

Puerarin promotes MIN6 cell survival by reducing cellular reactive oxygen species

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Abstract. Type 1 diabetes is caused by destruction of the pancreatic β -cells and, to date, no cure has been developed. Promoting the survival of pancreatic β -cells may be beneficial for patients with type 1 diabetes. Puerarin is an estrogen analogue that been demonstrated in previous studies to be able to decreased blood glucose in patients with type 1 diabetes. Similar results were demonstrated in previous studies which additionally demonstrated that puerarin was able to decreased blood glucose in type 1 diabetic mice by protecting pancreatic β -cells. However, the mechanism underlying the function of puerarin in pancreatic β -cells remains unclear. Therefore, the present study sought to investigate the detailed function of puerarin in pancreatic β -cells. In the present study, H_2O_2 was used to induce apoptosis. It was observed that puerarin significantly decreased H_2O_2 -induced apoptosis in mouse insulinoma MIN6 cells. It was additionally observed that puerarin decreased the levels of intracellular reactive oxygen species and mitochondrial superoxide in MIN6 cells. The protective effect of puerarin was markedly decreased by 6-aminonicotinamide, an inhibitor of glucose-6-phosphate dehydrogenase (G6PD). In conclusion, the results of the present study suggested that puerarin may increase the activity of G6PD, decreased the level of oxidative stress in MIN6 cells, protect mitochondria and promote MIN6 cell survival. Investigating the mechanism underlying the effect of puerarin in MIN6 cells may provide a novel approach for development of a cure for type 1 diabetes.

Introduction

Diabetes is a chronic disease that is common worldwide. According to its pathogenesis, diabetes can be broadly divided into type 1 and type 2 diabetes (1). Type 1 diabetes is caused by the destruction of β -cells, resulting in an inefficient level of insulin secretion, while type 2 diabetes is due to insulin resistance (2). There are a number of potential causes of pancreatic β -cell damage, including hypoxia, oxidative stress, glycosylation end-products and autoimmune reactions (3-6). However, the detailed mechanism underlying pancreatic β -cell injury in diabetes remains unclear. Therefore, the present study aimed to investigate a novel drug which may protect pancreatic β -cells against metabolic stress. Decreasing pancreatic β -cell damage is an essential part of treating type 1 diabetes.

Puerarin is an analogue of estrogen that was first isolated from *Pueraria lobata*. Previous studies have demonstrated that puerarin effectively decreased blood glucose in rats with type 1 diabetes (7,8). An additional previous study demonstrated that estrogen was able to effectively promote the survival of the transplanted pancreatic β -cells (9-11). Since puerarin is an analogue of estrogen (12-14), and puerarin is able to decrease blood glucose in type 1 diabetic rats and mice (15), it was hypothesized that puerarin may be of benefit in protecting mouse insulinoma MIN6 cells from external stress, while promoting cell survival. However, the detailed mechanism underlying the role of puerarin in diabetes remains unclear. Investigation of the mechanism of puerarin is conducive to the treatment of type I diabetes.

Due to the requirement for pancreatic β -cells to synthesize a large amount of insulin, pancreatic β -cells are frequently exposed to oxidative stress (16,17). Therefore, oxidative stress was considered to be among the primary causes of pancreatic β -cell apoptosis. The present study used H_2O_2 to induce intracellular oxidative stress, in order to investigate the protective effect of puerarin on MIN6 cells. In H_2O_2 -induced apoptosis experiments, it was observed that puerarin significantly decreased apoptosis, and the levels of intracellular reactive oxygen species (ROS) and mitochondrial superoxide (MitoSOX). It was additionally observed that the ability of puerarin to protect MIN6 cells was decreased by 6-aminonicotinamide (6AN). Puerarin in MIN6 cells may promote the activity of glucose-6-phosphate dehydrogenase (G6PD), thereby reducing intracellular oxidative

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stress; puerarin may therefore protect against apoptosis induced by H₂O₂ in pancreatic β -cells.

