

Imperatorin possesses notable anti-inflammatory activity *in vitro* and *in vivo* through inhibition of the NF- κ B pathway

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Abstract. Imperatorin (IMT) is a furanocoumarin from the root of *Phlomis younghusbandii* (Lamiaceae) with various activities. In the present study, the anti-inflammatory effects of IMT were evaluated by examining dimethylbenzene-induced ear edema, acetic acid-induced vascular permeability and by performing cotton pellet granuloma assessments in mice. In addition, the expression of pro-inflammatory cytokines, including tumor necrosis factor (TNF)- α , interleukin (IL)-6 and IL-1 β , were detected using enzyme-linked immunosorbent assay kits in mice and using reverse transcription polymerase chain reaction analysis in RAW 264.7 cells. The expression levels of inducible nitric oxide synthase (iNOS), cyclooxygenase-2 (COX-2), nuclear p65, cytosolic p65 and inhibitor of nuclear factor (NF)- κ B (I κ B) in RAW 264.7 cells were determined using western blot analysis. The results showed that the oral administration of IMT significantly inhibited the inflammatory reactions and reduced the release of TNF- α , IL-6 and IL-1 β reactions and reduced and suppressed the mRNA expression of TNF- α expression, and the protein expression of iNOS and COX-2 in the RAW 264.7 cells. The results also indicated that IMT suppressed the activity of NF- κ B via upregulating p65 and I κ B in the cytoplasm and downregulating p65 in the nucleus. In conclusion, IMT possessed notable anti-inflammatory activities *in vitro* and *in vivo* through inhibiting the NF- κ B pathway.

Introduction

Inflammation (Latin, *inflammatio*) represents a complex biological response, which occurs in reaction to any type

of injury to the body, including damage to cells and from irritants or pathogens (1). Inflammation is a protective response, and immune cells, blood vessels and molecular mediators are involved in this process. The acute stage of inflammation is the initial response of the body to harmful stimuli and persists for a short-term in which it mediates the host defense against infections (2,3). The inflammatory response requires active termination when it is no longer required to prevent unnecessary 'bystander' damage to tissues. Failure to do so results in chronic inflammation and cellular destruction. Chronic inflammation is present in the long term and can predispose the host to various chronic illnesses, including cancer (4,5). For example, individuals who have chronic inflammatory bowel diseases are at high risk of developing colon cancer (4). Therefore, inhibiting inflammatory activity may be a target in the treatment of cancer and other diseases.

Imperatorin (IMT) is a bioactive furanocoumarin, the structure of which is shown in Fig. 1. IMT is an effective component extracted from traditional Chinese medicines and is widely distributed in the plant kingdom, particularly in *Angelica dahurica*, *A. archangelica*, *Peucedani*, *Notopterygium* and *Radix Glehniae* (6,7). It has been reported that IMT has a wide range of potent pharmacological activities, including antibacterial, antitumor, anticonvulsant, acute neurotoxic effects and abirritation (8-11). Of note, previous investigations have demonstrated that IMT possesses notable anti-inflammatory activity by inhibiting the production of nitric oxide (NO) and prostaglandin E₂ (PGE₂), and decreasing the expression of inducible nitric oxide synthase (iNOS), cyclooxygenase (COX)-2 and microsomal prostaglandin E synthase (12-14). However, few systemic investigations of the molecular mechanisms underlying the anti-inflammatory effects of IMT have been performed, which limits the clinical uses of this compound for treating inflammatory diseases. Therefore, due to the anti-inflammatory activity of IMT, the present study further evaluated the inhibitory effects of IMT against inflammation *in vitro* and *in vivo*, to understand the mechanism underlying its effects.

The present study detected the anti-inflammatory effects of IMT on dimethylbenzene-induced ear edema in mice, acetic acid-induced vascular permeability in mice and cotton pellet-induced granuloma in rats, and in LPS-induced RAW264.7 cells. In addition, the levels of tumor necrosis

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factor (TNF)- α , interleukin (IL)-6 and IL-1 β in LPS-induced endotoxemic mice and LPS-induced RAW264.7 cells were measured. The protein expression levels of iNOS, COX-2, nuclear p65 [p65 (N)], cytosolic p65 [p65 (C)] and I κ B (C) in LPS-induced RAW264.7 were also investigated to elucidate the anti-inflammatory mechanism of IMT.

Materials and methods

Chemicals. IMT was purchased from Shanghai Bomaide Biotech. Co., Ltd. (Shanghai, China); TNF- α , IL-6, and IL-1 β enzyme-linked immunosorbent assay (ELISA) kits were purchased from Invitrogen; Thermo Fisher Scientific, Inc. (Waltham, MA, USA); dimethyl benzene was purchased from the Sino Pharm. (Chengdu, China); Dulbecco's modified Eagle's medium (DMEM), fetal bovine serum (FBS) and trypsinase were from Gibco; Thermo Fisher Scientific, Inc.; the Cell Counting Kit-8, bicinchoninic acid (BCA) protein assay reagent and goat-anti-rabbit/rat horseradish-peroxidase (HRP)-conjugated secondary antibodies (cat. nos. A0208 and A0192) were purchased from Beyotime Institute of Biotechnology (Haimen, China); lipopolysaccharide (LPS), dimethyl sulfoxide (DMSO) and Evans blue were purchased from Sigma; Merck Millipore (Darmstadt, Germany); COX-2 (cat. no. sc-19999), p65 (cat. no. sc-56735), I κ B (cat. no. sc-945) and Histone H1 (cat. no. sc-8030) antibodies were purchased from Santa Cruz Biotechnology, Inc. (Santa Cruz, CA, USA); anti- β -actin antibody (cat. no. ab8226) and anti-iNOS antibody (cat. no. ab15323) were purchased from Abcam (Cambridge, MA, USA).

