

The protective effect of the TUG1/miR-197/MAPK1 axis on lipopolysaccharide-induced podocyte injury

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Abstract. The podocyte, a type of glomerular epithelial cell, is the key constituent of the filtration barrier layer in the kidney. Previous studies have shown that long non-coding RNA (lncRNA)-taurine-upregulated gene 1 (TUG1) served a protective role in diabetes-induced podocyte damage. The aim of the present study was to investigate the potential role of TUG1 in the progress of podocyte injury induced by lipopolysaccharide (LPS), and explore the underlying mechanisms. The results showed that TUG1 expression was suppressed in LPS-induced podocytes. Enhanced TUG1 expression by exogenous recombinant vector regulated the expression of podocyte associated proteins [Nephrin, Podocin and CCAAT/enhancer-binding protein (CHOP)]. A marked decrease was observed in the level the albumin influx in cells transfected with TUG1. Further study indicated that microRNA (miR)-197 is a potential target of TUG1. The enhanced level of miR-197 induced by LPS was inhibited in cells transfected with TUG1. The decreased Nephrin and Podocin expression, upregulated CHOP expression and the increased albumin influx were slightly enhanced by miR-197 mimic transfection, while markedly suppressed by miR-197 inhibitor transfection in LPS-induced podocytes. Mitogen-activated protein kinase (MAPK) protein was predicted as a potential target of miR-197. The downregulated expression of phosphorylated-MAPK/MAPK induced by LPS was significantly suppressed by TUG1 transfection in podocytes. In addition to this, autophagy was promoted by TUG1 transfection via the

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elevation of the Beclin1 and light chain (LC)3 II/LC3 I levels, and suppressing p62 expression. However, the p38 MAPK inhibitor SB203580 reversed the changes that TUG1 induced in the levels of Beclin1, LC3 II/LC3 I and p62. Taken together, these results demonstrated that LPS-induced podocyte injury could be alleviated by the TUG1/miR-197/MAPK1 axis.

Introduction

Podocytes are terminally differentiated glomerular cells with elaborate extensions of the cell body and play an important role in maintaining the structural and functional integrity of the filtration barrier (1). Studies have indicated that podocytes damage and the reduction in podocyte density are the main contributors to the development of proteinuria and glomerulosclerosis (2). Thus, podocytes serve as vital role in preventing proteinuria and glomerulosclerosis from getting worse.

Long non-coding RNAs (lncRNAs), a heterogeneous class of long (>200 nucleotides) 5 transcripts, is characterized by an apparent lack of protein-coding potential (3). IncRNAs have attracted much attention of scientists for its diagnostic, prognostic, and predictive potential (4). IncRNA taurine-upregulated gene 1 (TUG1) was firstly identified in a genomic screen as a part of photoreceptors and overexpressed in response to retinal development in mouse retinal cells (5). Researchers have indicated that TUG1 could be a predictive biomarkers of kidney neoplasms. Besides, accumulated studies have demonstrated that TUG1 contributed to the development of chronic kidney diseases (6). However, the underlying molecular mechanisms are not fully understood.

Researches have shown that TUG1 has moderation effects on various diseases via interacting with microRNAs (miRNAs or miRs), including miR-204-5p (7), miR-144-3p (8), miR-382 (9) and so on. Studies over the last decades have suggested that miRNAs served as pathogenic or therapeutic factor to be involved in the development of a variety of renal diseases, such as chronic kidney disease (10), kidney fibrosis (11), type 2 diabetic kidney disease (12) and glucose-induced podocyte apoptosis (13). According to miRDB, at least two binding sites were identified between miR-197 and TUG1. Besides, TargetScan and miRwalk have predicted the high binding possibility between MAPK1 and miR-197. As we all know, MAPK pathway could induce cell

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Abbreviations: MAPK, mitogen-activated protein kinase; LPS, lipopolysaccharide; lncRNAs, long non-coding RNAs; TUG1, taurine-upregulated gene 1; miRNA/miR, microRNA; TNF- α , tumor necrosis factor- α

autophagy in various types of cells (14), and autophagy played a positive role in defending podocyte against lipopolysaccharide (LPS)-induced damage (15). Thus, miR-197 may be a potential biomarker and a regulation factor in the treatment of podocyte injury.

In this study, we investigated the underlying mechanisms of TUG1 in treating LPS-induced podocytes injury. Our results strongly demonstrated that TUG1 could protect podocyte against LPS-induced injury by targeting miR-197 through activating MAPK pathway.

