

CLC-2 is a positive modulator of oligodendrocyte precursor cell differentiation and myelination

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Abstract. Oligodendrocytes (OLs) are myelin-forming cells that are present within the central nervous system. Impaired oligodendrocyte precursor cell (OPC) differentiation into mature OLs is a major cause of demyelination diseases. Therefore, identifying the underlying molecular mechanisms of OPC differentiation is crucial to understand the processes of myelination and demyelination. It has been acknowledged that various extrinsic and intrinsic factors are involved in the control of OPC differentiation; however, the function of ion channels, particularly the voltage-gated chloride channel (CLC), in OPC differentiation and myelination are not fully understood. The present study demonstrated that CLC-2 may be a positive modulator of OPC differentiation and myelination. Western blotting results revealed that CLC-2 was expressed in both OPCs and OLs. Furthermore, CLC-2 currents (I_{CLC-2}) were recorded in both types of cells. The inhibition of I_{CLC-2} by GaTx2, a blocker of CLC-2, was demonstrated to be higher in OPCs compared with OLs, indicating that CLC-2 may serve a role in OL differentiation. The results of western blotting and immunofluorescence staining also demonstrated that the expression levels of myelin basic protein were reduced following GaTx2 treatment, indicating that the differentiation

of OPCs into OLs was inhibited following CLC-2 inhibition. In addition, following western blot analysis, it was also demonstrated that the protein expression of the myelin proteins yin yang 1, myelin regulatory factor, Smad-interacting protein 1 and sex-determining region Y-box 10 were regulated by CLC-2 inhibition. Taken together, the results of the present study indicate that CLC-2 may be a positive regulator of OPC differentiation and able to contribute to myelin formation and repair in myelin-associated diseases by controlling the number and open state of CLC-2 channels.

Introduction

In the central nervous system (CNS), myelin forms a medullary sheath around the membrane of axons and is developed by oligodendrocytes (OLs). Adult OLs are derived from oligodendrocyte precursor cells (OPCs) (1). The survival and differentiation of OLs is essential in axon myelination and myelin repair in diseases of the CNS that are caused by demyelination, which include multiple sclerosis, periventricular leukomalacia and adrenoleukodystrophy (2-5). Investigation of the underlying molecular mechanisms that regulate OPC differentiation is important in understanding myelin formation and repair in demyelination disorders.

Advances have been made towards understanding how the OPC differentiation process is controlled by various extrinsic and intrinsic factors (6,7). Axon-derived ligands, including Jagged/Notch (8), polysialylated-neural adhesion molecule (9) and leucine-rich repeat and immunoglobulin-like domain-containing protein 1 (10), have been reported to prevent OL differentiation and/or myelination. Transcription factors such as OL transcription factor (Olig)1, Olig2, myelin regulatory factor (MRF), sex-determining region Y-box (Sox)10, Sox2, Sox6, inhibitor of DNA binding (Id)-2, Id4, yin yang 1 (YY1) and Smad-interacting protein 1 (Sip1) function in the maintenance of an undifferentiated state in OPCs as they suppress myelin gene expression that is required for the generation of mature, post-mitotic OLs (11-16). Wnt/ β -catenin signaling has been reported to be a major regulator of OL development (17). Our previous study also demonstrated that glycogen synthase kinase 3 α may promote the differentiation

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of OPCs through a β -catenin-mediated mechanism of transcriptional regulation (18).

There are very few reports regarding the roles of ion channels in OL differentiation and myelination, although numerous types of ion channels are expressed in OPCs and OLs. Williamson *et al* (19) identified voltage-activated tetrodotoxin-sensitive Na^+ currents, two classes of voltage-activated outward K^+ currents, an inactivating inwardly rectifying K^+ current, a voltage-activated Cl^- current and three classes of Ca^{2+} currents in OL progenitors. Furthermore, an inward rectifying potassium channel (Kir4.1)-specific antibody was detected in children with acquired demyelinating CNS diseases (20), and Kir4.1 has been reported to be abnormally expressed in focal areas of patients with multiple sclerosis (21). Additionally, Ca^{2+} transients are considerably larger in mature OLs compared with OPCs, indicating that Ca^{2+} may be a signal for the initiation of myelin formation (22). The inhibition of voltage-gated chloride channel (CLC)-2, by 4,4'-diisothiocyanostilbene-2,2'-disulfonic acid (DIDS), is able to protect developing OLs in the white matter following ischemia-hypoxia injury in neonatal rats (23). However, the function of CLC-2 on OPC differentiation and myelination is not fully understood.

In the present study, the results demonstrated that CLC-2 may serve important roles in OPC development and differentiation. The development of OPCs may be inhibited by treatment with the CLC-2 specific inhibitor GaTx2. In addition, the inhibition of CLC-2 affected expression of myelin-associated proteins and may be involved in myelin development in the CNS; if CLC-2 gene was mutant or CLC-2 channel was blocked, myelin-associated proteins would be inhibited, and myelin formation would be affected.

Materials and methods

Ethics statement. Animals were treated according to the standards of the Animal Care and Use Committee of Zhejiang University. All procedures in the present study were approved by the Ethics Committee of Zhejiang University (Hangzhou, China).

