

# Downregulation of miR-136-5p in hepatocellular carcinoma and its clinicopathological significance

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**Abstract.** The clinical significance of microRNA (miR)-136-5p in hepatocellular carcinoma (HCC) has not been verified. Therefore, in the current study, the authors aimed to explore miR-136-5p expression and its clinical significance in HCC, as well as to investigate its potential target genes function. The authors detected the levels of miR-136-5p in 101 pairs of HCC and para-cancer tissues via reverse transcription-quantitative polymerase chain reaction. Gene Expression Omnibus database and the Cancer Genome Atlas (TCGA) database were used to further verify the clinical significance of miR-136-5p expression in HCC. The target genes prediction analysis of miR-136-5p, natural language processing (NLP) analysis of HCC in PubMed and gene functional enrichment analysis were conducted. The miR-136-5p level was markedly downregulated in HCC tissue, compared to para-non-tumor tissue. MiR-136-5p expression decreased in HCC patients with metastasis ( $P=0.004$ ), advance TNM stage ( $P<0.001$ ), portal vein tumor embolus ( $P=0.007$ ) and vaso-invasion ( $P=0.003$ ), compared with those HCC patients with non-metastasis, early TNM stage, non-portal vein tumor embolus and non-vaso-invasion, respectively. In the TCGA database, downregulated miR-136-5p was also observed in

HCC tissue compared to normal liver tissue ( $P<0.001$ ). There were 178 genes obtained from the overlap between predicted targets and NLP analysis. GO and KEGG pathway analyses revealed some significant pathways related to cancers. Downregulation of miR-136-5p may be responsible for the carcinogenesis and aggressiveness of HCC. miR-136-5p may act as an anti-carcinoma miRNA, which is essential for HCC progression through the regulation of various signaling pathways. Thus, miR-136-5p interaction may provide a novel strategy for HCC treatment.

## Introduction

Hepatocellular carcinoma (HCC), as a common malignancy in China, has a high incidence and poor prognosis (1). Although great progress has been achieved in the treatment of HCC, including surgery, interventional therapy, radiotherapy, chemotherapy and molecular target treatment, the prognosis of HCC is still poor due to the metastasis and recurrence (2). Therefore, it is necessary to explore highly stable and specific molecular targets and provide the basis for early diagnosis of HCC and targeted therapy, in order to improve the prognosis of HCC.

Micro (mi)RNAs, ~22 nucleotides, belong to a large family of non-coding, single stranded RNAs. miRNAs participate in cell growth, hematopoiesis, apoptosis and cell proliferation (3-5). It has been reported that miRNAs contribute to carcinogenesis and progression in cancers by positive or negative regulation of tumor related genes (6,7). Previous studies demonstrated downregulated miR-136-5p expression in epithelial ovarian cancer and glioma cell lines (8,9). However, regarding to non-small cell lung cancer, miR-136-5p was upregulated (10). In addition, miR-136-5p can promote apoptosis of glioma cells induced by chemotherapy (8). As for the role of miR-136-5p in HCC, the only article available was based on an *in vitro* study (11), which presented that miR-136-5p could attenuate cell migration. However, no publication of the investigation of miR-136-5p expression in HCC tissue is accessible, and the clinical significance of miR-136-5p in HCC remains unknown.

In the present research, the authors explored the expression of miR-136-5p in HCC and its clinical significance for the diagnosis and prediction of prognosis in HCC. Moreover,

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the potential targets of miR-136-5p and gene functional enrichment analysis were involved to explore the prospective mechanisms in the pathogenesis of HCC.

## Materials and methods

**Patients and samples.** There were 101 HCC and matched adjacent non-tumor tissues selected from 76 men and 19 women at a mean age of 50. The patients, who had no chemotherapy or radiotherapy before the surgery from March 2010 to March 2012, were selected in the First Affiliated Hospital of Guangxi Medical University (Nanning, China). Two pathologists independently confirmed all pathological diagnosis. The clinicopathological factors are summarized in Table I. Written informed consent was gained for the usage of patients' tissue samples.

**Reverse transcription-quantitative polymerase chain reaction (RT-qPCR).** The processes of RT-qPCR were performed as described previously (12-15). Total RNA from HCC and non-cancerous liver tissues were extracted using the miRNeasy FFPE kit (Qiagen Benelux B.V., PL Venlo, The Netherlands). Reverse transcription was performed using the TaqMan MicroRNA Reverse Transcription kit (cat. no. 4366596; Applied Biosystems; Thermo Fisher Scientific, Inc., Waltham, MA, USA) and the LightCycler 480 (Roche Diagnostics GmbH, Mannheim, Germany) according to the manufacturer protocol. The following temperature protocol was applied: 16°C for 30 min, 42°C for 30 min and 85°C for 5 min. LightCycler 480 SYBR Green I Master (Roche Diagnostics GmbH) was used for qPCR, with the following thermocycling conditions: initial pre-denaturation for 5 min at 95°C, followed by 40 cycles of 95°C with 10 sec, 60°C for 10 sec and 72°C for 10 sec; evaluation of the solubility curve was performed at 95°C for 5 sec and 65°C for 1 min, which was followed by cooling at 40°C for 30 sec. Each experiment was repeated in triplicate. The primers used were provided by Thermo Fisher Scientific, Inc. and the detailed sequences are commercially confidential. A combined union of RUN6B and RUN48, which was demonstrated to be a stable internal control in our previous studies, served as the reference to the detect miR-136-5p level (16-18). The primers of RNU6B, RNU48 and miR-136-5p were contained in TaqMan MicroRNA Assays (cat. no. 4427975; Applied Biosystems; Thermo Fisher Scientific, Inc.), the sequences of each marker are as follows: miR-136-5p: ACUCCAUUUGUUUUGAUG AUGGA; RNU6B: CGCAAGGAUGACACGCAAAUUCGU GAAGCGUCCAUUUUUU; RNU48: GAUGACCCCAGG UAACUCUGAGUGUGUCGUGAUGCCAUCACCGCAGC GCUCUGACC. The expression level of miR-136-5p was calculated by using formula  $2^{-\Delta\Delta C_q}$  (19).

**The miR-136-5p expression in HCC in The Cancer Genome Atlas (TCGA).** The TCGA database was used to further explore miR-136-5p expression in HCC tissues. In addition, miR-136-5p expression in HCC patients with various clinicopathological characteristics was analyzed. In TCGA database, the data of level three of the miRNA profile of HCC were extracted. The HCC patients, who had no neoadjuvant treatment, radiation therapy and pharmaceutical TX adjuvant treatment, were included. Meanwhile, the corresponding

clinical data of HCC were obtained. Afterwards, the expression of miR-136-5p was extracted from the miRNA profile and the statistical analyses were conducted to estimate the expression of miR-136-5p in HCC tissues and non-cancerous tissues, as well as the relationships between miR-136-5p expression and various clinicopathological factors.

