Basaloid squamous cell carcinomas (BSccs) in oral lesions are extremely rare and the histology is not well understood. Histologically, they are often similar to conventional squamous cell carcinoma (SCC). The present study was designed with an aim to distinguish BScc from SCC using claudin-4, occludin, SrY-box 2 (SoX2) and proliferating cell nuclear antigen (Pcna) immunoreactivities and staining patterns. Three BSccs (with abundant, with moderate, and without squamous components) specimens and 20 SCC specimens were selected for comparison of their immunoreactivity. These specimens were stained with claudin-4, occludin, SoX2 and Pcna. In addition to histological analysis, the expression of claudin-4, occludin and PCNA was determined in oral cancer HSc2 and HSc3 cells with or without SoX2 overexpression, and cell proliferation was determined by XTT assay. Claudin-4 had strong and occludin had weak immunoreactivity as detected in the membrane of squamous components of BScc but not in cancer cells. No obvious detection of squamous components and cancer cells were observed in SCC. SoX2 and PCNA immunoreactivities in SCC had dot-like staining patterns in the nuclei of partial and marginal cancer cells. In contrast, in BSccs, SoX2 and PCNA had diffuse staining patterns in almost all cancer cells. SoX2 overexpression had little effect on the expression levels of claudin-4, occludin and PCNA. It also had little effect on the cell proliferation of HSC2 and HSC3 cells. Differences in immunoreactivity and staining pattern may be valuable to distinguish between BScc and SCC in diagnosis.

Introduction

Basaloid squamous cell carcinomas (BSccs) in oral lesions are extremely rare and are a variant of squamous cell carcinoma (SCC) (1-4). Although BSccs are reportedly aggressive, have early lymph node metastasis, and have worse prognosis, their biological features are similar to those of conventional SCC (1-4). Several studies have demonstrated that BSccs have grossly flat or slightly elevated mucosa and histologically small crowded cells with hyperchromatic nuclei and stromal hyalinization (1-4). BScc exhibits comedonecrosis and has a palisading pattern of basal cells with squamous components in the center, although not specifically (3,5,6). As the histological aspects of BScc are often similar to those of SCC, it is difficult to distinguish BScc from SCC for diagnosis, especially in biopsy specimens. It has also been reported that BSccs exhibit high proliferation involving proliferating cell nuclear antigen (PCNA) and Ki-67 immunoreactivity (7-9). Interestingly, cell adhesion molecule E-cadherin and b-catenin expression patterns reportedly differ between BScc and SCC (9). There are few reports that feature tight junction molecule analysis of claudin-4, occludin and PCNA (10,11). Their immunoreactivities were reported to be extremely weak in cancer cells (10,11). Whether claudin-4 and occludin are expressed in BScc, however, remains unknown. We hypothesized that cell proliferation, adhesion and tight junction markers facilitate a differential diagnosis. In the present study, the immunohistochemical detection of cell proliferation marker PCNA, tight junction markers claudin-4 and occludin, and stem cell marker SOX2 was conducted in BScc and SCC. Additionally, it was ascertained whether SOX2 overexpression affects the expression of these factors and cell proliferation.

Materials and methods

Tissue preparation. Histological specimens collected from January 2014 to December 2018 were retrieved from our hospital's archives according to the guidelines of the...
Japanese Society of Pathology. Informed consent was provided by each patient for the use of the clinical images. Two cases of BSCC and 20 cases of conventional SCC were collected from craniocervical lesions, and each case included lymph node metastasis. Specimens were obtained from the Department of Diagnostic Pathology, Wakayama Medical University. Another BSCC specimen was obtained from the Department of Pathology, Tsurumi University School of Dental Medicine. Diagnosis was performed by at least two pathologists. Clinical and pathological information is described below. Case 1 is a female patient, 81 years of age, with a clinically diagnosed mandibular malignant tumor; pathological diagnosis is BSCC with abundant squamous components. Case 2 is a male patient, 57 years of age, with a clinically diagnosed malignant tumor in the palatine tonsil; pathological diagnosis is BSCC with moderate squamous components and lymph node (LN) metastasis. Case 3 is a male patient, 68 years of age, with clinically diagnosed tongue cancer; pathological diagnosis is BSCC without squamous components. Case 4 is a female patient, 87 years of age, with clinically diagnosed gingival cancer; pathological diagnosis is well-differentiated SCC. Case 5 is a male patient, 78 years of age, with clinically diagnosed tongue cancer; pathological diagnosis is well-differentiated SCC and LN metastasis. Case 6 is a female patient, 74 years of age, with clinically diagnosed buccal cancer; pathological diagnosis is poorly differentiated SCC. The clinical information regarding the other SCC cases is documented in Table I. Partial resection and LN dissection specimens were used in cases 2 and 5, and biopsy specimens were used in the other cases.

