

# Analysis of virulence diversity of 73 *Helicobacter pylori* strains isolated in Guizhou province, China

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**Abstract.** The present study aimed to investigate the virulence diversity of *Helicobacter pylori* (*H. pylori*) in major ethnic groups residing in Guizhou province, China, and its association with clinical outcomes. Gastric mucosal biopsies were collected from the pylorus of patients with gastrointestinal disorders. *H. pylori* was identified by colonial morphology, Gram staining, a urease test and *H. pylori*-specific *16S rRNA* gene fragment PCR amplification. DNA was extracted from pure culture and used for virulence gene analysis. The cytotoxin associated gene A (*cagA*), vacuolating cytotoxin A (*vacA*) and induced by contact with epithelium gene A (*iceA*) genes were analyzed by polymerase chain reaction analysis. The *cagA* gene was further analyzed through sequencing of the C-terminal region containing EPIYA motifs, and phylogenetic analysis of the *cagA* C-terminal variable region was performed using MEGA 6.0 software. In the present study, 73 *H. pylori* strains were isolated from clinical samples. *cagA* genotypes were detected in all strains, namely *cagA*-AB, -ABC, -ABD and -BD genotypes were found in five (6.85%), three (4.11%), 63 (86.30%) and two (2.74%) isolates, respectively.

Phylogenetic analysis showed that there was a clustering association between the *cagA*-AB and *cagA*-ABC genotypes, and between the *cagA*-ABD and *cagA*-BD genotypes. In terms of the frequency of the four EPIYA or EPIYA-like motifs, the most predominant was EPIYA (92.92%), followed by EPIYT (3.77%), ESIYA (2.83%) and ESIYT (0.47%). The predominant *vacA* genotype was *slc/m2* (65.75%), and the predominant *iceA* genotype was *iceA1* (79.45%). There were no associations between the *H. pylori* *cagA*, *vacA* or *iceA* genotypes and clinical outcomes. No significant difference was found in the distribution of these genotypes according to the age, ethnicity or location of residence of patients. In conclusion, *H. pylori* isolated from patients in Guizhou region, China, showed a unique genotype, which was mainly East Asia-type *cagA* (ABD), *vacA* *slc/m2* genotype or *iceA1*-positive. These results provide important information on the distribution of *H. pylori* virulence genotypes in Guizhou province, China.

## Introduction

*Helicobacter pylori* (*H. pylori*) is a Gram-negative, spiral-shaped, microaerophilic bacterium that colonizes the human gastric mucosa (1). *H. pylori* infection can cause chronic gastritis, peptic ulcer disease, gastric carcinoma and mucosa-associated lymphoid tissue lymphoma (2-4). More than half of the world's population is infected with *H. pylori* (5). However, the majority of infected individuals remain asymptomatic. There is an interplay between host genetic susceptibility, environmental factors and bacterial virulence factors, which influences the outcome of *H. pylori*-associated diseases (6-8). Polymorphisms of several virulence genes, including cytotoxin-associated gene A (*cagA*), vacuolating toxin A (*vacA*), induced by contact with epithelium gene A (*iceA*), blood group antigen binding adhesin and duodenal ulcer promoting gene A are considered to increase the risk for the development of upper gastrointestinal diseases (9). Among these genes, *cagA* and *vacA* have been investigated extensively.

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**Abbreviations:** *H. pylori*, *Helicobacter pylori*; *cagA*, cytotoxin associated gene A; *vacA*, vacuolating cytotoxin A; *iceA*, induced by contact with epithelium gene A; PCR, polymerase chain reaction

**Key words:** *Helicobacter pylori*, virulence, cytotoxin associated gene A, vacuolating cytotoxin A, induced by contact with epithelium gene A

CagA is an oncogenic protein, and is a major virulence factor associated with gastric cancer (10,11). CagA is injected into host epithelial cells via the type IV secretion system encoded by cag pathogenicity island (cagPAI) following *H. pylori* infection (12). CagA undergoes phosphorylation at its EPIYA motif by the Abl and Src family tyrosine kinases (13). Once tyrosine is phosphorylated, the cagA EPIYA motifs serve as a recognition site for Src homology phosphotyrosine phosphatase 2 (SHP2), and activate an intracellular signal transduction pathway that leads to cytoskeletal rearrangement and cell elongation, known as the 'hummingbird' phenotype (14). This increases the risk of developing precancerous lesions (15).

The EPIYA motif is located within the C-terminal of cagA, and is formed of the conserved amino-acid residues Glu-Pro-Ile-Tyr-Ala. EPIYA can be further classified into four types of motif: EPIYA-A, -B, -C and -D, based on the flanking amino acid sequences (16). *H. pylori* is classified as East Asian-type cagA and western-type cagA, according to the composition of the EPIYA-A, -B, -C and D motifs. The western-type cagA is mainly cagA-ABC, whereas the East Asian-type is mainly cagA-ABD (17). The EPIYA-C and EPIYA-D motifs act as phosphorylation sites for SHP-2 (18). The EPIYA-D segment exhibits a greater degree of tyrosine phosphorylation and higher binding affinity to SHP2 than the EPIYA-C segment and shows higher virulence (2). East Asian-type cagA is associated with a higher risk of peptic ulcers or gastric cancer within the same geographical area compared with western-type cagA (19). Epidemiological surveys show that ~50-60% of *H. pylori* strains in western countries contain the cagA gene, which increases the risk of peptic ulcers and gastric cancer (20). Although 90-100% of *H. pylori* strains in East Asian countries are cagA-positive, cagA-positive strains may not be associated with clinical outcomes (21).