Animals and IMT treatment. A total of 150 Institute of Cancer Research (ICR) mice (weight, 20 ± 2 g; n=75 female; n=75 male) and 50 Sprague-Dawley rats (weight, 220 ± 20 g; n=25 female; n=25 male) were purchased from the Shanghai Laboratory Animal Centre (Shanghai, China). Each animal was housed under standard conditions ($21 \pm 1^\circ\text{C}$, 50-10% relative humidity, 12 h light/dark cycle) and had free access to food and water. The experimental protocols were approved by the Animal Care and Use Committee of The First Affiliated Hospital of Xinjiang Medical University (Urumuqi, China).

The anti-inflammatory activity of IMT *in vivo* was determined against dimethylbenzene-induced ear edema in mice, acetic acid-induced vascular permeability in mice and cotton pellet-induced granuloma in rats. A total of 50 mice or rats were randomly divided into five groups of 10 animals, including a control, positive control and three graded IMT treatments groups. IMT was administered orally at doses of 15, 30 or 60 mg/kg. The positive control group received indometacin (10 mg/kg/day) by intraperitoneal (i.p.) injection and the control group received an equal volume of vehicle (0.5% CMC-Na, 10 ml/kg/day), which had been used to dilute IMT. For the assessment of dimethylbenzene-induced ear edema and acetic acid-induced vascular permeability in mice, mice received one dose of the treatment (1 dose of 15, 30 or 60 mg/kg for IMT, 10 mg/kg for indometacin). To assess the cotton pellet-induced granuloma in rats, treatments were administered once daily for 7 consecutive days (15, 30 or 60 mg/kg/day for IMT, 10 mg/kg/day for indometacin).

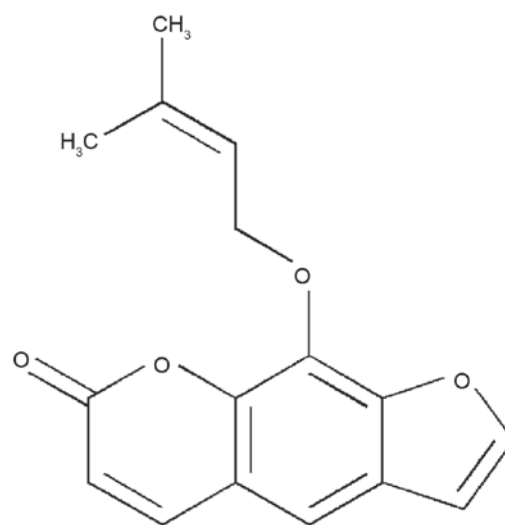


Figure 1. Chemical structure of imperatorin.

Assessment of dimethylbenzene-induced ear edema in mice.

The assessment of dimethylbenzene-induced ear edema in mice was performed according to the method described in a previous report (15). In brief, dimethyl benzene was applied locally to the right ear 1 h following final drug administration. The mice were sacrificed under anesthesia via sodium pentobarbital injection (40 mg/kg i.p.) 1 h following dimethylbenzene application. The right ear and left ear were then amputated at the same location. Finally, the ear edema was calculated by subtracting the weight of the left ear from that of the right ear.

Assessment of acetic acid-induced vascular permeability in mice.

The acetic acid-induced vascular permeability assessment in mice was performed according to a previously reported method with modification (15). At 1 h following final drug administration, each mouse was injected intravenously with Evans blue (2% in normal saline; 10 ml/kg) and was injected abdominally with acetic acid (0.7% in saline; 10 ml/kg). Following injection of acetic acid for 20 min, all the mice were sacrificed by cervical dislocation. The abdominal cavity was then washed several times with 5 ml of saline solution per mouse. The rinsed solution was collected and centrifuged for 15 min ($780 \times g$ at 4°C). Finally, the absorbance of Evans blue in the supernatant was determined at 630 nm with a spectrophotometer (Model 721; Shanghai Optical Instrument Factory Co., Ltd., Shanghai, China).

Assessment of cotton pellet-induced granuloma in rats.

The assessment of cotton pellet-induced granuloma in rats was performed to evaluate the chronic anti-inflammatory activity of IMT and was performed as described previously (16). In brief, a small section of sterile cotton pellet (50 ± 1 mg) was applied subcutaneously to the dorsolateral skin of the anesthetized rats (one on either side), and drugs were administered once daily for 7 days consecutively. On day 8, all rats were sacrificed and the pellets with granuloma were excised. The increments of the wet and dry weights of the pellets were used to evaluate granuloma formation.

Measurement of levels of TNF- α , IL-6 and IL-1 β in LPS-induced endotoxemic mice. The mice were injected with IMT (15, 30 and 60 mg/kg) for 24 h in the presence of LPS (2 mg/kg). Subsequently, the mice were sacrificed and serum was collected to measure blood levels of TNF- α , IL-6 and IL-1 β using ELISA kits, according to the manufacturer's protocols.

Cell culture. The murine macrophage RAW264.7 cell line was purchased from the Shanghai Cell Bank of the Chinese Academy of Sciences (Shanghai, China), and the cells were cultured in DMEM with 10% FBS, 1% penicillin and 1% streptomycin in a 5% CO₂ humidified atmosphere at 37°C.

