non-tumor tissues and HK-2 cells, respectively. The downregulation of miR-205 was demonstrated in breast cancer (18), prostate cancer (19) and osteosarcoma (20). However, in non-small cell lung cancer, miR-205 was demonstrated to be significantly upregulated in tumor tissues and in cell lines (21). Increased expression of miR-205 was additionally observed in a variety of types of human cancer, including nasopharyngeal carcinoma (22), laryngeal squamous cell carcinoma (23), endometrial cancer (24) and ovarian cancer (17). These previous conflicting results demonstrated that the expression of miR-205 may be altered in different tumors and may exhibit tissue specificity.

The abnormal expression of miR-205 was previously demonstrated to be associated with clinicopathological features and prognosis in cancer. In cervical cancer, miR-205 was upregulated, and increased miR-205 expression was associated with poor tumor differentiation, lymph node metastasis and increased tumor stage (25). In addition, Kaplan-Meier survival analysis demonstrated that patients with cervical cancer, with increased

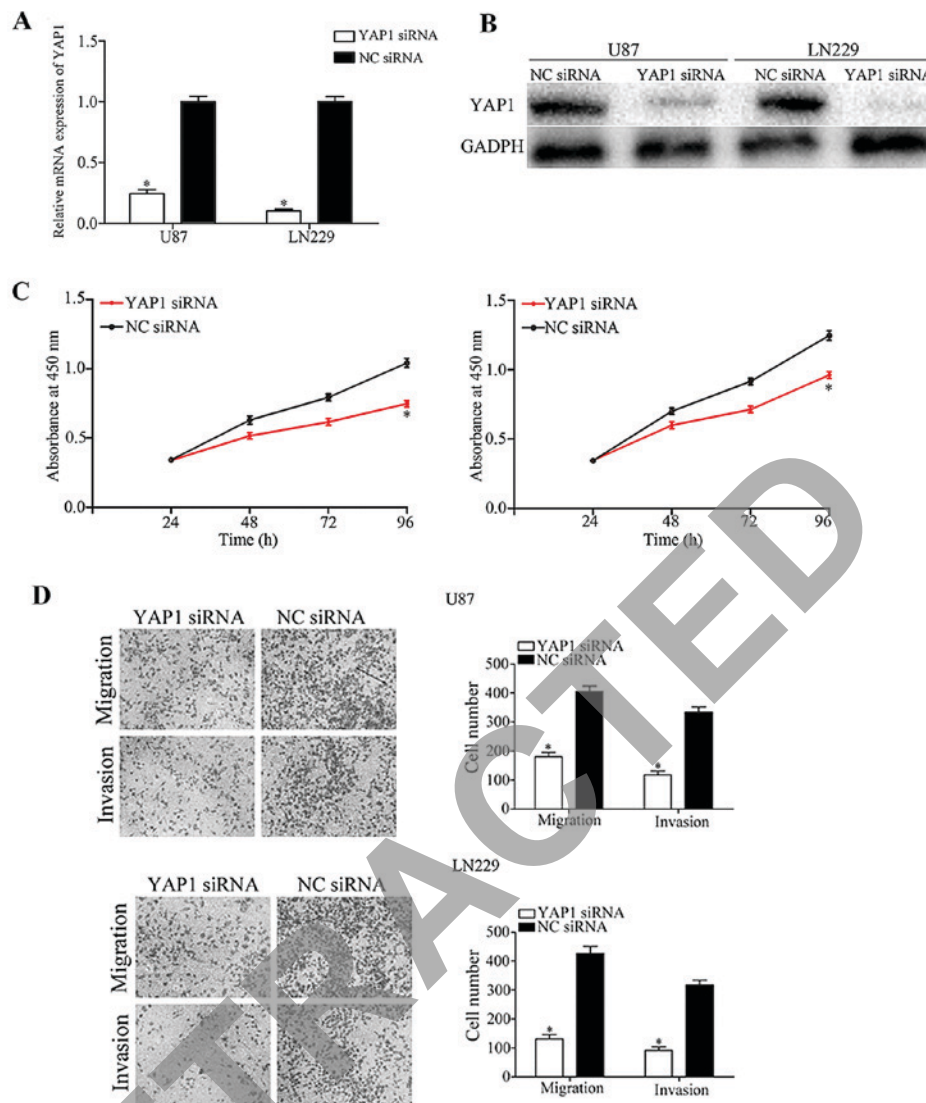


Figure 4. Effects of YAP1 knockdown on glioma cells. (A) YAP1 mRNA expression in U87 and LN229 cell lines was detected using reverse transcription-quantitative polymerase chain reaction, following transfection with YAP1 siRNA or NC siRNA. (B) YAP1 protein expression in U87 and LN229 cell lines was detected using western blotting, following transfection with YAP1 siRNA or NC siRNA. (C) Cell proliferation was determined using a Cell Counting kit 8 assay in U87 and LN229 cells transfected with YAP1 siRNA or NC siRNA. (D) The migratory and invasive capabilities of U87 and LN229 cells transfected with YAP1 siRNA or NC siRNA were determined using Transwell migration and invasion assays. Data are expressed as the mean \pm standard deviation. * $P < 0.05$ vs. NC. YAP1, yes associated protein 1; siRNA, small interfering RNA; NC, negative control.

miR-205 expression, tended to exhibit decreased overall survival times (25). Using multivariate Cox regression analysis, miR-205 was identified to be an independent prognostic marker in patients with cervical cancer (25). Hou *et al* (26) demonstrated that reduced miR-205 expression was associated with tumor grade and Karnofsky performance status score in patients with glioma. Survival analysis demonstrated that patients with glioma exhibiting decreased miR-205 expression presented with poorer overall survival and poorer disease-free survival compared with those exhibiting increased miR-205 expression. Multivariate Cox regression analysis further demonstrated that miR-205 expression was an independent prognostic indicator of the overall survival of patients with glioma (26). These previous studies suggested that miR-205 may be investigated as a useful prognostic marker in human cancer.

A number of studies have reported that miR-205 is involved in the malignant phenotype of cancers. Xu *et al* (27)