**The miR-136-5p expression in HCC in Gene Expression Omnibus (GEO).** In GEO database, including GEO dataset and GEO profile, the authors searched miR-136-5p expression in HCC using the following key words: 1) (hepatocellular OR liver OR hepatic OR HCC), 2) malignan\* OR cancer OR tumor OR tumour OR neoplas\* OR carcinoma, 3) microRNA OR miRNA OR miR. The microarrays, which investigated miR-136-5p expression in HCC tissues and non-cancerous tissues, or explored miR-136-5p expression in different clinicopathological features in HCC, were included.

The meta-analysis, to combine the data of miR-136-5p expression in RT-qPCR, microarray in GEO database and RNA sequencing in TCGA database in different tissues, was performed by Stata 12.0 (StataCorp LP, College Station, TX, USA). The pooled standard mean difference (SMD) with its 95% confidence interval (95% CI) was used to assess miR-136-5p expression in different tissues. Moreover, Q test (chi-squared test) and  $I^2$  statistic were performed to estimate heterogeneity among the eligible studies. A Mantel-Haenszel fixed-effect model was performed if there was no obvious heterogeneity among the pooled studies ( $I^2 < 50\%$  and  $P > 0.1$ ). However, when obvious heterogeneity was identified among the include studies ( $I^2 > 50\%$  or  $P < 0.1$ ), random-effect model was conducted (20).

**MiRNA target prediction.** A total of twelve databases were involved for the target prediction of miR-136-5p, including miRWalk, MicroT4, miRanda, mirbridge, miRDB, miRMap, miRNAMap, Pictar2, PITA, RNAhybrid, TargetsScan and miRWalk2.0 (21). Then, the union of target genes of miR-136-5p of human reported in more than five databases and the validated genes of miR-136-5p with strong evidence were gathered for further gene functional enrichment analysis.

**Natural language processing (NLP) analysis for HCC-related studies.** The authors searched the PubMed database by using the keywords: 1) Hepatocellular OR liver OR hepatic OR HCC, 2) malignan\* OR cancer OR tumor OR tumour OR neoplas\* OR carcinoma. All related genes were extracted in the studies by biomedical named entity recognizer (ABNER, <http://pages.cs.wisc.edu/~bsettles/abner/>). In NLP analysis, the frequency of each gene that merged in the eligible studies in PubMed was counted. The greater the frequency indicates the higher likelihood of the correlation between a certain gene and HCC. In PubMed, the number of the relevant studies in total was marked as 'N'. The frequency of related genes and HCC in PubMed was marked as 'm' and 'n'. Moreover, the simultaneous occurrence of the disease and a certain gene was marked as 'k'. The probability of frequency greater than 'k' co-citation was calculated with random conditions completely through hypergeometric distribution. It was considered the statistically significance with  $P < 0.01$ . The formula of the NLP analysis was as following:

$$P = 1 - \sum_{i=0}^{k-1} p(i|n, m, N); \quad p(i|n, m, N) = \frac{n!(N-n)!m!(N-m)!}{n-i!i!n-m!N-n-m+i!N!}$$

Table I. Expression of miR-136-5p and clinicopathological parameters in hepatocellular carcinoma in the 101 pairs detected by real time reverse transcription-quantitative polymerase chain reaction.

Clinicopathological features	n	miR-136-5p relevant expression ( $2^{-\Delta\Delta C_q}$ )		
		Mean $\pm$ standard deviation	t	P-value
Tissue				
Adjacent non-cancerous liver	101	4.8507 $\pm$ 3.0644	6.516	<0.001
Hepatocellular carcinoma	101	2.8460 $\pm$ 1.7090		
Age				
$\geq 50$	51	2.9133 $\pm$ 1.7885	0.398	0.692
<50	50	2.7774 $\pm$ 1.6393		
Gender				
Male	80	2.8011 $\pm$ 1.6987	0.813	0.609
Female	21	3.0171 $\pm$ 1.7797		
Differentiation				
High	7	2.1857 $\pm$ 1.3570	F=0.666 <sup>a</sup>	0.516
Moderate	64	2.8387 $\pm$ 1.6319		
Low	30	3.0157 $\pm$ 1.9416		
Size (cm)				
<5	21	3.1014 $\pm$ 1.9355	0.773	0.444
$\geq 5$	80	2.7790 $\pm$ 1.6513		
Tumor nodes				
Single	57	2.9779 $\pm$ 1.8381	0.882	0.380
Multiple	44	2.6752 $\pm$ 1.5296		
Metastasis				
Without metastasis	49	3.3429 $\pm$ 1.9272	2.943	0.004
With metastasis	52	2.3779 $\pm$ 1.3307		
TNM stage				
I~II	25	4.2680 $\pm$ 2.0246	4.380	<0.001
III~IV	76	2.3783 $\pm$ 1.2988		
Portal vein tumor embolus				
-	69	3.1146 $\pm$ 1.8511	2.774	0.007
+	32	2.2669 $\pm$ 1.1827		
Vaso-invasion				
-	63	3.2059 $\pm$ 1.8550	3.100	0.003
+	38	2.2495 $\pm$ 1.2418		
Tumor capsular infiltration				
With complete capsule	49	2.9510 $\pm$ 1.8003	0.597	0.552
No capsule or infiltration	52	2.7471 $\pm$ 1.6298		
AFP				
-	46	2.6313 $\pm$ 1.5243	1.444	0.152
+	39	3.1751 $\pm$ 1.9453		
Cirrhosis				
-	54	2.5633 $\pm$ 1.5825	1.802	0.075
+	47	3.1709 $\pm$ 1.8063		

miR, microRNA; AFP,  $\alpha$ -fetoprotein. <sup>a</sup>One-way analysis of variance analysis was performed.

**Gene functional enrichment analysis.** The database of DAVID was used for GO annotation, including biological process, molecular function and cellular component (22,23). Cytoscape

3.0 software (<http://www.cytoscape.org/>) with BINGO plugins were responsible for graphical display of the network of GO (24). In the networks, each node represents a biological

