Downregulation of lncRNA H19 inhibits migration and invasion of human osteosarcoma through the NF-κB pathway

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Abstract. The present study aimed to investigate the role of long non-coding RNA (lncRNA) H19 in the development of osteosarcoma and to determine the underlying mechanism involved. A total of 40 patients with osteosarcoma were selected and the expression level of H19 in tumor tissue and adjacent healthy tissue was detected by reverse transcription-quantitative polymerase chain reaction. Survival curves were plotted using the Kaplan-Meier method to investigate the prognostic value of H19 expression level for patients with osteosarcoma. H19 knockdown osteosarcoma cell lines were constructed using small interfering (si)RNA transfection. Cell migration and invasion abilities were measured by Transwell migration and invasion assays, respectively. Western blot analysis was performed to detect the expression levels of phosphatidylinositol 3-kinase (PI3K), phospho (p)-PI3K, RAC-alpha serine/threonine-protein kinase (AKT), p-AKT and NF- κ B inhibitor α (I κ B α) in osteosarcoma cells transfected with H19 siRNA. Expression level of H19 was significantly elevated in tumor tissue compared with adjacent healthy tissue. Expression level of H19 was positively associated with distant metastasis of osteosarcoma (P<0.01), but not with gender and age. Overall survival of patients with osteosarcoma with high H19 level was significantly shorter compared with patients with low H19 expression (P<0.05). H19 knockdown significantly reduced migration and invasion ability of osteosarcoma cells. Significantly decreased levels of p-PI3K and p-AKT, and elevated level of IkBa were observed in H19 knockdown osteosarcoma cells compared with control osteosarcoma cells, while no significant differences in levels

Correspondence to: Dr Shi-Tang Ma, Food and Drug College, Anhui Science and Technology University, 9 Donghua Road, Fengyang, Anhui 233100, P.R. China E-mail: shitangma123@gmail.com of PI3K and AKT were observed. Therefore, the present study demonstrated that knockdown of lncRNA H19 can inhibit migration and invasion of human osteosarcoma cells by inhibiting the nuclear factor- κ B pathway.

Introduction

Although osteosarcoma is a relatively rare type of cancer, it is considered to be one of the major causes of cancer-associated mortality among children and young adults (1). In the USA, ~1,000 new cases of osteosarcoma are reported each year (2). Young patients are typically diagnosed with primary conventional osteosarcoma, while secondary osteosarcoma is more common among elderly patients (2). Osteosarcoma primarily affects the long bones, but other bones in the body can also be affected (3). Similar to other tumors, development of osteosarcoma is a complex process with various signaling pathways involved (4). Therefore, in-depth analyses of signal transduction involved regulation in tumorigenesis may aid in elucidation of molecular mechanisms underlying the onset, development and progression of osteosarcoma.

The role of noncoding RNAs in the development of various pathophysiological conditions has been well characterized (5). Long non-coding RNAs (lncRNA) are a type of noncoding RNAs of >200 nucleotides in length (6). lncRNAs have roles in the onset and development of different human cancer types, including breast cancer, non-small-cell lung cancer and colorectal cancer (7-10). Expression of lncRNA H19 has been reported to be positively associated with progression of various malignant human tumors (11,12). It has been reported that H19 is frequently overexpressed during the development of osteosarcoma (13) and that H19 can promote migration and invasion of osteosarcoma cells (14). However, the molecular mechanism underlying the function of H19 in osteosarcoma remains to be elucidated.

In the present study, expression of H19 in osteosarcoma tumor tissue and adjacent healthy tissue was detected. Effects of H19 on migration and invasion of multiple osteosarcoma cells lines were determined. Association between H19 and the NF- κ B signaling pathway was investigated. In addition, prognostic value of expression of H19 for patients with osteosarcoma was also evaluated.