Analysis of cell viability using a Cell Counting kit-8 (CCK-8 assay). The effects of IMT on cellular viability were evaluated using a CCK-8 assay according to the manufacturer's protocol. The RAW264.7 cells (5x10³ cells/well) were seeded into 96-well plates and incubated with either increasing concentrations of IMT (18.5, 37, 74, 148, 296 and 592 μ M) or vehicle (DMSO) in the presence of LPS (100 ng/ml). Following treatment for 72 h, CCK-8 solution was added to each well and incubated for another 1 h in the incubator. The absorbance at 450 nm was then read using a 96-well plate reader (Bio-Rad Laboratories, Inc., Hercules, CA, USA). The results were determined as a percentage of the DMSO control cells.

Quantitative analysis of the mRNA expression levels of TNF- α , IL-6 and IL-1 β in RAW 264.7 cells. The effects of IMT on the mRNA expression levels of TNF- α , IL-6 and IL-1 β were determined using reverse transcription-quantitative polymerase chain reaction (RT-qPCR) analysis. The RAW264.7 cells at a density of 3x10⁵ cells/well were seeded into 6-well plates and incubated with either increasing concentrations of IMT (55.5, 111 and 222 μ M) or vehicle (DMSO) in the presence of 100 ng/ml LPS for 72 h at 37°C. Subsequently, the cells were collected and total RNA was extracted using an RNAiso Plus kit (Takara Biotechnology Co., Ltd., Dalian, Japan). The total RNA was then used to synthesize cDNA of TNF- α , IL-6, IL-1 β and β -actin using PrimeScript™ RT reagent kits (Takara Biotechnology Co., Ltd.). The reverse transcription temperature protocol was as follows: 37°C for 15 min and 85°C for 5 sec. The synthesized cDNAs were amplified using SYBR Green mix on a CFX96 Touch Real-Time PCR detection system (Bio-Rad Laboratories, Inc.). Each 10- μ l reaction mixture contained 5 μ l of SYBR Premix (Bio-Rad Laboratories, Inc.), 1 μ l of 2 μ M forward and reverse primer mix, 1 μ l of cDNA, and 3 μ l of ddH₂O. The PCR thermocycling conditions were as follows: 95°C for 10 min, followed by 39 cycles of 95°C for 15 sec and 60°C for 60 sec. All primers used for RT-qPCR analysis are shown in Table I. The relative mRNA expression levels of TNF- α , IL-6 and IL-1 β were assessed via 2^{- $\Delta\Delta C_q$} relative quantitative analysis (17) and all samples were analyzed in triplicate.

Western blot analysis. The effects of IMT on the protein expression levels of iNOS, COX-2, p65 (N), p65 (C) and I κ B (C) were measured in the LPS-induced RAW264.7 cells using western blot analysis. Following treatment with either graded concentrations of IMT (55.5, 111 and 222 μ M) or

Table I. Primers used for reverse transcription-quantitative polymerase chain reaction analysis.

Gene	Primer sequence (5'-3')
TNF- α	Forward: CAGGTTCTGTCCCTTTCACTCACT Reverse: GTTCAGTAGACAGAAGAGCGTGGT
IL-6	Forward: TGGAGTACCATAGCTACCTGGAGT Reverse: TCCT-TAGCCACTCCTTCTGTGACT
IL-1 β	Forward: ATGGCAACTGTTCTCTGAAGT Reverse: TTAGGAAGACACGGATTCCAT
β -actin	Forward: GGGAAATCGTGCGTGACATCAAAG Reverse: CATACCCAAGAAGGAAGGCTGGAA

TNF- α , tumor necrosis factor- α ; IL, interleukin.

vehicle (DMSO) in the presence of 100 ng/ml LPS for 72 h, the cells were collected and total protein was extracted using western blot and IP cell lysis buffer (Sangon Biotech Co., Ltd.). Subsequently, the concentration of total protein was determined using BCA protein assay reagent (Sangon Biotech Co., Ltd.). The total protein (40 μ g) in each sample was separated by 12% sodium dodecyl sulfate-polyacrylamide electrophoresis (SDS-PAGE) and blotted onto polyvinylidene difluoride (PVDF) filter membranes. The proteins on the PVDF membrane were then probed with anti-iNOS (1:1,000), anti-COX-2 (1:200), anti-p65 (1:200), anti-cytosolic I κ B (1:200), anti- β -actin (1:5,000) and anti-Histone H1 (1:500) antibodies at 4°C for 12 h, followed by incubation with corresponding HRP-conjugated secondary antibodies (1:7,000) for 2 h at 37°C. Finally, the immunoreactive bands were visualized using ECL-detecting reagents and optical density (OD) values were analyzed using Image J 2X software. To normalize for protein loading, the expression levels of iNOS, COX-2, p65 (C) and I κ B (C) were expressed as a relative value to that of β -actin, and the expression level of p65 (N) was expressed as a relative value to that of Histone H1.

Statistical analysis. All results are expressed as the mean \pm standard deviation of three independent experiments performed in triplicate. Statistical analyses were performed using the SPSS 19.0 software package (IBM SPSS, Armonk, NY, USA). One-way analysis of variance with Dunnett's test was used to compare the means between two groups. P<0.05 was considered to indicate a statistically significance difference.

Results

Assessment of dimethylbenzene-induced ear edema. The local and acute anti-inflammatory activities of IMT were evaluated by measuring its inhibition of dimethylbenzene-induced ear edema in mice. As shown in Fig. 2A, IMT had significant and dose-dependent inhibitory effects on ear edema at concentrations of 15, 30 and 60 mg/kg, compared with the control group (P<0.05, P<0.01 and P<0.01, respectively). At a dose of 60 mg/kg, the anti-inflammatory activity of IMT was comparable with that of indometacin at a dose of 10 mg/kg.

