

3,3'-Diindolylmethane mitigates lipopolysaccharide-induced acute kidney injury in mice by inhibiting NOX-mediated oxidative stress and the apoptosis of renal tubular epithelial cells

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Abstract. 3,3'-Diindolylmethane (DIM) is a naturally derived indole compound found in the Brassica family of vegetables. DIM has several beneficial effects, including anti-cancer, anti-inflammatory and anti-angiogenic functions. However, the effects of DIM on acute kidney injury (AKI) stimulated by lipopolysaccharide (LPS) are poorly studied. In this present study, male BALB/c mouse models of AKI were established using intraperitoneal injections of 10 mg/kg LPS. DIM (40 mg/kg) was administered intraperitoneally 24 and 2 h before LPS exposure. The results indicated that DIM significantly mitigated histopathological changes in the kidneys and improved the levels of blood urea nitrogen and serum creatinine. DIM also suppressed the LPS-induced production of reactive oxygen species and cell apoptosis. Furthermore, DIM treatment significantly decreased the expression of NADPH oxidase 2 (NOX2) and NOX4 in LPS-treated mice. Therefore, DIM may exert its renoprotective actions by inhibiting NOX-mediated oxidative stress and the apoptosis of renal tubular epithelial cells.

Introduction

Acute kidney injury (AKI), characterized by a sharp decline in renal function, is a severe complication with high morbidity and mortality rates and is commonly encountered in the intensive care unit (ICU) (1,2). The Finnish Acute Kidney Injury study showed that AKI affects 40% of critically ill patients (3). AKI can be caused by various events, such as sepsis, cardiac surgery, liver or kidney transplantation,

rheumatic fever, urinary tract obstruction, pharmacological toxins (4-6) and acute severe pancreatitis (7). Among these, endotoxic shock caused by lipopolysaccharide (LPS), which is the outer membrane component of gram-negative bacteria, is a common cause of AKI (8-10). Sepsis-associated AKI carries a particularly high mortality rate. One multicenter, international study involving an observational cohort of ICU patients demonstrated that the mortality rate of sepsis-related AKI patients was as high as 50% (2).

The mechanisms of sepsis-associated AKI are complex and not well understood. Although the mechanisms of sepsis-associated AKI are poorly understood, release of inflammatory factors, oxidative stress, apoptosis and micro-circulatory dysfunction are believed to play an important role (11-13). So far, there are no effective drugs for the treatment of AKI. Therefore, novel and effective therapies to reduce the mortality of AKI are urgently needed.

3,3'-Diindolylmethane (DIM), a natural compound derived from the acid-catalyzed self-condensation of indole-3-carbinol, is abundant in cruciferous vegetables including kale and broccoli (14,15). Studies have found that DIM can inhibit LPS-induced acute liver injury by regulating the expression of miRNAs (14,16). Other studies have shown that DIM has a protective effect on the LPS-induced damage of cardiomyocytes and brain inflammation by reducing the release of pro-inflammatory mediators and negative regulation of the NF- κ B signaling pathway (17,18). In addition, DIM may also exert its organ protective function by mitigating oxidative stress and apoptosis (19).

However, the potentially protective characteristics of DIM have not yet been tested in LPS-triggered AKI. To address this, an experiment was designed to evaluate how DIM modifies disease progression in LPS-induced AKI.

Materials and methods

Animal protocols. Research protocols were reviewed and approved by the Experimental Animal Ethics Committee of Chongqing Medical University, while all the handling and care of animals were performed in strict compliance with the U.S. National Institute of Health Guide for the Care and Use of Laboratory Animals (1996 revision). Mice (male, aged 8 weeks

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old, body weight 22–25 g) were bred in a specific pathogen-free laboratory and allowed free access to food and tap water. The animal room was kept on a 12 h light/dark cycle at a constant temperature (25°C) and relative humidity of 55±5% throughout the experimental period. Mice were divided into 4 groups of 10 mice (40 mice in total). The four groups comprised the control, LPS (10 mg/kg; Sigma Aldrich; Merck KGaA), DIM (40 mg/kg; MedChemExpress) and LPS + DIM (10 mg/kg LPS + 40 mg/kg DIM) groups. The choice of DIM concentration was based on a previous study (19) and incorporated similar DIM doses as administered in previous experiments (14,19). An AKI murine model was produced by intraperitoneally injecting LPS (10 mg/kg in 200 μ l saline solution) and allowing 24 h for renal damage to develop. Mice were administered two intraperitoneal injections of DIM (40 mg/kg). The first and second doses were administered 24 and 2 h prior to the LPS inoculation, respectively. The animals were sacrificed after 24 h exposure to LPS and blood and kidney samples were harvested.