VacA is a pore-forming toxin, which has several effects on epithelial cells. In addition to inducing vacuolation (22), vacA can induce membrane channel formation, which leads to the release of cytochrome *c* from mitochondria and results in apoptosis (23). Notably, it has immunomodulatory effects through inhibiting T-cell activation and proliferation (6,19). There are four sequence diversity regions of vacA closely associated with *H. pylori* vacuolating activity, namely signal region (s-), deletion region (d-), intermediate region (i-) and middle region (m-) (4). The cytotoxicity of vacA is determined by variability in the structure of the vacA gene (24). The s- and m- regions of vacA are the two main polymorphic regions and serve as markers of *H. pylori* virulence and the risk of associated diseases (25).

Several studies have shown that iceA has two main allelic variants, namely iceA1 and iceA2 (26). The expression of iceA1 is upregulated upon contact between *H. pylori* and human epithelial cells. The iceA1 genotype is linked with enhanced mucosa interleukin-8 expression and acute inflammation (27). Epidemiological data shows that the geographical distribution of iceA genotypes varies. The iceA1 strains mainly occur in Japan and South Korea, whereas iceA2 strains are predominant in the United States and Columbia (28). A previous meta-analysis showed that the prevalence of iceA1 was significantly higher in East Asian countries than in western countries, whereas the prevalence of iceA2 was higher in western countries than in East Asian countries (26). Functionally, the iceA1 genotype is associated with peptic ulcers (29) and the

iceA2 genotype with the occurrence of chronic gastritis (28). To date, the majority of studies have shown that the iceA gene is another virulence gene independent of cagA and vacA (30).

Guizhou, a province located in southwest China, is a multi-ethnic society. In particular, those in Qiannan, Buyei and Miao Autonomous Prefecture have a particular lifestyle and are different from other ethnic groups. The individuals living here often use herbal medicine to treat diseases, including stomach disorders. As Xie *et al* (31) reported, such traditional medicine may have an effect on bacteria in the stomach, including *H. pylori*. Although investigations on the role of *H. pylori* virulence cagA and vacA genotypes have been performed worldwide, the associations between *H. pylori* virulence genotype and gastroduodenal diseases, ethnicity or economic conditions in Guizhou province remain to be fully elucidated. Therefore, the present study aimed to investigate these associations, which may facilitate diagnosis and therapeutic strategies for gastroduodenal diseases.

## Materials and methods

**Clinical samples.** Gastric pylorus mucosa biopsy samples were obtained by endoscopy from patients with gastric disorders at the First Affiliated Hospital of Guizhou Medical University (Guizhou, China) and the People's Hospital of Qiannan Autonomous Prefecture (Duyun, China) between January and December 2016. Clinicopathological data were collected, and written informed consent was obtained from all patients. Patients with the following conditions were excluded from the present study: A tendency to bleed, lactating or pregnant women, the inability to undergo surgery to the UGI tract, and severe cardiovascular or hepatic disease. All protocols were approved by the Ethical Committee of the First Affiliated hospital of Guizhou Medical University. All procedures contributing to the study complied with the Declaration of Helsinki.

***H. pylori* isolation and bacterial DNA extract.** Gastric mucosa samples were cut into small sections, homogenized and smeared on the surface of Brain Heart Infusion agar with 10% sheep blood (Qingdao Hope Biol-Technology Co., Ltd., Qingdao, China) and antibiotic supplement (*H. pylori* selective supplement, Thermo Fisher Scientific Oxoid, Ltd., Basingstoke, UK). The plates were incubated at 37°C for 3-5 days under microaerophilic conditions. *H. pylori* was identified through colonial morphology, Gram staining, a urease test, and *H. pylori*-specific *16S rRNA* gene fragment polymerase chain reaction (PCR) amplification. The colony of *H. pylori* was smooth and translucent, and the morphology was Gram-negative with spiral-shaped bacilli. The confirmed colonies were subcultured to single colonies on fresh medium. Following incubation at 37°C for 3-5 days, the colonies were subjected to DNA extraction using an Ezup column bacteria genomic DNA purification kit (Sangon Biotech Co., Ltd., Shanghai, China), according to the manufacturer's protocol.

**PCR amplification and sequencing.** PCR assays to amplify cagA and sequence its C-terminal region were performed according to the report by Sicinschi *et al* (14). Primers (Sangon Biotech Co., Ltd., Shanghai, China) used for amplification of

Table I. Primer sequences and reaction conditions for polymerase chain reaction.

