

Role of catalpol in ameliorating the pathogenesis of experimental autoimmune encephalomyelitis by increasing the level of noradrenaline in the locus coeruleus

QIAN LI¹, TAO YANG¹, AN-CHEN GUO²⁻⁵ and YONG-PING FAN¹

¹Department of Chinese Medicine; ²Laboratory of Clinical Medical Research; ³Beijing Key Laboratory of Translational Medicine for Cerebrovascular Diseases; ⁴China National Clinical Research Center for Neurological Diseases; ⁵Beijing Institute for Brain Disorders, Beijing Tiantan Hospital, Capital Medical University, Beijing 100050, P.R. China

Received February 26, 2016; Accepted February 10, 2017

DOI: 10.3892/mmr.2018.8378

Abstract. The endogenous neurotransmitter, noradrenaline, exerts anti-inflammatory and neuroprotective effects *in vivo* and *in vitro*. Reduced noradrenaline levels results in increased inflammation and neuronal damage. The primary source of noradrenaline in the central nervous system is tyrosine hydroxylase (TH)-positive neurons, located in the locus coeruleus (LC). TH is the rate-limiting enzyme for noradrenaline synthesis; therefore, regulation of TH protein expression and intrinsic enzyme activity represents the central means for controlling the synthesis of noradrenaline. Catalpol is an iridoid glycoside purified from *Rehmannia glutinosa* Libosch, which exerts a neuroprotective effect in multiple sclerosis (MS). The present study used an experimental mouse model of autoimmune encephalomyelitis to verify the neuroprotective effects of catalpol. Significant improvements in the clinical scores were observed in catalpol-treated mice. Furthermore, catalpol increased TH expression and increased noradrenaline levels in the spinal cord. In primary cultures, catalpol exerted a neuroprotective effect in rat LC neurons by increasing the noradrenaline output. These results suggested that drugs targeting LC survival and function, including catalpol, may be able to benefit patients with MS.

Introduction

Multiple sclerosis (MS) is a chronic inflammatory disease of the central nervous system (CNS), which is characterized by

widespread regions of focal demyelination, loss of oligodendrocytes and axonal degeneration (1). It affects 0.1% people in the western world and results in chronic disability, primarily in young adults (2). The etiology and pathogenesis of the disease, however, remain unclear. Currently, MS is believed to be a complex multifactorial disease, involving genetic and environmental factors affecting the autoimmune responses that damage the myelin (3). Increasing evidence suggests that abnormalities of the locus coeruleus noradrenaline (LC-NA) system may be a contributing factor for myelin damage. A magnetic resonance study demonstrated an impairment of selective attention in MS, which was increased with axonal damage at the right LC (4). In mice immunized with myelin oligodendrocyte glycoprotein peptide MOG₃₅₋₅₅, cortical and spinal cord levels of noradrenaline were significantly reduced compared with control mice (5). The immunized mice additionally exhibited increased astrocyte activation and neuronal damage in the LC (5). The LC-selective neurotoxin, N-(2-chloroethyl)-N-ethyl-2-bromobenzylamine (DSP-4), (6) significantly increases the clinical severity of experimental autoimmune encephalomyelitis (EAE) in mice (7), which suggests that the functional status of LC influences EAE disease. Numerous studies have demonstrated that noradrenaline levels are altered in the CNS of patients with multiple sclerosis, and that noradrenaline levels are reduced in the brainstem and spinal cord of rats with EAE (5,8,9).

The endogenous neurotransmitter, noradrenaline, exerts anti-inflammatory and neuroprotective effects *in vivo* and *in vitro* (10,11). Noradrenaline acting at β 2-adrenergic receptors reduces the expression of numerous genes involved in inflammation, including those encoding adhesion molecules, chemokines, major histocompatibility class II molecules, inducible nitric oxide synthase (iNOS)2, interleukin (IL)-1 β and tumor necrosis factor (TNF)- α via the activation of cyclic AMP-signaling pathways in astrocytes (12-16). Another important target of noradrenaline signaling in the CNS are microglia, which are often considered to be the resident immune cells of the CNS. Noradrenaline regulates the three key microglia actions: Migration, proliferation and phagocytosis, via activation of the β 2-adrenergic receptor (17-19). As in astrocytes, noradrenaline reduces the expression of

Correspondence to: Dr An-Chen Guo, Laboratory of Clinical Medical Research, Beijing Tiantan Hospital, Capital Medical University, 6 Tiantan Xili, Dongcheng, Beijing 100050, P.R. China
E-mail: guoanchenphd@163.com

Dr Yong-Ping Fan, Department of Chinese Medicine, Beijing Tiantan Hospital, Capital Medical University, 6 Tiantan Xili, Dongcheng, Beijing 100050, P.R. China
E-mail: fanyaping@yeah.net

Key words: catalpol, experimental autoimmune encephalomyelitis, locus coeruleus, noradrenaline, tyrosine hydroxylase

proinflammatory cytokines in microglia (20). In addition to suppressing the production of proinflammatory cytokines, noradrenaline increases neurotrophin expression in glia cells, including brain-derived neurotrophic factor (BDNF), glial cell-derived neurotrophic factor and fibroblast growth factor-2 (21-23). Noradrenaline induces the production of the IL-1 receptor antagonist and IL-R2, which leads to an overall decrease in IL-1 signaling and IL-10 levels in the cortex and hippocampus. Noradrenaline additionally has beneficial effects on the maturation of oligodendrocyte progenitor cells, which may stimulate the myelination of axons and promote the recovery of MS (24). Therefore, a diminished noradrenaline level or perturbation of the noradrenaline-signaling system exacerbates neuro-inflammation in MS (25).

Increased levels of noradrenaline reduces neurotoxicity due to inflammatory or excitotoxic stimuli, or incubation with amyloid β . For example, using an α 2-adrenergic antagonist reduces neuronal NOS2 expression due to aggregated amyloid β (26). Selective noradrenaline reuptake inhibitors reduce CNS cytokine, chemokine and adhesion molecule expression following systemic endotoxin injection and increased anti-inflammatory cytokines (27,28); and a synthetic noradrenaline precursor reduces astrocyte activation in EAE (7).

The primary source of noradrenaline in the CNS is tyrosine hydroxylase (TH)-positive neurons, which are located in the LC (29). The LC is located at the lower corners of the fourth cerebral ventricle, and releases noradrenaline over almost the entire CNS via nonjunctional varicosities (30). Degeneration or damage of the LC decreases the levels of noradrenaline in its projection areas (31). As reduced noradrenaline levels may lead to increased inflammation and neuronal damage, and as the LC is the primary source of brain noradrenaline and the sole source of noradrenaline fibers to the hippocampus and neocortex (32), methods to raise noradrenaline levels or improve LC function may benefit patients with MS (5). However, a better understanding of the interactions between the LC-NA and immune systems is required to develop novel therapeutic approaches for the treatment of MS.

Catalpol is an important iridoid glycoside, which is purified from the roots of *Rehmannia glutinosa* Libosch. It has been demonstrated to exert anti-inflammatory, antioxidant, anti-apoptotic and other neuroprotective properties, and serves a role in neuroprotection against ischemic injuries and neurodegenerative diseases (33-36). It is additionally found to exert neuroprotective effects age-associated neuroplasticity loss (37). It was recently reported that catalpol may improve memory defects in animal neurodegenerative models (38,39). However, the effects and underlying mechanisms of catalpol in EAE were not investigated extensively. The present study investigated the effects of catalpol as a putative TH inducer by measuring TH protein concentration and activities in the LC.

Materials and methods

Animals. Female C57BL/6 mice of specific pathogen-free grade (age, 4-6 weeks; weight, 14-16 g; n=80) and timed-pregnant Sprague-Dawley rats bearing E15 pups (age, 8-10 weeks; weight, 250-300 g; n=30) were purchased from Vital River Laboratories [Beijing, China; certification

no. SCXK (JING) 2012-0001]. All animals had unrestricted access to food and water and were kept in a controlled 12 h light/dark environment. The animal facility was maintained at $21\pm 2^\circ\text{C}$ and $50\pm 10\%$ relative humidity. All animal experiments were approved by the Animal Experiments and Experimental Animals Management Committee and Animal Experiments and Experimental Animal Welfare Committee of Capital Medical University (Beijing, China; AEEI-2014-018).