Key words: osteosarcoma, long noncoding RNA H19, migration, invasion, phosphatidylinositol 3-kinase/phosphorylated-AKT pathway, nuclear factor-κB pathway

Materials and methods

Patients. A total of 40 patients with osteosarcoma were selected in Tianjin Medical University Cancer Institute and Hospital from January 2010 to January 2013. All patients were diagnosed based on pathological and imaging examinations. Inclusion criteria were: i) Patient pathologically diagnosed as osteosarcoma and ii) patients willing to cooperate with researchers. Exclusion criteria were: i) Patient with other malignancies; ii) patients with metal disorders; iii) patients who had received treatment prior to admission. Surgical resection was performed for all patients. The patients included 18 males and 22 females (age range 8-66 years; mean age of 31±8.1 years). Distant metastasis was observed in 22 patients. During surgical operations, cancer tissues and normal tissues within the region 0.5 cm around tumor were collected for subsequent experiments. The present study was approved by the Ethics Committee of Tianjin Medical University Cancer Institute and Hospital (Tianjin, China), and all patients signed informed consent. A follow-up study was performed for 48 months to monitor the survival.

Cell lines and cell culture. Human MG-63, U2OS and SAOS-2 osteosarcoma cell lines, and hFOB normal bone cell line, were purchased from American Type Culture Collection (Manassas, VA, USA). MG-63 and SAOS-2 cells were cultured with Eagle's minimum Essential medium (cat. no. 30-2003; American Type Culture Collection) containing 10% heat-inactivated fetal bovine serum (Thermo Fisher Scientific, Inc., Waltham, MA, USA). U2OS and SAOS-2 cells were cultured with McCoy's 5a modified medium (cat. no. 30-2007; American Type Culture Collection) containing 10% fetal bovine serum (Thermo Fisher Scientific, Inc.). All cells were cultured at 37°C and 5% CO₂. Cells were harvested during logarithmic growth phase for subsequent experiments.

Reverse transcription-quantitative polymerase chain reaction (RT-qPCR). TRIzol reagent (Invitrogen; Thermo Fisher Scientific, Inc.) was used to extract total RNA from tissues and cells. Concentration and quality of RNA samples were determined using a NanoDrop[™] 2000 Spectrophotometer (Thermo Fisher Scientific, Inc.). Only RNA samples with a ratio of A260/A280 between 1.8 and 2.0 were used for reverse transcription to synthesize cDNA using SuperScript III Reverse Transcriptase (Thermo Fisher Scientific, Inc.). Reaction conditions for reverse transcription were: 25°C for 5 min, 55°C for 20 min and 80°C for 10 min. SYBR®-Green Real-Time PCR Master Mix (Thermo Fisher Scientific, Inc.) was used for qPCR. The following primers were used in PCR reactions: 5'-TGAGCTCTCAGGAGGGAGGATGG-3' (forward) and 5'-TTGTCACGTCCACCGGACCTG-3' (reverse) for H19; 5'-GACCTCTATGCCAACACAGT-3' (forward) and 5'-AGT ACTTGCGCTCAGGAGGA-3' (reverse) for β -actin. PCR reactions were performed using CFX96 Touch™ Real-Time PCR Detection System (Bio-Rad Laboratories, Inc., Hercules, CA, USA). The following thermocycling conditions were used for the PCR: Initial denaturation at 95°C for 40 sec; 40 cycles of 95°C for 15 sec and 60°C for 45 sec. Relative expression was calculated using $2^{-\Delta\Delta Cq}$ method (15). Relative expression level of H19 was normalized to endogenous control β -actin.

Establishment of H19-silenced cell lines. H19 small interfering RNA (siRNA; cat. no. 1299001; Thermo Fisher Scientific, Inc.) and *Silencer*[™] Select Negative Control No. 1 siRNA (cat. no. 4390843; Thermo Fisher Scientific, Inc.) were used. Cells were cultured overnight to reach 80-90% confluence prior to transfection. Transfection (50 nM siRNA) with Lipofectamine[®] 2000 reagent (Invitrogen; Thermo Fisher Scientific, Inc.) was performed according to the manufacturer's protocol. Cells were cultured for another 6 h before subsequent experiments.

