

Role of PIM2 in allergic asthma

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Abstract. T cell-associated inflammation, particularly type 2 inflammation, has an important role in asthma pathogenesis, which is suppressed by regulatory T cells (Tregs). Proviral integration site for Moloney murine leukemia virus 2 (PIM2), a member off the serine/threonine kinase family, promotes the growth and survival of T cells and influences the function of Treg cells. However, whether PIM2 affects asthma pathogenesis remains unclear. Peripheral blood mononuclear cells and Treg cells from asthmatic and healthy subjects were obtained, and the expression level of PIM2 was measured by reverse transcription-quantitative polymerase chain reaction and immunocytochemistry. In addition, BALB/c female mice sensitized and challenged by ovalbumin were used as an asthma model, and PIM2 inhibitor was injected during the challenge period to observe the effect of PIM2 on asthma. The asthma symptoms were recorded, and airway hyper-responsiveness (AHR), expression levels of cytokines in the serum or bronchoalveolar lavage fluid (BALF), and the number of BALF leukocytes were evaluated. In addition, hematoxylin and eosin staining and immunohistochemistry of lung tissues was performed. The results demonstrated that PIM2 was overexpressed in patients with asthma in natural Treg cells. Inhibition of PIM2 attenuated asthma symptoms, and improved AHR and airway inflammation compared with asthmatic mice without inhibition of PIM2. In addition, expression levels of interleukin (IL)-10 and forkhead box protein 3 (FOXP3) in BALF were increased following PIM2 inhibition (IL-10, 470.3±21.78 vs. 533.7±25.55 pg/ml, P<0.05;

FOXP3, 259±4.68 vs. 279.3±3.68 pg/ml; asthma and PIM2 inhibition groups, respectively; P<0.05). In conclusion, PIM2 may exhibit an important role in asthma pathogenesis and exacerbate AHR, airway inflammation and asthma symptoms. These effects of PIM2 may be dependent on Treg cells and the secretion of IL-10 by Tregs. The results of the present study suggest that PIM2 may be a potential target molecule for asthma treatment.

Introduction

Asthma is a chronic, heterogeneous respiratory disease characterized by persistent airway inflammation and variable expiratory airflow limitation. It affects 1-18% of the global population and inhaled corticosteroids are the primary treatment (1). However, the mechanisms underlying the disease occurrence and progression remain to be fully elucidated. Type 2 inflammation, which is specifically caused by T helper (Th)2 cells, is thought to have a central role in asthma pathogenesis, however, recently the importance of the dysregulation of other types of immune cells has also been emphasized, including Th17, Th9 and regulatory T (Treg) cells (2).

As a type of CD4⁺ T cell, Treg cells are essential for immune tolerance and the prevention of excessive inflammation. Tregs are divided into two subsets: Natural Treg cells (nTregs), which are derived from the thymus and account for the largest population of Treg cells *in vivo*; and inducible Treg cells (iTregs), which are derived from peripheral naïve T cells under a certain microenvironment (3). Forkhead box protein 3 (FOXP3) expression in both Treg cell subsets is crucial for their immune suppressive function (4). Interleukin (IL)-10 and transforming growth factor (TGF)-β1 are the major cytokines that are produced by Treg cells. These cells limit the activation of proinflammatory cytokines, downregulate T cell-mediated inflammation and inhibit the proliferation of Th2 cells (5). The development of asthma is controlled by Treg cells as they inhibit the activation of Th2 cells, prevent the entry of effector immune cells into inflamed tissue, suppress IgE production and limit Th17-mediated inflammation (3), functions that are partially dependent on the cytokines secreted by Treg cells.

The proviral integration site for Moloney murine leukemia virus 2 (PIM2) is a type of serine/threonine kinase belonging to the PIM family, and has an important function in cellular proliferation, survival and differentiation (6). The majority of

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studies concerning PIM2 focus on its participation in hematologic malignancies and solid tumors, however, a number of studies have also indicated its role in regulating the immune system (7,8). These studies demonstrated that PIM2 induced by FOXP3 is essential for Treg cell expansion and, conversely, PIM2 also inhibits the suppressive function of Treg cells by phosphorylating FOXP3, which indicates the complex roles of PIM2 in the regulation of Treg cells. As Treg cells have an important role in asthma pathogenesis, PIM2 may also influence the development of asthma inflammation, however, to the best of our knowledge, no previous studies have investigated the effect of PIM2 on asthma pathogenesis.

Therefore, the present study investigated the role of PIM2 in asthma pathogenesis, and the results demonstrated that PIM2 was overexpressed in peripheral blood mononuclear cells (PBMCs), and specifically in nTreg cells, from patients with asthma. In addition, inhibition of PIM2 in asthmatic mice alleviated airway inflammation, airway hyper-responsiveness (AHR) and associated symptoms, and these effects of PIM2 may be caused by decreased expression levels of FOXP3 and IL-10 in Treg cells.

Patients and methods

Subjects. The population consisted of patients with asthma ($n=12$; 8 males and 4 females) aged between 16 and 65 years, who were recruited from Ruijin Hospital (Shanghai, China) between January 2015 and September 2015. Healthy subjects ($n=8$; 3 male and 5 female) were included in the present study and were recruited as volunteers from Ruijin Hospital and School of Medicine, Shanghai Jiao Tong University (Shanghai, China). The diagnosis of asthma was made by the pulmonary physicians, according to the Global Initiative for Asthma guidelines (1). Subjects were excluded if they had experienced an asthma exacerbation in the previous 4 weeks or a respiratory infection 1 week prior to enrollment. All subjects gave written informed consent, and the study was approved by the Ethics Committee of Ruijin Hospital.