Materials and methods

Cell culture. The mouse immortalized podocyte cell line MPC5 was purchased from National Infrastructure of Cell Line Resource. Cells were cultivated at 33°C with RPMI-1640 containing 10% FBS and recombinant mouse interferon- γ (50 U/ml, Toyobo Co., Osaka, Japan) to propagate cells. After reaching confluence, cells were incubated without interferon- γ to induce cell differentiation into the podocyte lineage at 37°C for more than 6 days. The culture medium was changed every three days. Then, cells were divided into different groups.

Cell transfection. SB203580 was purchased from Calbiochem (San Diego, CA, USA) and dissolved in dimethyl sulfoxide (DMSO). PcDNA-TUG1, pcDNA vector, miR-197 mimic and miR-197 inhibitor were purchased from Addgene (Cambridge, MA, USA). MPC5 cells were transfected with pcDNA-TUG1 (40 μ g/ml), pcDNA vector (40 μ g/ml), miR-197 mimic (80 nM) or miR-197 inhibitor (160 nM) using Lipofectamine[®] 2000 (Invitrogen; Thermo Fisher Scientific, Inc., Waltham, MA, USA) according to manufacturers instruction. 48 h after transfection, transfected MPC5 podocytes and normal MPC5 podocytes were treated with lipopolysaccharide (LPS; 1 μ g/ml) or the combination of LPS (1 μ g/ml) and SB203580 (10 μ M) for 24 h for further experiments.

Western blot assay. The total protein was isolated from MPC5 cells in different groups, and separated by SDS-PAGE and transferred onto polyvinylidene difluoride (PVDF) membranes (Millipore, Billerica, MA, USA). After blocking with 5% BSA and washing with 0.05% Tween-20, membranes were incubated with primary antibodies [Podocin, Nephrin, CCAAT/enhancer-binding protein (CHOP), p-MAPK, MAPK, Beclin1, p62, light chain (LC)3 and β -actin] (Abcam, Cambridge, UK) at 4°C overnight. Then, all membranes were incubation with horseradish peroxidase (HRP)-conjugated secondary antibodies for 1.5 h at room temperature. The bands were analyzed using ImageJ [National Institutes of Health (NIH), Sacaton, AZ, USA].

Reverse transcription-quantitative polymerase chain reaction (RT-qPCR) assay. Total RNAs were extracted from MPC5 podocytes in different groups using TRIzol reagent (Invitrogen; Thermo Fisher Scientific, Inc.) according to the manufacturer's instructions. RT-qPCR was conducted using Applied Biosystems StepOnePlus[™] system. The RT-PCR primers for Podocin, Nephrin, CHOP, TUG1 and miR-197 purchased from GeneCopoeia Inc. (Rockville, MD, USA). The specific primers were as follows: Podocin (forward: 5'-GCA TCAAGCCCTCTGGATTAG-3' and reverse: 3'-AGACGG AGATCAACCTTGTGATA-5'); Nephrin (forward: 5'-ATG GGAGCTAAGGAAGCCACA-3' and reverse: 3'-CCACAC CACAGCTTAACTGTC-5'), CHOP (forward: 5'-TTGCCC TCTTATTGGTCCAGC-3' and reverse: 3'-TAGCGACTGTTC TGTTCCCAC-5'). GAPDH was used as the internal control. The relative levels were measured using the $2^{-\Delta\Delta Cq}$ analysis method (16).

Albumin influx assay. Filtration barrier function of podocyte monolayers was assessed using albumin influx assay. Briefly, $5x10^3$ podocytes were seeded into a 3 μ m pores trans-well filter coated with collagen (Corning Incorporated, Corning, NY, USA), then incubated for 10 days. After that, MPC5 podocytes were serum-starved overnight and washed with PBS containing 1 mmol/l MgCl₂ and 1 mmol/l CaCl₂. Then, top chamber was filled with 0.15 ml RPMI-1640 and the bottom chamber with 1 ml RPMI-1640 containing 40 mg/ml of bovine serum albumin at 37°C with 5% CO₂. Cells were cultured for 24 h. Albumin concentration was measured using an ELISA kit (Thermo Fisher) according to the manufacturer's instructions.

Statistical analysis. SPSS 19.0 software (IBM Corp., Armonk, NY, USA) was used to analyze all the data. The data are presented as the mean \pm standard deviation. Analysis was performed using one-way analysis of variance followed by a Bonferroni post hoc test. Statistical significance was assigned at P<0.05.