Histological examination. Tissues from the right kidney were first fixed in 4% paraformaldehyde (4°C, 24 h) and embedded in paraffin. Sections of 4 μ m thickness were cut and processed and then stained with hematoxylin (0.2%) and eosin (1%) (H&E) at room temperature for 30 sec and 1 min respectively. Histological alterations in renal tissues and the degree of kidney injury were scored on a scale of 0–4, as previously described (20), upon visualization by light microscopy (magnification, x400; Olympus Corporation).

Evaluation of renal function. Blood samples were extracted via the retro-orbital venous plexus and processed to extract the serum. In this present study, serum creatinine (SCr) and blood urea nitrogen (BUN) levels were employed as markers of renal function and were analyzed with an AutoAnalyzer (Roche Diagnostics GmbH). SCr and BUN were quantified in accordance with the manufacturer's instructions.

TUNEL staining. Tissues of the right kidney were fixed in 4% paraformaldehyde, paraffin-embedded, and resected into 4 μ m-thick sections according to the aforementioned procedure. TUNEL staining was conducted with a commercially available kit (Roche Diagnostics GmbH) according to the manufacturer's instructions. Briefly, the dehydrated sections were treated with DNase-free proteinase K (37°C, 30 mins), followed by 3% H₂O₂ to quench endogenous peroxidase activity. Free 3'-OH termini were labeled with digoxigenin-dUTP for 1 h at 37°C using the TUNEL reaction mixture. Then, the sections were incubated with converter-POD at 37°C for 30 min. DAB chromogenic reagent was employed to develop the stain and hematoxylin was used to stain the nucleus. Finally, the samples were cleared in xylene, mounted with neutral balsam and coverslipped. Under optical microscopy (magnification, x400; Olympus Corporation), the number of TUNEL-positive cells in 400 histological fields were counted per kidney section.

Fluorescence microscopy analysis for kidney reactive oxygen species (ROS). ROS generated in the kidney were detected using dihydroethidium (DHE; Beyotime Institute of Biotechnology). The left kidney tissues were stored in liquid nitrogen before preparation into frozen sections. The kidney sections were washed

three times with ice-cold PBS, treated with DHE (10⁻⁵ M, final concentration) and incubated for 30 min at 37°C. After treatment, sections were again washed three times with ice-cold PBS. The excitation (480 nm) and emission (590 nm) wavelengths for these experiments were set by fluorescent microscopy. All mean fluorescence values were analyzed by comparing each group using the ZEN 2012 software (Carl Zeiss AG, Germany).

Measurement of kidney lipid peroxidation and glutathione (GSH) content. The malondialdehyde (MDA) level in kidney homogenates was analyzed by measuring the level of trimethine, a reaction product of MDA and thiobarbituric acid, spectrophotometrically at a wavelength of 535 nm. The reduced GSH level in the kidney homogenates was determined using 5,5'-dithiobis-2-nitrobenzoic acid (DTNB) at a wavelength of 412 nm. Thereafter, oxidized GSH (GSSG) is reduced to GSH by GSH reductase, total GSH (T-GSH) is measured by the same method. The level of GSSG can be calculated by subtracting GSH from T-GSH.