Chemicals. Antibodies against CLC-2 (cat. no. sc-20122), YY1 (cat. no. sc-7341), Sip1 (cat. no. sc-48789) and MRF (cat. no. sc-137607) were purchased from Santa Cruz Biotechnology, Inc. (Dallas, TX, USA). The antibody against platelet-derived growth factor (PDGF) α receptor (PDGF α R; cat. no. 558774) was obtained from BD Biosciences (Franklin Lakes, NJ, USA). Antibodies against myelin basic protein (MBP; cat. no. AB5864), Olig2 (cat. no. AB109186), myelin-associated glycoprotein (MAG; cat. no. ST1684), 2',3'-cyclic nucleotide 3'-phosphodiesterase (CNP; cat. no. MAB326), Sox2 (cat. no. AB5603), Sox10 (cat. no. AB5727) and GAPDH (cat. no. CB1001) were purchased from EMD Millipore (Billerica, MA, USA). The marker of proliferation Ki67 (Ki67) antibody (cat. no. ab15580) was obtained from Abcam (Cambridge, UK). Triiodothyronine (T3) was from Sigma-Aldrich (Merck KGaA, Darmstadt, Germany). Dulbecco's modified Eagle's medium (DMEM), neurobasal medium, B27 supplements, proteinase inhibitor cocktail, horseradish peroxidase-conjugated secondary antibodies for western blotting, DAPI and Alexa Fluor-conjugated secondary

antibodies (Goat anti-Rabbit IgG (H+L) Cross-Adsorbed Secondary Antibody, Alexa Fluor 568, cat. no. A-11011; Goat anti-Rabbit IgG (H+L) Cross-Adsorbed Secondary Antibody, Alexa Fluor 488, cat. no. A-11008; Goat anti-Rat IgG (H+L) Cross-Adsorbed Secondary Antibody, Alexa Fluor 555, cat. no. A-21434) were from Invitrogen (Thermo Fisher Scientific, Inc., Waltham, MA, USA). GaTx2 was purchased from Smartox Biotechnology (Grenoble, France). Recombinant PDGF AA (cat. no. AF 100 13A) was purchased from PeproTech, Inc. (Rocky Hill, NJ, USA). Bicinchoninic acid and enhanced chemiluminescence (ECL) kits were from Pierce (SuperSignal™ West Femto Maximum Sensitivity Substrate; cat. no. 34096; Thermo Fisher Scientific, Inc.). All other chemicals were purchased from OriGene Technologies, Inc. (Rockville, MD, USA) unless stated otherwise.

OPC culture. Primary OPC cultures were modified from a previous publication (24) in which postnatal day 2-4 CD rats were dissected out, minced and digested for 15 min at 37°C in S-MEM. In the present study, postnatal day 1-3 rat cortices were digested for 20 min at 37°C in DMEM. Cortical glial cell mixtures were obtained from 10 newborn Sprague Dawley rats provided by Animal Center of Zhejiang University, Hangzhou, China (n=80, male, passage 1-3, weight, 5-10 g). Rats were kept in temperature and humidity-controlled conditions (22°C, 55-60% humidity) under 12-h light/dark cycle with food and water available *ad libitum*. Cells from 10 rats (5×10^5 cells per flask) were seeded into five 75 cm² tissue culture flasks in DMEM with 20% fetal bovine serum (cat. no. 12483012; Thermo Fisher Scientific, Inc.) for 10 days at 37°C and 5% CO₂. The culture medium was replaced every 3 days. Cultures were shaken for 1-2 h at 200 rpm to remove microglia, incubated in fresh DMEM medium with 20% fetal bovine serum for 4 h and subsequently shaken at 260 rpm at 37°C for 16-18 h for OPC collection. Detached cell suspension was collected and left to adhere in uncoated Petri dishes for 1 h at 37°C and 5% CO₂, which allowed microglia and astrocytes to firmly attach, while OPCs were loosely attached and were collected by gently shaking the dishes. Collected OPCs were re-plated at 5×10^3 - 5×10^4 cells/cm² onto poly-D-lysine-coated plates, dishes or coverslips in neurobasal medium supplemented with 2% B27. To expand OPCs and to keep them undifferentiated, the culture medium was supplemented with PDGF-AA (10 nM). For differentiation assays, OPCs were cultured in a differentiation medium (neurobasal medium containing 40 ng/ml T3 and 2% B27) for 3 days.

Treatment of cells by GaTx2. OLs or OPCs were seeded at 5×10^4 cells- 1×10^5 cells/well. To expand OPCs and to keep them undifferentiated, the culture medium was supplemented with PDGF-AA (10 nM) for 4 days. For differentiation assays, OPCs were cultured in medium with PDGF-AA (10 nM) for 1 day, then changed with medium containing 40 ng/ml T3 for another 3 days. For the control, only T3 was added, but for the experimental groups, T3 together with 10, 5, 2, 1, 0.2 or 0.1 nM GaTx2 were added for different time points (12, 24 and 72 h).

Immunocytochemistry. OLs or OPCs were seeded at 5×10^4 cells in 100 μ l/well on coverslips in 24-well plates. For immunostaining, cells were washed twice with PBS, fixed with

4% paraformaldehyde in PBS for 15 min at room temperature, washed three times with PBS and preincubated in 10% bovine serum albumin (cat. no. A8806; Sigma-Aldrich; Merck KGaA) and 0.4% Triton X-100 in PBS for 1 h at room temperature, prior to being immediately exposed to primary antibodies overnight at 4°C. The primary antibodies used for immunostaining were anti-PDGFR (1:500), anti-MBP (1:5,000), anti-Olig2 (1:1,000) and anti-Ki67 (1:1,000). Cultures were rinsed in PBS and incubated with fluorophore-conjugated secondary antibodies for 1 h at room temperature. The sections were mounted using ProLong Gold Antifade Reagent with DAPI (cat. no. P36931; Invitrogen; Thermo Fisher Scientific, Inc.) and immediately covered with coverslips. Immunocytochemical images were obtained with a fluorescence microscope (magnification, x20; Olympus Corporation, Tokyo, Japan) at a resolution of 1,360 x 1,024 pixels. A total of 10 fields were analyzed with ImageJ software version 1.49v (National Institutes of Health, Bethesda, MD, USA). The parameters used for microscopy were consistent in all experiments.