Induction of EAE and DSP-4-EAE. C57BL/6 female mice were injected subcutaneously with an emulsion of 300 μg MOG₃₅₋₅₅ (catalog no. MEVGWYRSPFSRVVHLYRNGK; SciLight Biotechnology, Beijing, China) dissolved in 100 μl phosphate-buffered saline (PBS), mixed with 100 μl complete Freund's adjuvant containing 500 μg heat-inactivated Mycobacterium tuberculosis H37RA (Difco; BD Biosciences, Franklin Lakes, NJ, USA). Pertussis toxin (Sigma-Aldrich; Merck KGaA, Darmstadt, Germany; 200 ng/mouse) in 200 μl PBS was administered intraperitoneally (i.p.) on the day of immunization and repeated 2 days later. The mice received a booster MOG₃₅₋₅₅ injection 1 week later (40) (Fig. 1). Subcutaneous injection of animals was conducted following anesthetizing the mice with isoflurane.

C57BL/6 female mice were treated i.p. with 50 mg/kg DSP-4 (Sigma-Aldrich; Merck KGaA) dissolved in PBS once a week for 2 weeks (41). The mice were subsequently immunized with MOG₃₅₋₅₅ 1 week later (14 days following the first DSP-4 injection) and again 1 week later (Fig. 1).

Mice were weighed and evaluated daily from the day of first immunization. Clinical signs were scored by a scale from 0-15 as follows: Tail function: 0, no symptoms; 1, partial paralysis; 2, full paralysis. Hind or forelimbs (assessed separately): 1, weak and abnormal gait with the particular limb; 2, paresis (dragging movement); 3, full paralysis. Scores were calculated as the total of the tail and all four limbs (42,43). Based on this scoring system, a fully paralyzed quadriplegic animal would attain a score of 14 and mortality would be given a score of 15. Scoring was conducted by a blinded investigator.

Treatments. A total of 80 mice were randomly assigned to four groups: Control (n=15), DSP-4-EAE mice (n=25), vehicle-treated EAE mice (n=20) and catalpol-treated EAE mice (n=20). Catalpol (110808-201210; purity, >99%; assayed by high-performance liquid chromatography analysis) was purchased from the Chinese Institute for Drug and Biological Product Control (Beijing, China). Catalpol (10 mg/kg) was administered i.p. daily from the day prior to MOG₃₅₋₅₅ immunization and continued for 40 days (44). Catalpol was prepared by dissolving 2 mg catalpol in 1.0 ml PBS. The control mice, DSP-4-EAE mice and vehicle-treated EAE mice received PBS (i.p.) for 40 days.

LC cell culture and treatments. Culture plates (Corning Incorporated, Corning, NY, USA) were precoated with 100 $\mu\text{g}/\text{ml}$ poly-L-lysine (Sigma-Aldrich; Merck KGaA) in sterile calcium-magnesium-free PBS (pH 7.2) and stored at 37°C overnight in an air-humidified, high efficiency particulate arrestance-filtered incubator. The plates were rinsed twice with sterile double-distilled water, following which 500 μl

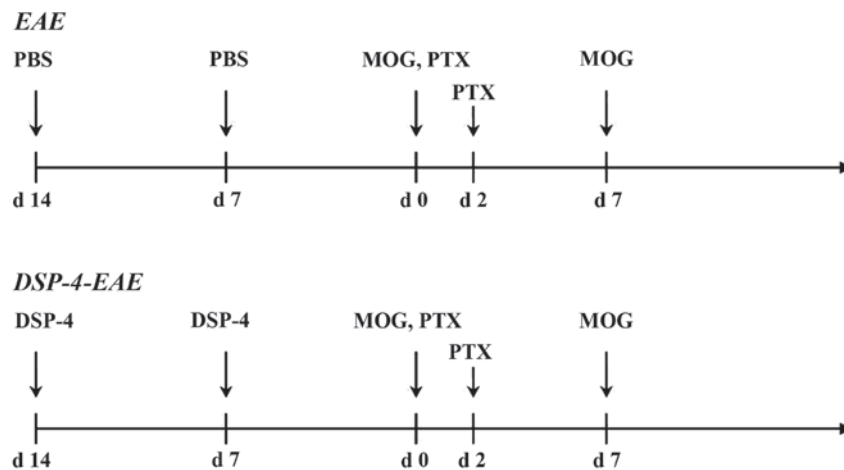


Figure 1. Induction of EAE and DSP-4-EAE. EAE, experimental autoimmune encephalomyelitis; DSP-4, N-(2-chloroethyl)-N-ethyl-2-bromobenzylamine; MOG, myelin oligodendrocyte glycoprotein; PTX, pertussis toxin.

culture media was added and the plates were incubated at 37°C in a 5% CO₂/95% air humidified incubator until further use.

Pregnant Sprague-Dawley rats were sacrificed at E15 by exposure to CO₂. The uterine horns were removed under sterile conditions and placed in a Petri dish containing cold Dulbecco's Modified Eagle's medium (DMEM; Gibco; Thermo Fisher Scientific, Inc., Waltham, MA, USA) on ice. The embryos were removed and further dissection was performed in the same medium. The area dissected corresponded to the proximal rhombencephalic ring between the distal part of the mesencephalic flexure and the proximal part of the pontine flexure (45). The tissues were completely dissociated by gentle trituration with fire-polished glass Pasteur pipettes. Cells were plated on previously prepared 24-well plates at a density of 2x10⁵ cells/well. Cells were cultured in culture medium consisting of DMEM and Ham's F-12 nutrient mixture [1:1 (v/v); Gibco; Thermo Fisher Scientific, Inc.], supplemented with 6 mg/ml glucose, 2 mmol/l glutamine, 5% horse serum (HyClone, GE Healthcare Life Sciences; Logan, UT, USA), 10% fetal bovine serum (Hyclone, GE Healthcare Life Sciences) and 50 U/ml penicillin and 50 µg/ml streptomycin (Invitrogen; Thermo Fisher Scientific, Inc.). Cells were cultured at 37°C in a water-saturated 5% CO₂ and 95% air incubator. At 6 h following plating, the original media was discarded and replenished with serum-free medium. The cells were maintained for 7 days in serum-free medium consisting of neurobasal medium with 1/50 B27 and bovine serum albumin (all purchased from Gibco; Thermo Fisher Scientific, Inc.) and penicillin-streptomycin, which yielded a relatively neuron-pure culture. The culture medium was replaced every other day.

Immunofluorescence staining. TH immunoreactivity was assessed in the LC as a marker of noradrenaline neurons. At 40 days following PBS or catalpol administration, mice were anesthetized with 10% chloral hydrate (5 ml/kg; Sinopharm Chemical Reagent Co., Ltd., Shanghai, China) and the thoracic cavity was opened, and mice were intra-cardially perfused with PBS (0.1 M, pH 7.4) at 4°C, followed by phosphate-buffered 4% paraformaldehyde solution. Directly following perfusion, the brains were removed, post fixed for further 48 h at 4°C

and dehydrated in 30% sucrose solution for 4-6 days. Coronal sections (25-µm thick) were cut from the blocks containing the LC using a Leica freezing microtome (Leica Microsystems GmbH, Wetzlar, Germany).

TH immunofluorescence detection was used to analyze TH-positive noradrenaline neurons. Following fixing with 4% paraformaldehyde for 15 min at 4°C, the cells were washed three times with PBS and the permeability of the cell membrane was increased with 0.2% Triton X-100. Subsequently, cells were blocked for 1 h with 10% goat serum (GS; Gibco, Thermo Fisher Scientific, Inc.) at room temperature and incubated overnight at 4°C with a rabbit polyclonal TH antibody (cat. no. IMG5070; Novus Biologicals, LLC, Littleton, CO, USA) diluted 1:400 in 10% GS. Subsequent incubations were performed at room temperature for 1 h with an Alexa Fluor 568 F (ab')₂ fragment of a goat anti-rabbit immunoglobulin (Ig)G secondary antibody (cat. no. A-11079; Invitrogen; Thermo Fisher Scientific, Inc.). Neurons were identified with mouse anti-β III tubulin primary antibody diluted 1:1,000 (cat. no. ab195879; Abcam, Cambridge, MA, USA) followed by incubation with an Alexa Fluor 488 F (ab')₂ fragment of a goat anti-mouse IgG secondary antibody (cat. no. A-11034; Invitrogen; Thermo Fisher Life Sciences, Inc.). Cells were imaged with the Leica Application Suite V4 software from the Leica Microsystems Framework using an inverted fluorescent microscope (DMI4000; Leica Microsystems GmbH).