Cell migration and invasion assay. Transwell cell migration assay (BD Biosciences, Franklin Lakes, NJ, USA) was performed to measure cell migration ability. Briefly, $5x10^4$ cells in 100 μ l serum-free Dulbecco's modified Eagle medium (Thermo Fisher Scientific) were transferred to the upper chamber, while the lower chamber was filled with RPMI-1640 medium (Thermo Fisher Scientific, Inc.) containing 20% fetal calf serum (Sigma-Aldrich; Merck KGaA, Darmstadt, Germany). Membranes were collected 24 h later and stained with 0.5% crystal violet at room temperature (Sigma-Aldrich; Merck KGaA) for 20 min. Stained cells were counted under a light microscope (Olympus Corporation, Tokyo, Japan). Invasion assay was performed using the same method with the upper chamber was pre-coated with Matrigel (EMD Millipore, Billerica, MA, USA).

Western blot analysis. Total protein extraction was performed using RIPA buffer (Thermo Fisher Scientific, Inc.), and protein concentration was measured using THE bicinchoninic acid method. Subsequently, 30 μ g of protein was subjected to 10% SDS-PAGE gel electrophoresis, and transferred to polyvinylidene fluoride membrane at 20 V for 1 h. Following blocking with 5% skimmed milk at room temperature for 1 h, membranes were washed with TBST (0.1% Tween 20) 3 times for 15 min each time. Membranes were subsequently incubated with primary antibodies, including rabbit anti-p-PI3K (1:2,000; cat. no. ab182651), anti-PI3K (1:2,000; cat. no. ab5451; Abcam), anti-p-AKT (1:2,000; cat. no. ab18206; Abcam), anti-AKT (1:2,000; cat. no. ab126811; Abcam), anti-NF-ĸB inhibitor a (IkBa; 1:1,000; cat. no. ab76429; Abcam) and anti-GAPDH (1:1,000; ab8245; all Abcam, Cambridge, UK) overnight at 4°C. Subsequently, membranes were washed three times with TBST, 15 min each time. Membranes were incubated with anti-rabbit immunoglobulin G-horseradish peroxidase conjugated secondary antibody (1:1,000; cat. no. MBS435036; MyBioSource, Inc., San Diego, CA, USA) at room temperature for 2 h. Following washing twice with TBST, 15 min each time, Enhanced Chemiluminescence detection reagent (Sigma-Aldrich; Merck KGaA) was used to detect the signals. Images were analyzed using Image J software version 1.8.0 (National Institutes of Health, Bethesda, MD, USA) to calculate relative expression level of each protein relative to endogenous control β -actin.

Statistical analysis. SPSS software (version 19.0; IBM Corp., Armonk, NY, USA) was used for all statistical analysis. Normal distribution data are presented as the mean ± standard deviation. Comparisons between two groups were performed using t test and comparisons among multiple groups were performed using one-way analysis of variance and least

	Group	Total no.	H19 exp		
Clinicopathological feature			High	Low	P-value
Sex	Male	18	8	10	0.79
	Female	22	12	10	
Age (years)	>25	13	6	7	0.72
	≤30	27	11	16	
Distant metastasis	Yes	22	17	5	< 0.01
	No	18	4	14	

Table I. Correlation between the expression level of H19 and clinicopathological feature	Table I.	Correlation	between the e	expression l	evel of	H19 an	d clinicopat	hological	features.
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Table II. Multivariate regression analysis of factors associated with mortality of patients with osteosarcoma.

Clinicopathological feature	Risk ratio	95% CI	P-value
Gender (male)	1.02	0.61-5.52	0.33
Age (>40)	3.83	2.12-7.73	0.04
Distant metastasis	6.92	2.85-12.82	0.01
High H19 expression level	6.53	2.27-9.58	0.01
CI, confidence interval.			

significant difference test. Data that were not normally distributed were analyzed using non-parametric Mann-Whitney U test. In multivariate logistic regression analysis, mortality was set as dependent variable, and gender (being a male), age (>40), distant metastasis and high H19 expression level were independent variables. Survival curves were plotted using Kaplan-Meier method and compared using log-rank test. With the median expression level of H19 in cancer tissues as the cutoff score, 40 patients with osteosarcoma were divided into high expression level group (n=20) and low expression level group (n=20), and the association between the expression level of H19 and clinicopathological features was analyzed by χ^2 test. P<0.05 was considered to indicate a statistically significant difference.