Materials and methods

Cell culture. MIN6 cells were purchased from ATCC (Manassas, VA, USA) and cultured in a 37°C incubator (Sanyo, Osaka, Japan) containing 5% CO₂. MIN6 cells were grown in high glucose Dulbecco's modified Eagle's medium (HyClone; GE Healthcare Life Sciences, Logan, UT, USA) containing 15% fetal bovine serum (HyClone; GE Healthcare Life Sciences), 1% penicillin-streptomycin (HyClone; GE Healthcare Life Sciences) and 0.2% β -mercaptoethanol (Sigma-Aldrich; Merck KGaA, Darmstadt, Germany). The cells were cultured overnight before treatment with puerarin, H₂O₂ and 6AN. Puerarin, 6AN and H₂O₂ were purchased from Sigma-Aldrich (Merck KGaA). Briefly, prior to experiments, MIN6 cells (1x10⁴ cells) were seeded in 96-well plates overnight. Then, puerarin (100 μ M) was used to pre-treat the cells for 6 h during the experiments. Finally, H₂O₂ (100 μ M) and 6AN (50 μ M) were used to treat the cells for 24 h at the same time and cultured in an incubator at 37°C.

Terminal deoxynucleotidyl transferase dUTP nick end labelling (TUNEL) assay and cell viability. The cells were homogeneously seeded in 24-well plates at 2x10⁵ cells per well with slides and cultured. Precooled 0.01 M PBS was used to wash the cells twice. MIN6 cells were fixed with 4% paraformaldehyde for 30 min at room temperature. Cells were subsequently permeabilized in 0.1% Triton-x 100 in 0.01 M PBS. TUNEL (Roche Diagnostics, Indianapolis, IN, USA) staining materials were mixed according to the manufacturer's protocol, and were subsequently added to the fixed cells in the incubator at 37°C for 60 min. The cells were washed three times with 0.01 M PBS. Finally, 0.3 mM DAPI was added for staining of the nuclei at room temperature for 3 min. Each sample was observed in 5 different fields, with a magnification of x400. The cells were washed three times with 0.01 M PBS. Cell viability was detected using a Cell Counting Kit-8 (CCK-8) assay (Dojindo Molecular Technologies, Inc., Kumamoto, Japan).

Cell ROS and MitoSOX detection. MIN6 cells were cultured in 24-well plates until 90% confluence, the cells were washed with 0.01 M PBS. The corresponding volumes (5 μ l) of CellROS (Invitrogen; Thermo Fisher Scientific, Inc., Waltham, MA, USA) and Hoechst (Guangzhou Ribobio Co., Ltd., Guangzhou, China) were added to MIN6 cells at 37°C for 30 min. Subsequently, the cells were washed three times with 0.01 M PBS, observed and photographed under a microscope, with a magnification of x400. MitoSOX (Invitrogen; Thermo Fisher Scientific, Inc.) and Hoechst were added to 24-well plates at 37°C for 15 min, and cells were washed three further times with 0.01 M PBS, observed and photographed under a microscope, with a magnification of x400.

Western blotting. MIN6 were cells grown in the 6-well plates and washed by precooled 0.01 M PBS. The cells were lysed with radioimmunoprecipitation assay lysis buffer (EMD Millipore, Billerica, MA, USA) and placed on ice for 30 min. Cells in lysis buffer were centrifuged at 15,000 x g and 4°C for 15 min. The