Immunohistochemistry. Claudin-4, occludin, SOX2 and PCNA expression in BSCC and SCC tissues were evaluated from serial deparaffinized sections. Our specimens had been fixed with formalin from 24 to 48 h at room temperature and treated with routine processing as in a previous study (12). Ten 4-mm serial sections were prepared for staining and were incubated with primary antibodies for 2 h. Immunohistochemistry was performed using a Discovery Auto-Stainer with automated protocols (Ventana Medical Systems, Inc.; Roche) as previously described (13). The average number of PCNA-positive cells from case 1 to case 23 were counted in 10 random microscopic fields at 400 magnification. The SOX2 intensity was determined by qualitative assessment of three levels as weak, 1; moderate, 2; and strong, 3. We defined a diffuse staining pattern as that positively detected in almost all cancer cells but a dot-like staining pattern as that positively detected in partial and marginal cancer cells.

Cell culture and treatment. HSC2 and HSC3 human oral squamous cell carcinoma cells, which do not have basoloid characteristics were obtained from the Health Science Research Resources Bank (Osaka, Japan). These cells were cultured in Dulbecco's modified Eagle's medium (DMEM) (Sigma Chemical Co.; Merck KGaA) supplemented with 10% fetal bovine serum and 1% penicillin streptomycin antibiotics. The construct expression vector for SOX2 was used for transfection. The human SOX2 plasmid (26817) was purchased from Addgene. Transient plasmid transfection was performed as previously described (14).

Western blot analysis. HSC2 cells were lysed using M-PER lysis buffer (Thermo Fisher Scientific, Inc.). Protein determination was performed using the bicinchoninic method. A total of 40 µg protein was loaded in each lane. The total cell lysates were run on 14% SDS-polyacrylamide gels followed by western blotting using standard procedures. The proteins were transferred on to PVDF membrane. For blocking, membranes were incubated with 5% skim milk for 60 min at room temperature after protein transfer. A WesternBright Sirius kit (Advansa) was used for antibody detection, and an AE-9300 Ez capture MG (ATTO) was used for image data capture. We repeated the western blot analysis three times and the results were similar.

Antibodies. The following commercial antibodies were purchased: claudin-4 (1:100, mouse monoclonal, sc-376643; Santa Cruz Biotechnology Inc.), occludin (1:100, mouse monoclonal, sc-132255; Santa Cruz Biotechnology), SOX2 (1:100, rabbit polyclonal, Ab97959, Abcam), PCNA (1:1,000, mouse monoclonal, sc-56; Santa Cruz Biotechnology) and actin (1:10,000, mouse monoclonal, A5441; Sigma Chemical Co.; Merck KGaA).

Real-time (quantitative) PCR (qPCR). We prepared three independent RNA samples from HSC3 cells. Total RNA was isolated and first-strand cDNA was synthesized as previously described (13). Real-time PCR was performed using SYBR Green Master Mix (Bio-Rad Laboratories, Inc.). The thermocycling conditions were the following: Initial denaturation at 95°C for 3 min, followed by 40 cycles of 95°C for 10 sec and 60°C for 30 sec. The 2$^{-\Delta\Delta C q}$ method was used for relative quantification (15). Amplification primer sequences were designed as follows: SOX2-F, 5'-GAA TGTCCTCAT GGTTGTGT-3' and R, 5'-TTGCTGATCTCCAGGTGTTG-3'; claudin-4-F, 5'-ATGGCCCTCATGCTGCTGCTA-3' and R, 5'-AGCGGAGTGCACACCTTGCA-3'; occludin-F, 5'-GGCG GATTGTTTATCTTGGGA-3' and R, 5'-CTGGATGACATG GCTGATGG-3'; PCNA-F, 5'-AGGTGTGGAGGCACTCAAG-3' and R, 5'-AGTCCATGCTCTGAGGT-3'; and 18S rRNA-F, 5'-GGCGCGCTAGAGGTGAAAT-3' and R, 5'-GAA AACATTCTTGGCAAAATGT-3'. Data were normalized using 18S rRNA. qPCR was repeated three times and the results were similar.