Cell viability assay. Survival of LC neurons was determined by measuring their metabolic activity using a Cell Counting Kit-8 (CCK-8) assay (Dojindo Molecular Technologies, Inc., Kumamoto, Japan). Cells were cultured at a density of 1x10⁵ in triplicate in a 96-well plate. A total of 7 days following plating, the culture media was replaced with fresh media (controls) or media containing 1, 5, 10 or 20 µM catalpol, followed by incubation for 24 h at 37°C. Oxidative damage was initiated by a water-soluble initiator 2,2'-azobis (2-amidinopropane) hydrochloride (AAPH; 20 µM) (46). Following incubation for 3 h at 37°C, cell viability was analyzed by CCK-8. The resultant orange formazan product indicative of metabolic activity of the cells was solubilized and quantified at a wavelength of 450 nm using the SpectraMax M5 multifunctional microplate

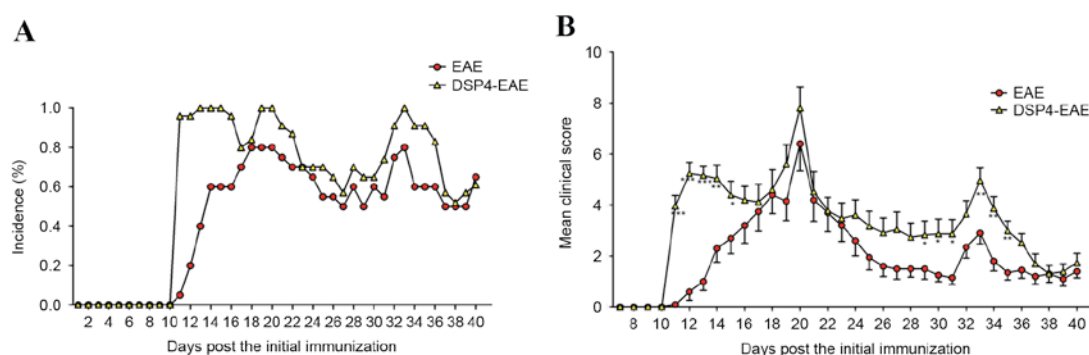


Figure 2. DSP-4 treatment exacerbates EAE. (A) Mean daily clinical scores and (B) mean daily incidence of disease for vehicle ($n=20$) and DSP-4 treated ($n=25$) mice. In the two groups, scores were significantly different from baseline values by day 11 post-immunization, and there were significant differences in the mean day of onset (13.5 ± 0.6 and 11.0 ± 0.08 for EAE and DSP-4-EAE groups, respectively). Additionally, there was a statistically significant difference between development of clinical scores in DSP-4 vs. vehicle groups ($F[33,1]=2.73$; $P<0.001$). Data are presented as the mean \pm standard error from two independent experiments. * $P<0.05$; ** $P<0.01$; *** $P<0.001$ vs. EAE group. EAE, experimental autoimmune encephalomyelitis; DSP-4, N-(2-chloroethyl)-N-ethyl-2-bromobenzylamine.

reader (Molecular Devices, LLC, Sunnyvale, CA, USA). Mean values were obtained from three replicates ($n=3$).

Enzyme-linked immunosorbent assay (ELISA). Cell lysates were prepared from samples of the cortex and spinal cord of control, vehicle-treated EAE and catalpol-treated EAE mice. Tissues (100 mg) were rinsed with PBS, homogenized in 1 ml PBS and stored overnight at -20°C . Following performing two freeze-thaw cycles to break the cell membranes, the homogenates were centrifuged at $5,000 \times g$ for 5 min at 4°C . The supernatant was collected and assayed immediately. ELISA for noradrenaline was performed using a Mouse Noradrenaline ELISA kit (cat. no. MBS701514; MyBioSource, Inc., San Diego, CA, USA) at a detection range of 7.5–300 pg/ml, according to the manufacturer's protocol.

Following drug treatment, the conditioned medium was collected and assayed immediately. Noradrenaline levels were assessed using a rat noradrenaline ELISA kit (cat. no. MBS701791; MyBioSource, Inc.) at a range of 0.49–2,000 pg/ml, according to the manufacturer's protocol.

Western blotting. Pons sections containing the LC of differently treated mice were homogenized in cell lysis buffer (cat. no. C3228; Sigma-Aldrich; Merck KGaA) supplemented with a protease inhibitor cocktail (cat. no. P3840; Sigma-Aldrich; Merck KGaA). The homogenate was subsequently centrifuged at $8,000 \times g$ for 15 min at 4°C . Similar to the LC of animals, the homogenates prepared from the LC neurons were centrifuged at $8,000 \times g$ for 15 min at 4°C . Protein concentration of the supernatants was quantified by the bicinchoninic acid assay kit (Thermo Fisher Scientific, Inc.). Proteins ($40 \mu\text{g}$) from each sample were separated by 10% SDS-PAGE and subsequently transferred onto polyvinylidene difluoride membranes (Merck KGaA) by a semi-dry electrophoretic transfer system (Bio-Rad Laboratories, Inc., Hercules, CA, USA). The membranes were blocked with 5% skimmed milk (BD Biosciences) in TBS with Tween-20 (0.1%, TBST) for 2 h at room temperature and subsequently incubated with a rabbit polyclonal TH or mouse monoclonal anti- β -actin (cat. no. AC-15; Sigma-Aldrich; Merck KGaA) antibody at 4°C overnight. The antibodies were used at a dilution of 1:1,000. The primary antibody

was removed and washed three times with TBST, following which the membranes were further incubated with horse-radish peroxidase-conjugated anti-mouse or anti-rabbit IgG secondary antibodies (1:2,000; Abmart Inc., Shanghai, China) for 1 h at room temperature. Following three washes with TBS with Tween-20, the bands were visualized using Luminol reagent (Santa Cruz Biotechnology Inc., Dallas, TX, USA) by FluorChem FC2 (ProteinSimple, San Jose, CA, USA), and were quantified using Quantity One image analysis software (Version, 4.62; Bio-Rad Laboratories, Inc.).

Statistical analysis. Statistical analyses were performed using GraphPad Prism software (version, 5.0; GraphPad Software, La Jolla, CA, USA). Data are presented as the mean \pm standard error. Repeated measures two-way analysis of variance (ANOVA) was performed to compare EAE score and body weight in treatment groups. Pair-wise comparison between the groups was performed by one-way ANOVA followed by Tukey's post hoc test. $P<0.05$ was considered to indicate a statistically significant difference.

Results

DSP-4 administration exacerbates EAE. DSP-4 is widely used to reduce brain noradrenaline levels to study the functions of the central noradrenergic system. Numerous studies have demonstrated that DSP-4 may produce long-lasting reduction of endogenous noradrenaline in the brain of mice and rats (47–49). Administration of 50 mg/kg DSP-4 has previously resulted in a significant reduction in noradrenaline levels in EAE animals (a reduction of 73% vs. vehicle) (50). In the present study, 2 weeks following the first DSP-4 injection, mice were immunized with a MOG_{35–55} peptide. The results indicated that in the EAE group, 16/20 (80%) mice developed clinical symptoms, and the mean day of onset was 13.5 ± 0.6 days post initial immunization. In the DSP-4-treated group, the incidence was 25/25 (100%), and the mean time of onset was at 11.0 ± 0.08 days post initial immunization, which was markedly different from the EAE group (Fig. 2A). In addition, the mean daily clinical scores were significantly increased in DSP4-EAE mice compared with the EAE group