| Primer                       | Gene              | Primer sequence                  | Size (bp) | Amplification condition                                      |
|------------------------------|-------------------|----------------------------------|-----------|--|
| <i>16S rRNA</i>              | <i>16S rRNA-F</i> | 5'-CTTGCTAGAGTGCTGATTA-3'        | 550       | 35 cycles: 94°C for 30 sec; 55°C for 30 sec; 72°C for 30 sec |
|                              | <i>16S rRNA-R</i> | 5'-TCCCACACTCTAGAAATAGT-3'       |           |  |
| cagA 5'-end conserved region | cagA F            | 5'-GATAACAGGCAAGCTTTTGAGG-3'     | 349       | 30 cycles: 94°C for 1 min; 55°C for 1 min; 72°C for 1 min    |
|                              | cagA R            | 5'-CTGCAAAAGATTGTTTGGCAGA-3'     |           |  |
| cagA 3'-end variable region  | cagA-VF           | 5'-ACCCCTAGTCGGTAATGGGTAA-3'     | 591-856   | 30 cycles: 94°C for 1 min; 50°C for 1 min; 72°C for 1 min    |
|                              | cagA-VR           | 5'-GTAAATTGTCTAGTTTCGC-3'        |           |  |
| cagPAI empty site            | Empty site-F      | 5'-ACATTTTGGCTAAATAAACGCTG-3'    | 535       | 30 cycles: 94°C for 1 min; 55°C for 1 min; 72°C for 1 min    |
|                              | Empty site-R      | 5'-GGTTGCACGCAATTTCCCTTAATC-3'   |           |  |
| iceA1                        | iceA1-F           | 5'-GCTTGTAACGATAAGAAACGCCAGAT-3' | 297       | 35 cycles: 94°C for 30 sec; 55°C for 30 sec; 72°C for 30 sec |
|                              | iceA1-R           | 5'-GGAATGAGCTTGTATTAGAGCCGAT-3'  |           |  |
| iceA2                        | iceA2-F           | 5'-GTTGGGTATATCACAAATTTAT-3'     | 229/334   | 30 cycles: 94°C for 30 sec; 52°C for 30 sec; 72°C for 45 sec |
|                              | iceA2-R           | 5'-TTRCCCTATTTTCTAGTAGGT-3'      |           |  |
| vacA-s1a                     | vacA-s1a-F        | 5'-CTCTCGCTTTAGTAGGAGC-3'        | 213       | 30 cycles: 94°C for 30 sec; 60°C for 30 sec; 72°C for 45 sec |
|                              | vacA-s1a-R        | 5'-CTGCTTGAATGCGCCAAAC-3'        |           |  |
| vacA-s1b                     | vacA-s1b-F        | 5'-AGCGCCATACCGCAAGAG-3'         | 187       |  |
|                              | vacA-s1b-R        | 5'-CTGCTTGAATGCGCCAAAC-3'        |           |  |
| vacA-s1c                     | vacA-s1c-F        | 5'-CTCTCGCTTTAGTGGGGYT-3'        | 213       |  |
|                              | vacA-s1c-R        | 5'-CTGCTTGAATGCGCCAAAC-3'        |           |  |
| vacA-s2                      | vacA-s2-F         | 5'-GCTAACACGCCCAATGATCC-3'       | 199       |  |
|                              | vacA-s2-R         | 5'-CTGCTTGAATGCGCCAAAC-3'        |           |  |
| vacA-m1a                     | vacA-m1a-F        | 5'-GGTCAAAATGCGGTCATGG-3'        | 290       |  |
|                              | vacA-m1a-R        | 5'-CCATTGGTACCTGTAGAAAC-3'       |           |  |
| vacA-m1b                     | vacA-m1b-F        | 5'-GGCCCCAATGCAGTCATGGAT-3'      | 291       |  |
|                              | vacA-m1b-R        | 5'-GCTGTAGTGCCTAAAGAAGCAT-3'     |           |  |
| vacA-m2                      | vacA-m2-F         | 5'-GGAGCCCCCAGGAAACATTG-3'       | 352       |  |
|                              | vacA-m2-R         | 5'-CATAACTAGCGCCTTGCAC-3'        |           |  |

cagA, cytotoxin associated gene A; iceA induced by contact with epithelium gene A; vacA, vacuolating cytotoxin A; F, forward; R, reverse.

Table II. Demographic characteristics of patients.

| Characteristic                | Patients (n) | Age (years) <sup>a</sup> | Range |
|-------------------------------|--------------|--------------------------|-------|
| Sex                           |              |                          |       |
| Male                          | 41           | 38.85±19.43              | 3-80  |
| Female                        | 32           | 39.94±18.93              | 4-66  |
| Ethnicity                     |              |                          |       |
| Han                           | 60           | 40.68±19.64              | 3-80  |
| Ethnic minority               |              |                          |       |
| Miao                          | 4            | 25.75±19.45              | 7-44  |
| Dong                          | 2            | 23±22.63                 | 7-39  |
| Tujia                         | 2            | 36.5±16.26               | 25-48 |
| Buyi                          | 2            | 42.5±0.71                | 42-43 |
| Bai                           | 2            | 36.5±2.12                | 35-38 |
| Yi                            | 1            | 50                       | -     |
| Place of residence            |              |                          |       |
| Guiyang city                  | 60           | 37.97±20.73              | 3-80  |
| Qiannan autonomous prefecture | 13           | 45.62±4.68               | 38-53 |
| Urban                         | 31           | 42.19±21.19              | 3-80  |
| Suburban                      | 42           | 36.41±19.49              | 4-72  |
| Clinical disease              |              |                          |       |
| Gastritis                     | 54           | 38.11±20.34              | 3-80  |
| Peptic ulcer                  | 19           | 42.79±14.90              | 9-60  |

<sup>a</sup>Age is expressed as the mean ± standard deviation.

the *cagA*, *vacA* and *iceA* genes in the present study are shown in Table I. Following DNA extraction from pure culture of *H. Pylori* isolates as previously mentioned, PCR assays were performed in a volume of 26 µl containing 1 µl forward primer, 1 µl reverse primer, 4 µl genomic DNA, 13 µl 2X Taq PCR Master Mix (Beijing Solarbio Science and Technology Co., Ltd., Beijing, China) and 7 µl ddH<sub>2</sub>O. Table I summarizes the expected size of the PCR products and cycling conditions for *cagA* gene, *vacA* subtype gene and *iceA* allelic genes (14,32-34). All runs included one negative (ddH<sub>2</sub>O) and one positive (NCTC 11637 or *H. pylori* 26695) DNA control, and DNA ladder markers (Tiangen Biotech Co., Ltd., Beijing, China). A total of 6 µl of amplified PCR products was then resolved by electrophoresis on 2% agarose gels run in acetate EDTA buffer, and stained with ethidium bromide. The PCR product was visualized under ultraviolet light. The *cagA* C-terminal PCR products were sent to Sangon Biotech Co., Ltd. for Sanger sequencing. The gene sequences were translated into amino acid sequences using Bioedit software (version 7.1.3.0; <https://www.bioedit.com/>). Phylogenetic tree cluster analysis of *cagA* carboxyl terminal variable region was constructed using MEGA software (version 6.0; <https://www.megasoftware.net/megamac.php>), based on neighbor joining. The control strain NCTC 11637 was obtained from the State Key Laboratory of Infectious Disease Prevention and Control (National Institute for Communicable Disease Control and Prevention, Beijing, China), and the amino acid sequences of *H. pylori* 26695 were obtained from GenBank (<https://www.ncbi.nlm.nih.gov/nuccore/CP003904.1>).