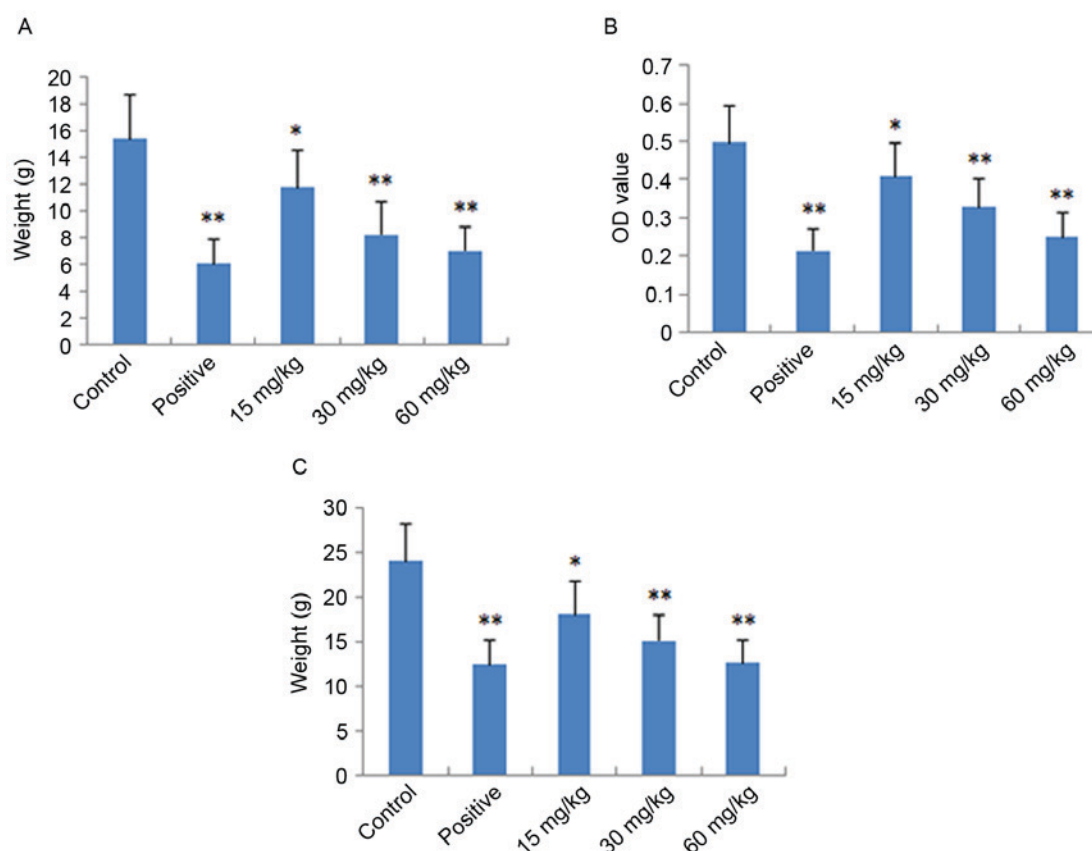


Figure 2. Anti-inflammatory effects of imperatorin on mice/rats. Indometacin (10 mg/kg) was used as the positive control drug. (A) Results of dimethylbenzene-induced ear edema assessment in mice; (B) results of acetic acid-induced vascular permeability assessment in mice; (C) results of cotton pellet-induced granuloma assessment in rats. Data are expressed as the mean \pm standard deviation (n=10). *P<0.05 and **P<0.01, compared with the control group. OD, optical density.

Assessment of acetic acid-induced vascular permeability. The acute anti-inflammatory activity of IMT was also detected by measuring its inhibition of acetic acid-induced vascular permeability in mice. As shown in Fig. 2B, IMT at doses of 15, 30 and 60 mg/kg significantly inhibited the increased vascular permeability induced by acetic acid in mice, which also occurred in a dose-dependent manner (P<0.05, P<0.01 and P<0.01, respectively), compared with the control group. The inhibitory effect of IMT on dye leakage at a dose of 60 mg/kg was comparable with that of indometacin at the dose of 10 mg/kg.

Cotton granuloma assessment. The chronic anti-inflammatory activity of IMT was performed by measuring its inhibition of cotton pellet-induced granuloma in rats, the results of which are shown in Fig. 2C. The anti-inflammatory effect of IMT was observed at the doses of 15, 30 and 60 mg/kg. Compared with the control group, IMT significantly reduced ball fralunoma weight at doses of 15, 30 and 60 mg/kg (P<0.05, P<0.01 and P<0.01, respectively). The positive control drug, indometacin (10 mg/kg), also manifested significant anti-inflammatory activity (P<0.01).

Effects of IMT on LPS-induced cell viability. The effects of IMT on LPS-induced cell viability were evaluated using CCK-8. As shown in Fig. 3, no significant effect was observed when the RAW 264.7 cells were treated with graded concentrations of

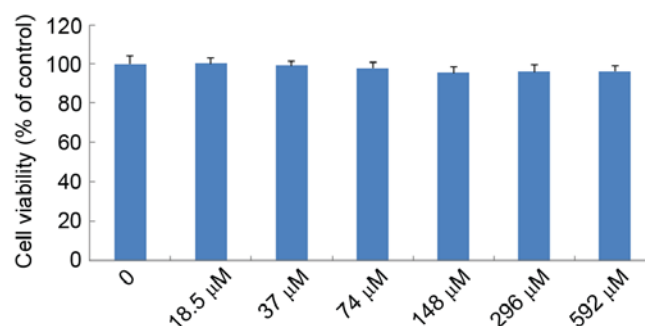


Figure 3. Effects of IMT on cell viability of RAW 264.7 cells. Following treatment with either increasing concentrations of IMT (18.5, 37, 74, 148, 296 and 592 μ M) or vehicle (DMSO, negative control) for 72 h, cell viability was determined using a Cell Counting Kit-8 assay. Data are expressed as the mean \pm standard deviation (n=6). IMT, imperatorin.