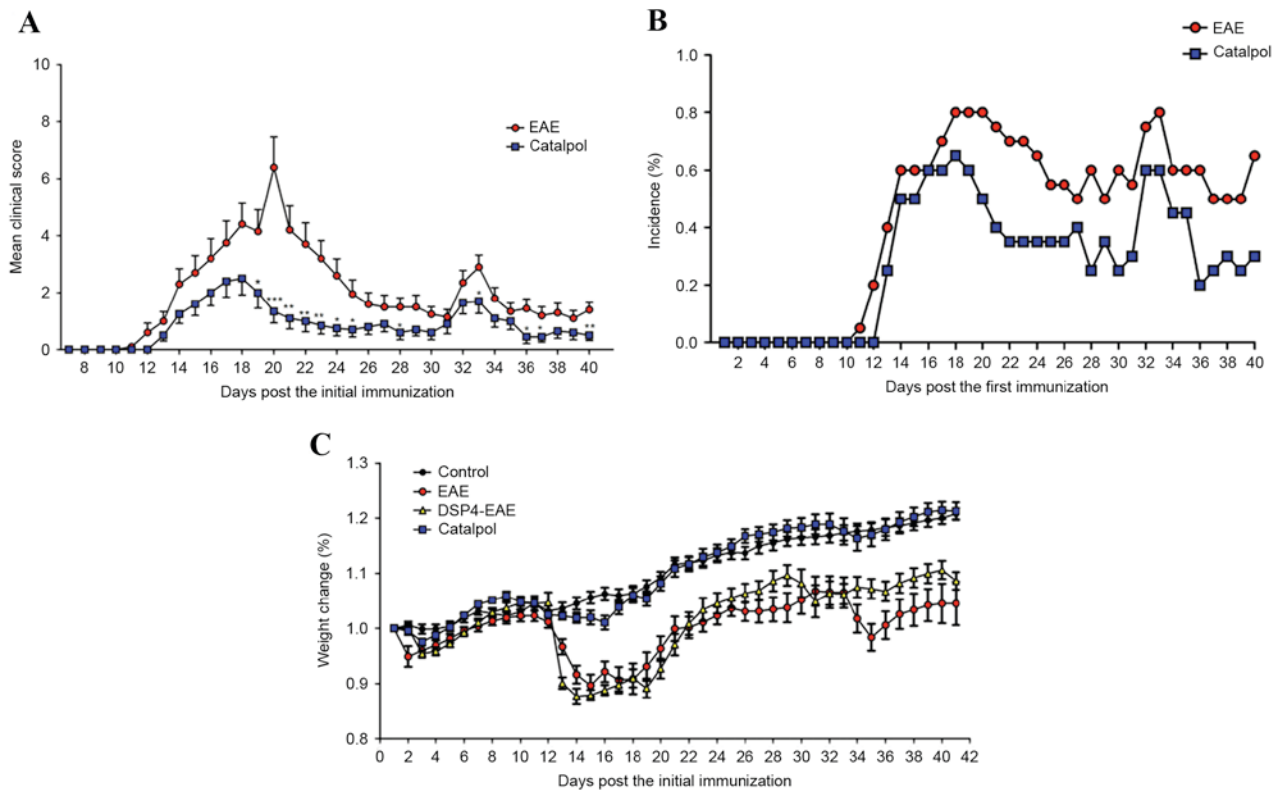


Figure 3. Catalpol treatment ameliorates EAE. (A) Mean daily clinical scores, (B) mean daily incidence of disease and (C) body weight alterations in vehicle- (n=20) and catalpol-treated (n=20) mice. There was a significant effect of catalpol on clinical scores from day 18 onwards (F[33,1]=3.17, $P<0.001$). Data are presented as the mean \pm standard error from two independent experiments. * $P<0.05$; ** $P<0.01$; *** $P<0.001$ vs. EAE group. EAE, experimental autoimmune encephalomyelitis; DSP-4, N-(2-chloroethyl)-N-ethyl-2-bromobenzylamine.

at the start and end time points, particularly at days 11, 12 and 13 ($P<0.001$) and days 33, 34 and 35 ($P<0.01$; Fig. 2B). These results suggested that selectively reducing noradrenaline levels in the CNS exacerbates EAE. Noradrenaline depletion significantly exacerbated the magnitude of EAE-induced clinical signs, and incidence was higher in DSP-4-treated group compared with the control group.

Catalpol treatment ameliorates EAE. The weight and disability scores of each mouse were obtained daily over the course of the 40-day experiment. The efficacy of catalpol on the neurological behaviors of EAE mice was investigated. Treatment with catalpol or vehicle was initiated the day prior to MOG₃₅₋₅₅ immunization and continued for 5 weeks. In the vehicle group, there was a significant increase in clinical scores vs. baseline at day 12, and the maximum mean clinical score (6.4 ± 0.10) was reached on day 20, which remained unaltered thereafter (Fig. 3A). From day 10 onwards, 3/20 mice in the vehicle group exhibited clinical improvement. In the catalpol group, clinical scores were significantly increased from baseline scores at day 13, and the maximum mean clinical score (2.5 ± 0.06) was reached on day 18, after which it began to decrease, reaching 0.5 ± 0.02 at the end of the study. From day 10 onwards, 10/20 catalpol-treated mice exhibited clinical improvement. Mean clinical scores of catalpol-treated EAE mice were significantly reduced compared with vehicle-treated EAE mice ($P<0.05$; Fig. 3A). Cumulative incidence scores of catalpol-treated EAE mice were also reduced compared with vehicle-treated EAE mice (Fig. 3B). The body weights of animals were monitored

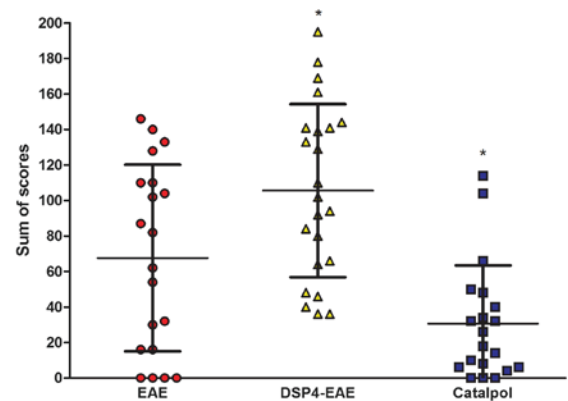


Figure 4. Catalpol treatment decreases disease burden in animals. The overall disease burden for vehicle- (n=20), catalpol- (n=25) and DSP-4-treated mice are represented as the sum of scores, which is the sum of the disability scores obtained daily over the course of the 40-day experiment. Data are presented as the mean \pm standard error. Catalpol vs. EAE, * $P<0.05$; DSP-4-EAE vs. EAE, * $P<0.05$. EAE, experimental autoimmune encephalomyelitis; DSP-4, N-(2-chloroethyl)-N-ethyl-2-bromobenzylamine.

daily to confirm the results of the clinical scores. As presented in Fig. 3C, mice demonstrated persistent loss of body weight with the progression of EAE, and catalpol treatment prevented body weight loss of EAE mice. No obvious adverse effects of catalpol were observed at a dose of 10 mg/kg/day in EAE mice 35 days following injection. The overall disease burden for each mouse is represented as the sum of the daily scores over the course of the 40-day experiment (42). There was a decrease

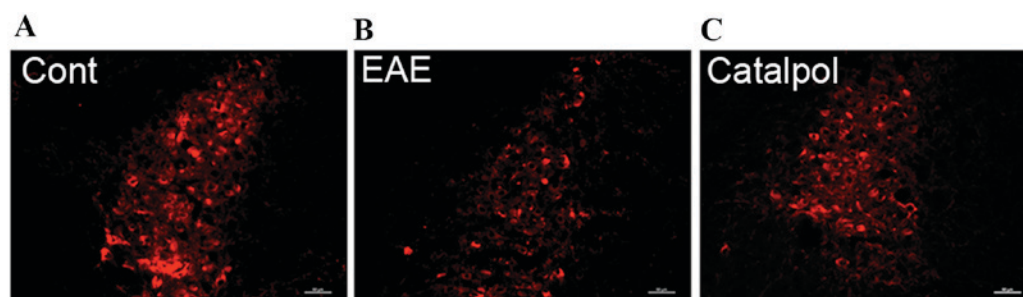


Figure 5. Sagittal sections through (A) control, (B) vehicle-treated and (C) catalpol-treated mice were stained for the presence of TH+ neurons. Images are representative of three mice in each group, and demonstrate an increase in TH + staining following catalpol treatment. TH, tyrosine hydroxylase; EAE, experimental autoimmune encephalomyelitis; DSP-4, N-(2-chloroethyl)-N-ethyl-2-bromobenzylamine; Cont, control.

in disease burden in catalpol-treated animals ($P < 0.05$; Fig. 4). These results indicated that catalpol suppressed the severity of EAE.