Results

TUG1 was downregulated in LPS impaired podocyte. To investigate whether TUG1 expression was related to LPS-induced damage, MPC5 podocytes were treated with LPS (1 μ g/ml) for 24 h. Results indicated that the protein expressions of Podocin and Nephrin were remarkably suppressed while CHOP significantly elevated in LPS group compared with control group (Fig. 1A, C and E). The mRNA expression was consistent with protein expression (Fig. 1B, D and F). LPS treatment result in a greater albumin influx across the podocyte monolayer compared with control (Fig. 1G). These results suggested that the function of MPC5 podocytes was impaired by LPS. Besides that, the relative mRNA expression of TUG1 was dramatically inhibited in LPS group in comparison with control group (Fig. 1H), indicating that TUG1 was downregulated in LPS damaged podocyte.

Overexpression of TUG1 alleviates LPS-induced podocyte injury. To explore whether TUG1 could attenuate the cell damage induced by LPS, MPC5 podocytes were transfected with pcDNA-TUG1. As shown in Fig. 2A, the relative expression of TUG1 was largely enhanced in TUG1 group, indicating the high transfection efficiency. The reduced Podocin and Nephrin expression along with the increased CHOP expression induced by LPS compared with control were significantly inhibited by TUG1 transfection (Fig. 2B-E). Moreover, the elevated albumin influx caused by LPS compared with control was remarkably repressed by





Figure 1. TUG1 is downregulated in LPS damaged podocyte. Podocytes were randomly divided into 2 groups: The control group consisted of cells without treatment; and the LPS group was comprised of cells that were treated with LPS. The protein and relative mRNA expressions of (A and B) Podocin, (C and D) Nephrin and (E and F) CHOP were detected using western blotting and RT-qPCR, respectively. (G) Albumin influx assay was applied to assess the glomerular filtration function. (H) The relative mRNA expression of TUG1 was also measured using RT-qPCR. The experiments were repeated at least 3 times, and data is presented as the mean ± standard deviation. ^{*}P<0.05 vs. control group. TUG1, taurine-upregulated gene 1; LPS, lipopolysaccharide; CHOP, CCAAT/enhancer-binding protein; RT-qPCR, reverse transcription-quantitative polymerase chain reaction.

TUG1 overexpression (Fig. 2F). These results suggested that LPS-induced podocyte injury could be alleviated by TUG1 overexpression.

miR-197 is a direct target of TUG1. The targeting relationship between miR-197 and MAPK1 was predicted by bioinformatics (Fig. 3A). To further evaluate the potential relationship between miR-197 and TUG1, RT-qPCR and western blotting were conducted in podocytes transfected with pcDNA-TUG1/pcDNA-vector/ miR-197 mimic/miR-197

inhibitor. As illustrated in Fig. 3B, the expression of miR-197 was significantly increased in LPS group compared with control group. The elevated level of miR-197 in cells treated with LPS was inhibited by transfection with TUG1 (Fig. 3C). Besides that, the reduced Nephrin and Podocin expression along with the enhanced CHOP expression were slightly enhanced by TUG1 mimic transfection while remarkably suppressed by TUG1 inhibitor transfection (Fig. 3D-G). Moreover, the increased albumin influx induced by LPS was slightly enhanced by TUG1 mimic transfection while



Figure 2. Overexpression of TUG1 alleviates LPS-induced podocyte injury. (A) Podocytes were randomly divided into 3 groups: Control group, cells without treatment; the mock group, cells transfected with pcDNA vector; and cells transfected with pcDNA-TUG1. The relative mRNA expression of TUG1 was assessed using reverse transcription-quantitative polymerase chain reaction. Podocytes were then randomly divided into 4 groups: Control group, cells without treatment; LPS group, cells induced by LPS; LPS+mock group, cells transfected with pcDNA vector that were induced by LPS; and LPS+TUG1 group, cells transfected with pcDNA-TUG1 that were induced by LPS. (B) The protein expressions of Nephrin, Podocin and CHOP were measured using western blot analysis. Data summary and analysis of the expression of (C) Nephrin, (D) Podocin and (E) CHOP in podocyte cells according to the results of western blotting. (F) Albumin influx assay was used to evaluate the glomerular filtration function. The experiments were repeated at least 3 times, and data is presented as the mean ± standard deviation. *P<0.05 vs. control group; #P<0.05 vs. TUG1, taurine-upregulated gene 1; LPS group. LPS, lipopolysaccharide; CHOP, CCAAT/enhancer-binding protein.

remarkably inhibited by TUG1 inhibitor transfection (Fig. 3H). These results indicated that miR-197 is a potential target of TUG1.

lncRNA TUG1 aggravates podocytes damage via regulating MAPK pathway. The target sequences of miR-197 in the 3'-UTR region of MAPK1 was predicted through bioinformatics analysis (Fig. 4A). To detect the regulatory relationship between TUG1 and MAPK1, MPC5 podocytes were transfected with pcDNA-TUG1/pcDNA-vector. As illustrated in Fig. 4B, the decreased expression of p-MAPK/MAPK induced by LPS compared with control was strongly