Sample preparation and T cell isolation. Peripheral blood (20 ml) was obtained in a sodium heparin vacuum tube. Subsequently, the blood sample was centrifuged (700 \times g for 20 min at 4°C), the supernatant was discarded and the same volume of PBS was then added. The sample was added onto the surface of lymphocyte separation medium, Ficoll-Paque PLUS (GE Healthcare Life Sciences, Little Chalfont, UK), and centrifuged at 700 \times g for 20 min at 4°C. PBMCs were collected from the middle layer and suspended at a density of 2×10^6 cells/ml in RPMI-1640 medium with GlutaMAX (Gibco; Thermo Fisher Scientific, Inc., Waltham, MA, USA) supplemented with 100 U/ml penicillin, 100 μ g/ml streptomycin and 10% heat inactivated fetal calf serum (Gibco; Thermo Fisher Scientific, Inc.). The cells were subsequently stained at 4°C for 30 min with the following fluorescently-labeled antibodies: CD4-fluorescein isothiocyanate (10 μ g/ml; 11-0049-41; eBioscience; Thermo Fisher Scientific, Inc.), CD25-PerCP-Cyanine5.5 (1.25 μ g/ml; 45-0259-42; eBioscience; Thermo Fisher Scientific, Inc.) and CD45RA-allophycocyanin (20 μ l/test; 550855; BD Biosciences, Franklin Lakes, NJ, USA) to sort for nTreg cells (CD4⁺ CD25⁺ CD45RA⁻ T cells) and naïve T cells (CD4⁺ CD25⁻ CD45RA⁺)

on a FACS ARIA II cell sorter. Prior to cell staining, Fc receptors were blocked by incubating cells with 10% normal human serum (ImmunoReagents, Inc., Raleigh, NC, USA) for 20 min at 4°C.

Induction of Th1, Th2 and iTreg cells. Naïve T cells (1×10^6 cells/ml) were stimulated with CD3 and CD28 antibody-coated beads (Dynabeads Mouse T-activator; Invitrogen; Thermo Fisher Scientific, Inc.) at a 1:1 cell-to-bead ratio for 3 days at 37°C to induce T cell differentiation. Cultures were supplemented with IL-12 (10 ng/ml), IL-2 (50 U/ml) and anti-IL-4 (10 μ g/ml; 556917) for Th1 induction, IL-4 (5,000 U/ml), IL-2 (100 U/ml), anti-interferon γ (10 μ g/ml; 554699) and anti-IL-12 (10 μ g/ml; 555065) for Th2 induction, and IL-2 (100 U/ml), TGF- β 1 (5 ng/ml), anti-IL-4 (10 μ g/ml), anti-IFN γ (10 μ g/ml) and anti-IL-12 (10 μ g/ml) for iTreg induction, at 37°C for 3 days. All antibodies and reagents were purchased from BD Biosciences.

RNA extraction and reverse transcription-quantitative polymerase chain reaction (RT-qPCR). Total RNA in total PBMCs and individual T cell subsets was isolated with TRIzol reagent (Invitrogen; Thermo Fisher Scientific, Inc.) and was reverse transcribed into cDNA with Reverse Transcription system (Promega Corporation, Madison, WI, USA), according to manufacturer's protocol. Primers were designed by Invitrogen and synthesized by BioTNT (Shanghai, China), according to the manufacturer's protocol. For amplification, a SYBR-Green I qPCR kit was used (BioTNT). The thermocycling conditions were as follows: 95°C for 5 min followed by 40 cycles of 95°C for 5 sec, 55°C for 20 sec and 72°C for 20 sec. Each reaction was performed in triplicate on the ABI ViiA 7 Real-Time PCR system (Invitrogen; Thermo Fisher Scientific, Inc.) and expression was normalized using the $2^{-\Delta\Delta C_q}$ method (9) to the expression of the housekeeping gene β -actin. The specific primers used were as follows: PIM2, 5'-ACTCCA GGTGGC CATCAAAG-3' (forward) and 5'-TCCATAGCA GTGCGA CTTTCG-3' (reverse); and β -actin, 5'-AAGGTGACA GCAGTC GGT-3' (forward) and 5'-TGTGTGGACTTGGGA GAG G-3' (reverse).

Immunocytochemistry. Human nTreg cells were obtained (1×10^4 - 1×10^5 cells/sample) as aforementioned, fixed in 4% paraformaldehyde (PFA) for 15 min at room temperature, permeabilized with 1% Triton-X-100 for 5 min at room temperature and incubated with 3% H₂O₂ for 20 min at room temperature. Cells were subsequently incubated with 5% BSA (Sigma Aldrich; Merck KGaA, Darmstadt, Germany) for blocking at room temperature for 20 min and then anti-PIM2 mouse antibody (0.5 μ g/ml; MAB4355; R&D Systems, Inc., Minneapolis, MN, USA) at 4°C overnight. Following three washes with PBS, cells were incubated with rabbit anti-mouse peroxidase secondary antibody (1:200 dilution; A9044; Sigma Aldrich; Merck KGaA) for 2 h at room temperature. Following an additional three washes with PBS, cells were stained with a 3,3'-diaminobenzidine color developing reagent kit, and nuclei were stained with 1% hematoxylin for 2 min at room temperature. Samples were examined on a Nikon Eclipse 50i microscope (Nikon Corporation, Tokyo, Japan) and the images were analyzed with Image-Pro Plus 6.0 software (Media Cybernetics, Inc., Rockville, MD, USA).

Animals and asthma model. A total of 90 Female BALB/c mice were purchased from Shanghai SLAC laboratory Co., Ltd. (Shanghai, China). Mice were aged between 6 and 8 weeks-old, weighing between 20 and 22 g, and were housed at 18-25°C, humidity 50-60%, 0.03% CO₂, 12/12 h light/dark cycle and food/water were available ad libitum and refreshed every 3 days. The mice received an intraperitoneal injection of 100 µg ovalbumin (OVA; Sigma Aldrich; Merck KGaA) and 2 mg alum (Sigma Aldrich; Merck KGaA) in PBS on days 0, 7 and 14. On days 25, 26 and 27, the mice were challenge with aerosolized 1% OVA in PBS for 30 min. Control animals received PBS intraperitoneally with alum on days 0, 7 and 14, and were challenged with aerosolized PBS on days 25, 26 and 27. The PIM2 inhibitor (5Z)-5-[[3-(Trifluoromethyl) phenyl] methylene]-2,4-thiazolidinedione (Z)-5-(3-trifluoromethylbenzylidene) thiazolidine-2,4-dione (5 µg/µl; Sigma Aldrich; Merck KGaA) was given 30 min prior to challenge with OVA by intraperitoneal injection to OVA-sensitized mice on days 25, 26 and 27.