Catalpol treatment effects on TH immunoreactivity in LC. Serial sections prepared from the LC of control, vehicle-treated and catalpol-treated mice were immunostained for TH. In the LC, where TH-immunoreactivity reflects noradrenergic neurons, examination of immunofluorescence in sections from control mice revealed numerous large spherical TH immunoreactivity neurons (Fig. 5A). In EAE mice (Fig. 5B), TH expression was greatly reduced compared with the control-group mice, whereas catalpol treatment (Fig. 5C) recovered this effect.

Effect of catalpol on the viability of LC cells. As assessed by CCK-8 assay, vehicle-treated LC cells demonstrated markedly decreased viability 3 h after treatment with DSP-4. Catalpol (10 and 20 μ M) significantly inhibited this effect (Fig. 6).

Catalpol increases noradrenaline levels in vivo and in vitro. Noradrenaline levels were determined by specific ELISA analyses in homogenates prepared from the cortex and spinal cords of control, vehicle-treated EAE and catalpol-treated EAE mice 20 and 40 days following immunization. In control mice, noradrenaline levels were increased in the cortex compared with the spinal cord. A significant decrease in noradrenaline levels was observed in the cortex (~38% reduction) and spinal cord (~62% reduction) of EAE mice compared with controls. Additionally, noradrenaline levels were significantly increased in catalpol-treated EAE mice (Fig. 7A).

Treatment with 10 μ M catalpol for 24 h markedly enhanced the amount of noradrenaline released into the medium compared with the control after DSP-4-induced damage (20 μ M AAPH) for 3 h (Fig. 7B).

Catalpol treatment effects on TH expression in vivo and in vitro. Mice were injected with PBS or catalpol for 40 days. The LC was dissected from the mouse brains, and the protein expression levels of TH protein were determined. Protein expression levels of TH were increased when the animals were chronically treated with 10 mg/kg catalpol for 40 days ($P < 0.01$; Fig. 8A). However, no significant differences were observed in TH protein expression levels between catalpol- and vehicle-treated LC neurons ($P > 0.05$; Fig. 8B).

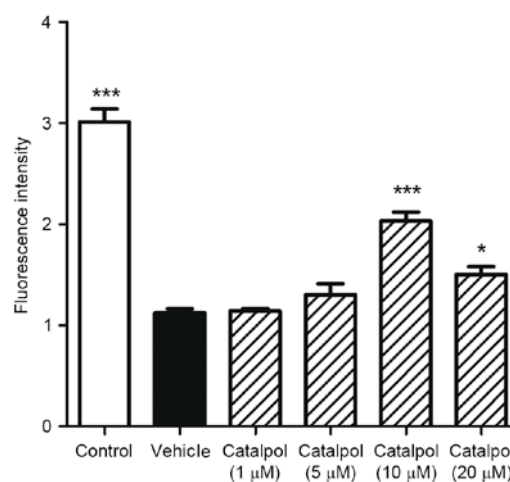


Figure 6. Catalpol protects against 2,2'-azobis (2-amidinopropane) hydrochloride-induced oxidative damage in locus coeruleus neurons. Cells were treated with 1, 5 10 or 20 μ M catalpol and incubated for 24 h. Oxidative damage was initiated by 20 μ M DSP-4 treatment for 3 h. Cell viability was analyzed using a Cell Counting Kit-8. Cell viability was significantly decreased in the vehicle-treated group 3 h following DSP-4 treatment. Catalpol (10 and 20 μ M) inhibited this effect. Data are presented as the mean \pm standard error ($n = 8$). * $P < 0.05$; *** $P < 0.001$ vs. vehicle. DSP-4, N-(2-chloroethyl)-N-ethyl-2-bromobenzylamine; Cont, control.

Discussion

Numerous studies have indicated that alterations in peripheral noradrenaline levels occur during MS and EAE (51-53). In contrast to its role in EAE and MS, the role of central noradrenaline remains to be fully elucidated. The results of the present study indicated that noradrenaline depletion significantly exacerbates clinical scores in EAE mice, and the incidence was increased in the DSP-4-treated group compared with the vehicle-treated EAE group. As DSP-4 has limited effects on peripheral noradrenaline levels (47), reduction in the noradrenaline levels in the CNS exacerbates the severity of EAE. These results were not completely consistent with previously published results, which reported that the incidence and average day of onset in the DSP-4-treated group were not significantly different from the control group (7).

The present study re-examined whether CNS noradrenaline level reduction occurs in EAE. Consistent with a previous study (5), it was observed that noradrenaline levels were decreased in the frontal cortex and spinal cord of EAE mice,

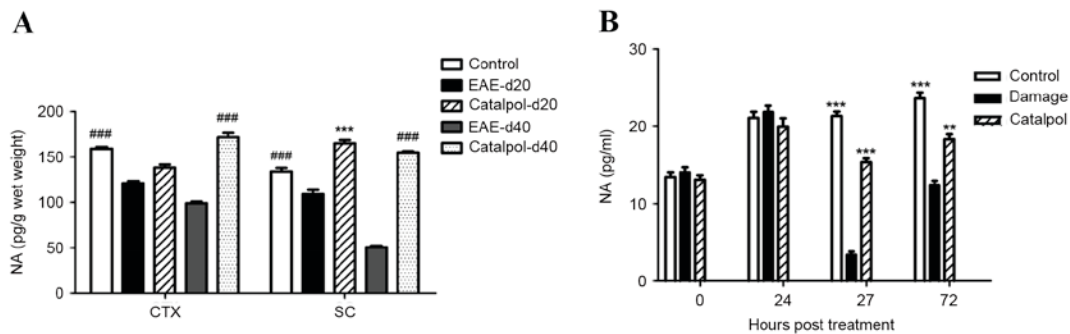


Figure 7. Catalpol increases noradrenaline levels *in vivo* and *in vitro*. (A) Noradrenaline levels were significantly decreased in the frontal cortex and spinal cord of EAE mice compared with controls. There were statistically significant differences between catalpol-treated and vehicle groups. Data are presented as the mean \pm standard error. *** $P < 0.001$ vs. EAE-d40; *** $P < 0.001$ vs. EAE-d20. (B) Catalpol treatment markedly increased the amount of noradrenaline released into the medium. Data are presented as the mean \pm standard error ($n = 6$). ** $P < 0.01$; *** $P < 0.001$ vs. damage. Damage, vehicle-treated neurons; NA, noradrenaline; CTX, cortex; SC, spinal cord; EAE, experimental autoimmune encephalomyelitis; d20, day 20 following immunization; d40; day 40 following immunization.

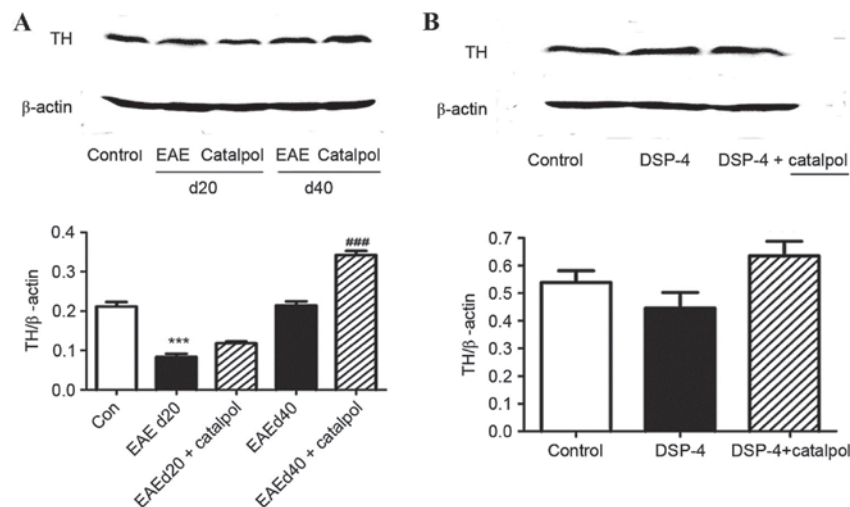


Figure 8. Representative western blotting images and quantification of TH protein expression levels in (A) C57BL/6 mice and (B) Sprague-Dawley rat embryos. β -actin served as an internal control. Data are presented as the mean \pm standard error ($n = 3$). *** $P < 0.001$ vs. control; *** $P < 0.01$ vs. EAE d40. Con, control; TH, tyrosine hydroxylase; EAE, experimental autoimmune encephalomyelitis; DSP-4, N-(2-chloroethyl)-N-ethyl-2-bromobenzylamine; d20, day 20 following immunization; d40; day 40 following immunization.