Figure 3. miR-197 is a direct target of TUG1. (A) The targeting association between miR-197 and TUG1 was predicted through bioinformatics. (B) The expression of miR-197 in podocytes treated with or without LPS was measured using RT-qPCR. (C) Podocytes were randomly divided into 4 groups: Control group, cells without treatment; LPS group, cells induced by LPS; LPS+mock group, cells transfected with pcDNA vector that were induced by LPS; and LPS+TUG1 group, cells transfected with pcDNA.TUG1 that were induced by LPS. The relative miR-197 expression was detected using RT-qPCR. (D) Podocytes were also randomly divided into 4 groups: Control group, cells without treatment; LPS group, cells induced by LPS; and LPS+tinhibitor group, cells transfected with miR-197 inhibitor that were induced by LPS; and LPS+inhibitor group, cells transfected with miR-197 inhibitor that were induced by LPS. The protein expressions of Nephrin, Podocin and CHOP were detected by western blot analysis. Data summary and analysis of the expression of (E) Nephrin, (F) Podocin and (G) CHOP in podocytes cells according to the results of western blotting. (H) Albumin influx assay was applied to assess the glomerular filtration function. The experiments were repeated at least 3 times, and data is presented as the mean ± standard deviation. *P<0.05 vs. control group; *P<0.05 vs. LPS group. TUG1, taurine-upregulated gene 1; LPS, lipopolysaccharide; RT-qPCR, reverse transcription-quantitative polymerase chain reaction; CHOP, CCAAT/enhancer-binding protein; miR, microRNA.



Figure 4. miR-197 aggravates podocyte damage by regulating the MAPK signaling pathway. (A) The targeting association between miR-197 and MAPK1 was predicted by bioinformatics. (B and C) Podocytes were randomly divided into 4 groups: Control group, cells without treatment; LPS group, cells induced by LPS; LPS+mock group, cells transfected with pcDNA vector that were induced by LPS; and LPS+TUG1 group, cells transfected with pcDNA-TUG1 that were induced by LPS. (B) The expression of p-MAPK/MAPK was measured using western blotting. (C) The levels of Beclin1, p62 and LC3 I/ LC3 II were evaluated using western blotting. (D) Podocytes were randomly divided into 4 groups: Control group, cells without treatment; LPS group, cells induced by LPS; LPS+TUG1 group, cells transfected with pcDNA-TUG1 that were induced by LPS; LPS+TUG1 group, cells transfected with pcDNA-TUG1 that were induced by LPS; LPS+TUG1 group, cells transfected with pcDNA-TUG1 that were induced by LPS; and LPS+TUG1+SB group, cells transfected with pcDNA-TUG1 were treated with LPS and SB. The expression level of Beclin1, p62 and LC3 I/ LC3 II were detected using western blot analysis. The experiments were repeated at least 3 times, and data is presented as the mean ± standard deviation. *P<0.05 vs. control group; #P<0.05 vs. LPS group; \$P<0.05 vs. LPS+TUG1 group. MAPK, mitogen-activated protein kinase; LPS, lipopolysaccharide; TUG1, taurine-upregulated gene 1; LC3, light chain 3; p-, phosphorylated; SB, SB203580 (inhibitor).

inhibited by TUG1 transfection. Besides, the decreased Beclin1, LC3 II/LC3 I along with upregulated p62 levels were vastly suppressed by TUG1 transfection, indicating TUG1 could regulate cell autophagy via regulating MAPK pathway (Fig. 4C). To further validate this result, SB203580 was added into cells to inactivate MAPK pathway. Results indicated that SB203580 could reverse all the changes in the expressions of Beclin1, LC3 II/LC3 I and p62 caused by TUG1 (Fig. 4D). These results suggested that lncRNA TUG1 could aggravate podocytes damage via regulating MAPK pathway indirectly.

Discussion

The podocyte is a kind of special glomerular epithelial cell and key constituent of the layer of the filtration barrier in the kidney (17). Intact kidney filtration barrier maintains the balance of proteins in the blood (18). If the podocytes are impaired, plasma albumin can leak into the urinary space, leading to the deterioration of kidney diseases and hypoalbuminemia. If the albuminuria persist and increase in patients, it will develop into systemic diseases, such as anemia, postural hypotension, bradyarrhythmia. Thus, treatment for podocyte damage is extremely important to the prevention of various renal diseases and related systemic diseases.