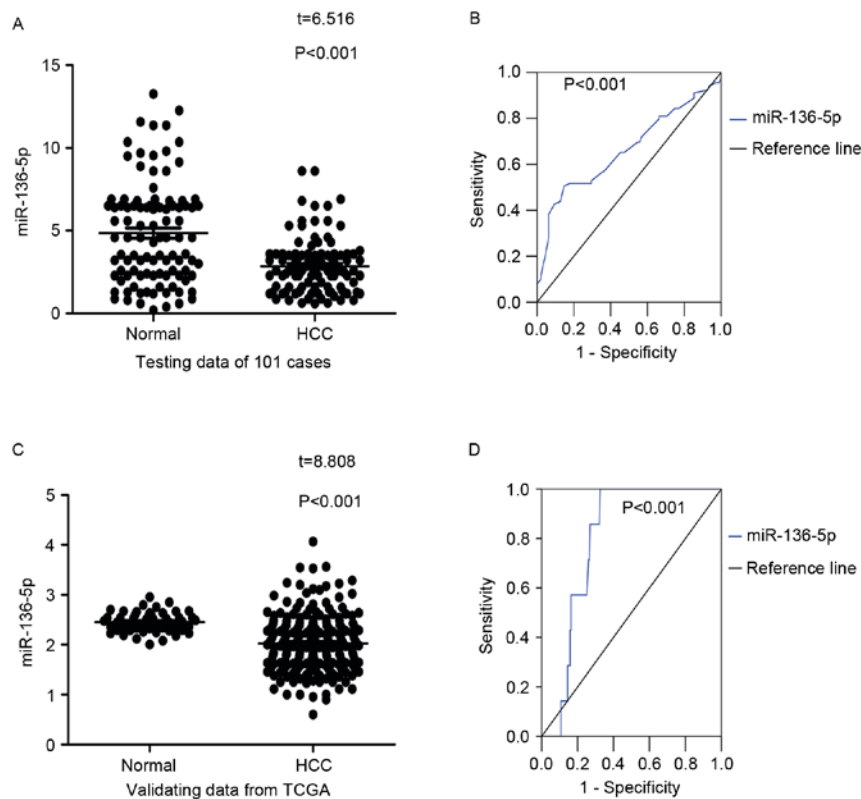


Figure 1. miR-136-5p expression in HCC tissues and adjacent non-tumor liver tissues. (A) 101 HCC tissues v. 101 adjacent non-tumor liver tissues. (B) The ROC curve of miR-136-5p to assess its diagnostic value in the 101 pairs; AUC=0.696. (C) The comparison of miR-136-5p levels in 215 HCC tissues and 50 adjacent non-tumor tissues in TCGA database. (D) The ROC curve of miR-136-5p expression in HCC performed by TCGA data; AUC=0.771. miR, microRNA; HCC, hepatocellular carcinoma; ROC, receiver operating characteristic; AUC, area under curve; TCGA, the Cancer Genome Atlas.

process. The bigger node means more genes participating in the biological process and the deeper color indicates the smaller P-value of the biological process. All colored nodes indicate statistical significance ( $P<0.05$ ). White nodes were only used to connect the biological processes and there is no statistical significance. The protein-to-protein network analysis was conducted by STRING (25). The database of Kyoto Encyclopedia of Genes and Genomes (KEGG) was also applied to reflect the prospective biological pathways involved in the targets of miR-136-5p in HCC.

**Statistical analysis.** The authors performed the statistical analyses by using SPSS software (version, 22; IBM SPSS, Armonk, NY, USA). Quantitative data of miR-136-5p expression were presented as mean  $\pm$  standard deviation. Student's *t* tests were performed for the analysis of miR-136-5p expression between two groups and when it exceeded two groups, one-way analysis of variance was employed. Additionally, the authors drew receiver operating characteristic (ROC) curve with purpose for evaluating the power of miR-136-5p in HCC diagnosis. In addition, Spearman analysis was conducted to estimate the associations between miR-136-5p and clinical factors.  $P<0.05$  was considered to indicate a statistically significant difference.

## Results

**miR-136-5p expression in HCC and its clinicopathological significance detected by RT-qPCR.** In contrast with the adjacent normal liver tissue ( $2.8460\pm1.7090$ ), miR-136-5p was

significantly underexpressed in HCC tissue ( $4.8507\pm3.0644$ ;  $P<0.001$ ; Table I; Fig. 1A). Meanwhile, in the ROC curve, an area under the curve (AUC) of 0.696 was observed ( $P<0.001$ ; Fig. 1B).

All clinicopathological factors are presented in Table I. When concerning the relationships between miR-136-5p and clinicopathological parameters in HCC, miR-136-5p expression was lower in tissues with metastasis ( $3.3429\pm1.9272$ ) than that of HCC tissues without metastasis ( $2.3779\pm1.3307$ ;  $P=0.004$ ; Fig. 2A). Moreover, in the group of III~IV clinical TMN stage, miR-136-5p was also downregulated significantly ( $2.3783\pm1.2988$ ), compared with the group of stage I~II ( $4.2680\pm2.0246$ ;  $P<0.001$ ; Fig. 2B). Furthermore, the HCC tissues with portal vein tumor embolus had a lower expression of miR-136-5p ( $2.2669\pm1.1827$ ), while the ones without portal vein tumor embolus held an oppositely higher expression of miR-136-5p ( $3.1146\pm1.8511$ ;  $P=0.007$ ; Fig. 2C). Meanwhile, it witnessed a lower expression in the vaso-invasion positive group ( $2.2495\pm1.2418$ ) compared with the negative ones ( $3.2059\pm1.8550$ ;  $P=0.003$ ; Fig. 2D). The rest of clinicopathological characteristics in HCC have no significant connections with miR-136-5p expression (Table I). In addition, Spearman analysis was used to assess the relationships between miR-136-5p and clinicopathological parameters, and it reported that negative correlations were discovered between miR-136-5p and metastasis ( $r=-0.284$ ;  $P=0.004$ ), TNM ( $r=-0.480$ ;  $P<0.001$ ), portal vein tumor embolus ( $r=-0.232$ ;  $P=0.02$ ), vaso-invasion ( $r=-0.272$ ;  $P=0.006$ ), however, the correlations were relatively low. No significant association between the rest clinicopathological



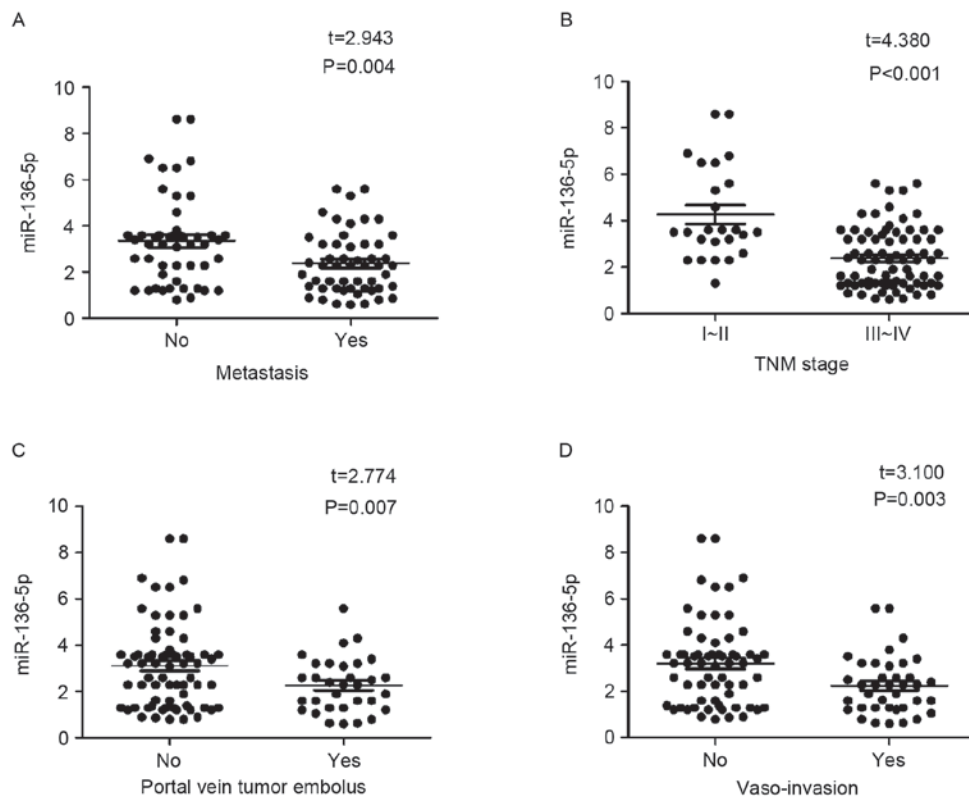


Figure 2. Expression of miR-136-5p in various clinicopathological factors of hepatocellular carcinoma using the reverse transcription-quantitative polymerase chain reaction. (A) Metastasis; (B) TNM stage; (C) portal vein tumor embolus and; (D) vaso-invasion. miR, microRNA.