IMT (18.5-592 μ M) in the presence of 100 ng/ml LPS for 72 h, compared with the control group (P>0.05). This indicated that IMT had no inhibitory effect on RAW 264.7 cell growth.

Effects of IMT on TNF- α , IL-6 and IL-1 β in vitro and in vivo. To further verify the anti-inflammatory effects of IMT *in vitro* and *in vivo*, the expression levels of pro-inflammatory cytokines (TNF- α , IL-6 and IL-1 β) were determined in LPS-induced endotoxemic mice and RAW 264.7 cells using ELISA and RT-qPCR analysis, respectively. As shown in Fig. 4, following

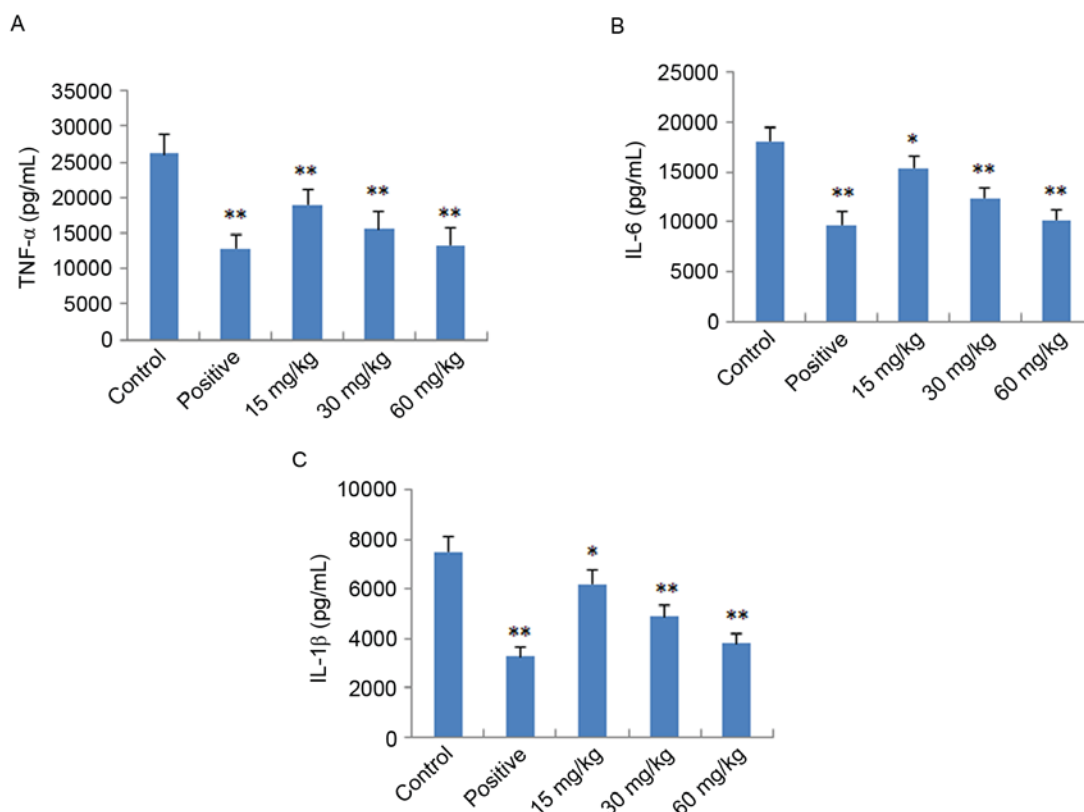


Figure 4. Effect of imperatorin on plasma levels of (A) TNF- α , (B) IL-6 and (C) IL-1 β in lipopolysaccharide-induced endotoxemic mice. Indometacin (10 mg/kg) was used as the positive control drug. Data are expressed as the mean \pm standard deviation (n=10). *P<0.05 and **P<0.01, compared with the control group. TNF- α , tumor necrosis factor- α ; IL, interleukin.

treatment with IMT (55.5, 111 and 222 μ M), the levels of TNF- α , IL-6 and IL-1 β induced by LPS were significantly reduced, compared with those in the LPS group (P<0.05) in the LPS-induced endotoxemic mice. In the LPS-induced RAW 264.7 cells, the mRNA expression levels of these pro-inflammatory cytokines were also significantly downregulated, compared with those in the LPS group (P<0.01; Fig. 5).

Protein expression levels of iNOS, COX-2, p65 and I κ B. According to the results described above, IMT not only inhibited topical ear edema, vascular permeability and granuloma *in vivo*, but also suppressed the expression levels of pro-inflammatory cytokines TNF- α , IL-6 and IL-1 β *in vitro* and *in vivo*. To further investigate the possible mechanisms underlying these changes, the protein expression levels of iNOS, COX-2, p65 (N), p65 (C) and I κ B (C) were measured using western blot analysis. As shown in Figs. 6 and 7, compared with the LPS group, the protein expression levels of iNOS, COX-2 and p65 (N) were markedly downregulated in the IMT groups, whereas the protein expression of p65 (C) and I κ B (C) were significantly upregulated in the LPS-induced RAW 264.7 cells.