Evaluation of AHR and asthma symptoms. AHR was assessed by measuring changes of dynamic lung compliance (C_{dyn}) in response to increasing doses (0-4 mg/ml) of acetylcholine (Ach; Shanghai Mengry Biotechnology Co., Ltd., Shanghai, China) injected into the tail intravenously in anesthetized and ventilated mice. AHR was assessed using the AniRes2005 animal lung function analysis system (Beijing Bestlab High-Tech Co., Ltd., Beijing, China) 24 h after the last OVA challenge. Asthma symptoms, including cyanosis, frequently scratching the mouth/nose/limbs (>10 scratches in 5 sec) and standing upright were observed during challenge on days 25, 26 and 27. Time to each symptom occurrence (indicated as T1, T2 and T3) were measured and symptom scores of each mouse on different days was calculated as [(30 - T1) + (30 - T2) + (30 - T3)]/3.

Bronchoalveolar lavage fluid (BALF), lung histology and immunohistochemistry (IHC). Experiments were performed 24 h after the final OVA challenge. Lungs were lavaged with 1 ml PBS through the trachea immediately following the assessment of AHR, and the number of total leukocytes was counted with a hemocytometer, and cell differentiation was performed on cytospin slides prepared with Wright-Giemsa staining to differentiate between leukocytes. Cells were seeded in PBS medium (Beyotime Institute of Biotechnology, Haimen, China) at 1x10⁵ cells/ml and stained with Fast Wright and Giemsa Stain kit (Nanjing Jiancheng Technology Co., Ltd., Nanjing, China), according to manufacturer's protocol. Percentages of each cell were counted per sample with a light microscope: Number of each cell type/every 200 cells x100%.

The left upper lobes of the lungs were fixed in 2 ml 4% PFA at 4°C for 12-24 h and embedded in paraffin. Prepared sections (4 µm) were stained with 0.5% hematoxylin (10 min) and 0.5% eosin (30 sec) at room temperature using standardized protocols and analyzed with a Nikon Eclipse 50i microscope. IHC was performed on 4 µm-PFA fixed sections. Antigen retrieval was performed with 0.01 M citric acid buffer (pH 6.0) for 15 min at 95°C prior to rehydration in descending alcohol series. Sections were subsequently blocked with 5% goat serum (Beyotime) for 20 min at room temperature. Endogenous peroxidase/phosphatase activity

was blocked with 3% hydrogen peroxide prior to incubation overnight at 4°C with anti-PIM2 (2 µg/ml; sc13514; Santa Cruz Biotechnology, Inc., Dallas, TX, USA) and anti-FOXP3 (1:500; SAB5300461; Sigma-Aldrich; Merck KGaA). Subsequently, sections were incubated for 30 min at 37°C with anti-mouse IgG-peroxidase secondary antibody (1:100; A0168; Sigma Aldrich; Merck KGaA). 3,3'-Diaminobenzidine was applied for chromogen detection. Sections were counterstained with 0.5% hematoxylin and eosin for 30 sec at room temperature and visualized with the Nikon Eclipse 50i upright microscope (magnification, x200). Lung inflammation was scored as described previously (10). Expression levels of PIM2 and FOXP3 in lung tissues were evaluated by Image-Pro Plus 6.0, and recorded as the mean optical density.

ELISA. Experiments were performed 24 h after the last OVA challenge. A total of 0.5-0.8 ml blood was drawn from each mouse. Serum was collected following centrifugation at 1,000 x g for 5 min at 4°C. Concentrations of cytokines and OVA specific IgE in the BALF and serum of mice were measured by ELISA, according to the protocols of each ELISA kit (FOXP3, MR45686; IL-4, MR63901; IL-5, MR63900; TRG-β, MR63871; IL-10, MR63912; OVA specific IgE, MR64027; Shanghai Mengry Biotechnology Co., Ltd.).

Statistical analysis. Data are presented as the mean ± standard deviation for continuous variants. All analyses were performed using SPSS (version 20; IBM Corp., Armonk, NY, USA). Student's t-test was used to determine differences between two groups. For comparisons between multiple groups, one-way analysis of variance (post hoc Tukey test) was used. Nonparametric analyses, using the Mann-Whitney U test or Kruskal-Wallis test (post hoc Dunn's test), were applied if the distributions of numerical data were not normal. Comparisons were made using Pearson's Chi-square test if the data were categorical variants. P<0.05 was considered to indicate a statistically significant difference.

Results

Baseline characteristics of subjects. Asthma patients (n=12) and healthy subjects (n=8) were included in the present study. The age of the asthma patients was 47±10 years, with no significant difference compared with healthy controls (38±9 years; P>0.05). A total of 8 patients with asthma were male, which was comparable to healthy subjects (3 males; P>0.05).

PIM2 is highly expressed in asthma patients and primarily located in Treg cells. PIM2 has been reported to be expressed ubiquitously, with the highest levels in the brain and lymphoid tissues (11). In addition, PIM2 is established to be upregulated in various hematologic malignancies, including acute or chronic myeloid leukemia, acute lymphoblastic leukemia, multiple myeloma and lymphoma (12), however, its expression pattern in human T lymphocytes is not clear. The present study isolated naïve T cells and nTreg cells from healthy human PBMCs and induced naïve T cells into Th1, Th2 and iTreg cells by different culture conditions, the results demonstrated that PIM2 was primarily expressed in Treg cells, particularly nTreg cells (Fig. 1A).