Western blot analysis. A protein extraction kit (RIPA Lysis Buffer, Beyotime Institute of Biotechnology) was used to isolate proteins from the left renal tissue samples. The bicinchoninic acid method was employed to quantify protein concentrations. Each protein sample of 20 μ g was separated via 10% SDS-PAGE gel electrophoresis and transferred to a PVDF membrane (EMD Millipore) and successively incubated with 5% non-fat milk at room temperature for 1.5 h. PVDF membranes were allowed to incubate overnight at 4°C with the indicated primary antibodies, including caspase-3 (1:500; cat. no. wl01927; Wanleibio Co., Ltd.), cleaved caspase-3 (1:500; cat. no. wl01992; Wanleibio Co., Ltd.), poly (ADP-ribose) polymerase (PARP; 1:500; cat. no. wl0326; Wanleibio Co., Ltd.), cleaved PARP (1:500; cat. no. wl01932; Wanleibio Co., Ltd.), NADPH oxidase-1, -2, -3 and -4 (NOX1, NOX2, NOX3 and NOX4, respectively; 1:500; sc-518023, sc-17844, sc-7662 and sc-20781, respectively; Santa Cruz Biotechnology, Inc., USA), and β -actin (1:1,000; cat. no. A0208; Beyotime Institute of Biotechnology). The following day, HRP labeled goat anti-rabbit IgG (H+L) (1:3,000; Beyotime Institute of Biotechnology) was added and further incubated at room temperature for 1 h. Immunoreactive bands were visualized using an ECL (cat. no. WLA003a, Wanleibio Co., Ltd.) system and quantified with using Image J software (version 1.8.0).

Statistical analysis. All experiments were conducted at least three times. All data are presented as the mean \pm standard error of the mean. All statistical analyses were performed using SPSS 17.0 statistical software (SPSS, Inc.). Statistical significance of multiple groups was determined by one-way ANOVA, followed by the least significant difference for multiple comparisons test. $P < 0.05$ was considered to indicate a statistically significant difference.

Results

DIM mitigates histopathological changes and restores renal function in LPS-induced AKI mouse models. To determine how DIM affected the progression of LPS-mediated AKI, the kidneys were examined for histopathological changes.