supernatant was collected and the concentration of the total protein in the supernatant was measured using a bicinchoninic acid kit (Thermo Fisher Scientific, Inc.). Loading buffer was added to the supernatant and boiled for 10 min. Protein samples (30 μ g per lane) were subjected to electrophoresis on a 10% SDS-PAGE gel, at 80 V and constant pressure. The total proteins in the gel were transferred onto polyvinylidene fluoride (PVDF) membranes (EMD Millipore) at 300 mA constant current for 90 min. The PVDF membranes were blocked in 5% non-fat milk (Cell Signaling Technology, Inc., Danvers, MA, USA) for 1 h at room temperature. The primary antibodies against cleaved caspase3 (cat. no. 9664; Cell Signaling Technology, Inc.), β -actin (cat. no. 3700; Cell Signaling Technology, Inc.) and G6PD (cat. no. 12263; Cell Signaling Technology, Inc.) were diluted 1:1,000. All the primary antibodies were incubated at 4°C for 8 h. The secondary antibodies Goat Anti-Rabbit IgG (cat. no. A9169; Sigma-Aldrich; Merck KGaA) and Goat Anti-Mouse IgG (cat. no. A8924; Sigma-Aldrich; Merck KGaA) were diluted 1:10,000. The secondary antibodies (HRP conjugated) were incubated at 25°C for 1 h. Enhanced chemiluminescence liquid (Lulong, Inc., Xiamen, China) was added to the membranes and images were captured. The results of Western blots were quantified by densitometry using ImageJ software version 1.41 (National Institutes of Health, Bethesda, MA, USA).

Statistical analysis. All results were analyzed using GraphPad Prism version 5.0 software (GraphPad Software, Inc. La Jolla, CA, USA). All experiment results were analyzed using a one-way analysis of variance followed by a post hoc Bonferroni test for multiple comparisons. The results are expressed as the mean \pm standard error of the mean. P<0.05 was considered to indicate a statistically significant difference.

Results

H₂O₂ reduces MIN6 cell viability. In MIN6 cells, the effects of different concentrations of H₂O₂ on the viability of the cells were measured. The results demonstrated that the viability of MIN6 cells was significantly decreased as the concentration of H₂O₂ increased (Fig. 1A). Similarly, we also examined the effects of different concentrations of puerarin on the viability of MIN6 cells. Puerarin exerted no apparent effects on the viability of MIN6 cells under normal conditions (Fig. 1B). Puerarin neither promoted nor inhibited cell viability.

Puerarin decreases H₂O₂-induced cellular apoptosis. According to the results of the CCK-8 assay, a concentration of 100 μ M H₂O₂ was selected for use in subsequent experiments (Fig. 2). MIN6 cells were treated with 100 μ M H₂O₂ and different concentrations of puerarin were added to the cells. Cell viability was detected using a CCK-8 assay. It was observed that low concentrations of puerarin were not able to restore cell viability. A concentration of 100 μ M puerarin markedly restored the vitality of the cells (Fig. 2C). The effect of puerarin on the apoptosis of MIN6 cells was additionally examined. H₂O₂ induced apoptosis in MIN6 cells and puerarin decreased cellular apoptosis (Fig. 2A). The results of the present study indicated that a high concentration of puerarin decreased H₂O₂-induced apoptosis and restored cell viability (Fig. 2B).