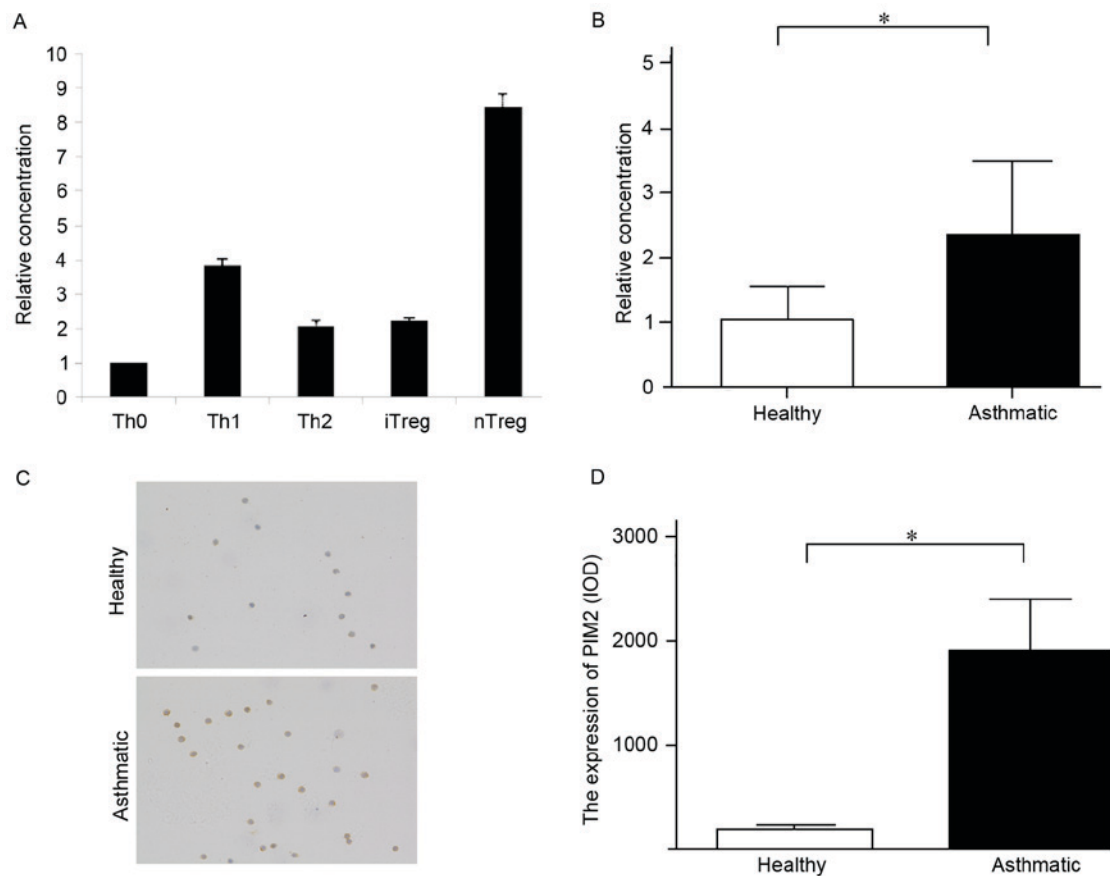


Figure 1. The expression of PIM2 in PBMCs and Treg cells from patients with asthma was higher compared with healthy subjects. (A) Expression levels of PIM2 mRNA in different types of T cells in healthy subjects. (B) Expression level of PIM2 mRNA in PBMCs. (C) Representative images (magnification, $\times 200$) of PIM2 expression in nTreg cells from patients with asthma and healthy subjects, as measured by immunocytochemistry. (D) Quantified PIM2 expression levels in Treg cells, as measured by immunocytochemistry. * $P < 0.05$, as indicated. PIM2, proviral integration site for Moloney murine leukemia virus 2; PBMCs, peripheral blood mononuclear cells; Tregs, T regulatory cells; Th, T helper cells; iTregs, inducible Tregs; nTregs, natural Tregs; IOD, integrated optical density.

To the best of our knowledge, no previous study has investigated the PIM2 expression level in patients with asthma, and the present study demonstrated that the mRNA and protein expression levels of PIM2 in PBMCs and nTregs, respectively, were higher in patients with asthma compared with healthy subjects (Fig. 1B-D), indicating that PIM2 may have an important role in asthma pathogenesis.

PIM2 is essential for airway inflammation and AHR. As PIM2 was highly expressed in patients with asthma, an asthma mouse model and PIM2 inhibitor were used to investigate the role of PIM2 in asthma pathogenesis. The mice were sensitized and challenged with OVA, and the PIM2 inhibitor was added during the challenge period. Serum levels of PIM2 were determined in control, OVA and PIM2 inhibitor mice, and levels were significantly increased in the OVA treatment group compared with the control group, while levels were significantly decreased in the OVA + PIM2 inhibitor group compared with the OVA-only group (Fig. 2A). In addition, the serum levels of OVA-specific IgE were measured, with levels significantly increased in the OVA group compared with controls. However, although PIM2 inhibitor reduced the levels compared with the OVA-only group, this reduction was not statistically significant (Fig. 2B). Furthermore, the results indicated that PIM2 inhibition improved AHR (indicated as Cdyn)

and asthma-associated symptoms significantly, compared with OVA-treated mice only (Fig. 2C and D).

Additionally, increases in BALF cell numbers induced by OVA were marginally decreased following PIM2 inhibition (Fig. 2E), and the percentages of BALF eosinophils and neutrophils were significantly reduced following PIM2 inhibition in OVA-treated mice (Fig. 2F). In addition, H&E staining of lung tissues from mouse models demonstrated that PIM2 inhibition alleviated airway inflammation compared with the OVA-only group (Fig. 3A and B). These results indicate an important role of PIM2 in inducing airway inflammation and AHR, however, the underlying mechanism was yet to be determined.