Results

Comparison of expression levels of H19 in cancer tissues and adjacent healthy tissues. Expression levels of H19 in cancer tissues and adjacent healthy tissues of 40 patients with osteosarcoma were detected using RT-qPCR. Expression levels of H19 were significantly increased in cancer tissues of all 40 patients compared with the respective adjacent healthy tissues, indicating the potential role of H19 in development of osteosarcoma (Fig. 1).

Association between expression of H19 and clinicopathological features, and the prognostic values. Univariate analysis demonstrated that the expression level of H19 was positively associated with distant metastasis (P<0.01), not with gender and age (Table I). Multivariate regression analysis



Figure 1. Expression of H19 in tumor tissues and adjacent healthy tissues. *P<0.05 vs. the respective adjacent healthy tissue. Inc, long noncoding RNA.



Figure 2. Overall survival curves plotted using Kaplan-Meier method for patients with high and low expression level of H19.

demonstrated that aging, distant metastasis and high expression level of H19 were associated with mortality of patients with osteosarcoma (P<0.05; Table II). Survival curves were plotted using the Kaplan-Meier method to evaluate the prognostic value of H19 for osteosarcoma. Survival curves were



Figure 3. Effects of H19 knockdown on osteosarcoma cell migration and invasion. (A) Expression of H19 in different cell lines following different treatments. (B) Effects of H19 knockdown on osteosarcoma cell migration (magnification, x20). (C) Effects of H19 knockdown on osteosarcoma cell invasion (magnification, x20). Relative cell number was normalized to the control group, which was set as 100. Scale bar=20 μ m. *P<0.05. NC, negative control; siRNA, small interfering RNA.



Figure 4. Effects of H19 knockdown on NF- κ B pathway-associated proteins. Effects of H19 knockdown on NF- $\kappa\beta$ pathway-associated proteins in (A) MG-63, (B) U2OS and (C) SAOS-2 cell lines. *P<0.05 vs. the control group. NF- κ B, nuclear factor- κ B; C, control; siRNA, small interfering RNA; p-, phospho-; PI3K, phosphatidylinositol 3-kinase; AKT, RAC-alpha serine/threonine-protein kinase; I κ B α , NF- κ B inhibitor α .

compared using log-rank test. The overall survival rate of patients with high expression level of H19 was significantly lower compared with patients with low expression level of H19 (P<0.05; Fig. 2).

Effects of H19 knockdown on osteosarcoma cell migration and invasion. Expression level of H19 was significantly increased in human osteosarcoma cell lines MG-63, U2OS and SAOS-2 compared with normal bone cell line hFOB (all P<0.05; Fig. 3A). Following siRNA transfection, expression level of H19 was significantly reduced in all three osteosarcoma cell lines and in the normal hFOB cell line compared with the controls (all P<0.05). Following siRNA H19 transfection, migration ability was significantly reduced in all osteosarcoma cell lines compared with control cells (all P<0.05). Similar results were observed in the invasion assay, where invasion ability of all osteosarcoma cell lines was significantly reduced following siRNA transfection (all P<0.05). However, no significant differences in migration and invasion abilities were identified in hFOB cells with and without H19 knockdown (Fig. 3B and C).