It has been suggested and supported by researchers that lncRNAs have extensive biological functions. Abnormal expression of lncRNAs is linked to a variety of diseases, such as cancer as well as neurological, cardiac and renal diseases (19,20). Among these lncRNAs, Long and colleagues observed that the expression of TUG1 was remarkably inhibited in the podocytes of diabetic mice. Podocyte-specific overexpression of TUG1 could elevate PGC-1 α expression, resulting in the improvement of mitochondrial bioenergetics (21). Similarly, in our research, overexpression of TUG1 could protect podocyte from LPS-induced damage, evidences were the reduced Nephrin, Podocin expression, elevated CHOP expressions along with the enhanced albumin influx were vastly inhibited by TUG1 transfection. However, how did TUG1 worked need further exploration.

Previous studies have shown that TUG1 acted as a regulation role in a variety of diseases through interacting with miRNAs. miRNAs, existed in eukaryotic cells, could regulate various physiological biological metabolism through combining with 3'-UTR of target genes mRNA, thereby promoting mRNA cleavage or suppressing the translation of mRNA (22). More and more researchers have demonstrated that many miRNAs were involved in the development of multiple kidney diseases. For example, miR-21 acts as an oncogene in renal cancer via inducing cell proliferation and invasion (23). miR-214, -132, -21 and -15b have been testified to participate in the renal fibrosis process (10). It is predicted by TargetScan and miRwalk that miR-197 has a close relationship to TUG1, at least two binding sites were found between them. Our research further provided evidences that miR-197 may be a target of TUG1 through detecting the miR-197 content in podocytes transfected with TUG1. In addition to this, the decreased Nephrin, Podocin expression, upregulated CHOP expressions along with the increased albumin influx induced by LPS were slightly enhanced by miR-197 mimic transfection while significantly suppressed by miR-197 inhibitor transfection in podocytes.

Apoptotic cell death is a genetically programmed mechanism that allows the cell to commit suicide (24), and plays a vital role in the development of multiple organ dysfunction in LPS-induced sepsis (25). miRNAs serve as the useful laboratory markers of organ failure during sepsis and have various effects on the processes of pro-apoptosis or anti-apoptosis (26,27). According to the previous report, miR-197 induced apoptosis and suppressed tumorigenicity in multiple myeloma xenograft models (28). Report also indicate that the level of miR-197 was elevated in severe acute viral hepatitis with coagulopathy (29). Thus, cell apoptosis regulated by miR-197 may be involved in the pathological process of LPS-induced podocytes injury. Autophagy serves a vital role in keeping the balance between anabolism and catabolism, maintaining protein quality control and removing impaired proteins, organelles and lipids (30). The results obtained from the current studies have demonstrated that the dysfunction of podocyte autophagy taken responsibility for the development of various kidney diseases, including diabetic nephropathy (31), immunoglobulin A Nephropathy (32) and proteinuric kidney disease (33). Besides, Tan et al (15) pointed out that the podocytes autophagy was remarkably suppressed by LPS, which result in LPS-induced injury of podocytes. The inducing autophagy ability of MAPK pathway was demonstrated by many researchers in multiple diseases (34,35). Thus, MAPK pathway may be a major target in treating LPS-induced podocytes injury. According to the prediction of TargetScan and miRwalk, miR-197 may inactivate MAPK pathway via binding to MAPK1, and then restrain the autophagy of podocyte. Considering that miR-197 is a target of TUG1, TUG1 might induce the autophagy of podocytes via motivating MAPK pathway indirectly. Our study demonstrated that TUG1 transfection could remarkably inhibit the increased p-MAPK/MAPK level in LPS-induced podocytes. Besides that, the reduced Beclin1, p62 and LC3 II/I and increased p62 expression caused by LPS were remarkably inhibited by TUG1 transfection. Moreover, the p38 MAPK inhibitor SB203580 could reverse all the changes caused by TUG1 in LPS-induced podocytes.

All in all, our study indicated that TUG1 level was downregulated in podocytes impaired by LPS. Elevated TUG1 by transfection suppressed the LPS-induced alteration of podocyte injury related proteins (Nephrin, Podocin and CHOP) expression and elevated albumin influx. Further research demonstrated that miR-197 was a direct target of TUG1. TUG1 was confirmed to restore the function of podocytes via sponging miR-197 to enhance the level of p-MAPK/MAPK, thereby inducing autophagy. In summary, the TUG1-miR-197-MAPK pathway may act as a potential new target for LPS-induced podocytes injury therapy.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

DZ analyzed and interpreted the data regarding cell culture, transfection and western blot analysis, and conducted the statistical analysis. ZL was involved in RT-qPCR analysis. HZ was responsible for the Albumin influx assay, as well as the study design and drafting of the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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