Western blotting. To determine the expression of certain proteins, OPC and OLs cultures were lysed in radioimmuno-precipitation assay buffer (cat. no. P0013C; Beyotime Institute of Biotechnology, Shanghai, China) containing a proteinase inhibitor cocktail and harvested. Following incubation at 4°C for 30 min, the lysates were centrifuged at 8,050 x g at 4°C for 30 min. Protein concentrations were measured in each sample using a BCA assay. Protein denaturation was subsequently performed in a sample buffer containing 2-mercapto ethanol and bromophenol blue for 10 min at 95°C. Equal quantities of protein (10 µg) were resolved by 10% SDS-PAGE and transferred onto polyvinylidene difluoride membranes (EMD Millipore). The membranes were blocked with 5% non-fat milk in TBS containing 0.1% Tween-20 for 1 h at room temperature and incubated at 4°C overnight with the indicated antibodies (anti-CLC-2, 1:200; anti-MBP, 1:5,000; anti-CNP, 1:10,000; anti-MAG, 1:1,000; anti-YY1, 1:500; anti-MRF, 1:1,000; anti-Sip1, 1:2,000; anti-Sox2, 1:2,000; and, anti-Sox10, 1:1,000). After washing with PBS, membranes were incubated with secondary antibodies at dilutions of 1:10,000 at room temperature for 1 h. The proteins were subsequently visualized using the ECL kit and membranes were exposed to X-ray films. The blotting signals were digitally scanned and quantified using ImageJ version 1.49v software (National Institutes of Health, Bethesda, MD, USA). The immunoreactivity of GAPDH (1:2,000) was set as the loading control. All data were normalized to the corresponding control.

Electrophysiology. Whole-cell currents in OPCs and OLs were measured using tight-seal whole-cell voltage-clamp techniques, as previously described (25). Data acquisition and command potentials were controlled by pCLAMP version 10.0 software (Molecular Devices, LLC, Sunnyvale, CA, USA). Whole-cell currents were filtered at 1 kHz and sampled at 5 kHz. Recording pipettes were prepared from borosilicate glass electrodes (1.5 mm outside diameter) with tip resistance of 1-3 MΩ when filled with pipette solutions. After a tight seal between the cell membrane and the pipette tip (seal resistance >10 GΩ) had been formed, the membrane patch was ruptured with brief additional suction. The capacitive transients elicited

by symmetrical 5-mV steps from 0 mV were recorded at 100 kHz for subsequent calculation of capacitance and access resistance. Input resistance was defined as the amplitude of the steady state voltage response divided by the current amplitude, and input conductance as the reciprocal value of input resistance. To account for differences in cell size, whole-cell currents were normalized to cell capacitance and the average data were reported as current densities (pA/pF). To isolate CLC-2 currents, cells were perfused with a Na⁺- and K⁺-free extracellular solution containing 145 mM NaCl, 3 mM KCl, 2 mM CaCl₂, 10 mM HEPES and 10 mM glucose [310 osmole (Osm); pH 7.4 adjusted with NaOH]. The pipette solution contained an intracellular solution containing 135 mM Cs methanesulfonate, 10 mM CsCl, 0.2 mM EGTA, 10 mM HEPES, 4 mM Na-ATP and 0.4 mM Na-GTP (310 Osm; pH adjusted to 7.3 with KOH). To elicit CLC-2 currents, voltage steps from +40 up to -140 mV from a holding potential of 0 mV were used. A final 50 msec voltage step at +40 mV was applied before returning to the holding potential. Cell cultures were stimulated with voltage pulses between -140 and +60 mV and the currents were recorded as the control. Experiments were conducted at room temperature. Averaging, normalizing and subtracting trace profiles were performed off-line using Clamp Fit version 10.2 (Mapu Scientific LLC, Guangzhou, China). For trace subtraction, current traces recorded from GaTx2 application were subtracted ('Arithmetic tool') from the respective currents from control condition to examine CLC-2-specific current/capacitance profiles in OPCs and OLs.

Statistical analysis. Data analysis was performed using Excel 2003 (Microsoft Corporation, Redmond, WA, USA), Igor Pro version 6.0 (Wave Metrics, Lake Oswego, OR, USA), and SPSS version 16.0 (SPSS, Inc., Chicago, IL, USA). Statistical differences were determined by Student's t-test for two-group comparisons or one-way analysis of variance followed by Tukey's post-hoc test for multiple comparisons. P<0.05 was considered to indicate a statistically significant difference. Data in the text and figures are presented as the mean ± standard error of the mean.

Results

Assessment of cultured OPCs and OLs. To examine the purity of OPCs and OLs, cells were immune stained with antibodies against PDGFR, an OPC specific marker, and Olig2, which is expressed in both OLs and OPCs. As demonstrated in Fig. 1A, the purity of oligodendrocytes (OPCs+OLs) was >85% (PDGFR+Olig2⁺/DAPI). To assess the differentiation state induced by T3 treatment for 3 days, the expression levels of the mature OL marker protein, MBP, was determined. It was demonstrated that MBP was expressed primarily in mature OLs, while Olig2 was present in OPCs and OLs (Fig. 1B). These results indicated that T3-induced differentiation of OPCs was successful.

Effects of CLC-2 inhibitor GaTx2 on CLC-2 currents. The presence of CLC-2 protein in OPCs and OLs was initially determined. Western blotting revealed that CLC-2 was expressed in both OPCs and mature OLs (Fig. 2A). The effects of CLC-2 inhibition in OPCs and OLs were subsequently