and that the noradrenergic neurons in LC were additionally damaged, which confirmed the association between abnormalities of the LC-NA system with exacerbation of neuroinflammation and neural damage. However, decreased synthesis, increased metabolism or increased re-uptake may account for reduced noradrenaline levels, inflammation in the LC, and stress in TH-positive neurons. The present study demonstrated that noradrenaline metabolism in the cortex was increased in EAE mice compared with controls. Catecholamine-O-methyl transferase and monoamine oxidase are ubiquitous metabolic enzymes of noradrenaline; the two have been demonstrated to be increased in neurological diseases (54,55). Noradrenaline increases the expression of numerous neurotrophins, which are known to be critical for LC survival and maturation. These findings suggested that diminished LC function and reduced noradrenaline levels may contribute to damage by reducing neurotrophic support from glial cells and LC neurons.

Numerous tests to increase CNS noradrenaline levels have been demonstrated to provide benefit for MS and EAE. Noradrenaline levels may be increased by the use of

selective α_2 -adrenoceptor antagonists, selective noradrenaline reuptake inhibitors (NARIs) and noradrenaline precursors. α_2 -adrenoceptors are inhibitory pre-synaptic neuronal auto-receptors and are primarily located in the noradrenergic neurons of LC. α_2 -antagonists increase noradrenaline release from LC noradrenergic neurons, reduce excitotoxicity (56), promote neuronal survival (57) and inhibit inflammatory responses (26) *in vivo*. However, their clinical use has been limited. Selective NARIs, including the non-tricyclic antidepressant atomoxetine, exhibit anti-inflammatory effects on microglial cells (58) and increase BDNF expression in the hippocampus (59). However, treatment with atomoxetine did not promote recovery in EAE mice (7), thereby suggesting that although extracellular noradrenaline levels may be increased by NARIs, the reduced endogenous noradrenaline levels are not sufficient to exert anti-inflammatory and neuroprotective effects. Noradrenaline levels may be directly increased by treatment with the synthetic noradrenaline precursor, L-threo-3,4-dihydroxyphenylserine (L-DOPS), which is converted to noradrenaline via decarboxylation by dopamine

decarboxylase (60). In contrast, treatment with L-DOPS reduced clinical scores in EAE mice (7). Furthermore, treatment with L-DOPS and atomoxetine resulted in a synergistic effect on EAE mice (7). It was reported that co-treatment with L-phenylalanine and the NARI lofepramine improved clinical symptoms and reduced lesion volumes in MS patients (61,62).

TH catalyzes the rate-limiting and committed steps in the biosynthesis of noradrenaline (63). Therefore, regulation of TH protein expression levels and intrinsic enzyme activity may represent a key pathway for controlling the synthesis of noradrenaline. Chronic neuroinflammation leads to reduced TH enzyme levels in the LC and decreased noradrenaline levels in LC projections, including the cortex and hippocampus (64). Noradrenaline acts as an anti-inflammatory molecule, stimulating the production of BDNF, and supporting neurogenesis in the hippocampus. Therefore, cognitive decline may be associated with decreased noradrenaline levels in the cortex and hippocampus (65).

LC stress and damage are primary causes of noradrenaline dysfunction (66). In this regard, drugs that selectively increase LC neuronal viability or activity may be of benefit. A previous study reported that vindeburnol, a structural analog of the cerebral vasodilator vincamine, increased TH activity and protein expression in the LC (67). However, it was demonstrated that vindeburnol increases the expression of genes involved in LC survival and maturation, and increases central noradrenaline levels (66). Therefore, in relation to the present study, it may be hypothesized that binding of vindeburnol to α_2 -adrenergic receptors lead to a sustained increase in noradrenaline release and ultimately a requirement for *de novo* noradrenaline synthesis and increased TH expression.

Numerous approved first-line drugs, including interferon- β , glatiramer acetate, mitoxantrone and natalizumab, are either immunoregulators or immunosuppressants, and have significant adverse effects associated with long-term therapy, including infection, cardiotoxicity, anemia, nausea and depression (68). However, there are limited treatment options that reduce or inhibit the neurodegeneration, promote remyelination and improving neuron survival, which determines the outcome and prognosis of the disease.

Catalpol is widely used as a traditional Chinese herbal medicine for the treatment of various neurodegenerative diseases, including Alzheimer's, Parkinson's and ischemic diseases. Catalpol may additionally cross the blood-brain barrier (68). Furthermore, catalpol may enhance neuronal axon growth (69), implicating a potential role for the treatment of MS. Catalpol has been demonstrated to protect dopaminergic neurons from LPS-induced neurotoxicity (70). The present study used the most commonly used model for MS to confirm the neuroprotective effects of catalpol. In mice treated with catalpol, a significant improvement in the clinical scores was observed in EAE. Catalpol exerts neuroprotective effects in cortical neurons (35); however, its role in exerting similar effects on LC cells, the primary source of noradrenaline in the CNS, remains unclear. The present study tested the effects of catalpol on LC neurons. In primary LC neuron cultures, catalpol exerted a neuroprotective effect and enhanced the generation of noradrenaline following DSP-4-induced neuronal damage. Furthermore, when the cultures were incubated with catalpol alone, there was no alteration in the

production of noradrenaline, which may account for the fact that catalpol had fewer side effects at 10 μ M. These results confirmed that catalpol serves as a potential therapeutic drug and may be useful for the treatment of MS.

In conclusion, these data suggested that catalpol treatment exerted positive effects on the synthesis of noradrenaline and LC physiology. However, as the study was restricted to the CNS, further investigation into whether catalpol is involved in regulation of peripheral lymphocytes and macrophage activation is required.

Acknowledgements

The present study was supported by the National Natural Science Foundation of China (grant nos. 81072765 and 81273742) and the Beijing Natural Science Foundation (grant no. 7142053).

References

1. McQualter JL and Bernard CC: Multiple sclerosis: A battle between destruction and repair. *J Neurochem* 100: 295-306, 2007.
2. Deckx N, Lee WP, Berneman ZN and Cools N: Neuroendocrine immunoregulation in multiple sclerosis. *Clin Dev Immunol* 2013: 705232, 2013.
3. Alvarez JI, Cayrol R and Prat A: Disruption of central nervous system barriers in multiple sclerosis. *Biochim Biophys Acta* 1812: 252-264, 2011.
4. Gadea M, Martinez-Bisbal MC, Marti-Bonmati L, Espert R, Casanova B, Coret F and Celda B: Spectroscopic axonal damage of the right locus coeruleus relates to selective attention impairment in early stage relapsing-remitting multiple sclerosis. *Brain* 127: 89-98, 2004.
5. Polak PE, Kalinin S and Feinstein DL: Locus coeruleus damage and noradrenaline reductions in multiple sclerosis and experimental autoimmune encephalomyelitis. *Brain* 134: 665-677, 2011.
6. Fritschy JM and Grzanna R: Experimentally-induced neuron loss in the locus coeruleus of adult rats. *Exp Neurol* 111: 123-127, 1991.
7. Simonini MV, Polak PE, Sharp A, McGuire S, Galea E and Feinstein DL: Increasing CNS noradrenaline reduces EAE severity. *J Neuroimmune Pharmacol* 5: 252-259, 2010.
8. White SR, Bhatnagar RK and Bardo MT: Norepinephrine depletion in the spinal cord gray matter of rats with experimental allergic encephalomyelitis. *J Neurochem* 40: 1771-1773, 1983.
9. Krenger W, Honegger CG, Feurer C and Cammisuli S: Changes of neurotransmitter systems in chronic relapsing experimental allergic encephalomyelitis in rat brain and spinal cord. *J Neurochem* 47: 1247-1254, 1986.
10. Marien MR, Colpaert FC and Rosenquist AC: Noradrenergic mechanisms in neurodegenerative diseases: A theory. *Brain Res Brain Res Rev* 45: 38-78, 2004.
11. Madrigal JL, Kalinin S, Richardson JC and Feinstein DL: Neuroprotective actions of noradrenaline: Effects on glutathione synthesis and activation of peroxisome proliferator activated receptor delta. *J Neurochem* 103: 2092-2101, 2007.
12. Frohman EM, Vayuvegula B, Gupta S and van den Noort S: Norepinephrine inhibits gamma-interferon-induced major histocompatibility class II (Ia) antigen expression on cultured astrocytes via beta-2-adrenergic signal transduction mechanisms. *Proc Natl Acad Sci USA* 85: 1292-1296, 1988.
13. Benveniste EN, Huneycutt BS, Shrikant P and Ballester ME: Second messenger systems in the regulation of cytokines and adhesion molecules in the central nervous system. *Brain Behav Immun* 9: 304-314, 1995.
14. Feinstein DL: Suppression of astroglial nitric oxide synthase expression by norepinephrine results from decreased NOS-2 promoter activity. *J Neurochem* 70: 1484-1496, 1998.
15. Nakamura A, Johns EJ, Imaizumi A, Abe T and Kohsaka T: Regulation of tumour necrosis factor and interleukin-6 gene transcription by beta2-adrenoceptor in the rat astrocytes. *J Neuroimmunol* 88: 144-153, 1998.