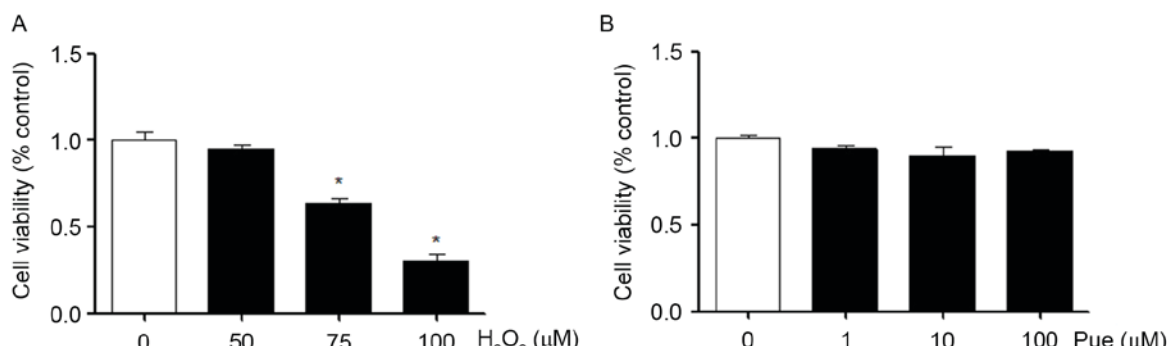


Figure 1. H₂O₂ inhibits MIN6 cells viability. (A) H₂O₂ and (B) puerarin was used to treat MIN6 cells in different concentrations. Cell viability was measured using a Cell Counting Kit-8 assay. Each experiment was repeated six times. *P<0.05 vs. control. Pue, puerarin.

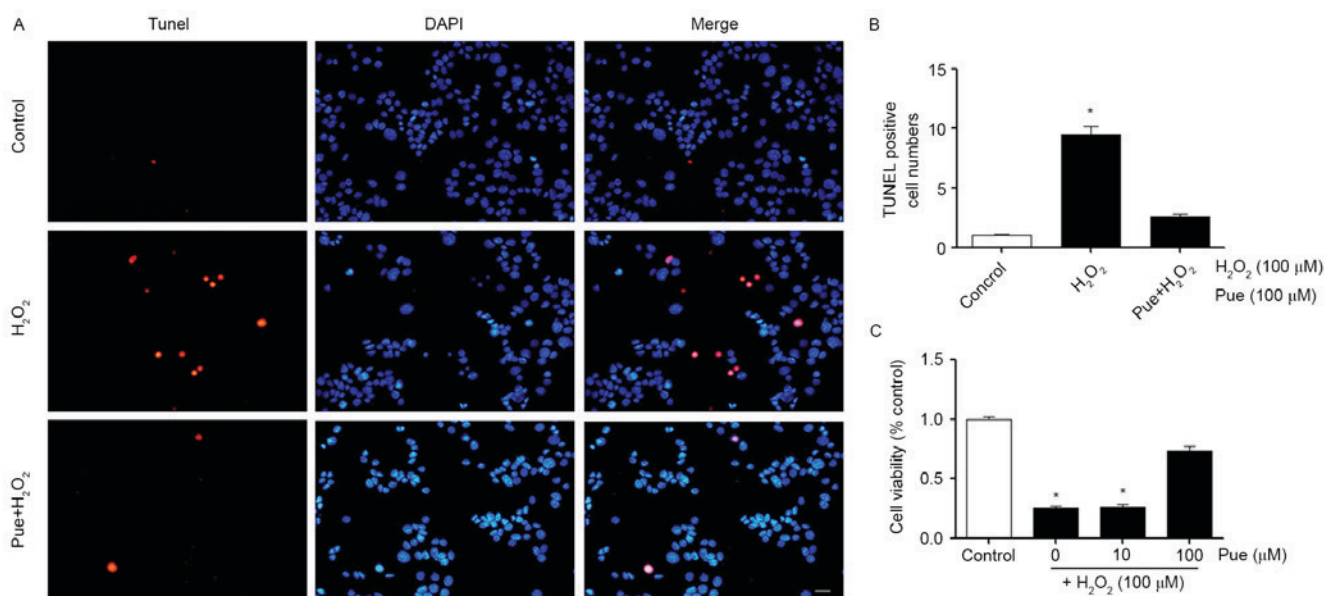


Figure 2. Puerarin protects MIN6 cells against H₂O₂ induced apoptosis. (A) Puerarin significantly reduced MIN6 cellular apoptosis induced by H₂O₂. (B) The positively-stained cells were counted using Image J software. (C) The effect of puerarin in different concentrations was measured in MIN6 cells exposed to oxidative stress. Each experiment was repeated six times. *P<0.05 vs. control. Scale bar, 20 μ m. Pue, puerarin; TUNEL, terminal deoxynucleotidyl transferase dUTP nick end labelling.