Cell proliferation assay. HSC2 and HSC3 cells were seeded into 96-well plates. After transfection, 50 µl of XTT kit reaction solution was added to each well (XTT-based) (Biological Industries) and the cells were incubated at 37°C for an additional 2 h according to the manufacturer's instructions. Absorbance at optical density at 480 nm (OD$_{480}$) and at (OD$_{690}$) was measured using a 96-well microplate reader (SH-9000; Hitachi).

Statistical analysis. Statistical analysis with Dunnett’s test was performed using JMP Pro software version 13.0 (SAS Institute Japan, Tokyo, Japan). The data are expressed as the mean value ± SE (bars) of three independent samples. *P<0.05
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was considered to indicate a statistically significant result, and ANOVA was used followed by Dunnett's test.

Results

Macroscopic and radioscopic aspects in BSCC. Representative macroscopic and radioscopic aspects in case 1 of BSCC are shown in Fig. 1. Fig. 1A shows flat appearance of gingival mucosa in a right anterior lesion. Fig. 1B shows the huge mandibular swelling that was found. Diffuse swelling was found in the palatine tonsil or tongue in case 2 and 3, respectively (data not shown). An orthopantomograph of the BSCC at first medical examination showed a multilocular cyst surrounded by well-defined radiolucent borders (Fig. 1C, left panel). The patient was treated with radiotherapy and the orthopantomograph at 8 months after the first examination showed a decreased multilocular cyst with calcification (Fig. 1C, right panel). This indicated that radiotherapy was effective. The other two patients with BSCC were not treated with radiotherapy.

Histological aspects of BSCC and SCC. Fig. 2A shows representative histological hematoxylin and eosin (H&E)-stained images from case 1, 2 and 3 of BSCC. Fig. 2B shows representative histological images from case 4, 5 and 6 of SCC.

Immunohistochemical detection of claudin-4, occludin, SOX2 and PCNA in BSCC and SCC. Representative images of claudin-4, occludin, SOX2 and PCNA immunoreactivities for case 1, 2 and 3 of BSCC are shown in Fig. 3. Claudin-4 immunoreactivity in case 1 was strongly detected in the cell membrane of squamous components, whereas no obvious detection was observed in cancer cells. Claudin-4 immunoreactivity in case 2 of the primary tumor was weakly detected in the membrane of squamous components, whereas no obvious detection was observed in the cancer cells. No obvious detection of claudin-4 immunoreactivity in case 2 was observed in cancer cells of LN metastasis. Claudin-4 immunoreactivity in case 3 was not detected in the cancer cells. Occludin immunoreactivity was similar to claudin-4 as above, but in case 1 it was weakly detected when compared to claudin-4. We defined diffuse staining patterns as positively detected in almost all cancer cells but dot-like staining patterns as positively detected in partial and marginal cancer cells. SOX2 in case 1 had a diffuse staining pattern in the nucleus, cytoplasm and in the membrane of squamous components. SOX2 immunoreactivity

Table I. Immunohistochemical detection of SOX2 and PCNA in BSCC and SCC specimens.