Discussion

In the present study, comprehensive analyses of the acute and chronic anti-inflammatory activities of IMT were performed using mouse and rat models of inflammation. In addition, the *in vitro* anti-inflammatory effect of IMT was demonstrated using LPS-stimulated RAW264.7 macrophages.

IMT significantly inhibited the levels of pro-inflammatory cytokines TNF- α , IL-6 and IL-1 β *in vitro* and *in vivo*. The expression levels of pro-inflammatory enzymes COX-2 and iNOS were downregulated in the LPS-stimulated RAW264.7 macrophages treated with IMT.

The anti-inflammatory effect of IMT was evaluated using three popular *in vivo* models: Dimethylbenzene-induced ear edema, acetic acid-induced vascular permeability, and cotton pellet-induced granuloma (18,19). The assessment of dimethylbenzene-induced ear edema in mice is a useful model for preliminary experiments of acute anti-inflammatory activity. In the inflammatory response, there is increased vascular permeability; therefore, vascular permeability was assessed to further demonstrate the anti-inflammatory effects of IMT. In addition, cotton pellet-induced granuloma was assessed to evaluate the chronic anti-inflammatory activity. The results of the present study showed that IMT at doses of 15, 30 and 60 mg/kg led to topic inhibition of dimethylbenzene-induced ear edema in mice in a dose-dependent manner. At the same doses, IMT exhibited similar inhibitory effects on the increased vascular permeability of mice. IMT also significantly and dose-dependently reduced the weights of the cotton pellet granuloma in the cotton pellet-induced model of chronic inflammation in rats. These data suggested that IMT had potential anti-inflammatory activity, therefore; the effect of IMT on the release of inflammatory cytokines and mediators was also examined to elucidate its anti-inflammatory mechanism.

Inflammation is a local response to irritants, tissue injury and infection. During inflammation, circulating

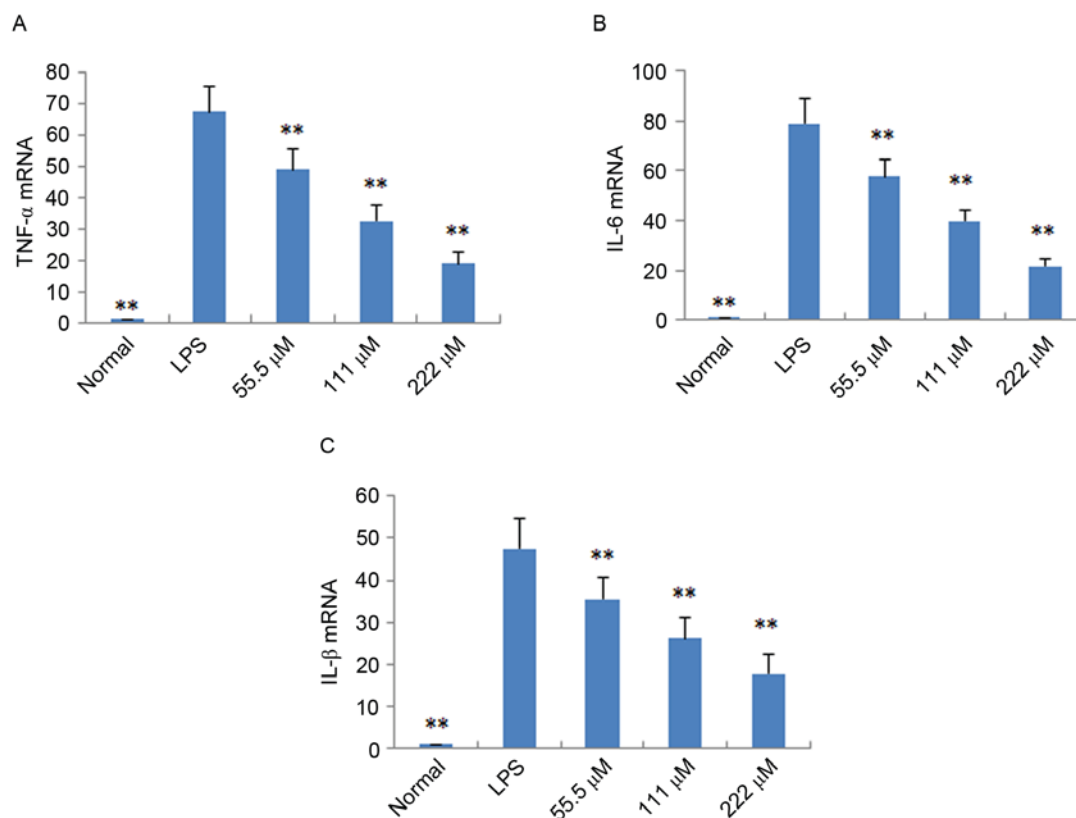


Figure 5. Effect of imperatorin on (A) TNF- α , (B) IL-6 and (C) IL-1 β in RAW 264.7 cells induced by LPS. Data are expressed as the mean \pm standard deviation (n=10). **P<0.01, compared with the control group. LPS, lipopolysaccharide; TNF- α , tumor necrosis factor- α ; IL, interleukin.