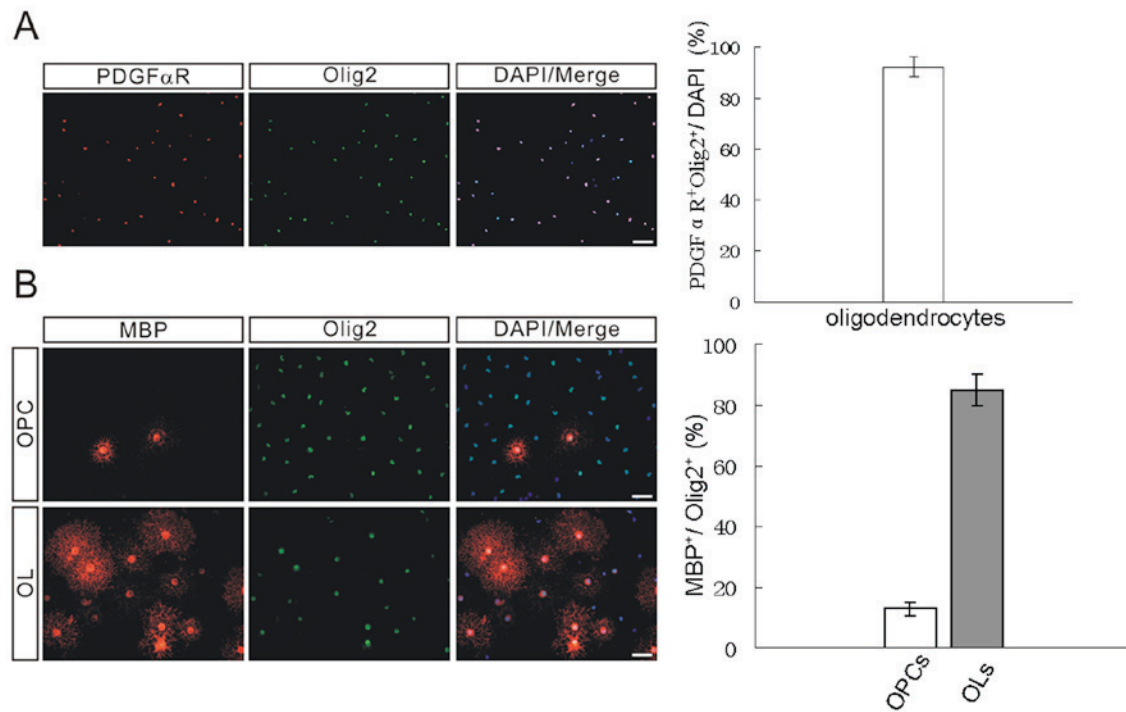


Figure 1. Assessment of cultured OPCs and OLs. (A) OPCs and OLs were immunostained with antibodies against PDGFR α and Olig2 to examine their expression levels in cultured cells. The purity of cultured OPCs reached >85%. Scale bar, 40 μ m. The purity of cultured OPCs was determined by the percentage of PDGFR α ⁺Olig2⁺ cells among DAPI-stained cells, as demonstrated in the bar graphs to the right (n=10). (B) Differentiation induced by T3 for 3 days as measured by MBP and Olig2 immunostaining. In OPCs, MBP⁺/Olig2⁺ is 13 \pm 2%, but in OLs, MBP⁺/Olig2⁺ is 85 \pm 5%. Scale bar, 20 μ m. OPCs, oligodendrocyte precursor cells; OLs, oligodendrocytes; PDGFR α , platelet-derived growth factor α receptor; Olig2, oligodendrocyte transcription factor 2; T3, triiodothyronine MBP; myelin basic protein.

examined using electrophysiological experiments. Cell cultures were stimulated with voltage pulses between -140 and +60 mV and the currents were recorded as the control (Fig. 2B). I_{CLC-2} were recorded in OPCs and OLs (left panels of Fig. 2C and D, respectively). GaTx2 (10 nM) was added to cells for 4 h to suppress CLC-2 activity. The alteration of currents in the control and GaTx2 treatment groups were presented in Fig. 2C and D. The left panels in Fig. 2C and D represented control CLC-2 currents. The middle panels represented the CLC-2 currents recorded after supplementing with 10 nM GaTx2. The right panels represented the currents of the middle panels subtracted from the left panels. The results indicated that residual currents following inhibition of CLC-2 with GaTx2 were higher in OLs compared with OPCs, indicating that CLC-2 may serve roles in OL differentiation. The curves of current and voltage association of OPCs and OLs were presented in Fig. 2E.

Effective concentration of GaTx2 in the inhibition of OL activity. To determine the most effective concentration and duration of GaTx2 treatment in OLs and OPCs, OL cultures were treated with different concentrations of GaTx2 (0.1, 0.2, 1, 2 and 5 nM) for different durations (12, 24 and 72 h). Western blot analysis demonstrated that MBP levels were decreased significantly at 12 and 24 h after 2 and 5 nM GaTx2 treatment compared with the control group, but no significant differences were observed at 72 h (Fig. 3A), which indicated that the effect of GaTx2 may be sustained only for 24 h. Therefore, the authors of the present study decided to add GaTx2 into cultured OPCs at the same

concentration (2 nM) every 24 h, and 40 ng/ml T3 was added for 72 h to induce differentiation. Immunofluorescence results demonstrated that following 5 nM GaTx2 treatment, the number of Olig2-positive cells decreased significantly compared with the control group, while 2 nM GaTx2 treatment had no effect on cell number, which indicated that 2 nM GaTx2 treatment had little effect on cell viability (Fig. 3B).

Effects of GaTx2 on OPC differentiation. As GaTx2 decreased CLC-2 currents in OPCs and OLs, it was investigated whether CLC-2 inhibition affected OPC differentiation by measuring the levels of myelin-associated proteins, including MBP, MAG and CNP. Control cells were treated with 10 nM PDGF-AA for 1 day and 40 ng/ml T3 was added for 3 days to induce differentiation. The experimental group was treated with T3 together with 2 nM GaTx2 for 72 h and GaTx2 was added every 24 h. The results revealed that CLC-2 inhibition with 2 nM GaTx2 substantially decreased the expression levels of MBP, while CNP was unchanged, compared with the control group (Fig. 4A). Following 2 nM GaTx2 treatment, myelin related glycoprotein (MAG) expressed as dMAG, a form of lower molecular weight without a protein modification function. Since the expression of Ki67 protein is closely associated with cell proliferation (26), Ki67 immunostaining was used to determine whether GaTx2 affected the proliferation of OPCs. The immunocytochemical results demonstrated that GaTx2 treatment decreased the percentage of MBP-positive cells (Fig. 4B), but not Ki67-positive cells (Fig. 4C), which indicated that GaTx2 may inhibit OPC differentiation but had no effect