**Statistical analysis.** All data were analysed using SPSS 19.0 (IBM SPSS, Armonk, NY, USA),  $\chi^2$  test and Fisher's exact test were used for the analysis of categorical data.  $P < 0.05$  was considered to indicate a statistically significant difference.

## Results

**Clinical and pathological information.** A total of 73 *H. pylori* strains were isolated from patients with upper gastroduodenal disorders. The demographics of these patients are shown in Table II. Among the 73 cases, 41 were men with a mean age of 38.85±19.43 years, and range of 3-80 years, and 32 were women with a mean age of 39.94±18.93 years, and range of 4-66. A total of 60 strains were isolated from Han groups, and 13 from other minority ethnic groups, including Miao, Dong, Tujia, Buyi, Bai and Yi groups. A total of 60 strains were isolated from Guiyang city, and 13 from Qiannan autonomous prefecture. A total of 31 cases were from urban populations, and 42 were from suburban populations. Finally, 54 strains were isolated from patients with gastritis and 19 from patients with peptic ulcers, the latter comprising eight with gastric ulcer and 11 with duodenal ulcer. PCR products of 550 bp represented *H. pylori*-specific 16S rRNA, as shown in Fig. 1A.

**Polymorphism of *cagA*.** The *cagA* N-terminal conserved region (*cagA* 5'-end) and C-terminal variable region (*cagA* 3'-end) were amplified by PCR using the specific primers (Table I). PCR products of 349 bp represented the

Table III. Distribution of *cagA* genotypes in Guizhou province.

| Genotype                        | n (%)      | Guiyang city strains (n) | Qiannan autonomous prefecture strains (n) |
|---------------------------------|------------|--------------------------|---|
| Western-type <i>cagA</i> -AB    | 5 (6.85)   | 5                        | 0   |
| Western-type <i>cagA</i> -ABC   | 3 (4.11)   | 3                        | 0   |
| East Asia-type <i>cagA</i> -ABD | 63 (86.30) | 50                       | 13  |
| East Asia-type <i>cagA</i> -BD  | 2 (2.74)   | 2                        | 0   |
| <i>vacA</i> s1c/m1b             | 18 (24.66) | 13                       | 5   |
| <i>vacA</i> s1c/m2              | 48 (65.75) | 41                       | 7   |
| <i>vacA</i> s1c/m1b/m2          | 7 (9.59)   | 6                        | 1   |
| <i>iceA</i> 1                   | 58 (79.45) | 49                       | 9   |
| <i>iceA</i> 2                   | 2 (2.74)   | 1                        | 1   |
| <i>iceA</i> 1+ <i>iceA</i> 2    | 13 (17.81) | 10                       | 3   |

*cagA*, cytotoxin associated gene A; *vacA*, vacuolating cytotoxin A; *iceA*, induced by contact with epithelium gene A.