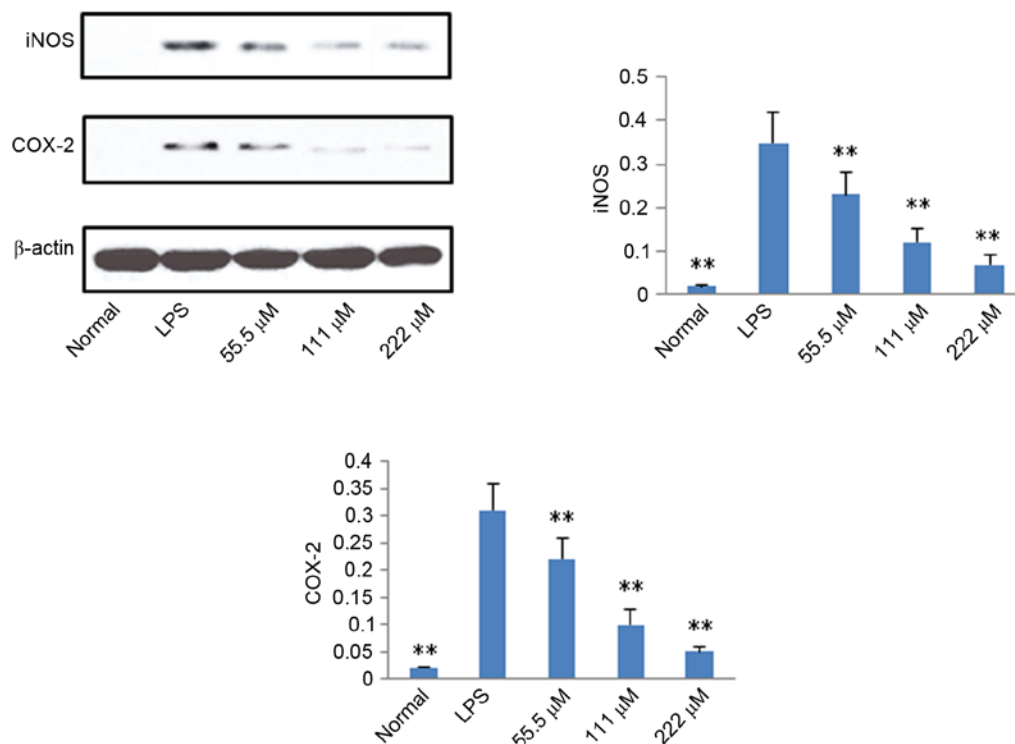


Figure 6. Effects of imperatorin on expression levels of iNOS and COX-2 in RAW 264.7 cells induced by LPS. Data are expressed as the mean \pm standard deviation (n=10). **P<0.01, compared with the control group. LPS, lipopolysaccharide; iNOS, inducible nitric oxide synthase; COX-2, cyclooxygenase-2.

macrophages, mast cells and neutrophils produce cytokines, including TNF- α , IL-6 and IL-1 β , which further exaggerate

inflammatory responses (20). TNF- α , IL-6 and IL-1 β are important in mediating acute inflammatory reactions. IL-1 β