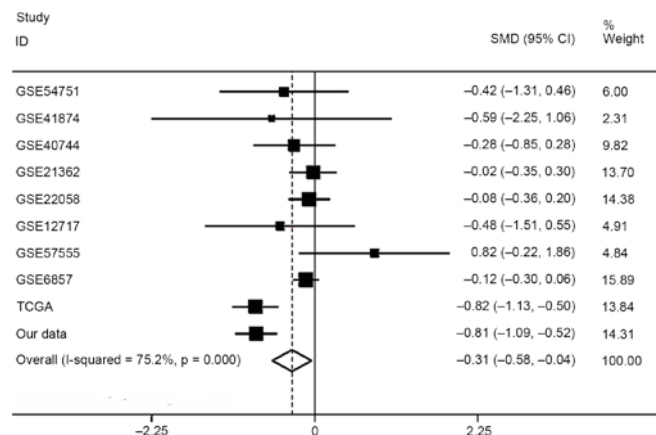


Figure 3. Forest of the meta-analysis for microRNA-136-5p expression in hepatocellular carcinoma in GEO database. Nine studies were included in GEO database and pooled SMD was -0.092 (95% CI: 0.221-0.036,  $P=0.159$ ) with a fixed effects model. GEO, Gene Expression Omnibus; SMD, standard mean difference; CI, confidence interval.

factors and miR-136-5p was noted. Besides, HCC patients with lower expression of miR-136-5p (lower than the mean value) shared a trend of longer recurrence time ( $37.507 \pm 13.074$ ) than the ones with higher miR-136-5p expression ( $30.478 \pm 14.768$ ). However, the difference had no statistical significance, as the P-value was 0.568 (data not shown).

**Validation of miR-136-5p expression in HCC in TCGA.** In consistent with the results from RT-qPCR, TCGA data indicated that significantly lower miR-136-5p levels were detected

in 215 HCC tissues compared to 50 cases of normal liver tissue ( $P<0.001$ ; Fig. 1C) with a AUC of 0.771 ( $P<0.001$ ; Fig. 1D). There was no significantly differential expression of miR-136-5p in the rest clinicopathological factors. The results regarding the associations between miR-136-5p and the clinicopathological characteristics from TCGA are presented in Table II.

**Meta-analysis of miR-136-5p expression in HCC.** Regarding miR-136-5p expression between HCC and non-tumor liver tissue, the data of RT-qPCR of this current study, eight studies of microarrays in the GEO database (GSE54751, GSE41874, GSE40744, GSE21362, GSE22058, GSE12717, GSE57555, GSE6857) and the RNA sequencing data from TCGA, which involved 793 HCC and 614 non-tumor liver tissues, were included for this meta-analysis. The pooled SMD of miR-136-5p was -0.313 (95% CI, -0.583- (-0.044);  $P=0.023$ ; Fig. 3) by the random effects model and the P value of the heterogeneity test was  $<0.001$  ( $I^2=75.2\%$ ).

**Prediction of the target genes of miR-136-5p and GO analysis.** A total of 12 databases were searched for the prediction of the potential targets of miR-136-5p. A total of 13,857 genes were identified in these databases and three genes (MTDH, BCL2 and PPP2R2A) were involved with strong evidence, as they have been confirmed by reporter assay, western blotting or qPCR. Following that, the genes reported in more than five databases combined with the gene with strong evidence were selected, and finally 1,766 genes were applied to functional enrichment, including 1,765 genes reported in more than five predicted databases and one gene reported with strong evidence (two genes in

Table II. Expression of miR-136-5p and clinicopathological parameters in HCC in the Cancer Genome Atlas.

Clinicopathological features	n	lg(miR-136-5p) expression		
		Mean $\pm$ standard deviation	t	P-value
Tissue				
Adjacent non-cancerous liver	50	2.4517 $\pm$ 0.2036	8.808	<0.0011
HCC	215	2.0279 $\pm$ 0.5652		
Age				
<60	103	2.1058 $\pm$ 0.5637	2.000	0.047
$\geq$ 60	111	1.9520 $\pm$ 0.5602		
Gender				
Male	156	1.9845 $\pm$ 0.5384	1.842	0.067
Female	59	2.1427 $\pm$ 0.6209		
AJCC pathologic T				
T0 TX	1	2.3263	F=0.231 <sup>a</sup>	0.749
T1	130	2.0382 $\pm$ 0.5771		
T2-T4	82	2.0040 $\pm$ 0.5532		
AJCC pathologic N				
N0	151	2.0358 $\pm$ 0.5933	0.314	0.745
NX	64	2.0093 $\pm$ 0.4966		
AJCC pathologic M				
M0	156	2.0428 $\pm$ 0.6010	F=0.355 <sup>a</sup>	0.702
M1	1	1.6721		
MX	58	1.9939 $\pm$ 0.4617		
Grade				
I~II	20	2.1286 $\pm$ 0.5371	0.871	0.631
III~IV	4	2.2691 $\pm$ 0.4566		
Cirrhosis				
-	203	2.0335 $\pm$ 0.5514	1.132	0.259
+	3	1.6682 $\pm$ 0.8406		
Pathologic stage				
I~II	175	2.0404 $\pm$ 0.5639	1.576	0.117
III~IV	27	1.8546 $\pm$ 0.6092		
Vaso-invasion				
-	129	2.0159 $\pm$ 0.5331	0.076	0.939
+	72	2.0223 $\pm$ 0.6249		

<sup>a</sup>One-way analysis of variance analysis was performed. HCC, hepatocellular carcinoma.

strong evidence were included in 1,765 predicted target genes). The flow chart of this prediction analysis is reported in Fig. 4.

In GO analysis in DAVID, there were 175 significant annotation information of biological process ( $P<0.05$ ) and some of them were involved in transcription, positive regulation of transcription, positive regulation of RNA metabolic process, positive regulation of macromolecule biosynthetic process and so on (Table III). With regard to the cellular component of GO analysis, 63 significant terms were observed ( $P<0.05$ ) and some of them correlated with insoluble fraction, membrane fraction, endomembrane system, synapse and vesicular fraction (Table III). In the molecular function of GO analysis, 66 significant terms were obtained ( $P<0.05$ ) and some of them were contributed to transcription regulator activity, transcription

factor activity, transcription repressor activity, protein tyrosine phosphatase activity and phosphoprotein phosphatase activity (Table III).