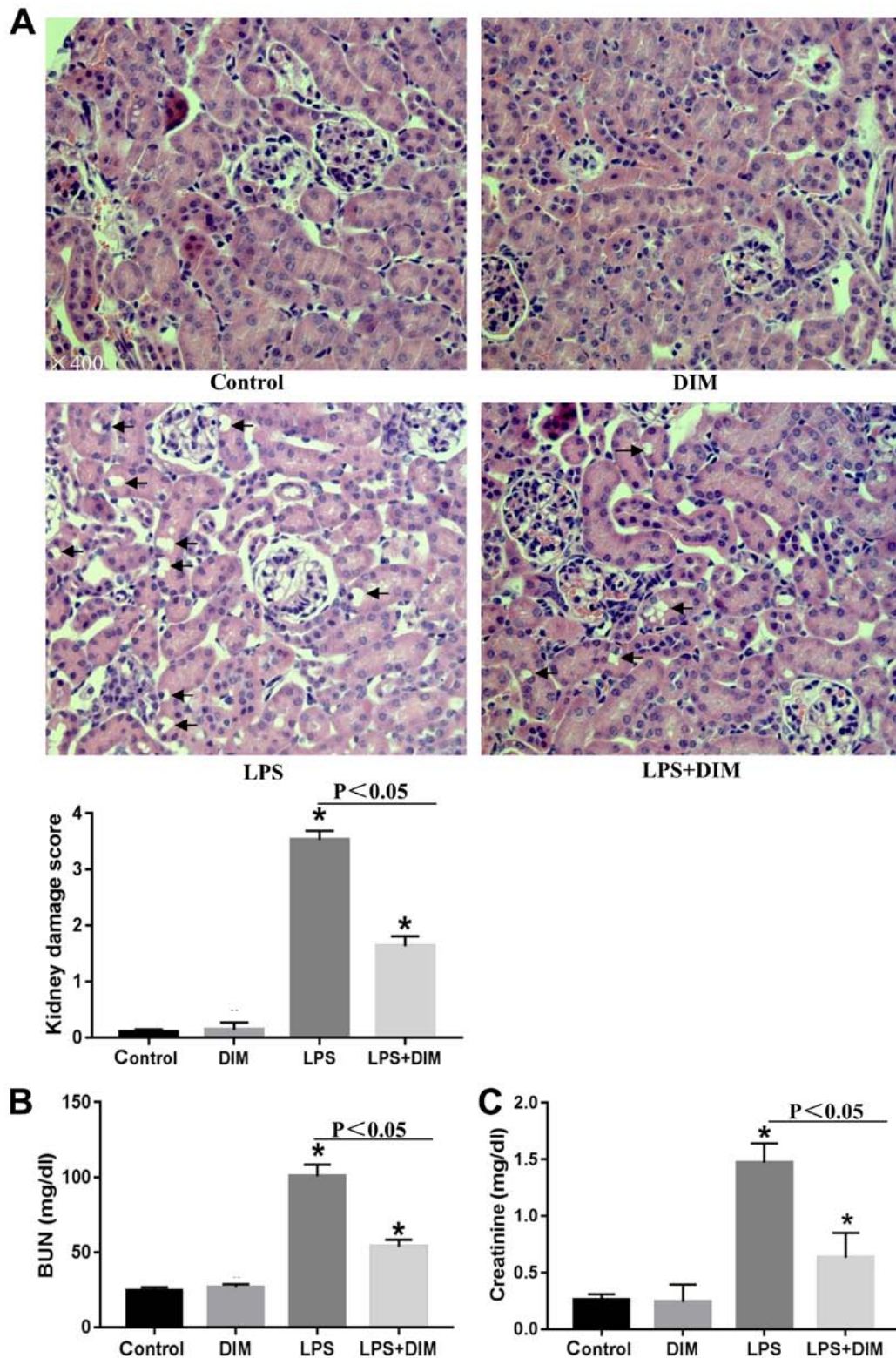


Figure 1. DIM mitigates histopathological changes and restores renal function in an LPS-induced acute kidney injury mouse model. (A) Mice were exposed to LPS (10 mg/kg), DIM (40 mg/kg), or LPS + DIM (10 mg + 40 mg) for 24 h; histological changes in kidneys were observed by hematoxylin and eosin staining. (B) BUN levels were determined after the mice were exposed to LPS, DIM or LPS + DIM for 24 h. (C) Blood creatinine levels were determined after the mice were exposed to LPS, DIM or LPS + DIM for 24 h. The arrows indicate damaged tubular epithelial cells. The results are presented as the mean \pm SEM. * $P < 0.05$ vs. control. DIM, 3,3'-diindolylmethane; LPS, lipopolysaccharide; BUN, blood urea nitrogen.

Mice that were exposed to LPS exhibited several indicators of AKI, including destroyed renal tubules, aberrant renal tubular epithelial cells, interstitial edema in renal epithelial cells and renal tubule dilation (Fig. 1A). H&E staining showed that the

epithelial cell structure and the glomerular membrane were normal in the control and DIM groups. AKI-related kidney lesions appeared to be attenuated in AKI mouse models treated with DIM.