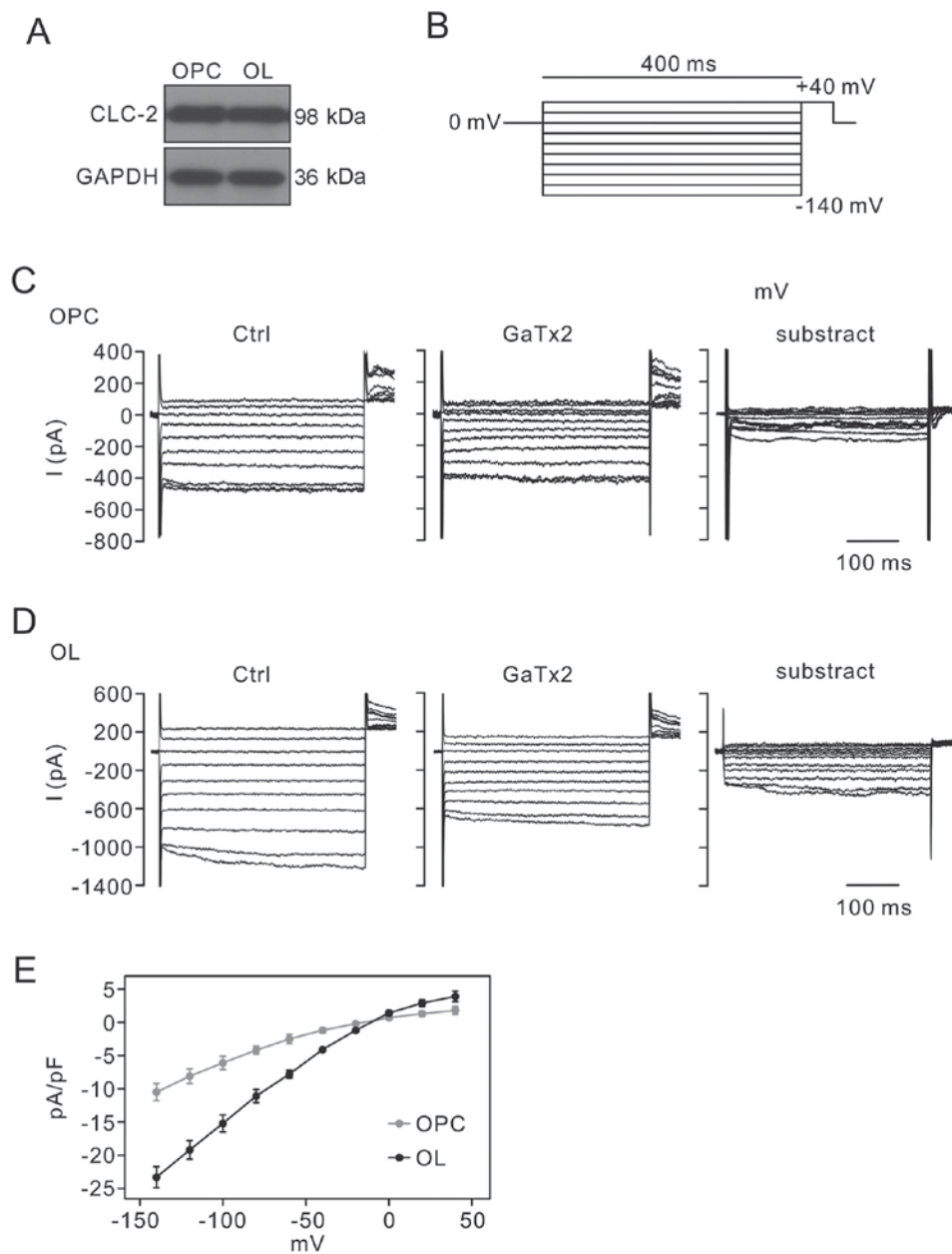


Figure 2. Effect of GaTx2 on CLC-2 current (I_{CLC-2}) in OPCs and OLs. (A) CLC-2 protein expression levels in OPCs and OLs. GAPDH served as the internal control. Bands were analyzed with ImageJ version 1.49v software (National Institutes of Health, Bethesda, MD, USA). The densities of the bands were: CLC2 in OPC (779.07 ± 21), OL (772.44 ± 19), GAPDH in OPC (628.09 ± 12) and OL (631.48 ± 9). (B) Schematic diagram of voltage stimulation mode. (C) OPCs and (D) OLs were stimulated with voltages between -140 and +60 mV in the three groups. I_{CLC-2} (right panel) was calculated by subtracting GaTx2 (10 nM)-sensitive currents (middle panel) from the control current (left panel). (E) Current to voltage association curves of I_{CLC-2} in OPCs and OLs. OPCs, oligodendrocyte precursor cells; OLs, oligodendrocytes; CLC-2, voltage-gated chloride channel 2.

on their proliferation. These data suggested that the inhibition of OPC differentiation by 2 nM GaTx2 is not mediated via altered cell survival or proliferation.

Effects of CLC-2 inhibition on transcriptional factors of OL differentiation. The present study demonstrated that MBP was significantly decreased by GaTx2 treatment. As an important myelin-associated protein in OLs, the expression of MBP is regulated by various transcriptional factors, including YY1, MRF, Sip1, Sox2 and Sox10. Therefore, the expression levels of several transcriptional factors following GaTx2 treatment were also measured by western blotting. The results

demonstrated that GaTx2 treatment decreased the levels of YY1, MRF, Sip1 and Sox10 proteins, which positively regulate MBP transcription (Fig. 5). However, the expression of Sox2, a negative transcriptional regulator of MBP expression, was unaltered (Fig. 5). These results indicated that inhibition of OPC differentiation by GaTx2 may be due to reduced expression levels of these transcription factors.