and Yin *et al* (28) observed that upregulation of miR-205 suppressed the proliferation, migration, invasion and epithelial-mesenchymal transition (EMT) of gastric cancer cells. In breast cancer, the restoration of miR-205 expression inhibited cell growth, colony-formation capacity, and motility, and promoted radiosensitivity (18,29,30). Mao *et al* (22) reported that miR-205 acted as an oncogene in nasopharyngeal carcinoma, by promoting tumor cell proliferation, migration and invasion. A study by Yang *et al* (20) demonstrated that miR-205 overexpression decreased the capacity for cell proliferation, invasion and migration, and enhanced G0/G1 growth arrest and apoptosis, in osteosarcoma cells. Lei *et al* (21) demonstrated that enforced miR-205 expression in non-small cell lung cancer promoted cell growth, metastasis, and improved chemoresistance. In the present study, miR-205 overexpression inhibited cell proliferation, migration and invasion. The results of the present study suggested that miR-205 was involved

in tumor progression, and provided a potential therapeutic strategy for cancer treatment in the future.

A number of target genes of miR-205 have been previously identified, including zinc finger E-box binding homeobox 1 (27) in gastric cancer, tumor protein p53 inducible nuclear protein 1 in prostate cancer (31), transforming growth factor- α in osteosarcoma (20), cyclic AMP responsive element binding protein 1 in colorectal cancer (32), cyclin dependent kinase 1 associated protein 1 in laryngeal squamous cell carcinoma (23) and angiominin in breast cancer (30). However, no target of miR-205 has been identified in glioma. The present study demonstrated that the tumor suppressive roles of miR-205 in glioma cells were potentially mediated via negative regulation of the expression of the novel identified target, YAP1. The YAP1 gene is located on chromosome 11q22.1 (33). A previous study demonstrated that YAP1 was upregulated or mutated in the majority of pancreatic cancer cases (34). In glioma, YAP1 was demonstrated to be upregulated in infiltrating astrocytomas and oligodendrogliomas, and YAP1 mRNA expression levels were associated with aggressive molecular subsets of glioma (35). YAP1, a member of the Hippo signaling pathway, may negatively regulate cell proliferation, invasion, EMT, metastasis, differentiation and survival (36). Therefore, YAP1 may be investigated as an effective therapeutic target for patients with glioma, by identifying the disease at an earlier stage (37). In the present study, it was observed that miR-205 targeted YAP1, and inhibited glioma cell growth and metastasis. miR-205/YAP1-based targeted therapy may be a promising therapeutic method for glioma.

In conclusion, the results of the present study provided evidence that miR-205 is involved in glioma tumorigenesis and progression. It was additionally demonstrated that miR-205 acts as a tumor suppressor in glioma, potentially sue to negative regulation of the novel target, YAP1.