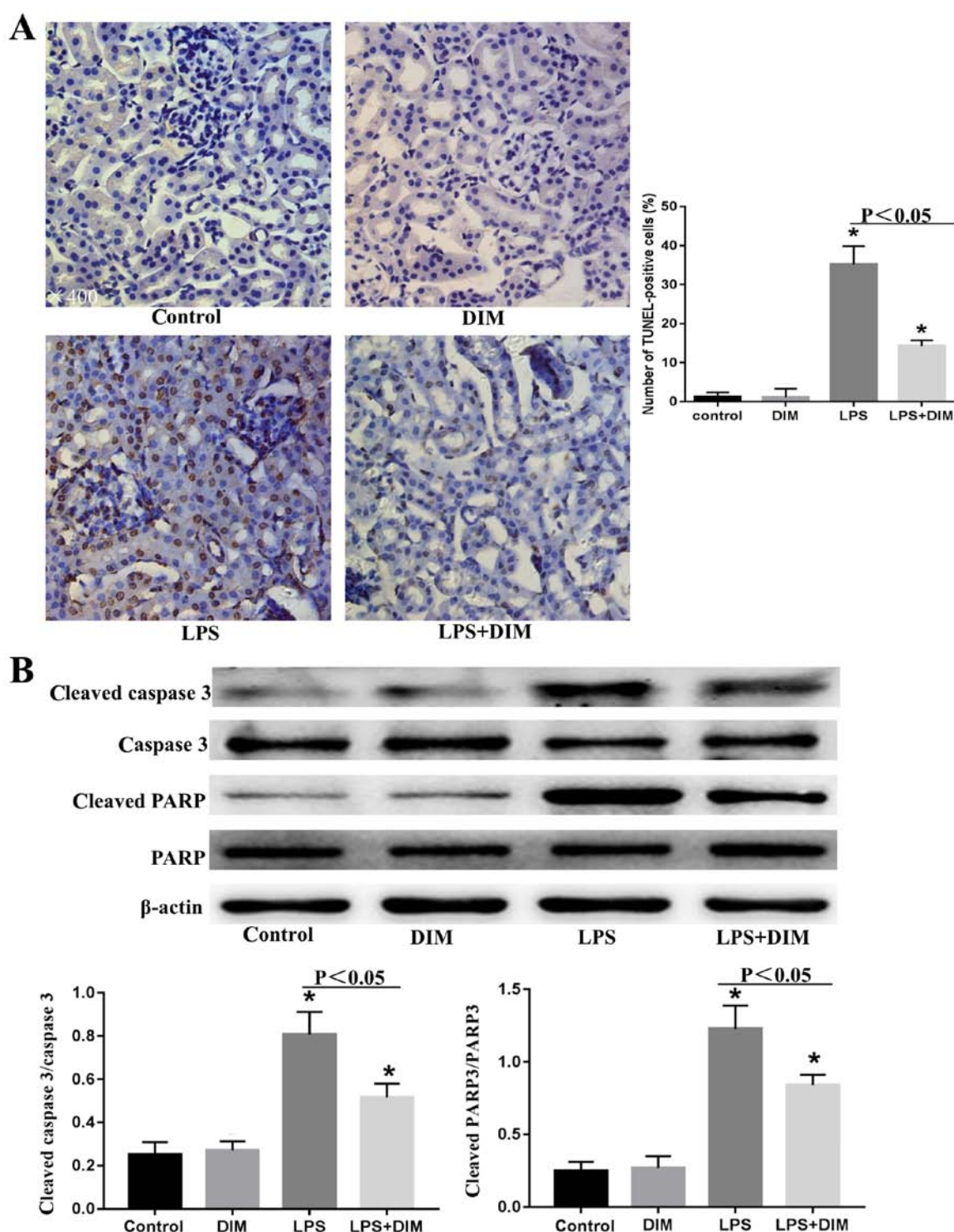


Figure 2. DIM treatment inhibits the apoptosis of renal tubular epithelial cells in mice with LPS-triggered acute kidney injury. Mice were exposed to LPS (10 mg/kg), DIM (40 mg/kg) or LPS + DIM (10 mg + 40 mg) for 24 h after which (A) the apoptosis of renal tubular epithelial cells was determined by TUNEL staining and (B) the expression levels of cleaved caspase-3 and cleaved PARP were determined by western blot analysis. * $P < 0.05$ vs. control. DIM, 3,3'-diindolylmethane; LPS, lipopolysaccharide; PARP, poly (ADP-ribose) polymerase.

To further confirm the results from the H&E staining, changes in SCr and BUN levels were also examined. As shown in Fig. 1B and C, DIM significantly improved the renal function of AKI mice.

DIM treatment inhibits the apoptosis of renal tubular epithelial cells in mice with LPS-triggered AKI. As the apoptosis of renal

tubular epithelial cells is an important mechanism leading to AKI, the effects of DIM on LPS-induced tubular epithelial cell apoptosis were examined.

As shown in Fig. 2A, numerous TUNEL-positive renal tubular epithelial cells were observed in the kidney tissues of the LPS-induced AKI mouse models. However, fewer TUNEL-positive tubular epithelial cells were observed in