Puerarin decreases H₂O₂-induced intracellular ROS levels.

It is established that H₂O₂-induced apoptosis is primarily caused by intracellular oxidative stress. Puerarin can reduce H₂O₂-induced apoptosis, suggesting that puerarin can reduce oxidative stress in MIN6 cells. The stress induced by exposure to H₂O₂ was detected by measuring the intracellular levels of ROS, and it was observed that intracellular ROS levels were increased in MIN6 cells following treatment with H₂O₂ (Fig. 3). Puerarin markedly decreased ROS levels in MIN6 cells. A small amount of ROS is beneficial for cells (18), although excessive production of ROS may cause cell damage, including protein abnormalities, DNA damage and mitochondrial damage (19-21). Following examination of the production of MitoSOX, it was observed that H₂O₂ caused an increase in MitoSox levels (Fig. 4A). ROS may cause mitochondrial damage. The expression of cleaved caspase 3 was subsequently detected, due to mitochondrial injury-induced caspase 3 activation. Consistent with expectations, puerarin decreased the expression of cleaved caspase 3 (Fig. 4B). Puerarin prevented mitochondrial damage induced by H₂O₂ in MIN6 cells.

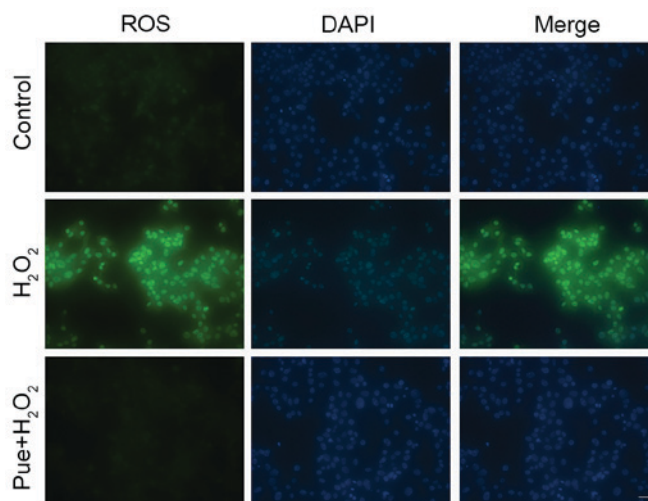


Figure 3. Puerarin decreases H₂O₂-induced cellular ROS generation. H₂O₂ increased MIN6 cellular ROS, while puerarin diminished cellular ROS. Each experiment was repeated three times. *P<0.05 vs. control. Scale bar, 20 μ m. Pue, puerarin; ROS, reactive oxygen species.