<table>
<thead>
<tr>
<th>Case</th>
<th>Age</th>
<th>Sex</th>
<th>Lesion</th>
<th>Diagnosis</th>
<th>SOX2 intensity</th>
<th>PCNA-positive cells</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>81</td>
<td>F</td>
<td>Mandibular</td>
<td>BSCC</td>
<td>3</td>
<td>113</td>
</tr>
<tr>
<td>2</td>
<td>57</td>
<td>M</td>
<td>Tonsil</td>
<td>BSCC</td>
<td>3</td>
<td>358.6</td>
</tr>
<tr>
<td>3</td>
<td>68</td>
<td>M</td>
<td>Tongue</td>
<td>BSCC</td>
<td>3</td>
<td>224.2</td>
</tr>
<tr>
<td>4</td>
<td>87</td>
<td>F</td>
<td>Gingiva</td>
<td>SCC (w)</td>
<td>2</td>
<td>48.2</td>
</tr>
<tr>
<td>5</td>
<td>78</td>
<td>M</td>
<td>Tongue</td>
<td>SCC (w)</td>
<td>3</td>
<td>51.2</td>
</tr>
<tr>
<td>6</td>
<td>74</td>
<td>F</td>
<td>Buccal</td>
<td>SCC (p)</td>
<td>1</td>
<td>33.4</td>
</tr>
<tr>
<td>7</td>
<td>87</td>
<td>M</td>
<td>Gingiva</td>
<td>SCC (w)</td>
<td>2</td>
<td>62.4</td>
</tr>
<tr>
<td>8</td>
<td>85</td>
<td>M</td>
<td>Gingiva</td>
<td>SCC (p)</td>
<td>2</td>
<td>37.3</td>
</tr>
<tr>
<td>9</td>
<td>57</td>
<td>M</td>
<td>Tongue</td>
<td>SCC (w)</td>
<td>1</td>
<td>27.6</td>
</tr>
<tr>
<td>10</td>
<td>86</td>
<td>M</td>
<td>Gingiva</td>
<td>SCC (m)</td>
<td>2</td>
<td>41.4</td>
</tr>
<tr>
<td>11</td>
<td>85</td>
<td>M</td>
<td>Gingiva</td>
<td>SCC (m)</td>
<td>2</td>
<td>41.2</td>
</tr>
<tr>
<td>12</td>
<td>69</td>
<td>M</td>
<td>Maxillary</td>
<td>SCC (p)</td>
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<td>50.9</td>
</tr>
<tr>
<td>13</td>
<td>64</td>
<td>M</td>
<td>Tongue</td>
<td>SCC (m)</td>
<td>3</td>
<td>47.6</td>
</tr>
<tr>
<td>14</td>
<td>73</td>
<td>F</td>
<td>Tongue</td>
<td>SCC (m)</td>
<td>2</td>
<td>73</td>
</tr>
<tr>
<td>15</td>
<td>87</td>
<td>M</td>
<td>Oral floor</td>
<td>SCC (m)</td>
<td>2</td>
<td>52.4</td>
</tr>
<tr>
<td>16</td>
<td>89</td>
<td>F</td>
<td>Gingiva</td>
<td>SCC (w)</td>
<td>3</td>
<td>56.7</td>
</tr>
<tr>
<td>17</td>
<td>69</td>
<td>M</td>
<td>Tongue</td>
<td>SCC (w)</td>
<td>2</td>
<td>47</td>
</tr>
<tr>
<td>18</td>
<td>90</td>
<td>F</td>
<td>Gingiva</td>
<td>SCC (w)</td>
<td>3</td>
<td>41.9</td>
</tr>
<tr>
<td>19</td>
<td>53</td>
<td>M</td>
<td>Tongue</td>
<td>SCC (w)</td>
<td>3</td>
<td>94.8</td>
</tr>
<tr>
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<td>66</td>
<td>F</td>
<td>Tongue</td>
<td>SCC (m)</td>
<td>2</td>
<td>81.2</td>
</tr>
<tr>
<td>21</td>
<td>60</td>
<td>M</td>
<td>Oral floor</td>
<td>SCC (m)</td>
<td>3</td>
<td>68.4</td>
</tr>
<tr>
<td>22</td>
<td>69</td>
<td>M</td>
<td>Oral floor</td>
<td>SCC (m)</td>
<td>3</td>
<td>77.2</td>
</tr>
<tr>
<td>23</td>
<td>65</td>
<td>M</td>
<td>Oral floor</td>
<td>SCC (p)</td>
<td>3</td>
<td>77</td>
</tr>
</tbody>
</table>