References

- Di Stefano AL, Enciso-Mora V, Marie Y, Desestret V, Labussière M, Boisselier B, Mokhtari K, Idbaih A, Hoang-Xuan K, Delattre JY, *et al*: Association between glioma susceptibility loci and tumour pathology defines specific molecular etiologies. *Neuro Oncol* 15: 542-547, 2013.
- Zhu GY, Shi BZ and Li Y: FoxM1 regulates Sirt1 expression in glioma cells. *Eur Rev Med Pharmacol Sci* 18: 205-211, 2014.
- Marumoto T and Saya H: Molecular biology of glioma. *Adv Exp Med Biol* 746: 2-11, 2012.
- Schwartzbaum JA, Fisher JL, Aldape KD and Wrensch M: Epidemiology and molecular pathology of glioma. *Nat Clin Pract Neurol* 2: 494-516, 2006.
- Stupp R, Mason WP, van den Bent MJ, Weller M, Fisher B, Taphoorn MJ, Belanger K, Brandes AA, Marosi C, Bogdahn U, *et al*: Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. *N Engl J Med* 352: 987-996, 2005.
- Wiedemeyer R, Brennan C, Heffernan TP, Xiao Y, Mahoney J, Protopopov A, Zheng H, Bignell G, Furnari F, Cavenee WK, *et al*: Feedback circuit among INK4 tumor suppressors constrains human glioblastoma development. *Cancer Cell* 13: 355-364, 2008.
- Lefranc F, Rynkowski M, DeWitte O and Kiss R: Present and potential future adjuvant issues in high-grade astrocytic glioma treatment. *Adv Tech Stand Neurosurg* 34: 3-35, 2009.
- Jiang Y, Yin L, Jing H and Zhang H: MicroRNA-219-5p exerts tumor suppressor function by targeting ROBO1 in glioblastoma. *Tumour Biol* 36: 8943-8951, 2015.
- Feng YA, Liu TE and Wu Y: microRNA-182 inhibits the proliferation and migration of glioma cells through the induction of neuritin expression. *Oncol Lett* 10: 1197-1203, 2015.
- Lu J, Getz G, Miska EA, Alvarez-Saavedra E, Lamb J, Peck D, Sweet-Cordero A, Ebert BL, Mak RH, Ferrando AA, *et al*: MicroRNA expression profiles classify human cancers. *Nature* 435: 834-838, 2005.
- Yates LA, Norbury CJ and Gilbert RJ: The long and short of microRNA. *Cell* 153: 516-519, 2013.
- Calin GA, Sevignani C, Dumitru CD, Hyslop T, Noch E, Yendamuri S, Shimizu M, Rattan S, Bullrich F, Negrini M and Croce CM: Human microRNA genes are frequently located at fragile sites and genomic regions involved in cancers. *Proc Natl Acad Sci USA* 101: 2999-3004, 2004.
- Zhou Y, Wu D, Tao J, Qu P, Zhou Z and Hou J: MicroRNA-133 inhibits cell proliferation, migration and invasion by targeting epidermal growth factor receptor and its downstream effector proteins in bladder cancer. *Scand J Urol* 47: 423-432, 2013.
- Jiang H, Hua D, Zhang J, Lan Q, Huang Q, Yoon JG, Han X, Li L, Foltz G, Zheng S and Lin B: MicroRNA-127-3p promotes glioblastoma cell migration and invasion by targeting the tumor-suppressor gene SEPT7. *Oncol Rep* 31: 2261-2269, 2014.
- Lee HK, Bier A, Cazacu S, Finniss S, Xiang C, Twito H, Poisson LM, Mikkelsen T, Slavin S, Jacoby E, *et al*: MicroRNA-145 is downregulated in glial tumors and regulates glioma cell migration by targeting connective tissue growth factor. *PLoS One* 8: e54652, 2013.
- Livak KJ and Schmittgen TD: Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) Method. *Methods* 25: 402-408, 2001.
- Niu K, Shen W, Zhang Y, Zhao Y and Lu Y: MiR-205 promotes motility of ovarian cancer cells via targeting ZEB1. *Gene* 574: 330-336, 2015.
- Wang Z, Liao H, Deng Z, Yang P, Du N, Zhanng Y and Ren H: miRNA-205 affects infiltration and metastasis of breast cancer. *Biochem Biophys Res Commun* 441: 139-143, 2013.