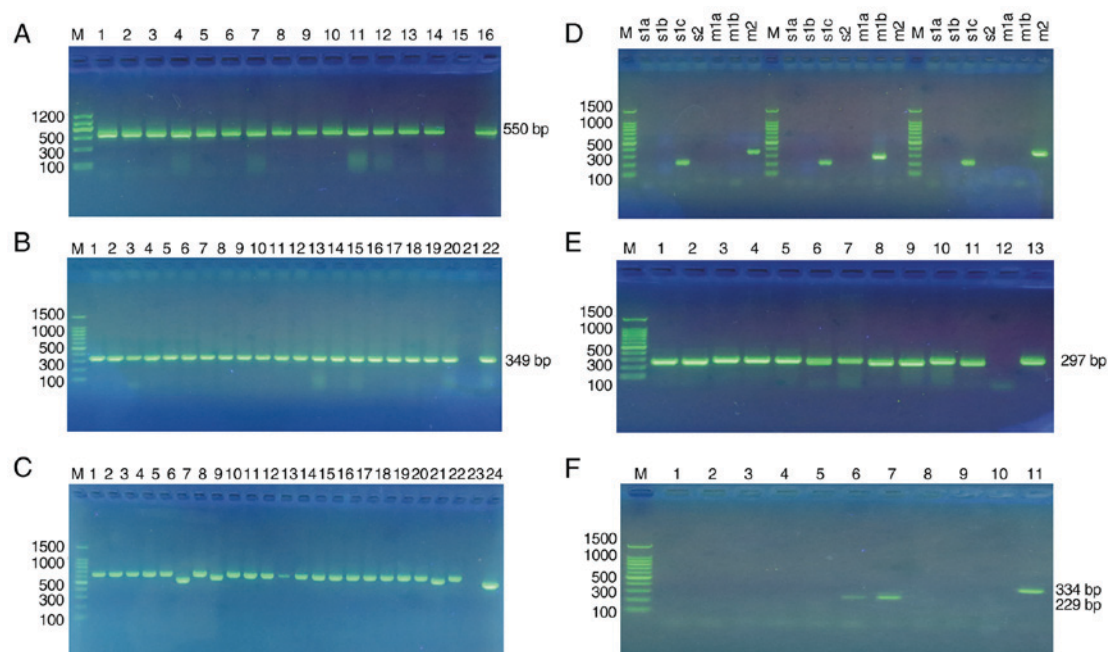


Figure 1. PCR amplification of *H. pylori* *cagA*, *vacA* and *iceA* genes. (A) Gel electrophoresis of genus-specific *16S rRNA* PCR products from *H. pylori* isolates. M, Marker II; 1-14, positive sample for *16S rRNA* PCR; 15, negative control; 16, positive control (NCTC 11637). (B) Gel electrophoresis of clinical samples with different *cagA* N-terminals. M, 100 bp DNA ladder marker; 1-20, clinical sample; 23, negative control; 24, positive control (NCTC 11637). (C) Gel electrophoresis of clinical samples with different *cagA* EPIYA patterns. M, 100 bp DNA ladder marker; 1-22, clinical sample; 23, negative control; 24, positive control (NCTC 11637). (D) Gel electrophoresis of *H. pylori* genotyping of *vacA* alleles. M, 100 bp DNA ladder marker; s1a, s1b, s1c and s2, *vacA* signal region; m1a, m1b and m2, *vacA* middle region. (E) Gel electrophoresis of *H. pylori* genotyping of *iceA*1 alleles. M, 100 bp DNA ladder marker; 1-11, clinical sample; 12, negative control; 13, positive control (*H. pylori* 26695). (F) Gel electrophoresis of *H. pylori* genotyping of *iceA*2 alleles. M, 100 bp DNA ladder marker; 1-5 and 8-9, negative clinical sample; 6-7, positive clinical sample (*iceA*2-229 bp); 10, negative control; 11, positive control (*iceA*2-334 bp). *H. pylori*, *Helicobacter pylori*; *cagA*, cytotoxin associated gene A; *vacA*, vacuolating cytotoxin A; *iceA* induced by contact with epithelium gene A; PCR, polymerase chain reaction.

5'-end fragments, as shown in Fig. 1B, and were present in all *H. pylori* isolates (n=73, 100%). *CagPAI* empty site PCR produced negative results in all of these strains, indicating the absence of *cagA*-negative strains in the mixture. PCR products within the range of 450-650 bp represented the 3'-end fragments, as shown in Fig. 1C. All *H. pylori* isolates were found to contain the 3'-end variable region expressing various EPIYA motifs. Notably, four distinct PCR products (500, 600, 610 and 550 bp amplicons) were obtained from the various *H. pylori* isolates (Fig. 1C).

**Polymorphism of *vacA*.** The *vacA* gene polymorphism was also analyzed. The *vacA* gene has variation regions including signal (s-) and middle (m-) regions (24). Usually, alleles are further divided into sub-alleles, including s1a, s1b, s1c, s2, m1a, m1b and m2 (34). The *H. pylori* isolates were screened for all sub-alleles by the PCR assay, as shown in Fig. 1D. *vacA* s1c/m1b and s1c/m2 genotypes were identified in 24.66 and 65.75%, respectively, whereas the s1c/m1b/m2 mixed genotype was only identified in 9.59%, as shown in Table III. The distribution of *vacA* genotypes in Guizhou province is shown

Table IV. Distribution of virulence genotypes by clinical disease and age of patients.

| Genotype        | Gastritis (n=54) | Peptic ulcer (n=19) | P-value | Age <18 years (n=16) | Age >18 years (n=57) | P-value |
|-----------------|------------------|---------------------|---------|----------------------|----------------------|---------|
| cagA-AB         | 5                | 0                   | 0.417   | 0                    | 4                    | 0.180   |
| cagA-ABC        | 3                | 0                   |         | 2                    | 1                    |         |
| cagA-ABD        | 44               | 19                  |         | 14                   | 50                   |         |
| cagA-BD         | 2                | 0                   |         | 0                    | 2                    |         |
| vacA slc/m1b    | 13               | 5                   | 1.000   | 5                    | 13                   | 0.605   |
| vacA slc/m2     | 36               | 12                  |         | 9                    | 39                   |         |
| vacA slc/m1b/m2 | 5                | 2                   |         | 2                    | 5                    |         |
| iceA1           | 41               | 17                  | 0.146   | 12                   | 46                   | 0.679   |
| iceA2           | 1                | 1                   |         | 0                    | 2                    |         |
| iceA1+iceA2     | 12               | 1                   |         | 4                    | 9                    |         |

cagA, cytotoxin associated gene A; vacA, vacuolating cytotoxin A; iceA, induced by contact with epithelium gene A.

in Table III. No significant association between the vacA genotypes and clinical outcomes was identified ( $P=1.000$ ). There was also no significant association between the vacA genotypes and age, place of residence (Guiyang city vs. Qiannan autonomous prefecture, urban vs. suburban), or ethnic group ( $P=0.605$ ,  $P=0.400$ ,  $P=0.718$  and  $P=0.210$ , respectively), as shown in Tables IV and V.

**Polymorphism of iceA.** The polymorphism of iceA genes was also assessed. Gel electrophoresis genotyping of *H. pylori* iceA1 and iceA2 alleles is shown in Fig. 1E and F, respectively. Among the 73 cases, the iceA1 and iceA2 genotypes were present in 79.45% (58/73) and 2.74% (2/73), respectively. The iceA1/iceA2 mixed genotype occurred in 17.81% (13/73) of cases. The distribution of iceA genotypes in Guizhou province is shown in Table III. No significant association was found between iceA genotypes and disease outcomes, age, place of residence (Guiyang city, vs. Qiannan autonomous prefecture, urban vs. suburban), or ethnic group ( $P=0.146$ ,  $P=0.679$ ,  $P=0.304$ ,  $P=0.884$  and  $P=0.621$ , respectively), as shown in Tables IV and V.