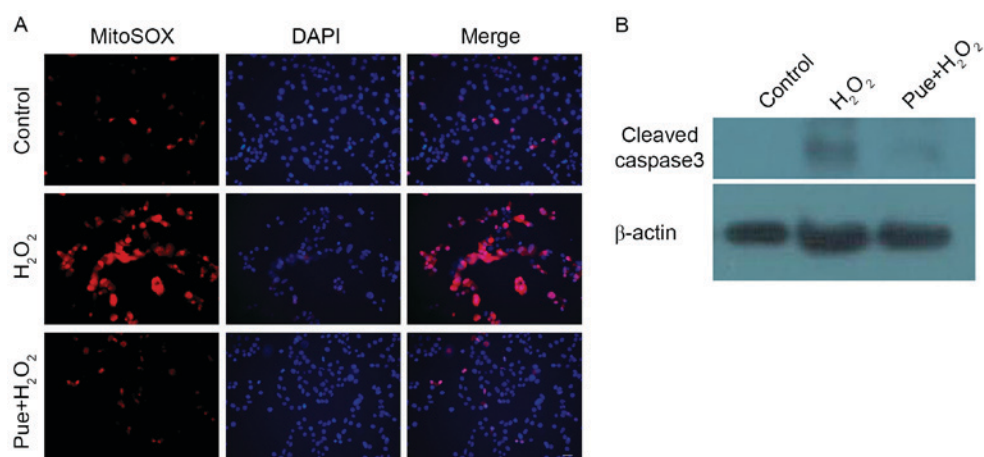


Figure 4. Puerarin protects mitochondria against H₂O₂. (A) MitoSOX was accumulated following treatment with H₂O₂, and puerarin decreased the levels of MitoSOX. (B) Cleaved caspase3 expression was detected via western blotting. Each experiment was repeated three times. Scale bar, 20 μ m. Pue, puerarin; MitoSOX, mitochondrial superoxide.

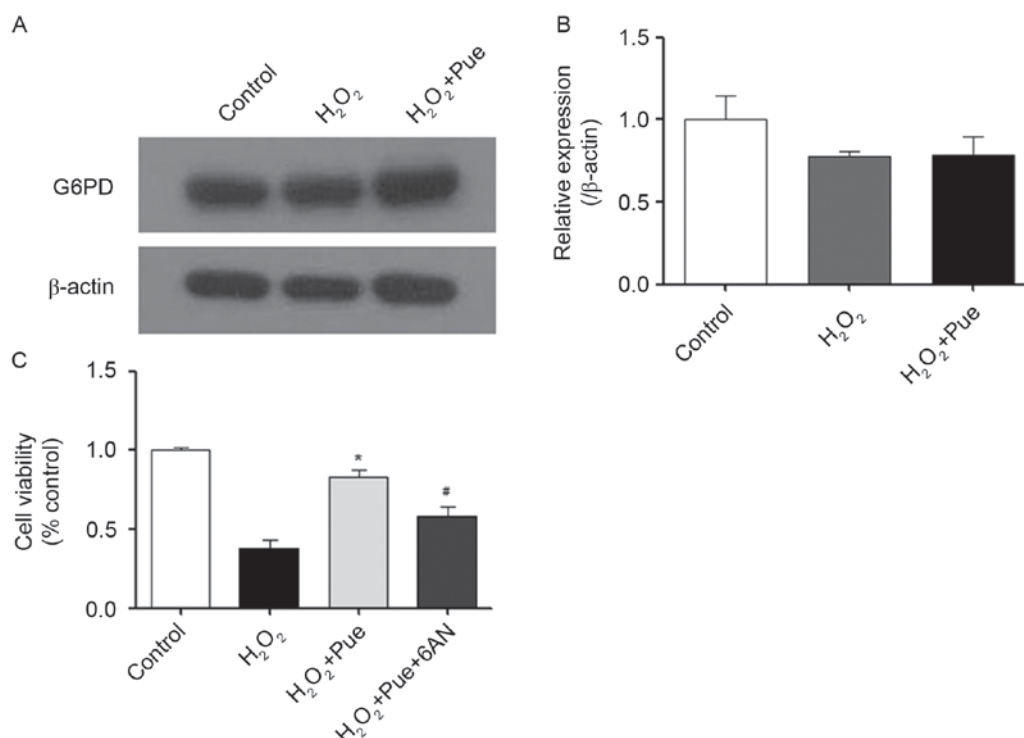


Figure 5. Puerarin protects MIN6 cell survival by increasing G6PD activity. (A) The expression of G6PD was decreased following treatment with H₂O₂, and puerarin did not influence the expression of G6PD. (B) The relative expression of G6PD was measured using Image J software. (C) The protective effect of puerarin in MIN6 cells was diminished by 6AN. Each experiment was repeated six times. * $P < 0.05$ vs. H₂O₂; # $P < 0.05$ vs. the H₂O₂ + Pue group. Scale bar, 20 μ m. Pue, puerarin; 6AN, 6-aminonicotinamide; G6PD, glucose-6-phosphate dehydrogenase.