After gene functional enrichment by GO analysis in DAVID, the authors performed BINGO analysis in order to display the network of the results in GO analysis. The networks of BINGO analysis, including three parts (biological process, cellular component and molecular function), are presented in Figs. 5-7, respectively.

*The overlap of predicted target genes of miR-136-5p and NLP analysis and its GO analysis.* There were 1,800 significant HCC-related genes in NLP analysis. Following uniting the genes of the predicted targets of miR-136-5p and NLP,

GO ID	Term	Count	P-value
Biological process			
0006350	Transcription	269	1.90x10 <sup>-8</sup>
0045449	Regulation of transcription	314	3.54x10 <sup>-7</sup>
0045893	Positive regulation of transcription, DNA-dependent	79	7.34x10 <sup>-7</sup>
0051254	Positive regulation of RNA metabolic process	79	1.03x10 <sup>-6</sup>
0010557	Positive regulation of macromolecule biosynthetic process	98	3.13x10 <sup>-6</sup>
Cellular component			
0005626	Insoluble fraction	118	1.59x10 <sup>-7</sup>
0005624	Membrane fraction	113	4.40x10 <sup>-7</sup>
0012505	Endomembrane system	107	2.49x10 <sup>-6</sup>
0045202	Synapse	58	4.69x10 <sup>-6</sup>
0042598	Vesicular fraction	44	6.59x10 <sup>-6</sup>
Molecular function			
0030528	Transcription regulator activity	187	5.24x10 <sup>-5</sup>
0003700	Transcription factor activity	129	5.32x10 <sup>-5</sup>
0016564	Transcription repressor activity	51	1.81x10 <sup>-4</sup>
0004725	Protein tyrosine phosphatase activity	23	2.33x10 <sup>-4</sup>
0004721	Phosphoprotein phosphatase activity	30	7.40x10 <sup>-4</sup>

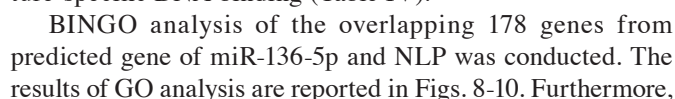
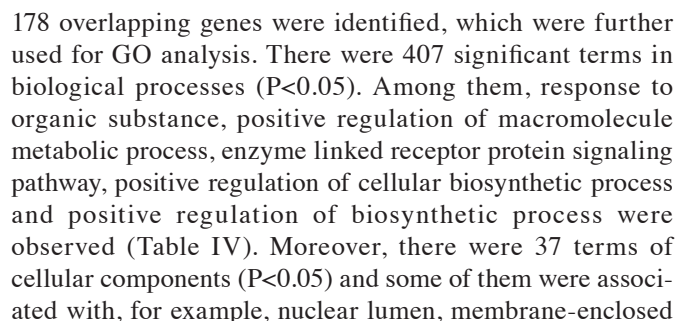


Table IV. GO analysis of the overlap between predicted target genes of miR-136-5p and NLP.

GO ID	Term	Count	P-value
Biological process			
0010033	Response to organic substance	34	$3.83 \times 10^{-11}$
0010604	Positive regulation of macromolecule metabolic process	37	$4.82 \times 10^{-11}$
0007167	Enzyme linked receptor protein signaling pathway	23	$1.96 \times 10^{-10}$
0031328	Positive regulation of cellular biosynthetic process	32	$2.20 \times 10^{-10}$
0009891	Positive regulation of biosynthetic process	32	$3.14 \times 10^{-10}$
Cellular component			
0031981	Nuclear lumen	35	$1.38 \times 10^{-5}$
0031974	Membrane-enclosed lumen	41	$1.47 \times 10^{-5}$
0043233	Organelle lumen	40	$2.23 \times 10^{-5}$
0045121	Membrane raft	10	$2.86 \times 10^{-5}$
0005887	Integral to plasma membrane	30	$3.33 \times 10^{-5}$
Molecular function			
0030528	Transcription regulator activity	39	$6.38 \times 10^{-6}$
0003690	Double-stranded DNA binding	9	$2.12 \times 10^{-5}$
0003700	Transcription factor activity	28	$3.55 \times 10^{-5}$
0003702	RNA polymerase II transcription factor activity	13	$3.82 \times 10^{-5}$
0043566	Structure-specific DNA binding	10	$6.26 \times 10^{-5}$

In the GO analysis of the overlap between NLP and predicted target genes of miR-136-5p, there were 407 available biological processes, 37 cellular components, 49 molecular functions ( $P < 0.05$ ). In this table, the first five terms of the GO analysis are presented. GO, Gene Ontology; NLP, natural language processing; miR, microRNA.

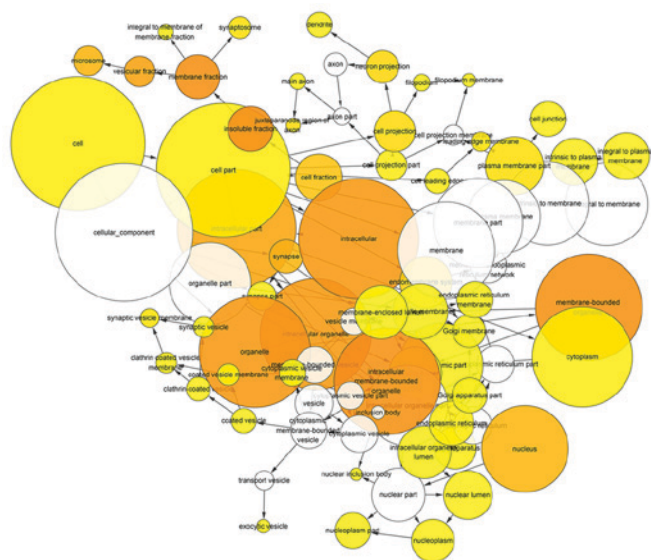


Figure 6. Hierarchical Network in BINGO analysis of predicted target genes of miR-136-5p of cellular component. Each node represents a biological process. The bigger node means more genes participating in the process. All colored nodes indicate statistical significance ( $P < 0.05$ ). The deeper color indicates the smaller P-value of the biological process. White colored nodes were used to connect the biological processes without statistical significance.

hub genes analysis was conducted by the protein-protein interaction and the results are presented in Fig. 11. In the network of the overlap, several genes demonstrated higher connectivity, including BCL2, PTEN, MMP2 and FOXO1.



Figure 7. Hierarchical Network in BINGO analysis of predicted target genes of miR-136-5p of molecular function. Each node represents a biological process. The bigger node means more genes participating in the process. All colored nodes indicate statistical significance ( $P < 0.05$ ). The deeper color indicates the smaller P-value of the biological process. White colored nodes were used to connect the biological processes without statistical significance.