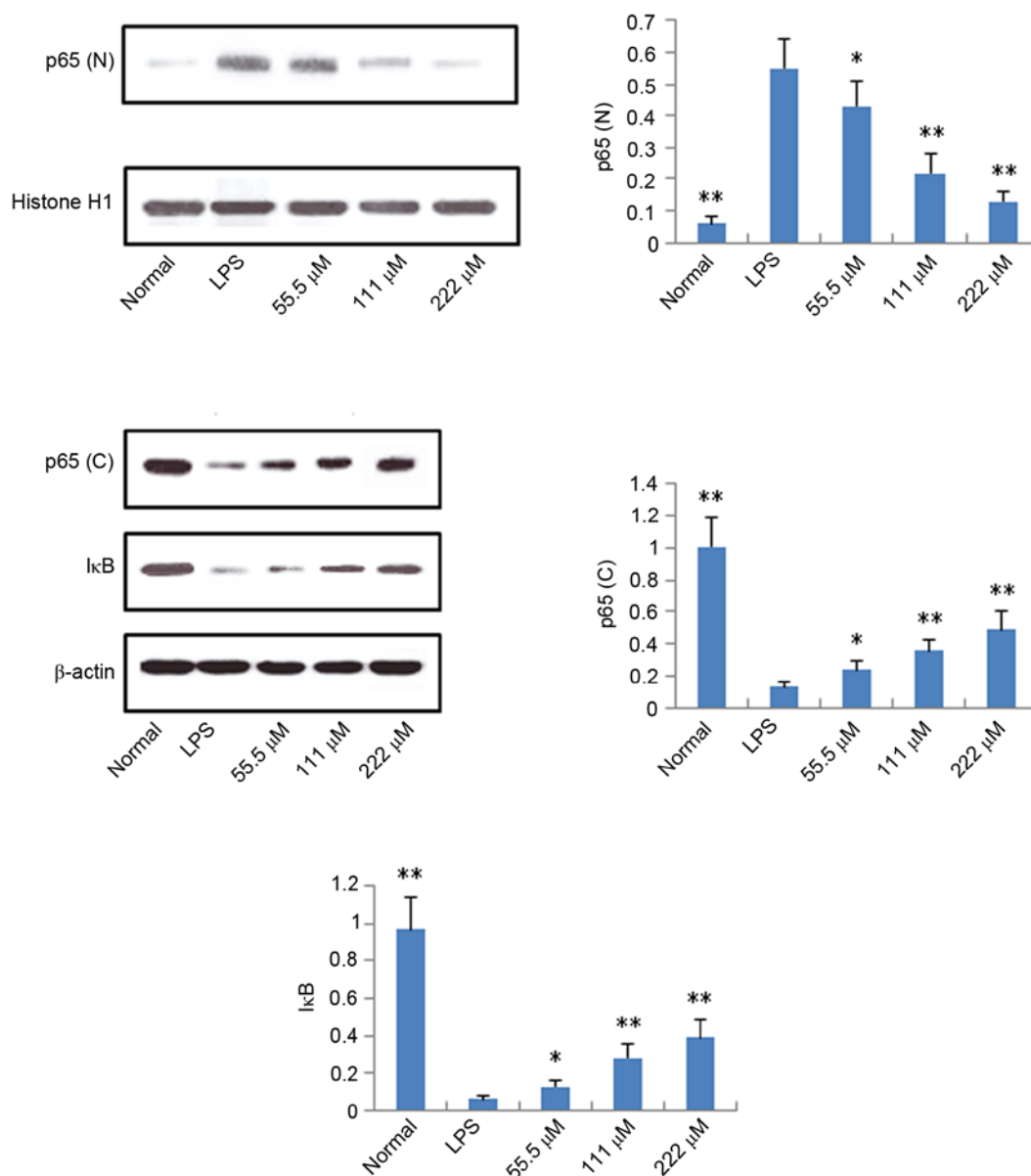


Figure 7. Effects of imperatorin on expression levels of p65 (N), p65 (C) and I κ B (C) in RAW 264.7 cells induced by LPS. Data are expressed as the mean \pm standard deviation (n=10). *P<0.05 and **P<0.01, compared with the control group. LPS, lipopolysaccharide; p65 (N), nuclear p65; p65 (C), cytosolic p65; I κ B, inhibitor of nuclear factor- κ B.

can induce fever through enhancing the synthesis of PGE₂. As with IL-1, TNF- α can trigger fever, either directly by inducing PGE₂ synthesis or indirectly by stimulating the release of IL-1. Subsequently, TNF- α and IL-1 trigger secondary inflammatory effects by inducing the synthesis of IL-6 (21). Therefore, inhibition of these pro-inflammatory cytokines is an important treatment strategy for inflammatory diseases. In the present study, IMT suppressed the production of TNF- α , IL-6 and IL-1 β in Raw264.7 cells and LPS-induced endotoxemic mice in a dose-dependent manner. These results indicated that IMT in LPS-stimulated macrophages and LPS-stimulated mice exerted anti-inflammatory effects by inhibiting the expression and secretion of pro-inflammatory cytokines.

In addition to cytokines, the inflammatory response induced by inflammatory mediators is generated through the upregulation of several inducible genes, including iNOS and COX-2. iNOS is expressed in macrophages under normal and

pathological conditions. In infectious and with pro-inflammatory stimuli, a high protein expression level of iNOS is induced to produce NO, which is an important messenger molecule with a critical function in the host immune defence (22). COX is also a molecular target for anti-inflammatory treatments, which have been used for hundreds of years; for example, aspirin, a selective COX-2 inhibitor and nonsteroidal anti-inflammatory drug, has been used globally since 1899 (23). It is well known that the COX enzyme consists of at least two isoforms, COX-1 and COX-2. In humans, COX-1 protein is expressed at relatively stable levels in several normal tissues, whereas the protein expression of COX-2 is low in the majority of normal mammalian tissues in response to stimuli, including LPS insult. In macrophages, the LPS-induced induction and activation of signal transduction pathways results in activation of the gene expression of the COX-2 (22). In the present study, LPS significantly elevated the protein expression levels

of iNOS and COX-2 in macrophages, and these levels were markedly suppressed by IMT in a dose-dependent manner. These results further supported the ability of IMT to inhibit inflammation, and this inhibitory effect may be associated with suppressing the production of inflammatory mediators iNOS and COX-2.

It is well known that NF- κ B is a major regulator of pathogen- and inflammatory cytokine-inducible gene regulation (24). Several stimuli, including pro-inflammatory cytokines, can activate NF- κ B, and activated NF- κ B can induce the protein expression of pro-inflammatory cytokines, COX-2 and iNOS (25). NF- κ B is a family of inducible dimeric transcription factors, which includes five members: Rel (c-Rel), RelA (p65), RelB, NF- κ B1 (p50/p105) and NF- κ B2 (p52/p100) (26). In unstimulated cells, the NF- κ B family members (homodimers or heterodimers) are bound to ankyrin rich regions of I κ B, an inhibitor of NF- κ B protein, which serves to retain NF- κ B dimers in the cytoplasm and thus inhibiting initiation of target gene transcription (26). As I κ B dissociates from NF- κ B, activated NF- κ B is translocated to the nucleus and exerts its function as a transcription factor. In the present study, it was found that, following LPS stimulation of RAW 264.7 cells, the protein expression of p65 (N) was higher, compared with that of the control group, whereas the protein expression levels of p65 (C) and I κ B (C) were significantly downregulated, which indicated that LPS led to the activation of NF- κ B. IMT significantly increased the protein expression levels of p65 and I κ B in the cytoplasm of LPS-triggered RAW 264.7 cells, and decreased the protein expression of p65 (N). These data indicated that IMT inhibited the LPS-induced activity of NF- κ B in macrophages, and then suppressed the production of pro-inflammatory cytokines and mediators.

In conclusion, IMT exhibited significant anti-inflammatory effects *in vitro* and *in vivo*. IMT reduced the release of proinflammatory cytokines (TNF- α , IL-6 and IL-1 β), inhibited the expression of inducible enzymes (iNOS and COX-2), and suppressed the activity of NF- κ B via upregulation of the expression of p65 (C) and I κ B (C), and downregulation of the expression of p65 (N).

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