16. Galea E, Heneka MT, Dello RC and Feinstein DL: Intrinsic regulation of brain inflammatory responses. *Cell Mol Neurobiol* 23: 625-635, 2003.
17. Fujita H, Tanaka J, Maeda N and Sakanaka M: Adrenergic agonists suppress the proliferation of microglia through beta 2-adrenergic receptor. *Neurosci Lett* 242: 37-40, 1998.
18. Heneka MT, Nadrigny F, Regen T, Martinez-Hernandez A, Dumitrescu-Ozimek L, Terwel D, Jandanhazi-Kurutz D, Walter J, Kirchhoff F, Hanisch UK and Kummer MP: Locus ceruleus controls Alzheimer's disease pathology by modulating microglial functions through norepinephrine. *Proc Natl Acad Sci USA* 107: 6058-6063, 2010.
19. Kong Y, Ruan L, Qian L, Liu X and Le Y: Norepinephrine promotes microglia to uptake and degrade amyloid beta peptide through upregulation of mouse formyl peptide receptor 2 and induction of insulin-degrading enzyme. *J Neurosci* 30: 11848-11857, 2010.
20. Campbell A: Inflammation, neurodegenerative diseases, and environmental exposures. *Ann N Y Acad Sci* 1035: 117-132, 2004.
21. Zafra F, Lindholm D, Castrén E, Hartikka J and Thoenen H: Regulation of brain-derived neurotrophic factor and nerve growth factor mRNA in primary cultures of hippocampal neurons and astrocytes. *J Neurosci* 12: 4793-4799, 1992.
22. Remy S, Naveilhan P, Brachet P and Neveu I: Differential regulation of GDNF, neurturin, and their receptors in primary cultures of rat glial cells. *J Neurosci Res* 64: 242-251, 2001.
23. Kajitani N, Hisaoka-Nakashima K, Morioka N, Okada-Tsuchioka M, Kaneko M, Kasai M, Shibasaki C, Nakata Y and Takebayashi M: Antidepressant acts on astrocytes leading to an increase in the expression of neurotrophic/growth factors: Differential regulation of FGF-2 by noradrenaline. *PLoS One* 7: e51197, 2012.
24. Ghiani CA, Eisen AM, Yuan X, DePinho RA, McBain CJ and Gallo V: Neurotransmitter receptor activation triggers p27 (Kip1) and p21 (CIP1) accumulation and G1 cell cycle arrest in oligodendrocyte progenitors. *Development* 126: 1077-1090, 1999.
25. Kaneko YS, Mori K, Nakashima A, Sawada M, Nagatsu I and Ota A: Peripheral injection of lipopolysaccharide enhances expression of inflammatory cytokines in murine locus coeruleus: Possible role of increased norepinephrine turnover. *J Neurochem* 94: 393-404, 2005.
26. Kalinin S, Polak PE, Madrigal JL, Gavrilyuk V, Sharp A, Chauhan N, Marien M, Colpaert F and Feinstein DL: Beta-amyloid-dependent expression of NOS2 in neurons: Prevention by an alpha2-adrenergic antagonist. *Antioxid Redox Signal* 8: 873-883, 2006.
27. O'Sullivan JB, Ryan KM, Curtin NM, Harkin A and Connor TJ: Noradrenaline reuptake inhibitors limit neuroinflammation in rat cortex following a systemic inflammatory challenge: Implications for depression and neurodegeneration. *Int J Neuropsychopharmacol* 12: 687-699, 2009.
28. O'Sullivan JB, Ryan KM, Harkin A and Connor TJ: Noradrenaline reuptake inhibitors inhibit expression of chemokines IP-10 and RANTES and cell adhesion molecules VCAM-1 and ICAM-1 in the CNS following a systemic inflammatory challenge. *J Neuroimmunol* 220: 34-42, 2010.
29. Benarroch EE: The locus coeruleus norepinephrine system: Functional organization and potential clinical significance. *Neurology* 73: 1699-1704, 2009.
30. Szabadi E: Functional neuroanatomy of the central noradrenergic system. *J Psychopharmacol* 27: 659-693, 2013.
31. Iversen LL, Rossor MN, Reynolds GP, Hills R, Roth M, Mountjoy CQ, Foote SL, Morrison JH and Bloom FE: Loss of pigmented dopamine-beta-hydroxylase positive cells from locus coeruleus in senile dementia of Alzheimer's type. *Neurosci Lett* 39: 95-100, 1983.
32. Berridge CW and Waterhouse BD: The locus coeruleus-noradrenergic system: Modulation of behavioral state and state-dependent cognitive processes. *Brain Res Brain Res Rev* 42: 33-84, 2003.
33. Li DQ, Bao YM, Li Y, Wang CF, Liu Y and An LJ: Catalpol modulates the expressions of Bcl-2 and Bax and attenuates apoptosis in gerbils after ischemic injury. *Brain Res* 1115: 179-185, 2006.
34. Tian YY, Jiang B, An LJ and Bao YM: Neuroprotective effect of catalpol against MPP(+)-induced oxidative stress in mesencephalic neurons. *Eur J Pharmacol* 568: 142-148, 2007.
35. Jiang B, Du J, Liu JH, Bao YM and An LJ: Catalpol attenuates the neurotoxicity induced by beta-amyloid (1-42) in cortical neuron-glia cultures. *Brain Res* 1188: 139-147, 2008.
36. Liang JH, Du J, Xu LD, Jiang T, Hao S, Bi J and Jiang B: Catalpol protects primary cultured cortical neurons induced by Abeta(1-42) through a mitochondrial-dependent caspase pathway. *Neurochem Int* 55: 741-746, 2009.
37. Liu J, He QJ, Zou W, Wang HX, Bao YM, Liu YX and An LJ: Catalpol increases hippocampal neuroplasticity and up-regulates PKC and BDNF in the aged rats. *Brain Res* 1123: 68-79, 2006.
38. Zhang XL, An LJ, Bao YM, Wang JY and Jiang B: d-galactose administration induces memory loss and energy metabolism disturbance in mice: Protective effects of catalpol. *Food Chem Toxicol* 46: 2888-2894, 2008.
39. Xia Z, Zhang R, Wu P, Xia Z and Hu Y: Memory defect induced by beta-amyloid plus glutamate receptor agonist is alleviated by catalpol and donepezil through different mechanisms. *Brain Res* 1441: 27-37, 2012.
40. Feinstein DL, Galea E, Gavrilyuk V, Brosnan CF, Whitacre CC, Dumitrescu-Ozimek L, Landreth GE, Pershadsingh HA, Weinberg G and Heneka MT: Peroxisome proliferator-activated receptor-gamma agonists prevent experimental autoimmune encephalomyelitis. *Ann Neurol* 51: 694-702, 2002.
41. Heneka MT, Galea E, Gavrilyuk V, Dumitrescu-Ozimek L, Daeschner J, O'Banion MK, Weinberg G, Klockgether T and Feinstein DL: Noradrenergic depletion potentiates beta-amyloid-induced cortical inflammation: Implications for Alzheimer's disease. *J Neurosci* 22: 2434-2442, 2002.
42. Giuliani F, Metz LM, Wilson T, Fan Y, Bar-Or A and Yong VW: Additive effect of the combination of glatiramer acetate and minocycline in a model of MS. *J Neuroimmunol* 158: 213-221, 2005.
43. Weaver A, Goncalves da Silva A, Nuttall RK, Edwards DR, Shapiro SD, Rivest S and Yong VW: An elevated matrix metalloproteinase (MMP) in an animal model of multiple sclerosis is protective by affecting Th1/Th2 polarization. *FASEB J* 19: 1668-1670, 2005.
44. Jiang B, Zhang H, Bi J and Zhang XL: Neuroprotective activities of catalpol on MPP+/MPTP-induced neurotoxicity. *Neurol Res* 30: 639-644, 2008.
45. Holm PC, Rodriguez FJ, Kresse A, Canals JM, Silos-Santiago I and Arenas E: Crucial role of TrkB ligands in the survival and phenotypic differentiation of developing locus coeruleus noradrenergic neurons. *Development* 130: 3535-3545, 2003.
46. Flueraru M, So R, Willmore WG, Poulter MO, Durst T, Charron M and Wright JS: Cytotoxicity and cytoprotective activity of naphthalenediols in rat cortical neurons. *Chem Res Toxicol* 19: 1221-1217, 2006.
47. Jonsson G, Hallman H, Ponzio F and Ross S: DSP4 (N-(2-chloroethyl)-N-ethyl-2-bromobenzylamine)-a useful denervation tool for central and peripheral noradrenaline neurons. *Eur J Pharmacol* 72: 173-188, 1981.
48. Hallman H and Jonsson G: Pharmacological modifications of the neurotoxic action of the noradrenaline neurotoxin DSP4 on central noradrenaline neurons. *Eur J Pharmacol* 103: 269-278, 1984.
49. Landa ME, Rubio MC and Jaim-Etcheverry G: The neurotoxic compound N-(2-chloroethyl)-N-ethyl-2-bromobenzylamine hydrochloride (DSP4) depletes endogenous norepinephrine and enhances release of [3H]norepinephrine from rat cortical slices. *J Pharmacol Exp Ther* 231: 131-136, 1984.
50. Cassano T, Gaetani S, Morgese MG, Macheda T, Laconca L, Dipasquale P, Taltavull J, Shippenberg TS, Cuomo V and Gobbi G: Monoaminergic changes in locus coeruleus and dorsal raphe nucleus following noradrenaline depletion. *Neurochem Res* 34: 1417-1426, 2009.
51. Dowdell KC, Gienapp IE, Stuckman S, Wardrop RM and Whitacre CC: Neuroendocrine modulation of chronic relapsing experimental autoimmune encephalomyelitis: A critical role for the hypothalamic-pituitary-adrenal axis. *J Neuroimmunol* 100: 243-251, 1999.
52. Cosentino M, Zaffaroni M, Marino F, Bombelli R, Ferrari M, Rasini E, Lecchini S, Ghezzi A and Frigo G: Catecholamine production and tyrosine hydroxylase expression in peripheral blood mononuclear cells from multiple sclerosis patients: Effect of cell stimulation and possible relevance for activation-induced apoptosis. *J Neuroimmunol* 133: 233-240, 2002.
53. Rajda C, Bencsik K, Vécsei LL and Bergquist J: Catecholamine levels in peripheral blood lymphocytes from multiple sclerosis patients. *J Neuroimmunol* 124: 93-100, 2002.