*Pathway analysis of the overlapping genes between predicted genes of miR-136-5p and NLP in KEGG.* The overlapping 178 genes between the predicted genes and NLP were also used to pathway analysis in KEGG. There were 33 available pathways and 17 signaling pathways that were statistically significant ( $P < 0.01$ , Table IV), including pathways in cancer (FGF19, E2F1, E2F3, XIAP, KITLG, FOXO1, FASLG, TCF7L2, PTEN, MMP2, TPM3, CDC42, FOS, BCL2, ITGAV, RAC1, FIGF,



Table V. Pathway analysis of the overlap of natural language processing analysis and predicted target genes of microRNA-136-5p.

Title	Count	P-value	Genes
Pathways in cancer	31	$1.03 \times 10^{-13}$	FGF19, E2F1, E2F3, XIAP, KITLG, FOXO1, FASLG, TCF7L2, PTEN, MMP2, TPM3, CDC42, FOS, BCL2, ITGAV, RAC1, FIGF, FGF2, AKT2, FN1, PRKCA, RET, AR, MSH3, BRAF, CBL, CDK6, CDK4, MAPK1, ETS1, PTCH1
Neurotrophin signaling pathway	13	$3.19 \times 10^{-6}$	YWHAZ, BRAF, FASLG, IRS1, MAPK1, CDC42, BDNF, PSEN1, BCL2, RAC1, NTRK2, NGFR, AKT2
Prostate cancer	11	$5.85 \times 10^{-6}$	E2F1, MAPK1, E2F3, AR, BRAF, BCL2, CREB1, FOXO1, PTEN, TCF7L2, AKT2
Melanoma	10	$6.45 \times 10^{-6}$	E2F1, FGF19, MAPK1, E2F3, BRAF, CDK6, CDK4, FGF2, PTEN, AKT2
Pancreatic cancer	10	$7.26 \times 10^{-6}$	E2F1, CDC42, MAPK1, E2F3, BRAF, RAC1, CDK6, CDK4, FIGF, AKT2
Glioma	9	$2.14 \times 10^{-5}$	PRKCA, E2F1, MAPK1, E2F3, BRAF, CDK6, CDK4, PTEN, AKT2
Focal adhesion	15	$2.22 \times 10^{-5}$	PRKCA, CDC42, MAPK1, CAV1, FLT1, BRAF, XIAP, CCND2, BCL2, ITGAV, RAC1, FIGF, PTEN, FN1, AKT2
Small cell lung cancer	10	$2.60 \times 10^{-5}$	E2F1, E2F3, XIAP, BCL2, ITGAV, CDK6, CDK4, PTEN, FN1, AKT2
Non-small cell lung cancer	8	$6.29 \times 10^{-5}$	PRKCA, E2F1, MAPK1, E2F3, BRAF, CDK6, CDK4, AKT2
Bladder cancer	7	$1.24 \times 10^{-4}$	E2F1, MAPK1, E2F3, BRAF, CDK4, FIGF, MMP2
MAPK signaling pathway	15	$4.72 \times 10^{-4}$	PRKCA, FGF19, BRAF, MAP2K4, FASLG, CDC42, MAPK1, FOS, DUSP4, BDNF, RAC1, NTRK2, GADD45B, FGF2, AKT2
Chronic myeloid leukemia		$5.09 \times 10^{-4}$	E2F1, MAPK1, E2F3, BRAF, CBL, CDK6, CDK4, AKT2
Colorectal cancer	8	$1.01 \times 10^{-3}$	MAPK1, FOS, MSH3, BRAF, BCL2, RAC1, TCF7L2, AKT2
Renal cell carcinoma	7	$2.021 \times 10^{-3}$	CDC42, MAPK1, BRAF, ETS1, RAC1, FIGF, AKT2
Thyroid cancer	5	$2.04 \times 10^{-3}$	MAPK1, RET, BRAF, TCF7L2, TPM3
Cell cycle	9	$2.47 \times 10^{-3}$	E2F1, CDC6, E2F3, YWHAZ, CCND2, PRKDC, CDK6, CDK4, GADD45B
p53 signaling pathway	6	$9.20 \times 10^{-3}$	CCND2, CDK6, CDK4, GADD45B, CCNG2, PTEN

The pathway analysis was performed in Kyoto Encyclopedia of Genes and Genomes database and there were 33 available pathways. Among them, 17 signaling pathways were significant ( $P < 0.01$ ).

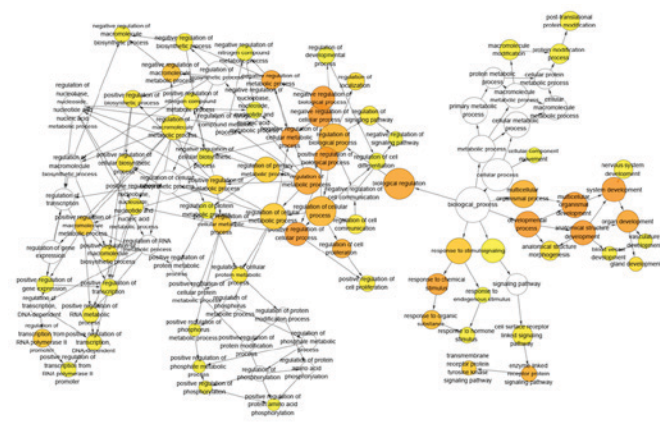


Figure 8. Hierarchical Network in BINGO analysis of the overlap between the predicted target genes of miR-136-5p and natural language processing analysis of biological process (GO). Each node represents a biological process. The bigger node means more genes participating in the process. All colored nodes indicate statistical significance ( $P < 0.05$ ). The deeper color indicates the smaller P-value of the biological process. White colored nodes were used to connect the biological processes without statistical significance.

FGF2, AKT2, FN1, PRKCA, RET, AR, MSH3, BRAF, CBL, CDK6, CDK4, MAPK1, ETS1, PTCH1,  $P = 1.03 \times 10^{-13}$ ), the neurotrophin signaling pathway (YWHAZ, BRAF, FASLG, IRS1, MAPK1, CDC42, BDNF, PSEN1, BCL2, RAC1, NTRK2, NGFR, AKT2,  $P = 3.19 \times 10^{-6}$ ), focal adhesion (PRKCA, CDC42, MAPK1, CAV1, FLT1, BRAF, XIAP, CCND2, BCL2, ITGAV, RAC1, FIGF, PTEN, FN1, AKT2,  $P = 2.22 \times 10^{-5}$ ), the MAPK signaling pathway (PRKCA, FGF19, BRAF, MAP2K4, FASLG, CDC42, MAPK1, FOS, DUSP4, BDNF, RAC1, NTRK2, GADD45B, FGF2, AKT2,  $P = 4.72 \times 10^{-4}$ ), the cell cycle (E2F1, CDC6, E2F3, YWHAZ, CCND2, PRKDC, CDK6, CDK4, GADD45B,  $P = 0.00246723$ ) and the p53 signaling pathway (CCND2, CDK6, CDK4, GADD45B, CCNG2, PTEN,  $P = 0.009198449$ ) (Table V).