PIM2 inhibits IL-10 production by regulating FOXP3. A previous study demonstrated the role of PIM2 in regulating the function of Treg cells by mediating FOXP3 phosphorylation (8), however, its influence on FOXP3 expression and downstream cytokines, including TGF- $\beta 1$ and IL-10, in asthma mouse models has not previously been investigated. The present study demonstrated that PIM2 inhibition led to increased expression levels of FOXP3 in lung tissues (Fig. 3A and C) and BALF compared with OVA-only treated mice (Fig. 4A), indicating that PIM2 may downregulate FOXP3 expression in allergic asthma.

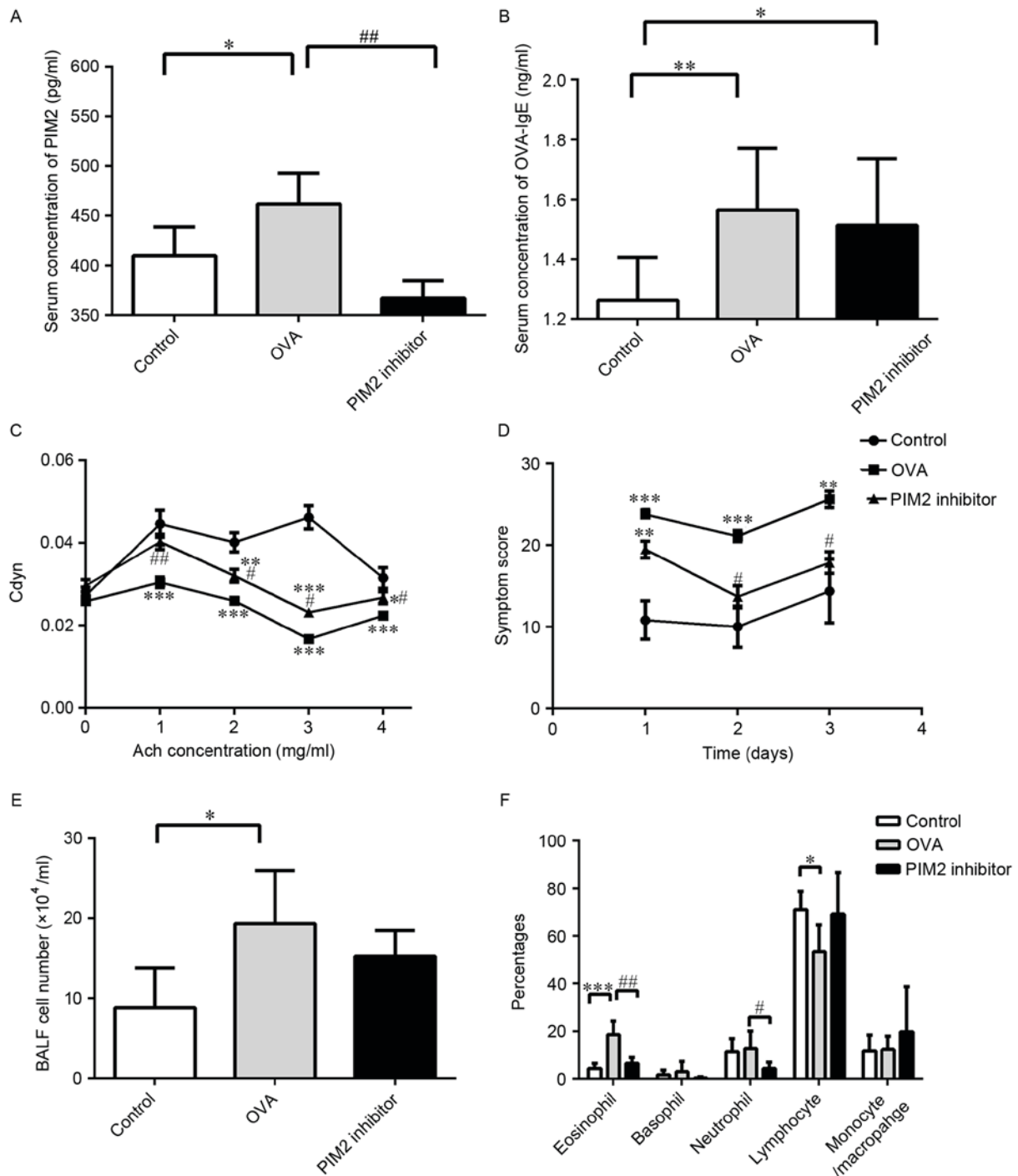


Figure 2. PIM2 inhibition resulted in attenuated asthma symptoms, improved airway hyper-responsiveness and reduced airway-infiltrated eosinophils and neutrophils in asthmatic mice. Serum concentrations of (A) PIM2 and (B) OVA-specific IgE were determined in the indicated groups. (C) Changes in lung compliance, indicated as Cdyn, in response to increased doses of Ach. (D) Asthma symptom score during the 3 challenge days. (E) Total number of leukocytes in BALF. (F) Percentages of different leukocyte types in BALF. * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$ vs. control group; # $P < 0.05$ and ## $P < 0.01$ vs. OVA group. $n \geq 5$ per group. PIM2, proviral integration site for Moloney murine leukemia virus 2; OVA, ovalbumin; Cdyn, dynamic lung compliance; Ach, acetylcholine; BALF, bronchoalveolar lavage fluid.

Additionally, BALF and serum IL-10 levels were increased following PIM2 inhibition in OVA-treated mice (Fig. 4B and C), however, the expression levels of TGF- β 1 in BALF and serum were not altered by PIM2 inhibition in asthmatic mice (Fig. 4D and E). These results indicate that PIM2 may inhibit IL-10 production, and not TGF- β 1 production, to suppress FOXP3 expression. Additionally, the expression levels of Th2 cytokines in the BALF and serum, including IL-4 and IL-5,

were not altered following PIM2 inhibition in OVA-treated mice (Fig. 5).