Discussion

CLC-2 is broadly expressed on the plasma membrane of various types of cells in the brain and epithelia (27). In the

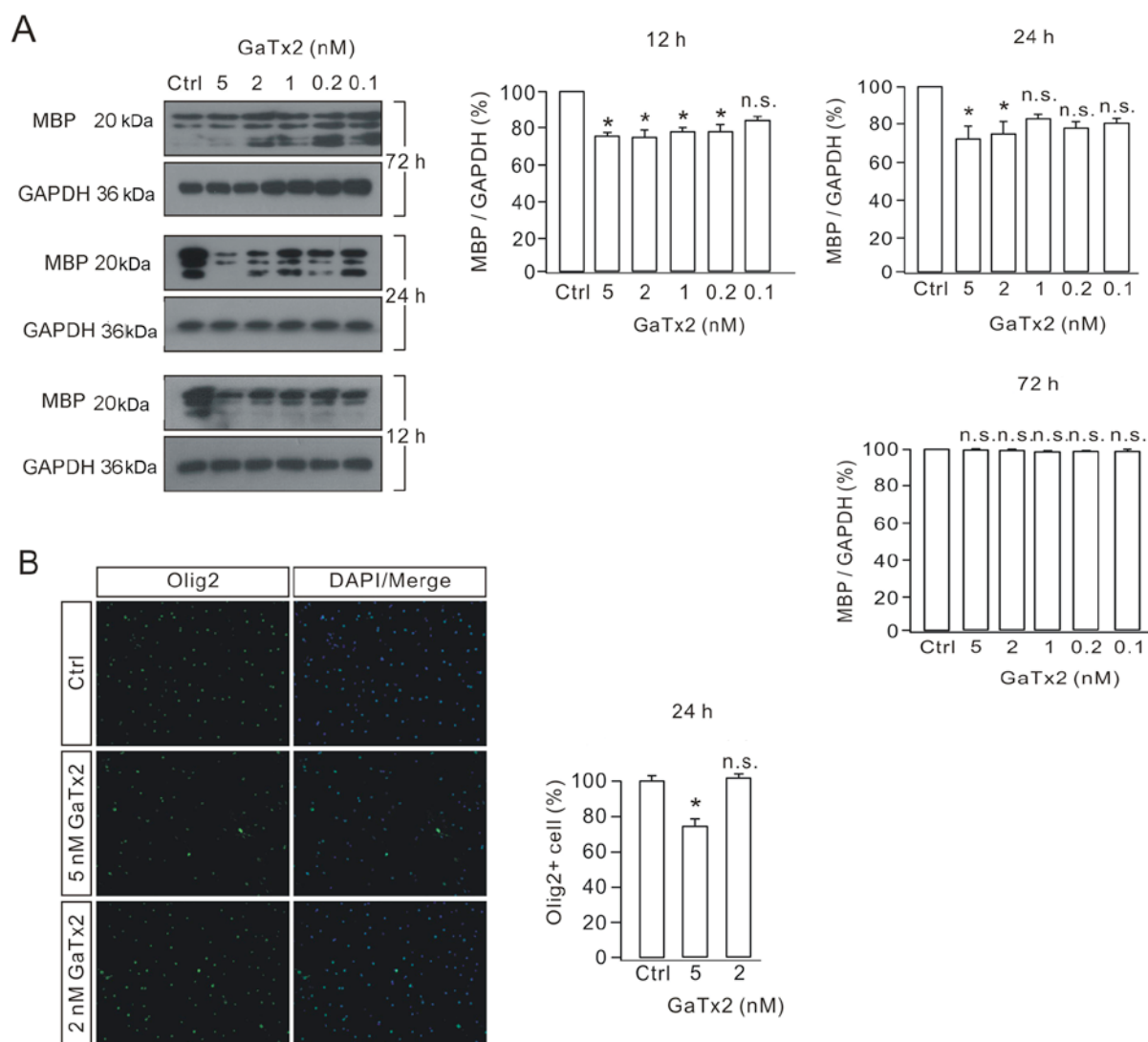


Figure 3. Effective concentration of GaTx2 in the inhibition of OL activity. (A) OL cultures were treated with different concentrations of GaTx2 for different durations. MBP expression levels were measured by western blotting and densitometric analysis was performed to quantify protein levels. (B) Olig2 immunostaining following 2 or 5 nM GaTx2 treatment for 24 h, indicating the difference compared with the control group. The experiment was repeated four times. Scale bar, 40 μ m. * P <0.05 vs. Ctrl. OLs, oligodendrocytes; MBP, myelin basic protein; Olig2, oligodendrocyte transcription factor 2; Ctrl, control; n.s., not significant.

CNS, CLC-2 is present on the endfeet of astrocytes and hippocampal neurons and is implicated in chloride homeostasis and Cl^- movements (28). The present study demonstrated that functional CLC-2 channels were observed in OPCs and mature OLs, and may be associated with the differentiation of OPCs and myelin gene expression. This conclusion was corroborated by results that were obtained following the pharmacological inhibition of CLC-2 in the present study. It was also revealed that the effect of CLC-2 inhibition on OPC differentiation was not a result of decreased proliferation levels (as measured by Ki67 expression). By contrast, activation of CLC-2 was sufficient to promote the differentiation of OPCs. Finally, it was demonstrated in the present study that CLC-2 inhibition reduced the expression levels of the transcription factors YY1, MRF, Sox10 and Sipl1, which are necessary for myelin protein expression (7,13,29). The aforementioned results of the present study indicated that CLC-2 may positively regulate the differentiation of

OPCs through regulation of the transcription of OPC differentiation factors.