**EPIYA motifs of cagA.** As described above, the EPIYA motif patterns were determined for each strain. A comparison of the results indicated that the EPIYA genotypes of the 500, 600, 610 and 550 bp amplicons were these of the cagA-AB, -ABC, -ABD and -BD genotypes, respectively. The structural polymorphism of the cagA amino acid sequence is shown in Fig. 2. It was noted that cagA EPIYA-ABD (63/73, 86.30%) was the predominant genotype in the present study, followed by cagA-AB (5/73, 6.85%), cagA-ABC (3/73, 4.11%) and cagA-BD (2/73, 2.74%). The distribution of cagA genotypes in Guizhou province is shown in Table III. However, statistical analysis revealed no significant correlation between the disease outcome and the cagA genotypes ( $P=0.417$ ). There was also no significant correlation between the cagA genotypes and age, place of residence (Guiyang city, vs. Qiannan autonomous prefecture, urban vs. suburban), or ethnic group ( $P=0.180$ ,  $P=0.845$ ,  $P=0.602$  and  $P=1.000$ , respectively), as shown in Tables IV and V.

The results of phylogenetic tree cluster analysis of the cagA carboxyl terminal variable region is shown in Fig. 3.

There was a clustering association between western-type cagA-AB and cagA-ABC; there was also a clustering association between East Asian-type cagA-ABD and cagA-BD. All five strains with western-type cagA-AB and three strains with western-type cagA-ABC were isolated from patients with chronic gastritis; 19 strains with East Asian-type cagA-ABD were isolated from patients with peptic ulcers, and a further 46 with East Asia-type cagA were isolated from patients with chronic gastritis. As mentioned above, it appeared that East Asia-type cagA strains may exhibit higher virulence than western-type cagA strains.

The frequencies of the four EPIYA or EPIYA-like motifs are shown in Table VI. In total, 212 EPIYA motifs were obtained from 73 cagA sequences. The most frequent EPIYA motif was EPIYA (197/212, 92.92%), followed by EPIYT (8/212, 3.77%), ESIYA (6/212, 2.83%) and ESIYT (1/212, 0.47%). The EPIYA-B motif had a high degree of variation in the five amino acids (EPIYA, EPIYT, ESIYA or ESIYT). In addition, 15 EPIYA-like motifs (EPIYT, ESIYA, ESIYT) were isolated from patients with gastritis.

## Discussion

*H. pylori* is well known for its genetic diversity and geographical differences, which may be associated with compound clinical disease outcomes (7). In the present study, the epidemiological characteristics of major virulence genes of *H. pylori* were investigated in Guizhou province. *H. pylori* was cultured from clinical samples, and virulence genotypes of cagA, vacA and iceA were examined by PCR assays. The results showed the prevalent strains of *H. pylori* in Guizhou province were cagA-positive, vacA slc/m2-positive and iceA-positive. The results describe the prevalence of *H. pylori* strains in Guizhou province between January 2015 and December 2016, and provides an experimental basis for molecular epidemiological investigations of *H. pylori* in this region.

Epidemiological investigations have shown that the cagA genotype varies markedly worldwide. The prevalence of cagA among *H. pylori* in different regions varies between 50 and 60% in certain western countries to almost 100% in East Asia (19). In the present study, it was shown that cagA genotypes were

Table V. Distribution of *cagA* genotypes by place of residence and ethnicity of patients.

| Genotype        | Place of residence |                               |         |       |          |         | Ethnic group |          |         |
|-----------------|--------------------|-------------------------------|---------|-------|----------|---------|--------------|----------|---------|
|                 | Guiyang city       | Qiannan autonomous prefecture | P-value | Urban | Suburban | P-value | Han          | Minority | P-value |
| CagA-AB         | 5                  | 0                             | 0.845   | 3     | 2        | 0.602   | 4            | 1        | 1.000   |
| CagA-ABC        | 3                  | 0                             |         | 2     | 1        |         | 3            | 0        |         |
| cagA-ABD        | 50                 | 13                            |         | 25    | 38       |         | 51           | 12       |         |
| CagA-BD         | 2                  | 0                             | 0.400   | 1     | 1        | 0.718   | 2            | 0        | 0.210   |
| vacA slc/m1b    | 13                 | 5                             |         | 7     | 11       |         | 13           | 5        |         |
| vacA slc/m2     | 41                 | 7                             |         | 22    | 26       |         | 42           | 6        |         |
| vacA slc/m1b/m2 | 6                  | 1                             |         | 2     | 5        |         | 5            | 2        |         |
| iceA1           | 49                 | 9                             | 0.304   | 24    | 34       | 0.884   | 46           | 12       | 0.621   |
| iceA2           | 1                  | 1                             |         | 1     | 1        |         | 2            | 0        |         |
| iceA1/iceA2     | 10                 | 3                             |         | 6     | 7        |         | 12           | 1        |         |

*cagA*, cytotoxin associated gene A; *vacA*, vacuolating cytotoxin A; *iceA*, induced by contact with epithelium gene A.