SOX2 intensity was classified as follows: 1, weak; 2, moderate; 3, strong. PCNA-positive cells were calculated as the average number of PCNA-positive cells in 10 random microscopic fields at 400x magnification. BSCC, basaloid squamous cell carcinoma; SCC, squamous cell carcinoma; PCNA, proliferating cell nuclear antigen; SOX2, SRY-box 2; w, well differentiated; m, moderately differentiated; p, poorly differentiated.
Figure 1. Clinical features of BSCC. (A) Appearance of flat gingival mucosa in the right mandibular (arrows). (B) Mandibular swelling. (C) The radiographical presentation is that of an extensive and well-defined multilocular radiolucency (left panel, black arrows) in right anterior lesion of mandibular. An orthopantomograph at the second medical examination (8 months after the initial examination) showing decreased multilocular cyst with calcification (right panel, white arrows). BSCC, basaloid squamous cell carcinoma.

Figure 2. Histological features of BSCC and SCC. (A) Representative H&E images of case 1, 2 and 3 of BSCC. Case 1, BSCC with abundant squamous components. Case 2, BSCC with moderate squamous components. Case 2 includes primary tumor and lymph node (LN) metastasis. Case 3, BSCC without squamous components. Top panels shows x100 magnification and bottom panels show x200 magnification (scale bars, 200 mm). Square inset shows a larger image. Black arrows show a palisading pattern in the basal cells, and the gray arrow shows large atypical nucleus. (B) Representative H&E images of case 4, 5, and 6 of SCC. Case 4, SCC with abundant squamous components. Case 5, SCC with abundant squamous components of primary tumor and LN metastasis. Case 6, SCC without squamous components. There are no palisading pattern in case 6, but a large atypical nucleus is observed (black arrow). Top panel shows x100 magnification and bottom panel shows x200 magnification (scale bars, 200 mm). BSCC, basaloid squamous cell carcinoma; SCC, squamous cell carcinoma; H&E, hematoxylin and eosin.
in cases 2 and 3 of the primary tumor and LN metastasis exhibited a diffuse staining pattern in the nucleus and some positive staining in squamous components was observed. The PCNA immunoreactivity of the primary tumor and LN metastasis in case 1, 2 and 3 also had diffuse staining in the nucleus, but there was no obvious detection observed in squamous components.

Representative images of claudin-4, occludin, SOX2 and PCNA immunoreactivities in SCC are shown in Fig. 4A. No positive immunoreactivities for claudin-4 and occludin in the primary tumor and LN metastasis from cases 4, 5 and 6 were detected in either cancer cells or squamous components. SOX2 immunoreactivities from case 4, 5 and 6 were dot-like staining patterns in the nucleus of cancer cells. No obvious detection was observed in squamous components. PCNA immunoreactivity from case 4, 5 and 6 of the primary tumor was similar to that of SOX2. There appeared to be weaker detection of PCNA immunoreactivity in case 6 compared with case 4 and 5. SOX2 intensities and PCNA-positive cells of BSCC and SCC cases are provided in Table I and Fig. 4B. The numbers of PCNA-positive cells were higher in BSCC compared to SCC. SOX2 intensities in BSCC were all strong, but they exhibited a variation in SCC.

**SOX2 overexpression exhibits little effect on the expression of claudin-4, occludin and PCNA.** SOX2 expression was strongly detected in all cases of BSCC. Therefore, it was ascertained whether SOX2 overexpression affects expression of claudin-4, occludin and PCNA using oral SCC cancer HSc2 cells. SOX2 overexpression had little effect on the protein expression of occludin and PCNA (Fig. 5A). Endogenous protein expression of claudin-4 was not detected in HSc2 cells, thus expression levels were further examined by qPCR in HSc3 cells. SOX2 overexpression had little effect on the mRNA expression of claudin-4, occludin and PCNA in HSc3 cells (Fig. 5B).