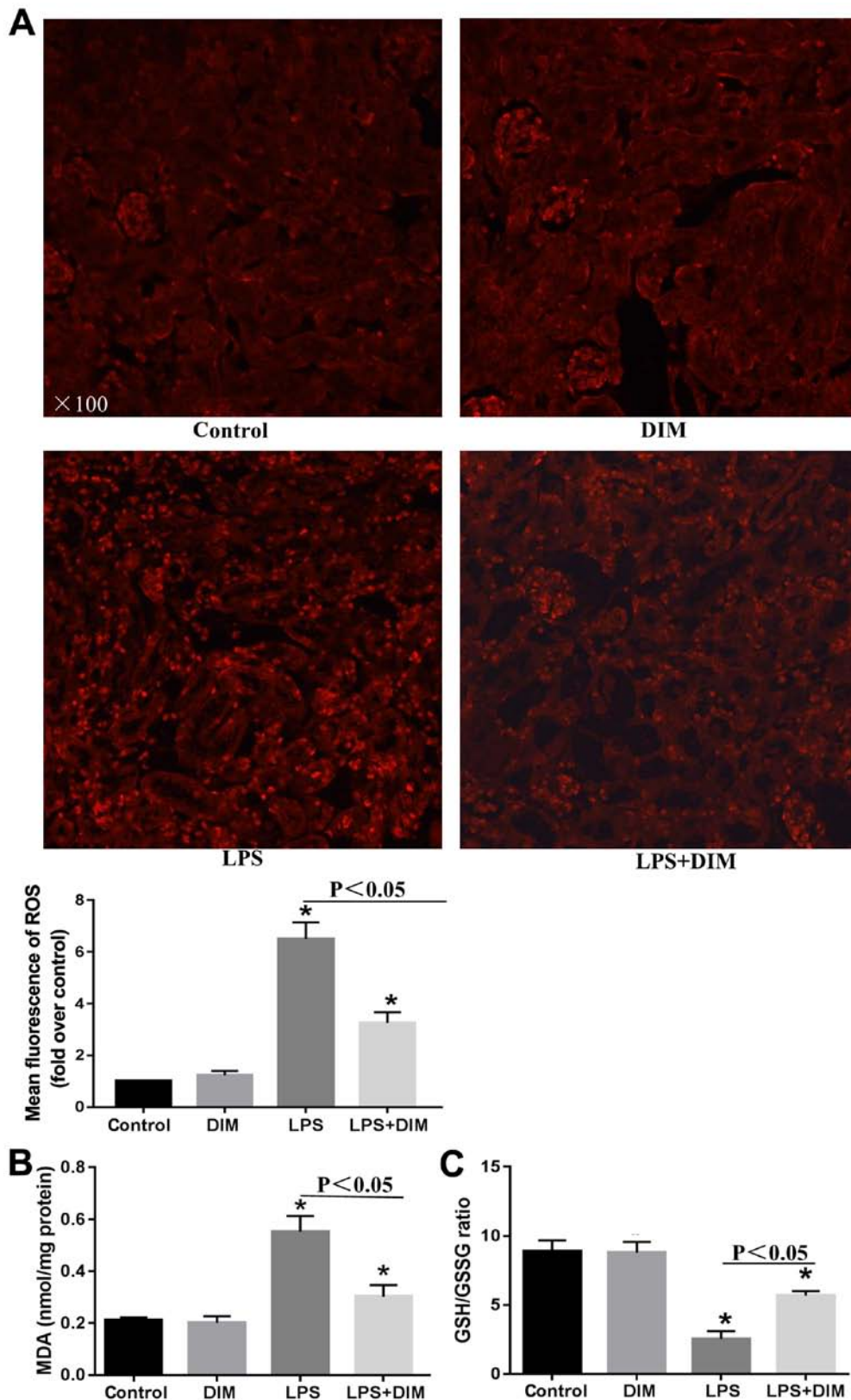


Figure 3. DIM treatment alleviates oxidative stress in mice with LPS-triggered acute kidney injury. Mice were exposed to LPS (10 mg/kg), DIM (40 mg/kg) or LPS + DIM (10 mg + 40 mg) for 24 h after which (A) ROS production was observed by dihydroethidium, (B) MDA levels in the kidney tissues were determined and (C) the GSH/GSSG ratio in the kidney tissues was calculated. * $P < 0.05$ vs. control. DIM, 3,3'-diindolylmethane; LPS, lipopolysaccharide; ROS, reactive oxygen species; MDA, malondialdehyde; GSH/GSSG, glutathione/oxidized glutathione.

the kidney tissues of the DIM-treated AKI mice. Consistent with this, western blot analysis revealed that the expression levels of apoptotic markers, including cleaved-caspase-3

and cleaved-PARP, were significantly increased in the LPS-induced AKI mouse model and were attenuated by DIM pre-treatment (Fig. 2B).