## Discussion

As a common cancer, HCC attracts more attention by the clinicians and researchers worldwide because of the increasing

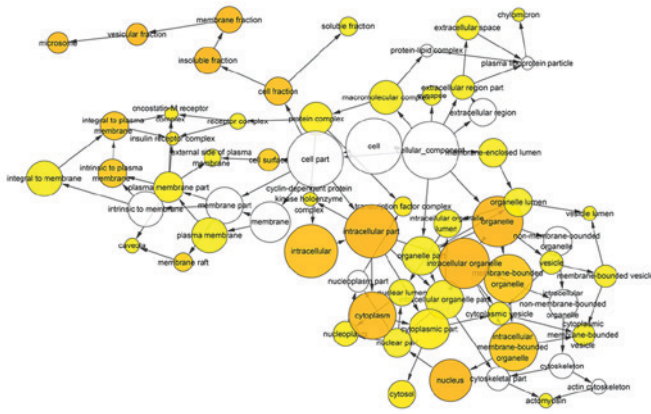


Figure 9. Hierarchical Network in BINGO analysis of the overlap between the predicted target genes of miR-136-5p and natural language processing analysis cellular component (GO). Each node represents a biological process. The bigger node means more genes participating in the process. All colored nodes indicate statistical significance ( $P < 0.05$ ). The deeper color indicates the smaller P-value of the biological process. White colored nodes were used to connect the biological processes without statistical significance.

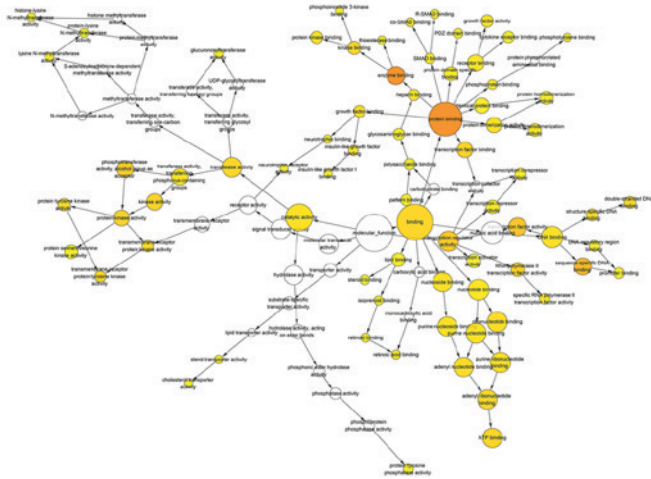


Figure 10. Hierarchical Network in BINGO analysis of the overlap between the predicted target genes of miR-136-5p and natural language processing analysis molecular function (GO). Each node represents a biological process. The bigger node means more genes participating in the process. All colored nodes indicate statistical significance ( $P < 0.05$ ). The deeper color indicates the smaller P-value of the biological process. White colored nodes were used to connect the biological processes without statistical significance.

incidence and poor survival despite all the advances in diagnosis and therapeutic techniques (26). miR-136-5p is a member of the miRNAs, a family of noncoding RNA, which can regulate the target genes through binding to 3'-untranslated regions of target mRNA, thereby inhibiting or promoting oncogenesis and progression (27). Thus, it is essential to explore and clarify the function of miR-136-5p in HCC in order to identify novel potential targets for HCC diagnosis, prognosis and treatment.

There were a few previous studies, which investigated the expression of miR-136-5p in cancers. Shen *et al* (10) demonstrated that miR-136 (miR-136-5p) was overexpressed in lung cancer. However, Yang *et al* (8) discovered that miR-136 (miR-136-5p) was downregulated in glioma. Momen-Heravi *et al* (28) confirmed miR-136 (miR-136-5p) was underexpressed in the patients with oral squamous

cell carcinoma through the genome wide high-throughput miRNA microarray. In addition, Zhao *et al* (9) discovered that decreased miR-136 (miR-136-5p) expression were determined in chemo-resistant epithelial ovarian cancer. According to the aforementioned studies, miR-136-5p may be heterogeneously expressed in different cancers. By far, only one study explored miR-136-5p expression in liver biopsy samples infected with HCV but the patients did not develop into HCC (29). To the best of the authors' knowledge, there is no study investigating the miR-136-5p expression in the tissues of HCC. Therefore, it is of great significance to explore miR-136-5p expression in neoplastic tissue of HCC and analyze its clinical value.

In this present study, the authors were the first group to focus on the expression of miR-136-5p in HCC with clinical parameters. They observed strikingly reduced levels of miR-136-5p in HCC tissues compared with the normal ones by using RT-qPCR ( $P < 0.001$ ) and consistent results were reported in TCGA database ( $P < 0.001$ ), which was in accordance with the results in glioma and giant cell tumor of bone (8,30). Moreover, miR-136-5p may be a potential biomarker for HCC diagnosis, due to two AUC values of 0.696 in RT-qPCR and 0.771 in TCGA data validation (both  $P < 0.001$ ). Furthermore, the meta-analysis based on the microarray data from GEO, RT-qPCR and TCGA indicated a positive SMD ( $P = 0.023$ ). Taken together, the above results confirmed the role of miR-136-5p in the carcinogenesis of HCC and the clinical value for HCC diagnosis. However, further investigations *in vivo* and *in vitro* are needed to clarify the underlying mechanism of miR-136-5p in occurrence of HCC.

Regarding the associations between miR-136-5p expression and clinicopathological factors in HCC and the potential regulatory mechanisms of miR-136-5p in HCC, several studies have demonstrated the relative results in cancers. Mosakhani *et al* (30) confirmed that miR-136 (miR-136-5p) expression was decreased in metastatic giant cell tumor of bone (GCTB) compared to non-metastatic GCTB. In the glioma, decreased miR-136 (miR-136-5p) expression was associated with high malignancy (Grade III-IV) and miR-136 (miR-136-5p) can target the oncogene (E2F transcription factor 1) to enhance chemo-sensitivity to cisplatin treatment (31). As a similar trend, in epithelial ovarian cancer, insufficiency of miR-136 (miR-136-5p) can induce chemo-resistance by attenuating apoptosis and promoting rehabilitation of DNA damage induced by cisplatin (9). In lung cancer cells, there were evidences to demonstrated that increased miR-136 (miR-136-5p) contributed to mobility and invasiveness through inhibiting epithelial-mesenchymal transition via by targeting Smad2 and Smad3 (32). *In vitro*, Zhao *et al* (11) demonstrated that miR-136 (miR-136-5p) was down-regulated by hepatitis B viral X protein in HCC and decreased miR-136-5p expression can upregulate astrocyte elevated gene-1 to promote cell migration (11). Taken together, miR-136-5p may act as an anti-oncogene role in various cancers. In this current study, concerning the clinicopathological characteristics of HCC development, such as TNM stage, portal vein tumor embolus, vaso-invasion and metastasis, the authors noticed that these factors were negatively relevant to the expression of miR-136-5p. According to the consistent expression of miR-136-5p, the similar regulative mechanisms of miR-136-5p might exist in HCC and further