Protective role of puerarin is inhibited by 6AN. Puerarin protected MIN6 cells against oxidative stress caused by H₂O₂. The present study further explored the mechanism underlying the effect of puerarin against oxidative stress. NADPH is considered to be the major intracellular reductive force. NADPH is produced by G6PD (22). Therefore, the effect of puerarin on G6PD expression was examined. It was observed that the expression of G6PD did not alter following treatment with puerarin (Fig. 5). Puerarin did not promote the expression of G6PD protein levels. 6AN was used to inhibit G6PD activity (23). The ability of puerarin to resist oxidative stress

was reduced significantly by 6AN (Fig. 5C). Therefore, it may be hypothesized that puerarin serves a role in the promotion of G6PD enzymatic activity, which may facilitate resistance to intracellular oxidative stress.

Discussion

Puerarin is able to effectively decreased blood glucose in type 1 diabetic mice and rats (8,15). However, the detailed mechanism underlying the protective effect of puerarin in pancreatic β -cells remains to be elucidated. Therefore, the present study

investigated the mechanism of puerarin in MIN6 cells treated with H_2O_2 . Using H_2O_2 to induce oxidative stress in MIN6 cells, it was observed that puerarin significantly decreased intracellular ROS levels and superoxide in the mitochondria. The protective effect of puerarin was attenuated by 6AN. It was determined that puerarin was able to decrease the intracellular ROS level, in part by regulating the activity of G6PD.

Type 1 diabetes is a disorder caused by damage to pancreatic β -cells, primarily. There have been a number of hypotheses formulated to explain the pathophysiology of pancreatic β -cell injury; these include hypoxia, oxidative stress and autoimmunity. However, the detailed mechanism through which pancreatic β -cells become damaged remains unclear. Ameliorating pancreatic β -cell damage may be beneficial in treating the development and course of type 1 diabetes. Previously, it has been demonstrated that puerarin may decrease blood glucose in type 1 diabetic rats (7). A previous study additionally demonstrated that puerarin may decrease blood glucose in type 1 diabetic mice and decrease hypoxia-induced MIN6 cell damage via activity in the phosphatidylinositol 3-kinase/RAC- α serine/threonine protein kinase signaling pathway (15). However, the detailed mechanism underlying the function of puerarin in promoting MIN6 cell survival during treatment with H_2O_2 remains unclear. The present study demonstrated that puerarin promoted the activity of G6PD and decreased the apoptosis of MIN6 cells induced by H_2O_2 . The results of the present study suggested that the promotion of G6PD activity in MIN6 cells or additional NADPH expression in MIN6 cells may reduce cell damage. At present, there is no cure for type 1 diabetes except islet transplantation; the limitation of islet transplantation is largely due to the number of available donors required to provide islets. In islet cells transplantation, the use of puerarin as an adjuvant may prove useful in promoting the survival of viable transplant pancreatic β -cell tissue.

In addition, when G6PD activity was inhibited, puerarin continued to exert a partial protective effect on MIN6 cells. This result suggested that puerarin may exhibit other functions in addition to promoting G6PD activity. Puerarin may promote other intracellular oxidoreductase activity or expression. Detection of the influence of puerarin on intracellular oxidoreductases may enhance the understanding of puerarin, which may serve as a means to provide a novel method for protecting pancreatic β -cells. In particular, stimulating the activity of a different variety of reductase within cells may effectively ameliorate the damage to pancreatic β -cells. Autophagy may also decrease the levels of intracellular ROS. Puerarin may promote autophagy to decrease intracellular oxidative stress. A more detailed mechanism of puerarin in pancreatic β -cells requires further investigation. Additionally, the function of puerarin in type 2 diabetes remains unknown, and future studies may examine this in more detail.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

TW, SL and SY conceived and designed the study. YL, HM, CH, WW and ZX performed the experiments. TW and HM wrote the paper. XL and SL analyzed the raw data. XL, SL and SY reviewed and edited the manuscript. SY agrees to be accountable for all aspects of the work. All authors read and approved the final manuscript.

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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