54. Helkamaa T, Reenilä I, Tuominen RK, Soinila S, Väänänen A, Tilgmann C and Rauhala P: Increased catechol-O-methyltransferase activity and protein expression in OX-42-positive cells in the substantia nigra after lipopolysaccharide microinfusion. *Neurochem Int* 51: 412-423, 2007.
55. Samantaray S, Knaryan VH, Butler JT, Ray SK and Banik NL: Spinal cord degeneration in C57BL/6N mice following induction of experimental parkinsonism with MPTP. *J Neurochem* 104: 1309-1320, 2008.
56. Martel J, Chopin P, Colpaert F and Marien M: Neuroprotective effects of the alpha2-adrenoceptor antagonists, (+)-efaroxan and (+/-)-idazoxan, against quinolinic acid-induced lesions of the rat striatum. *Exp Neurol* 154: 595-601, 1998.
57. Veyrac A, Didier A, Colpaert F, Jourdan F and Marien M: Activation of noradrenergic transmission by alpha2-adrenoceptor antagonists counteracts deafferentation-induced neuronal death and cell proliferation in the adult mouse olfactory bulb. *Exp Neurol* 194: 444-456, 2005.
58. Hashioka S, Klegeris A, Monji A, Kato T, Sawada M, McGeer PL and Kanba S: Antidepressants inhibit interferon-gamma-induced microglial production of IL-6 and nitric oxide. *Exp Neurol* 206: 33-42, 2007.
59. Russo-Neustadt A, Beard RC and Cotman CW: Exercise, antidepressant medications, and enhanced brain derived neurotrophic factor expression. *Neuropsychopharmacology* 21: 679-682, 1999.
60. Goldstein DS: L-Dihydroxyphenylserine (L-DOPS): A norepinephrine prodrug. *Cardiovasc Drug Rev* 24: 189-203, 2006.
61. Puri BK, Bydder GM, Chaudhuri KR, Al Saffar BY, Curati WL, White SJ, Mitchell L, Hajnal JV and Horrobin DF: MRI changes in multiple sclerosis following treatment with lofepramine and L-phenylalanine. *Neuroreport* 12: 1821-1824, 2001.
62. Loder C, Allawi J and Horrobin DF: Treatment of multiple sclerosis with lofepramine, L-phenylalanine and vitamin B (12): Mechanism of action and clinical importance: Roles of the locus coeruleus and central noradrenergic systems. *Med Hypotheses* 59: 594-602, 2002.
63. Stjarne L, Lishajko F and Roth RH: Regulation of noradrenaline biosynthesis in nerve tissue. *Nature* 215: 770-772, 1967.
64. Bardou I, Kaercher RM, Brothers HM, Hopp SC, Royer S and Wenk GL: Age and duration of inflammatory environment differentially affect the neuroimmune response and catecholaminergic neurons in the midbrain and brainstem. *Neurobiol Aging* 35: 1065-1073, 2014.
65. Grudzien A, Shaw P, Weintraub S, Bigio E, Mash DC and Mesulam MM: Locus coeruleus neurofibrillary degeneration in aging, mild cognitive impairment and early Alzheimer's disease. *Neurobiol Aging* 28: 327-335, 2007.
66. Polak PE, Kalinin S, Braun D, Sharp A, Lin SX and Feinstein DL: The vincamine derivative vindeburnol provides benefit in a mouse model of multiple sclerosis: Effects on the Locus coeruleus. *J Neurochem* 121: 206-216, 2012.
67. Weissmann D, Labatut R, Gillon J, Richard F and Pujol J: Measurement of brain tyrosine hydroxylase concentrations by quantitative autoradiography after transfer of soluble proteins to nitro-cellulose. *C R Acad Sci III* 306: 457-460, 1988 (In French).
68. He Y, Zhu H, Li W, Chen G, Li Z and Xu X: HPLC determination of catalpol in cerebrospinal fluid of rats. *Zhong Guo Zhong Yao Za Zhi* 34: 1717-1719, 2009 (In Chinese).
69. Wan D, Zhu HF, Luo Y, Xie P and Xu XY: Study of catalpol promoting axonal growth for cultured cortical neurons from rats. *Zhongguo Zhong Yao Za Zhi* 32: 1771-1774, 2007 (In Chinese).
70. Tian YY, An LJ, Jiang L, Duan YL, Chen J and Jiang B: Catalpol protects dopaminergic neurons from LPS-induced neurotoxicity in mesencephalic neuron-glia cultures. *Life Sci* 80: 193-199, 2006.



This work is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International (CC BY-NC-ND 4.0) License.