Discussion

PIM2 has been primarily recognized as an oncogene since its discovery, however, its role in regulating the immune system has not been extensively investigated. The present study

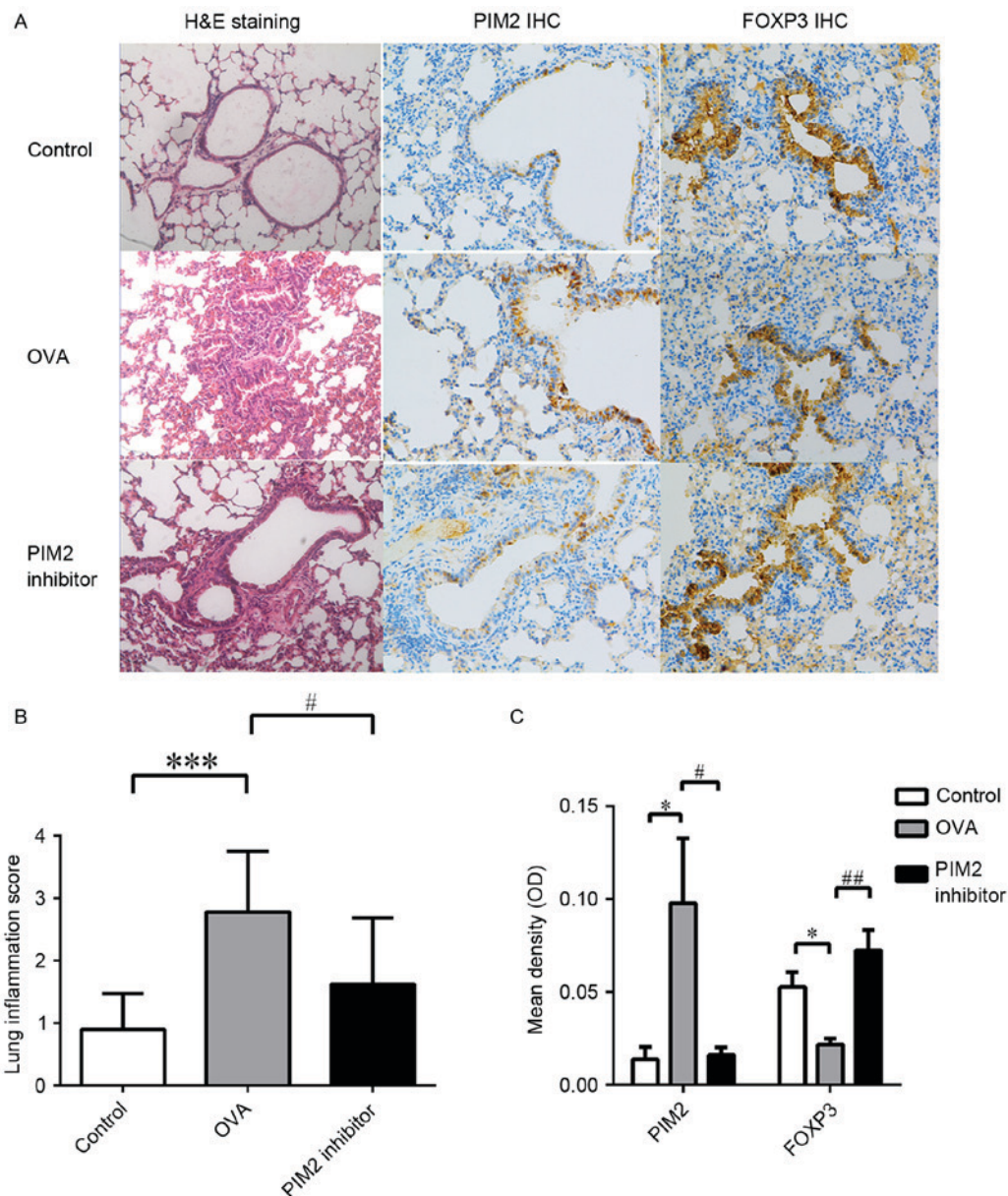


Figure 3. H&E staining and IHC of lung tissues from each group. (A) Representative images of lung tissue following H&E staining (magnification, x100) and IHC (magnification, x200) for PIM2 and FOXP3. (B) Quantitative results of H&E staining evaluating lung tissue inflammation. (C) Quantitative results of IHC for PIM2 and FOXP3. * $P < 0.05$ and *** $P < 0.001$ vs. control group; # $P < 0.05$ and ## $P < 0.01$ vs. OVA group. $n \geq 5$ per group. H&E, hematoxylin and eosin; IHC, immunohistochemistry; PIM2, proviral integration site for Moloney murine leukemia virus 2; FOXP3, forkhead box protein 3; OVA, ovalbumin; OD, optical density.

investigated its role in asthma development and demonstrated that PIM2 may have an important role in asthma pathogenesis. In patients with asthma, PIM2 expression levels were elevated, particularly in nTreg cells. Furthermore, results from an animal model of asthma further confirmed the critical role of PIM2 in promoting airway inflammation (both eosinophilic and neutrophilic) and AHR, which may occur via the regulation of FOXP3 and IL-10 expression in Treg cells.

Although no previous study has elucidated the role of PIM2 in asthma pathogenesis, the following observations have indicated the potential of a role for PIM2 in asthma: PIM2 is induced upon IL-4 release to promote T cell growth and survival, indicating its role in regulating adaptive immune system (8); it has been reported that PIM1, which is 61% homologous to PIM2 (11), has the ability to affect airway inflammation, AHR and the production of type 2 cytokines

in asthmatic mice, indicating that PIM2 may exhibit a similar effect on asthma development; PIM2 has been recognized as a molecule that may lead to mammalian target of rapamycin complex 1 (mTORC1) activation, which inhibits Treg differentiation (7,13,14); and, conversely, the inhibition of PIM2 was demonstrated to enhance the function of Treg cells *in vitro* (8). It is established that Treg cells are important regulators in excessive immune responses, such as persistent airway inflammation in patients with asthma, and these reports have indicated that PIM2 may have an important function in asthma pathogenesis by regulating the proliferation, differentiation and function of Treg cells.