Table VI. Frequencies of EPIYA motifs.

| Motif type           | EPIYA | EPIYT | ESIYA | ESIYT | Total |
|----------------------|-------|-------|-------|-------|-------|
| All <i>cagA</i> type |       |       |       |       |       |
| All motifs           | 197   | 8     | 6     | 1     | 212   |
| A motifs             | 71    | 0     | 0     | 0     |       |
| B motifs             | 59    | 7     | 6     | 1     |       |
| C motifs             | 2     | 1     | 0     | 0     |       |
| D motifs             | 65    | 0     | 0     | 0     |       |
| Western-type-ABC     |       |       |       |       |       |
| All motifs           | 5     | 4     | 0     | 0     | 9     |
| A motifs             | 3     | 0     | 0     | 0     |       |
| B motifs             | 0     | 3     | 0     | 0     |       |
| C motifs             | 2     | 1     | 0     | 0     |       |
| Eastern-type-ABD     |       |       |       |       |       |
| All motifs           | 181   | 2     | 5     | 1     | 189   |
| A motifs             | 63    | 0     | 0     | 0     |       |
| B motifs             | 55    | 2     | 5     | 1     |       |
| D motifs             | 63    | 0     | 0     | 0     |       |
| Western-type-AB      |       |       |       |       |       |
| All motifs           | 7     | 2     | 1     |       | 10    |
| A motifs             | 5     | 0     | 0     |       |       |
| B motifs             | 2     | 2     | 1     |       |       |
| Eastern-type-BD      |       |       |       |       |       |
| All motifs           | 4     | 0     | 0     |       | 4     |
| B motifs             | 2     | 0     | 0     |       |       |
| D motifs             | 2     | 0     | 0     |       |       |

*cagA*, cytotoxin associated gene A.

detected in all strains in Guizhou province. In another study in this region, Zhou *et al* reported that 30 *H. pylori* strains isolated from gastric cancer were *cagA*-positive, with *cagA*

having a positive rate of 100% (35), which was consistent with the results of the present study. The *vacA* genotypes also vary geographically. The *vacA* *slc* genotype is common in South Asia, the *slb* genotype is common in Latin America and Africa, and *slc* is common in East Asia (24,36,37). The *vacA* *m1* genotype is common in North Asian countries, including Japan and South Korea, whereas the *m2* genotype is predominant in Southeast Asia, including Taiwan, China and Vietnam (20). The results of the present study showed that the predominant *vacA* genotype was *slc/m2* in Guizhou province. Notably, the *iceA* genotype also varies among different regions. A previous meta-analysis showed that the prevalence of *iceA1* was significantly higher in East Asian countries than in western countries, whereas the prevalence of *iceA2* was higher in western countries than in East Asian countries (26). Consistent with this finding, the *iceA1* genotype was predominant in Guizhou province in the present study.

Although Guizhou province is a multi-ethnic society, there was no ethnic specificity of *H. pylori* infection. This differs from Malaysia, which has three major ethnic groups, namely, Malay, Chinese and Indian; *vacA* *slc/m2* has been detected in all of these ethnic groups, whereas *vacA* *slc/m2* was only isolated from Chinese patients (38). **Therefore, *vacA* genotypes were associated with ethnic groups in Malaysia.** In the present study, as no predictive value of these *H. pylori* virulence genes was identified in Guizhou province, there may be no need to classify patients according to ethnicity during treatment, with patients of different ethnicities offered the same diagnosis and treatment strategies.

*H. pylori* infection can result in compound gastroduodenal diseases, including gastritis, peptic ulcer and gastric carcinoma. The present study did not find an association between *H. pylori* virulence diversity and clinical disease outcomes in the Guizhou region. Zhou *et al* reported that *H. pylori* infection may induce the demethylation of lactate dehydrogenase, dihydrolipoamide dehydrogenase and calmodulin genes, and increase methylation of the Ran-specific GTPase-activating protein gene, which leads to dysfunctional gene expression in gastric cancer tissues and cells (35). A number of reports have shown that infection with

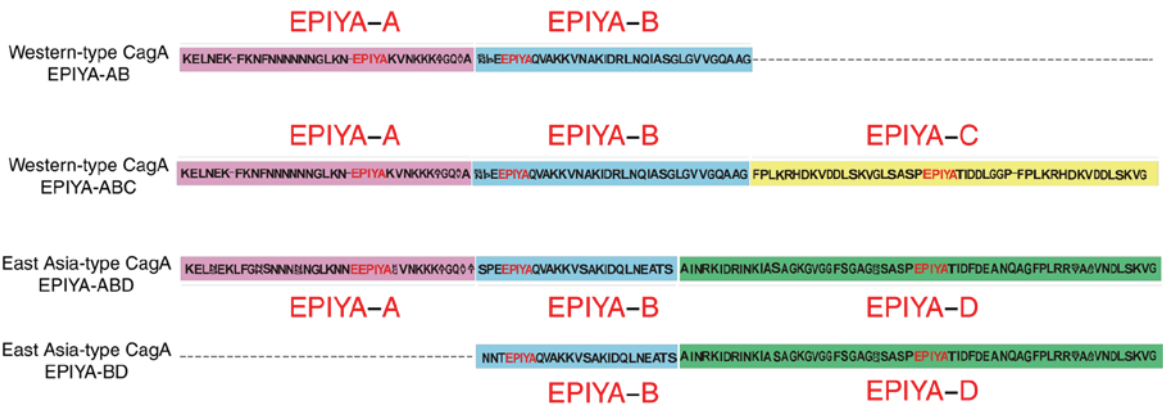


Figure 2. Structural polymorphism in the cagA amino acid sequence of East Asian-type and western-type cagA. cagA, cytotoxin associated gene A.