It has been previously demonstrated that CLC-2 channels are involved in various cellular functions, including ion homeostasis, cell volume regulation, transepithelial transport, regulation of electrical excitability, cell migration and cell proliferation (30). Furthermore, CLC-2 channels have been indicated to be involved in the cell proliferation and invasive cell migration of primary brain tumors cells, particularly human glioma cells (31). As the majority of primary brain tumors arise from glial cells, glioma cell migration resembles progenitor cell migration during embryonic brain development (32,33). In the CNS, OLs are responsible for the formation of myelin, and OLs are produced following OPC differentiation and maturation (7,34). The present study demonstrated that OPCs successfully differentiated into mature OLs with T3 supplement; however, inhibition of CLC-2 channels repressed OPCs differentiation, which was

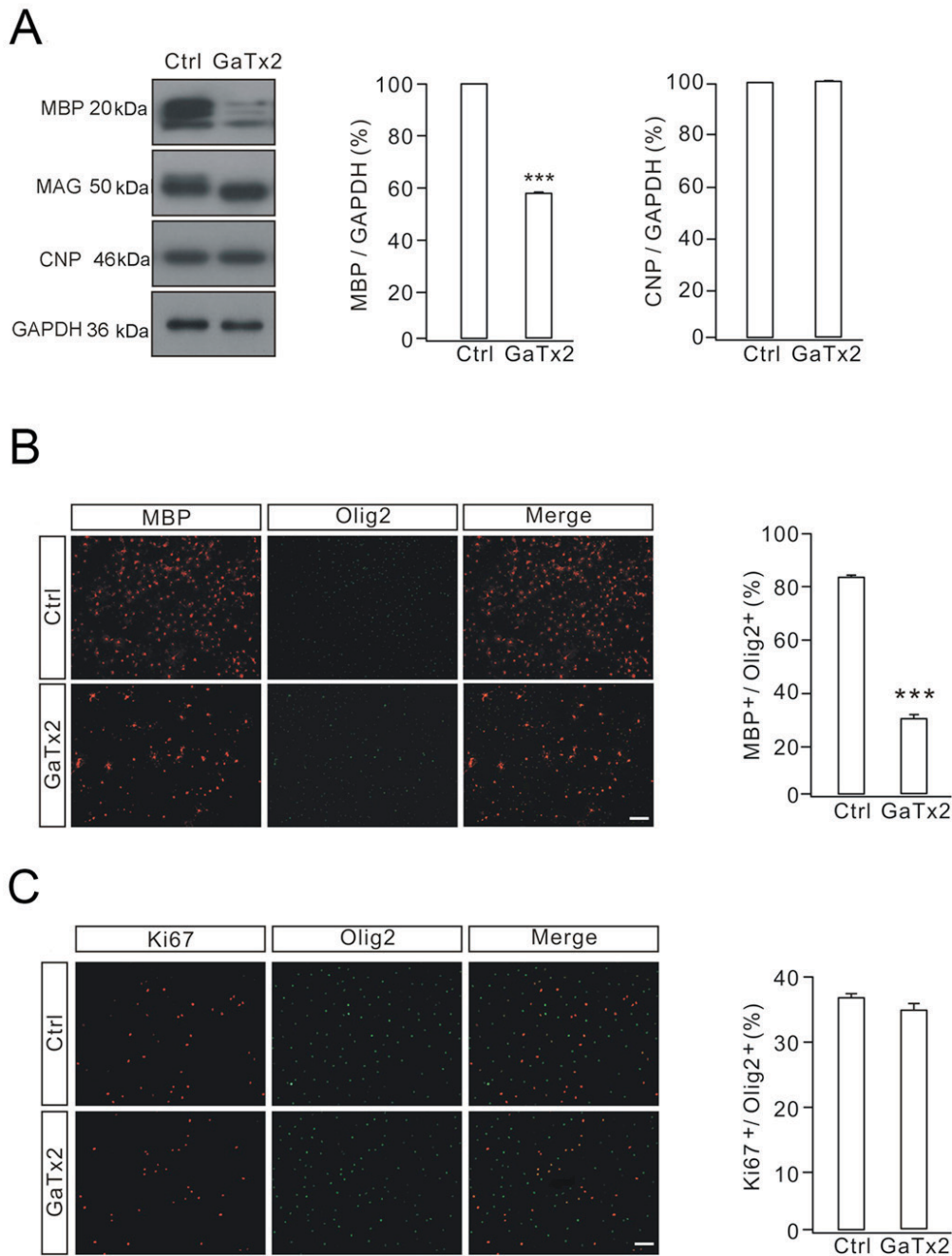


Figure 4. Effects of GaTx2 on OL differentiation. (A) MBP, MAG and CNP protein expression levels were assessed by western blotting. Control cells were treated with PDGF-AA for 1 day and T3 was added in for 3 days. In the experimental group, cells were treated with T3 together with GaTx2 (2 nM for 72 h). (B) Immunocytochemical staining of MBP and Olig2 indicated that GaTx2 may inhibit OL differentiation. (C) Immunocytochemical staining of Ki67 indicated that GaTx2 had no effect on OPC proliferation. The experiment was repeated four times. Scale bar, 40 μ m. ***P<0.001 vs. Ctrl. OLs, oligodendrocytes; MBP, myelin basic protein; MAG, myelin-associated glycoprotein; CNP, 2',3'-cyclic nucleotide 3'-phosphodiesterase; T3, triiodothyronine; Olig2, oligodendrocyte transcription factor 2; Ki67, marker of proliferation Ki67; OPCs, oligodendrocyte precursor cells; Ctrl, control.

consistent with a previous report where mice lacking CLC-2 (*Clcn2*^{-/-} mice) exhibited abnormal morphology of the white matter and resembled the pathology of patients with megalencephalic leukoencephalopathy with subcortical cysts (35). By contrast, the excessive activation of CLC-2 may lead to cell damage. A report concerning the white matter of neonatal rats demonstrated that ischemia-hypoxia elevated the activation of CLC-2 and subsequently initiated apoptosis in OLs (23). He *et al* (36) reported that, in the white matter of newborn rats

that presented with gestational diabetes mellitus, activation of CLC-2 resulted in apoptosis and myelination inhibition via repression of the phosphatidylinositol 3-kinase-protein kinase B signaling pathway. DIDS was demonstrated to prevent high glucose-induced damage to white matter damage, which may be due to the fact that following the differentiation of OPCs into early OLs, these immature OLs are susceptible to injury factors, leading to excessive activation of CLC-2 channels and enhanced Cl⁻ currents (36).