Overexpression of PIM2 is commonly reported in hematologic and solid malignancies, including acute or chronic leukemia, lymphomas, and prostate and liver cancers, however, no previous study has measured its expression level in patients

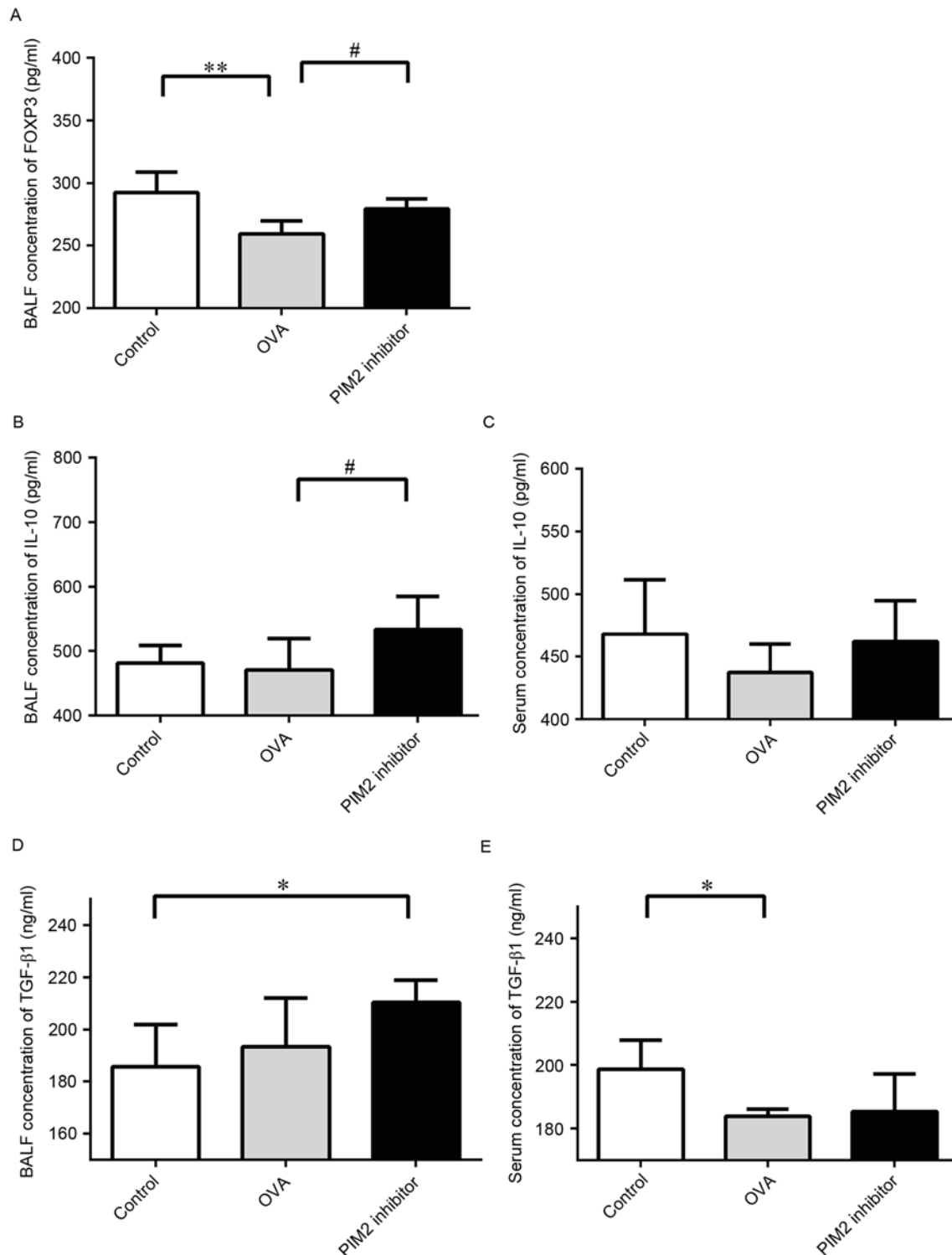


Figure 4. PIM2 inhibition increased the expression levels of FOXP3 and IL-10. (A) BALF concentrations of FOXP3 in each group. Concentration of IL-10 in (B) BALF and (C) serum in each group. Concentration of TGF-β1 in (D) BALF and (E) serum in each group. * $P < 0.05$ and ** $P < 0.01$ vs. control group; # $P < 0.05$ vs. OVA group. $n \geq 5$ per group. PIM2, proviral integration site for Moloney murine leukemia virus 2; FOXP3, forkhead box protein 3; IL, interleukin; BALF, bronchoalveolar lavage fluid; TGF, transforming growth factor; OVA, ovalbumin.

with asthma (11,15). In healthy subjects, PIM2 is constitutively expressed in lymphoid cells (16,17), and a previous report demonstrated high expression of PIM2 in nTreg cells (18). Therefore, the present study collected PBMCs from asthmatic and healthy subjects in order to measure the difference in PIM2 expression level between asthmatic and healthy subjects. The results demonstrated that PIM2 was overexpressed in patients

with asthma compared with healthy subjects in nTreg cells, which was consistent with our hypothesis that PIM2 may influence asthma development by regulating Treg cells.