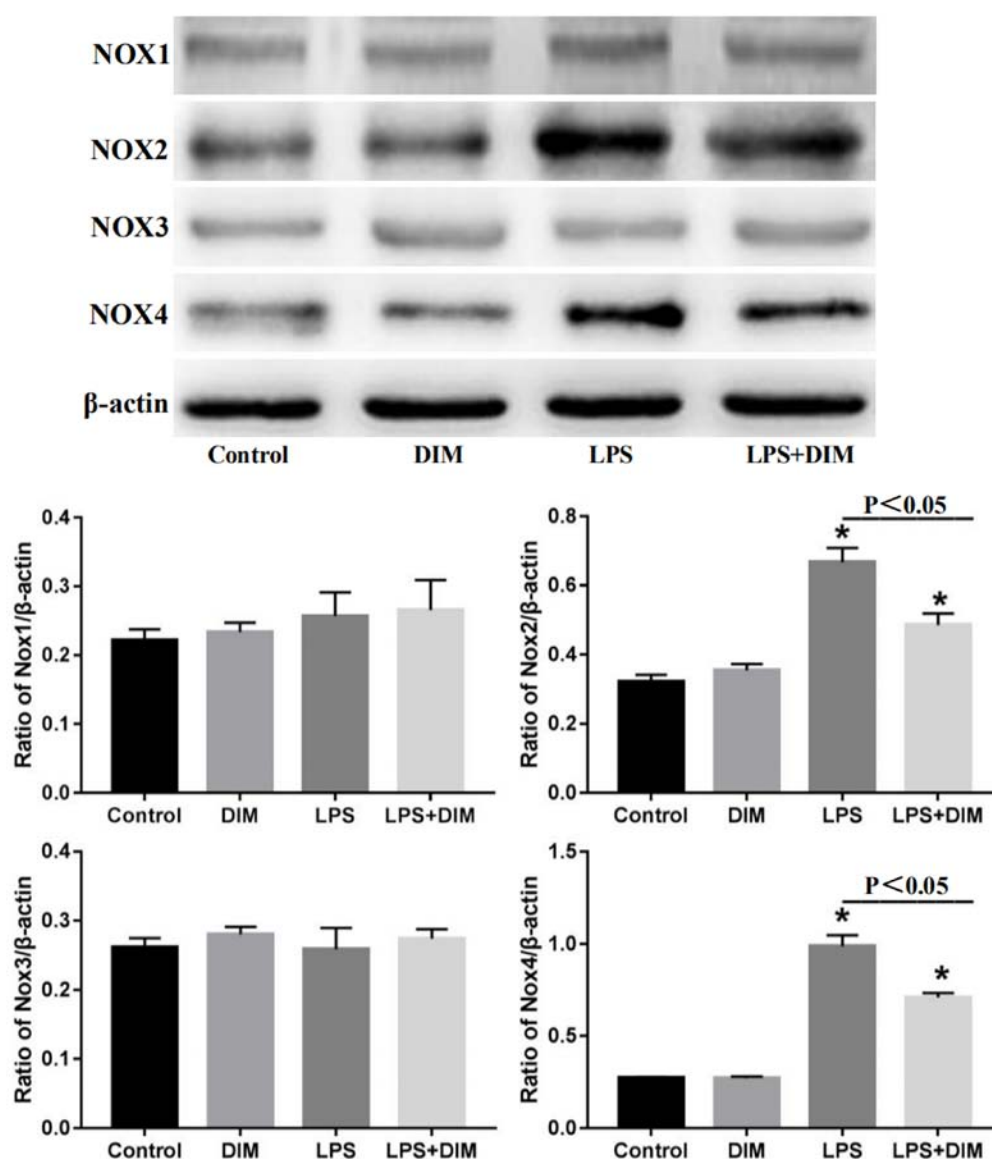


Figure 4. DIM treatment suppresses the expression of NOX2 and NOX4 in mice with LPS-triggered acute kidney injury. Mice were exposed to LPS (10 mg/kg), DIM (40 mg/kg) or LPS + DIM (10 mg + 40 mg) for 24 h after which the expression levels of NOX1, NOX2, NOX3 and NOX4 were determined by western blot analysis. * $P < 0.05$ vs. control. DIM, 3,3'-diindolylmethane; NOX, NADPH oxidase; LPS, lipopolysaccharide.

DIM treatment alleviates oxidative stress in mice with LPS-triggered AKI. To further elucidate the mechanisms allowing DIM to mediate its anti-apoptotic effects on LPS-induced cell apoptosis, alterations in oxidative stress were investigated. Mice that received LPS had elevated levels of ROS and MDA and a decreased GSH/GSSG ratio in comparison with the control and DIM groups (Fig. 3). The alterations in oxidative stress indexes significantly improved when treated with DIM.