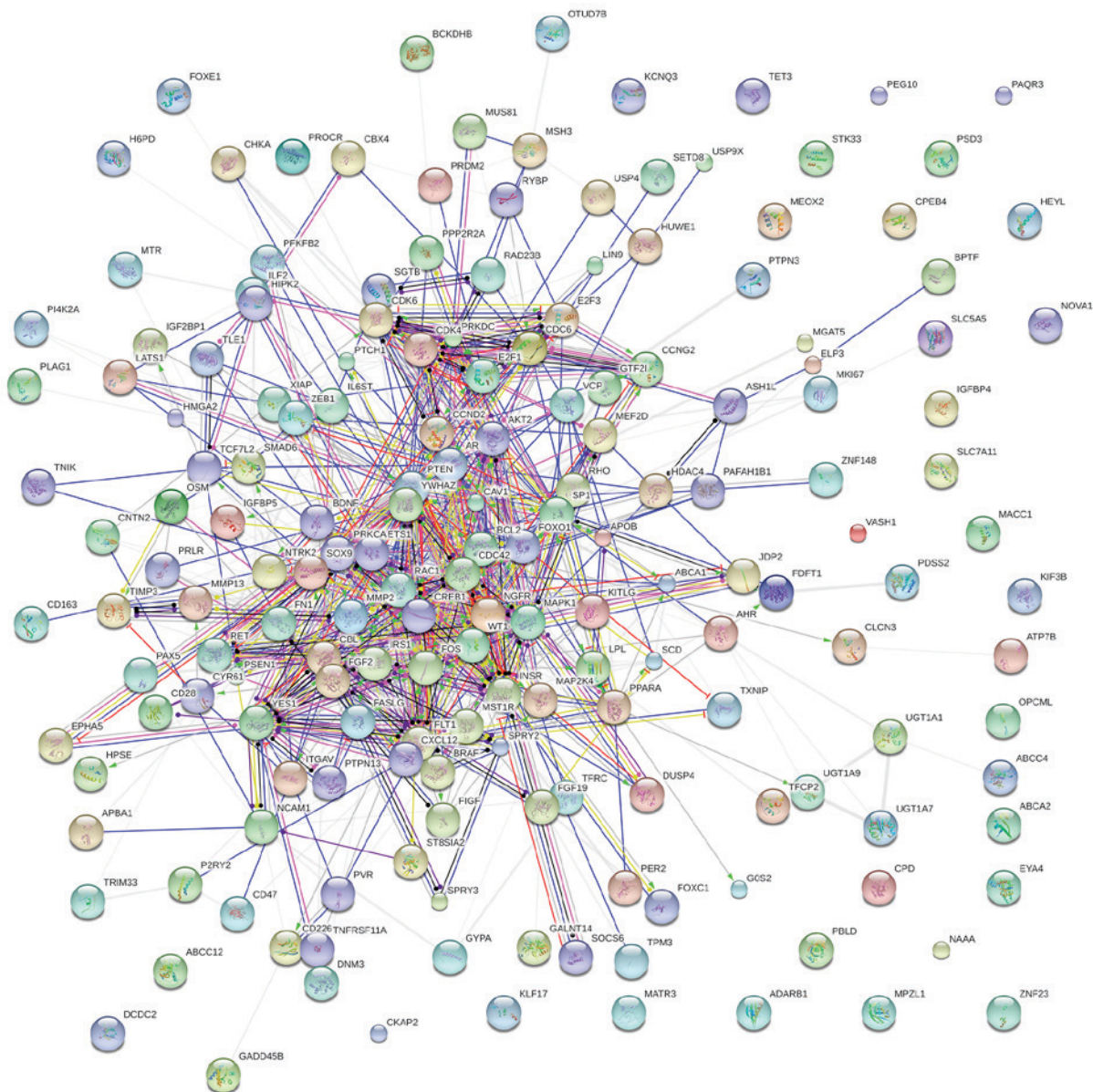


Figure 11. Protein-protein interactions of the overlapping genes between the predicted target genes of miR-136-5p and NLP analysis. The protein-to-protein network analysis of the 178 overlapping genes of predicted targets of miR-136-5p and NLP analysis was performed by the STRING website. The nodes represent for proteins, and each color corresponds to a cluster. The edges indicate the predicted functional associations, which are colored by the types of predicted associations using different evidences. The red lines, green lines, blue lines, purple lines, yellow lines, light blue lines and black lines indicate fusion evidence, neighborhood evidence, co-occurrence evidence, experimental evidence, text mining evidence, database evidence and co-expression evidence, respectively. The line thickness is an indicator for the strength of the evidence. miR, microRNA; NLP, natural language processing.

molecular experiments *in vitro* and *in vivo* are demanded for the confirmation.

To further clarify the potential mechanisms of miR-136-5p in the carcinogenesis and progression in HCC, gene functional enrichment analysis was performed. In gene functional enrichment analysis, NLP analysis was responsible for HCC-related genes and the prediction databases embodied miR-136-5p target genes. Following the combination of the two parts, the overlapping genes may be involved in the regulative mechanisms of miR-136-5p in HCC. The results of protein-protein network of the overlap showed several hub genes of the network, including BCL2, PTEN, MMP2, FOXO1. Further experiments *in vivo* and *in vitro* are needed to confirm the relationships between miR-136-5p and the hub genes in the

pathogenesis of HCC. In the GO analysis of the genes in the overlap, some target genes were correlated with positive regulation of macromolecule metabolic process, enzyme linked receptor protein signaling pathway, positive regulation of cellular biosynthetic process, transcription regulator activity, double-stranded DNA binding, transcription factor activity and structure-specific DNA binding. The relative function of the genes in the overlap mentioned above may be essential to the pathogenesis and progression of HCC (33-35). Further experiments are also necessary in order to demonstrate the function of these target genes.

In the pathway analysis, 17 significant signaling pathways of the gene in the overlap were obtained, including the MAPK and p53 signaling pathways. Among them,

several dysfunctional signaling pathways have been demonstrated to contribute to the oncogenesis and progression of HCC (36-40). Further *in vitro* and *in vivo* investigation of miR-136-5p's function in those signaling pathways in HCC is necessary to confirm and illuminate the regulative mechanisms of miR-136-5p in HCC.

In conclusion, the current study is a primary research to focus on the relationships between miR-136-5p expression and HCC tissues. miR-136-5p may serve a crucial function of the pathogenesis and development in HCC as a tumor suppressor miRNA via the regulation of transcription, macromolecule metabolic process, enzyme linked receptor protein signaling pathway, and several important pathways for carcinogenesis and progression, including the MAPK and p53 signaling pathways. Further investigations are necessary to confirm the relationships between miR-136-5p and some key hub genes (BCL2, MMP2, PTEN and FOXO1) in the pathogenesis of HCC. Thus, miR-136-5p interaction may provide a novel strategy for HCC treatment.

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