To further confirm our hypothesis, animal experiments were performed. The results demonstrated that PIM2 inhibition alleviated airway inflammation and AHR, and reversed the downregulated FOXP3 and IL-10 expression levels in

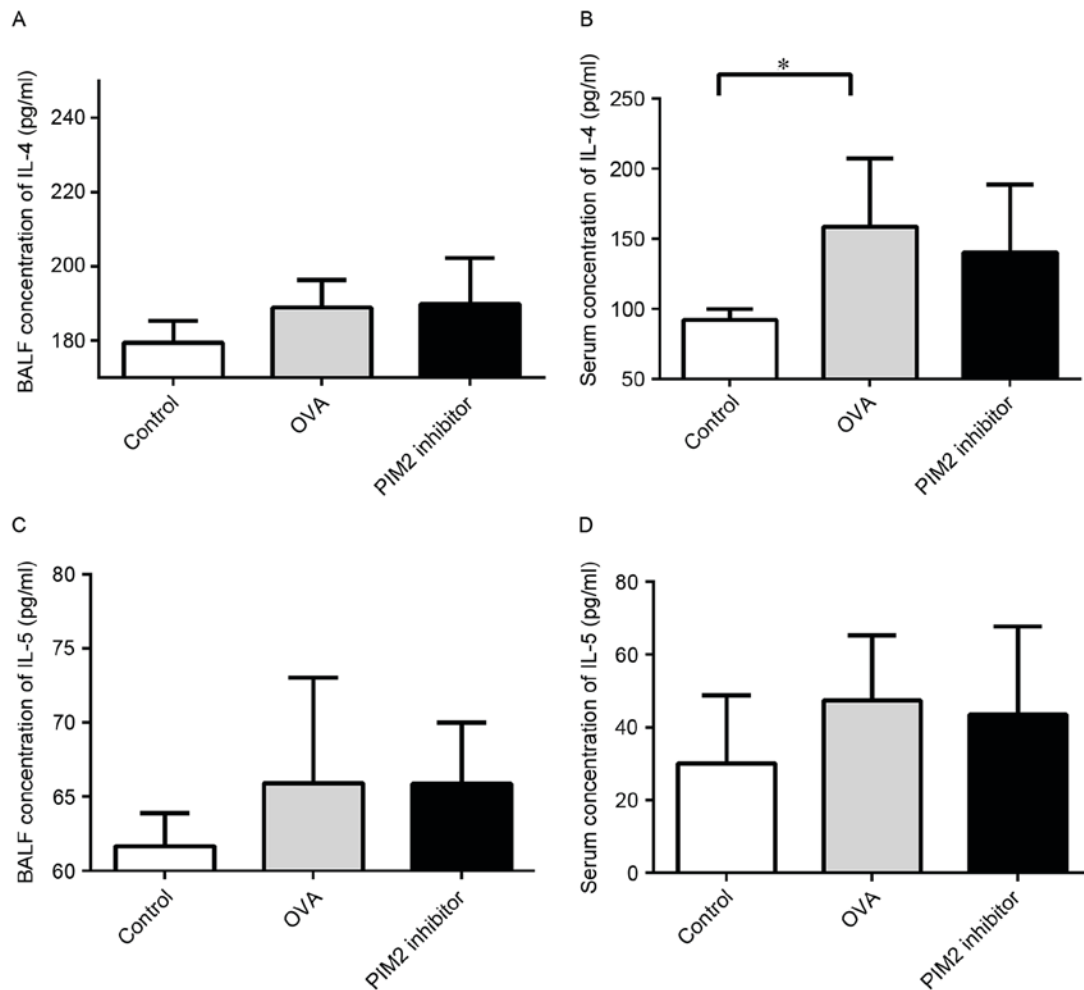


Figure 5. Expression of IL-4 and IL-5 was not affected by PIM2 inhibition in asthmatic mice. Concentration of IL-4 in (A) BALF and (B) serum in each group. Concentration of IL-5 in (C) BALF and (D) serum in each group. * $P < 0.05$ vs. control group. $n \geq 5$ per group. IL, interleukin; PIM2, proviral integration site for Moloney murine leukemia virus 2; BALF, bronchoalveolar lavage fluid; OVA, ovalbumin.

asthmatic mice. Unlike PIM1, which is associated with the survival of eosinophils, PIM2 has not been previously reported to influence eosinophil survival *in vitro* or *in vivo* (19). Therefore, the reduced percentage of eosinophils in BALF following PIM2 inhibition in asthmatic mice may be the result of enhanced Treg cell function or cytokine secretion, which may inhibit the function of type 2 cytokines, including IL-4 and IL-5, but not the expression levels of these cytokines, as PIM2 inhibition appeared to exert no direct influence on the expression levels of IL-4 and IL-5 in the present study.

Although the association between PIM2 and Treg cells is established, the molecular mechanisms underlying this association remain unclear. A previous study reported that PIM2 suppressed the function of Treg cells by phosphorylating the FOXP3 N-terminal domain (8), however, this may be only one of several mechanisms. For example, PIM2 is also reported to upregulate mTORC expression (20), which has a negative effect on Treg cell function and differentiation (4). In addition, increased expression of PIM2 was also demonstrated to be associated decreased phosphatase and tensin homolog deleted on chromosome 10, which is highly expressed in Treg cells and regulates their differentiation (4,21). The present study has demonstrated that PIM2 may influence the expression of FOXP3 and IL-10, both of which represent the function of Treg

cells, however, the precise underlying molecular mechanisms behind this phenomenon remain unclear, and further studies are required. In addition, previous studies have indicated the important role of TGF- β 1 in asthma pathogenesis and airway remodeling (22,23), however, in the current study, its expression level was increased marginally in BALF and decreased significantly in the serum of asthmatic mice compared with the control group. This result may be due to relative acute and early phase of asthma in this mouse model, in which airway remodeling was not predominant. Increased eosinophils in the airway may secrete small amounts of this cytokine leading to the increased expression of this cytokine observed in the BALF. Conversely, systemic functional deficiency of Treg cells may be responsible for the reduced secretion of TGF- β 1 observed in the serum.

In conclusion, the present study is the first to demonstrate the critical role of PIM2 in asthma pathogenesis, and this effect may be dependent on Treg cells and the secretion of IL-10 by Tregs, which suppresses excessive airway inflammation in patients with asthma. As the number of studies that have investigated the association between asthma pathogenesis and PIM2 is limited, further studies should be performed to determine the mechanisms underlying the association and any therapeutic potential of this kinase.

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