Effects of H19 knockdown on the NF-κB signaling pathway. The PI3K/AKT signaling pathway can activate the NF-κB pathway by regulating degradation of IκBα. No significant differences in levels of PI3K and AKT were identified between the control and H19 knockdown groups of MG-64 (Fig. 4A), U2OS (Fig. 4B) and SAOS-2 (Fig. 4C) cells. Levels of p-PI3K and p-AKT were decreased significantly in cells transfected with siRNA compared with control cells (P<0.05), suggesting inactivation of the PI3K/AKT signaling pathway following H19 knockdown. In addition, the level of IκBα protein increased significantly in cells transfected with siRNA compared with control cells (NAC) (Fig. 4B) and SAOS-2 (Fig. 4C) (F

Discussion

The role of lncRNA in the development of osteosarcoma has been demonstrated by numerous studies (14,16). Sun *et al* (14) reported that hepatocellular carcinoma up-regulated long non-coding RNA (HULC) was upregulated in osteosarcoma tissues compared with adjacent healthy tissues, and increased expression level of HULC was associated with a shorter overall survival of patients with osteosarcoma. By contrast, downregulation of HULC significantly reduced proliferation, migration and invasion of in vitro cultured osteosarcoma cells (14). lncRNA taurine up-regulated 1 (TUG1) was also demonstrated to inhibit apoptosis and promote proliferation of osteosarcoma cells, and TUG1 may be a novel target for treatment of osteosarcoma (16). H19 is an oncogenic lncRNA and was demonstrated to be involved in the progression of various types of human cancer (11,12). A previous study reported that activation of the Hedgehog signaling pathway upregulated the expression of H19, which in turn promoted tumorigenesis (13). Consistent with previous studies, in the present study, the expression level of H19 in 40 osteosarcoma patients was increased markedly in tumor tissue compared with adjacent healthy tissue. The results suggest that H19 may be involved in development of osteosarcoma. In addition, the expression level of H19 was positively associated with the occurrence of distant metastasis of osteosarcoma, but not with gender or age, and the overall survival of patients with osteosarcoma exhibiting high H19 expression significantly shorter compared with patients with low expression H19. Thus, the results of the present study suggest that increased expression of H19 may have a prognostic role for patients with distant metastasis of osteosarcoma.

lncRNA H19 serves a role in the progression of different types of cancer primarily by promoting tumor metastasis. Shi *et al* (17) reported that miR-675 derived from H19 can promote invasion of glioma cells. In another study, H19 promoted migration and invasion of human hepatocellular carcinoma cells by activating with the AKT/GSK-3 β /Cdc25A signal transduction pathway (18). In the present study, H19 knockdown reduced migration and invasion abilities of three osteosarcoma cell lines. The results of the present study suggest that expression level of H19 is positively associated with invasion and migration abilities of osteosarcoma cells.

NF-κB signaling has roles in numerous aspects of initiation and progression of tumorigenesis (19). Previous studies demonstrated that H19 can interact with the NF-κB pathway to perform its biological functions (20). It has also been demonstrated that the NF-κB pathway can be activated by the PI3K/AKT signaling pathway via the regulation of degradation of IκBα (21,22). In the present study, phosphorylation levels of PI3K and AKT decreased, and expression level of IκBα increased following knockdown of H19, while no significant differences in expression levels of PI3K and AKT were identified between osteosarcoma cells with and without H19 knockdown. The above results suggest that knockdown of H19 can mediate inactivation of the NF-κB pathway by inhibiting the activation PI3K/AKT pathway.

In conclusion, expression level of H19 increased in osteosarcoma tissues compared with adjacent healthy tissues. Expression level of H19 was positively associated with the occurrence of distant metastasis, and elevated expression level of H19 indicated poor prognosis of patients with osteosarcoma. Expression of H19 affected migration and invasion of osteosarcoma by activating the NF- κ B pathway. Future studies should focus on identifying targets of H19 to further elucidate the

underlying mechanism of H19 in osteosarcoma. The present study was limited by small sample size. Further studies with larger sample sizes are necessary to confirm the conclusions of the present study.

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Availability of data and materials

All data generated or analyzed during this study are included in this published article.

Authors' contributions

MST designed the experiments. ZJ performed the experiments. MST and ZJ analysed the data. MST wrote the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The present study was approved by the Ethics Committee of Tianjin Medical University Cancer Institute and Hospital (Tianjin, China), and all patients signed informed consent.

Consent for publication

All participants signed informed consent.

Competing interests

The authors declare that they have no competing interests.

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