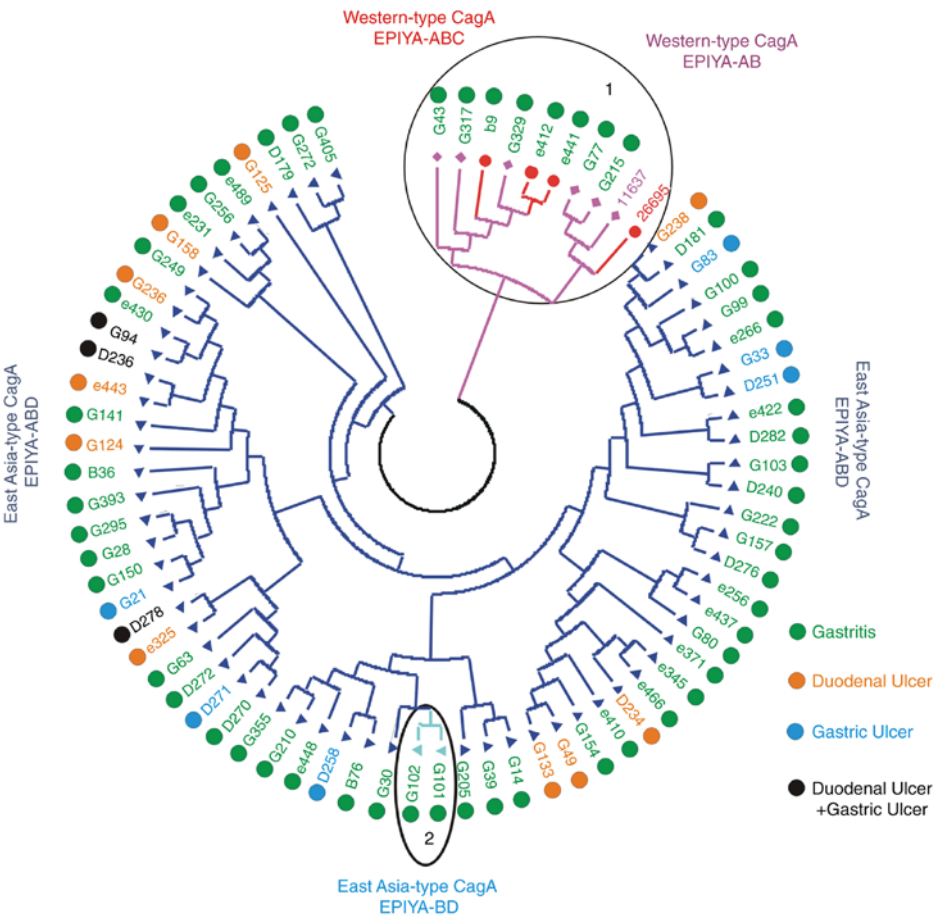


Figure 3. Phylogenetic tree cluster analysis of the cagA carboxyl terminal variable region. On the inner side of the phylogenetic tree, red branches are western-type cagA-ABC (indicated in circle 1); purple branches are western-type cagA-AB (circle 1); blue branches are East Asia-type cagA-ABD; cyan branches are East Asia-type cagA-BD (circle 2). On the outer side of the phylogenetic tree, green circles are gastritis; orange circles are for duodenal ulcer; blue circles are for gastric ulcer; black circles are for compound ulcer, including duodenal ulcer and gastric ulcer. cagA, cytotoxin associated gene A.

a higher number of cagA EPIYA-C motif strains was associated with increased gastric inflammation and atrophy in western countries (39-41). However, the genetic diversity of *H. pylori* virulence genes is not associated with disease progression in certain eastern countries (9,16). Matsunari *et al* reported that, in the Bhutan region, 209 isolate strains, in which >50% of the strains had multiple EPIYA motifs repeats in East Asian-type cagA-ABD, are associated with atrophic gastritis and gastric cancer (42). An association between vacA genotypes and gastric

precancerous or gastric carcinoma was observed in Brazil (39). However, no correlation has been found between vacA genotypes and diseases in Japan or Sweden (9,41). Additionally, an association has been reported between the iceA1 genotype and peptic ulcers, and the iceA2 genotype and gastritis (26). Among the 73 isolates in the present study, no association was found between the iceA1 or iceA2 genotypes and clinical disease outcomes.

Among the 73 patients, none were diagnosed with gastric cancer; only 14 gastric biopsy samples were obtained from

patients with gastric cancer at the First Affiliated Hospital of Guizhou Medical University and the People's Hospital of Qiannan Autonomous Prefecture between January and December 2016, however, *H. pylori* isolation was negative. This may be associated with reduced *Helicobacter* abundance and overrepresentation of bacterial genera.

Current knowledge of bacteria other than *H. pylori* in the human stomach remains limited. The investigation of the gastric microbiota in healthy individuals or disease states is lacking due to limitations of culture conditions and the collection of gastric mucosal specimens. With the development of molecular biology and bacterial *16S rRNA* gene identification technology, the constitution and diversity of gastric flora has been gradually identified. Notably, You *et al* reported a microarray used to compare the genomic profiles of strains isolated from patients with gastroduodenal diseases in the Heilongjiang province of China, and their findings may provide insight into novel biomarkers for the prediction of gastric diseases (43). Novel genetic variations may be examined in our future investigations using similar approaches. The gastric flora may be important roles in human health and disease, which remain to be elucidated. The composition of the gastric flora is dynamic and affected by several factors, including *H. pylori* infection and combination therapy with antibiotics and proton pump inhibitors. The gastric flora may also affect the development of gastric diseases following *H. pylori* colonization. *H. pylori* infection in Guizhou province may lead to various diseases without specificity. This may be explained by gastric flora in the stomach of local residents. Finally, the clarification of genotypes of gastric flora from different areas may facilitate clinical therapeutics.

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#### Availability of data and materials

All data generated or analyzed during this study are included in this published article.

#### Authors' contributions

ZC conceived, designed and supervised the experiments. LY, FL, CG and QW performed the experiments. KP, LX, YX and YC collected the samples. ZC and LY performed the data analysis and wrote the manuscript. All authors read and approved the final manuscript.

#### Ethics approval and consent to participate

All protocols were approved by the Ethical Committee of the First Affiliated hospital of Guizhou Medical University.

All procedures contributing to the study complied with the Declaration of Helsinki. Written informed consent was obtained from all patients.

#### Patient consent for publication

Not applicable.

#### Competing interests

The authors declare that they have no competing interests.

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