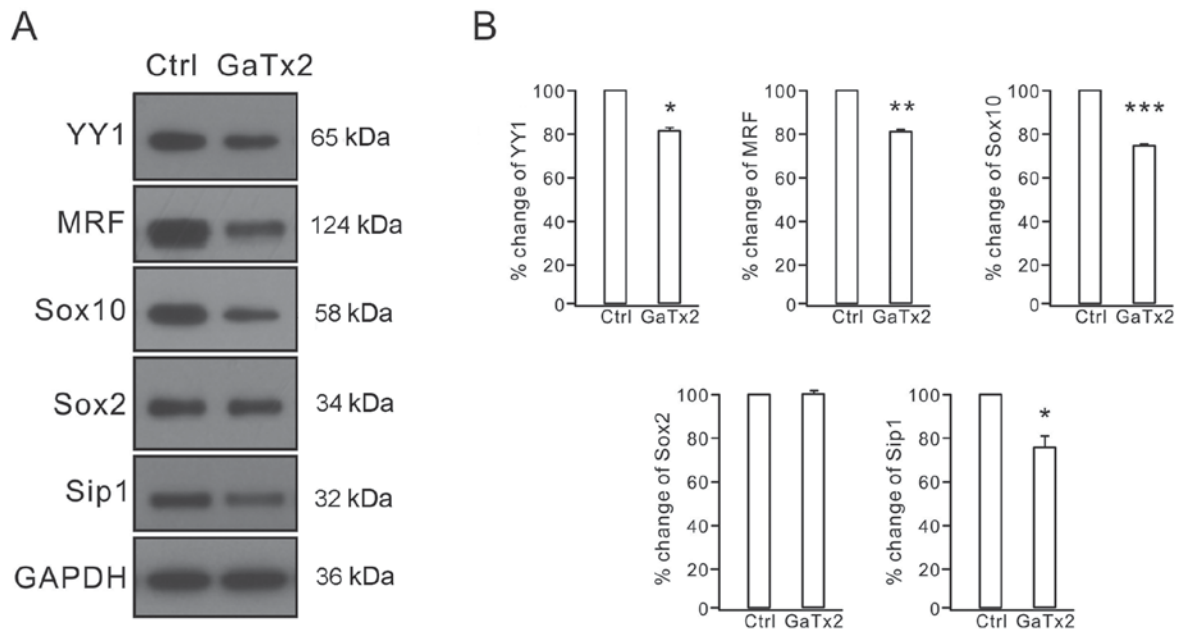


Figure 5. CLC-2 inhibition altered the protein expression levels of transcription factors involved in OL differentiation. (A) Protein expression levels of the transcription factors YY1, MRF, Sip1, Sox2 and Sox10 were measured in control and GaTx2 groups by western blotting. (B) Densitometric analysis was performed to quantify the protein levels following western blotting. Levels of transcription factors in the control were set at 100% and the corresponding percentage changes of these proteins are presented. The experiment was repeated three times. * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$ vs. Ctrl. CLC-2, voltage-gated chloride channel 2; OLs, oligodendrocytes; YY1, yin yang 1; MRF, myelin regulatory factor; Sip1, Smad-interacting protein 1; Sox, sex-determining region Y-box; Ctrl, control.

It has been demonstrated that altered expression of CLC-2 is associated with alterations in Cl^- currents (37). *In vitro* single cell recordings indicated that high concentrations of glucose led to enhanced opening of CLC-2 on cell membranes (38,39). In the present study, the patch-clamp recordings demonstrated that Cl^- currents in OLs were higher compared with those in OPCs, indicating that CLC-2 may serve roles in OL differentiation. Recordings from mouse astrocytes demonstrated that CLC-2-mediated current density was significantly lower in astrocytes isolated from immature brain and in reactive astrocytes within a lesion site but functional expressed in mature astrocytes isolated from adult mouse brain, indicating that as in oligodendrocytes, CLC-2 channels also served a role in astrocyte development and differentiation (40). In neurons, CLC-2 channels were reported to potentially constitute part of the background conductance serving as a safeguard mechanism to prevent chloride accumulation at synapses (41). It may be hypothesized that CLC-2 channels may have additional functions in mature OLs that do not occur in undifferentiated OPCs. Therefore, the role of CLC-2 in glial cells requires further investigation.

A previous study has demonstrated that the differentiation of OPCs is regulated by the induction of several transcription factors, most notably Sox10, Olig2, Sip1, MRF and YY1 (7), which are present in OPCs and in post-mitotic OLs, while Sox2 has been reported to function in the maintenance of an undifferentiated state and the repression of myelin gene expression in OPCs, which is consistent with the results of the present study. Specifically, in the current study, the inhibition of CLC-2 channels led to reduced expression levels of Sox10, Sip1, MRF and YY1, while the expression levels of Sox2, a transcriptional repressor, were unchanged.

Based on these results, enhancing the expression of positive transcription factors may be a major role of CLC-2 channels in OPC differentiation. However, the mechanism underlying the regulatory effect of CLC-2 on these transcription factors requires further investigation.

In conclusion, the present study demonstrated that voltage-gated CLC-2 was associated with OPC differentiation and myelin gene transcription factors during development. The effect of CLC-2 inhibition on OPC differentiation by its specific inhibitor GaTx2 was not a result of altered proliferation. Furthermore, CLC-2 inhibition reduced the expression of various myelin protein transcription factors, including YY1, MRF, Sox10 and Sip1. Taken together, the results of the current study indicated that CLC-2 may serve as an important positive regulator in OPC differentiation by regulating the expression of myelin gene transcription factors. The present study only performed *in vitro* experiments on cells. *In vivo* studies on animals are required to demonstrate the effects of CLC-2 on myelin development, injuries and repair. This will provide the clinical target for myelin-related diseases.

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