DIM treatment suppresses the expression of NOX2 and NOX4 in mice with LPS-triggered AKI. As the NOX enzymes are major source of ROS in the kidneys, the effect of DIM on the expression of NOX family members was tested. As shown in Fig. 4, LPS had no effects on the expression of NOX1 and NOX3, but significantly promoted the expression of NOX2 and NOX4. Nevertheless, DIM effectively blocked the LPS-induced expression of NOX2 and NOX4.

Discussion

AKI, which often arises in sepsis cases, is a common problem in critical patients and carries high morbidity and mortality rates (2,21). However, effective medical treatments for AKI are not commonly available. In the present study, the effects of DIM on LPS-induced AKI in mice was tested. DIM conferred protection against AKI induced by LPS, as demonstrated by the improvements in kidney function and the attenuation of the kidney damage score. These effects may be achieved by inhibiting the NOX-mediated oxidative stress and programmed cell death in renal tubular epithelial cells.

Apoptosis is an important pathological mechanism leading to AKI (13,22-25). As expected, the LPS-induced AKI mouse model showed increased apoptosis in renal tubular epithelial cells, as determined by the elevated expression levels of cleaved caspase-3 and cleaved PARP measured by western blot analysis. Consistent with the effects of DIM on renal

function in the AKI mouse model, DIM could decrease the levels of both cleaved caspase-3 and cleaved PARP in kidneys stimulated with LPS. TUNEL staining also showed that DIM could alleviate the level of apoptosis in LPS-induced tubular epithelial cells. Thus, DIM may mitigate LPS-induced AKI by reducing the level of apoptosis in renal tubular epithelial cells.

ROS play an important role in apoptosis induction under both physiological and pathological conditions (26). LPS is a strong inducer of ROS production, and the excessive production of ROS is closely related to the apoptosis of renal tubular cells by causing the release of apoptotic factors, including cytochrome C, and by destroying the permeability of the mitochondrial membrane. Aside from the actions on mitochondria, ROS may also activate sphingomyelinase-generating ceramide, an intracellular mediator of apoptosis in granulocytes (25-28). In the present study, mice challenged with LPS showed a significant increase in ROS and MDA (lipid peroxidation) levels coinciding with a rapid decrease in the GSH/GSSG ratio. Reduced GSH plays an important role as a free radical scavenger to counteract the deleterious effects of ROS (29). These changes were all attenuated by DIM. Thus, DIM may protect against LPS-induced apoptosis by reducing oxidative stress.

ROS are primarily generated in the mitochondria by NOX enzymes, cyclooxygenase and xanthine oxidase. The NOX family is the main source of ROS in the kidneys (30-32). To determine the mechanism by which DIM reduces ROS, the effect of DIM on the expression of NOX enzymes (NOX1-NOX4) was investigated. The data revealed that LPS promoted ROS production primarily through NOX2 and NOX4. The expression of NOX2 and NOX4 was decreased in the presence of DIM. Taken together, the data presented in this present study support the hypothesis that DIM suppresses LPS-triggered oxidative stress by inhibiting the NOX2 and NOX4 pathways.

A total of three major findings have been revealed in this present study. First, DIM significantly improved renal function as demonstrated by the lowering of SCr and BUN levels, as well as the attenuation of pathological kidney damage in the LPS-stimulated AKI mouse model. Secondly, DIM suppressed the pro-apoptotic effects of LPS. Finally, DIM may confer its protective benefits by inhibiting NOX/ROS signaling.

In conclusion, DIM is a potential therapeutic agent for use in AKI triggered by LPS. The possible mechanism through which DIM attenuates renal damage in the LPS-induced AKI mouse model is via the inhibition of NOX-mediated oxidative stress and programmed cell death of renal tubular epithelial cells. This present study focused on an animal model of AKI and was not sufficient to describe how DIM acts on AKI through the regulation of NOX-mediated oxidative stress and programmed cell death of renal tubular epithelial cells. Therefore, further studies using cells are required to elucidate the specific mechanism of DIM regulation.

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Availability of data and materials

The datasets generated and analyzed during the present study are available from the corresponding author on reasonable request.

Authors' contributions

JH and TH performed the experiments, data analysis and manuscript writing. LZ designed the experiments, participated in the generation of ideas and data interpretation and was responsible for the overall direction of this work. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The study protocol was reviewed and approved by the Animal Care and Use Committee of Chongqing Medical University.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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