- Verdoordt B, Neid M, Vogt M, Kuhn V, Liffers ST, Palisaar RJ, Noldus J, Tannapfel A and Mirmohammadsadegh A: MicroRNA-205, a novel regulator of the anti-apoptotic protein Bcl2, is downregulated in prostate cancer. *Int J Oncol* 43: 307-314, 2013.
- Yang G, Zhang P, Lv A, Liu Y and Wang G: MiR-205 functions as a tumor suppressor via targeting TGF- α in osteosarcoma. *Exp Mol Pathol* 100: 160-166, 2016.
- Lei L, Huang Y and Gong W: miR-205 promotes the growth, metastasis and chemoresistance of NSCLC cells by targeting PTEN. *Oncol Rep* 30: 2897-2902, 2013.
- Mao Y, Wu S, Zhao R and Deng Q: MiR-205 promotes proliferation, migration and invasion of nasopharyngeal carcinoma cells by activation of AKT signalling. *J Int Med Res* 44: 231-240, 2016.
- Zhong G and Xiong X: miR-205 promotes proliferation and invasion of laryngeal squamous cell carcinoma by suppressing CDK2AP1 expression. *Biol Res* 48: 60, 2015.
- Jin C and Liang R: miR-205 promotes epithelial-mesenchymal transition by targeting AKT signaling in endometrial cancer cells. *J Obstet Gynaecol Res* 41: 1653-1660, 2015.
- Ma Q, Wan G, Wang S, Yang W, Zhang J and Yao X: Serum microRNA-205 as a novel biomarker for cervical cancer patients. *Cancer Cell Int* 14: 81, 2014.
- Hou SX, Ding BJ, Li HZ, Wang L, Xia F, Du F, Liu LJ, Liu YH, Liu XD, Jia JF, *et al*: Identification of microRNA-205 as a potential prognostic indicator for human glioma. *J Clin Neurosci* 20: 933-937, 2013.
- Xu C, Li M, Zhang L, Bi Y, Wang P, Li J and Jiang X: MicroRNA-205 suppresses the invasion and epithelial-mesenchymal transition of human gastric cancer cells. *Mol Med Rep* 13: 4767-4773, 2016.
- Yin WZ, Li F, Zhang L, Ren XP, Zhang N and Wen JF: Down-regulation of microRNA-205 promotes gastric cancer cell proliferation. *Eur Rev Med Pharmacol Sci* 18: 1027-1032, 2014.
- Zhang P, Wang L, Rodriguez-Aguayo C, Yuan Y, Debeb BG, Chen D, Sun Y, You MJ, Liu Y, Dean DC, *et al*: miR-205 acts as a tumour radiosensitizer by targeting ZEB1 and Ubc13. *Nat Commun* 5: 5671, 2014.
- Zhang H and Fan Q: MicroRNA-205 inhibits the proliferation and invasion of breast cancer by regulating AMOT expression. *Oncol Rep* 34: 2163-2170, 2015.
- Wang W, Liu J and Wu Q: MiR-205 suppresses autophagy and enhances radiosensitivity of prostate cancer cells by targeting TP53INP1. *Eur Rev Med Pharmacol Sci* 20: 92-100, 2016.
- Li P, Xue WJ, Feng Y and Mao QS: MicroRNA-205 functions as a tumor suppressor in colorectal cancer by targeting cAMP responsive element binding protein 1 (CREB1). *Am J Transl Res* 7: 2053-2059, 2015.

33. Kapoor A, Yao W, Ying H, Hua S, Liewen A, Wang Q, Zhong Y, Wu CJ, Sadanandam A, Hu B, *et al*: Yap1 activation enables bypass of oncogenic Kras addiction in pancreatic cancer. *Cell* 158: 185-197, 2014.
34. Deng J, Lei W, Xiang X, Zhang L, Yu F, Chen J, Feng M and Xiong J: MicroRNA-506 inhibits gastric cancer proliferation and invasion by directly targeting Yap1. *Tumour Biol* 36: 6823-6831, 2015.
35. Orr BA, Bai H, Oda Y, Jain D, Anders RA and Eberhart CG: Yes-associated protein 1 is widely expressed in human brain tumors and promotes glioblastoma growth. *J Neuropathol Exp Neurol* 70: 568-577, 2011.
36. Zeng Q and Hong W: The emerging role of the hippo pathway in cell contact inhibition, organ size control, and cancer development in mammals. *Cancer Cell* 13: 188-192, 2008.
37. Liu YC and Wang YZ: Role of Yes-associated protein 1 in gliomas: Pathologic and therapeutic aspects. *Tumour Biol* 36: 2223-2227, 2015.

RETRACTED