**SOX2 overexpression exhibits little effect on cell proliferation.** Abnormalities in SOX2 expression affect proliferation, migration and apoptosis in various types of cancer cells. It was thus ascertained whether SOX2 overexpression affects cell proliferation of oral cancer HSc2 and HSc3 cells. SOX2 overexpression had little effect on the cell proliferation of HSc2 and HSc3 cells at 24 and 48 h (Fig. 5C).
Basaloid squamous cell carcinoma (BSCC) is regarded as a high-grade malignant tumor, yet it remains unclear how to distinguish it from squamous cell carcinoma (SCC). Oral BSCC is extremely rare, thus it is important to understand both the clinical and histological aspects. At our university, a diagnosed of oral BSCC has been made only in 2 cases during the past 20 years. Moreover, in another university, only 1 case of BSCC was found during the past 20 years. Therefore, more than 3 cases of BSCC could not be obtained, making additional cases difficult to find. Articles reporting on oral BSCC are quite rare, and there are certain studies that have used only 2 BSCC cases (16-18). We will attempt use more BSCC cases, collaborating with other universities in future research.

Clinically, BSCC is highly sensitive to radiotherapy (4,19). SCC, on the other hand, is not sensitive. In case 1, radiotherapy induced the suppression of tumor growth and calcification in the mandible. This clinical aspect is compatible with BSCC.

The histological aspects of BSCC are similar to SCC. Takubo et al (20) reported that a hyaline substance positively stained with periodic acid-Schiff (PAS) staining had been observed in BSCC of the esophagus. In the present study, however, we did not obviously observe positive staining of PAS in BSCC (data not shown). In addition, there are no critical markers for distinguishing between the two. Therefore, immunohistochemical detection of claudin-4, occludin, SOX2 and PCNA was conducted in BSCC compared to SCC. Overexpression of claudins has been reported in several types of cancer, and the overexpression or downregulation of

![Figure 4. Immunoreactivities of claudin-4, occludin, SOX2 and PCNA in SCC. (A) Representative images of the immunoreactivities of claudin-4, occludin, SOX2 and PCNA in case 4, 5 and 6 of SCC. Each panel shows x200 magnification (scale bars, 200 mm). Each square shows larger immunostaining images of SOX2 and PCNA. (B) The average number of PCNA-positive cells in cases 1-6 in 10 random microscopic fields at x400 magnification. SCC, squamous cell carcinoma; SOX2, SRY-box 2; PCNA, proliferating cell nuclear antigen.]
claudins and occludin are associated with tumor progression and recurrence (21-24). Therefore, whether claudin-4 and occludin are candidate markers for BS cc was ascertained. It is worth noting that the immunoreactivities of claudin-4 and occludin in BS cc were detected in the membrane of squamous components. They were not observed in cancer cells of BS cc without squamous components in case 3. This implies that claudin-4 and occludin play important roles in squamous components of BS cc. The squamous components are created by keratinization. Claudin-4 and occludin may therefore regulate the adhesion of keratinization in BS cc. On the contrary, no obvious detection of claudin-4 and occludin immunoreactivities in SCC were observed in either squamous components or cancer cells. It has been reported that claudin-4 immunoreactivity in SCC is weakly or not significantly detected in the membrane of cancer cells. Considering this, it was speculated that the functions of claudin-4 and occludin in squamous components of BS cc differ from those of SCC. The
differences in immunoreactivities between BSCC and SCC are valuable for differential diagnosis. However, these differences are not useful in BSCC without squamous components. Future research must clarify this possibility by using claudins with more cases of BSCC.

The staining patterns of PCNA immunoreactivity between BSCC and SCC were also found to be different. The PCNA labeling index is over 50% in BSCC and is associated with the malignant feature of BSCC (25). In SCC, the PCNA immunoreactivities exhibited dot-like staining patterns in nuclei, whereas in BSCC they appeared as diffuse staining patterns in the nuclei. Sampaio-Góes et al (7) and Yu et al (25) also reported findings that corroborate ours. Our specimens were fixed with formalin from 24 to 48 h. It has been reported that formalin fixation for over 24 and 48 h ensures better results for immunoreactivity (26). Thus, our samples used for immunoreactivity analysis were maintained in optimum conditions.

SOX2 overexpression is a poor prognostic factor and is associated with tumor progression (27-31). SOX2 regulates matrix metalloproteinase (MMP)-2, MMP-3, vimentin, slug, E-cadherin and β-catenin (31-33). Whether SOX2 plays a significant role in oral cancer, however remains unclear. We found that SOX2 immunoreactivity in SCC was presented in a dot-like staining pattern, but in BSCC this was exhibited as a diffuse staining pattern. SOX2 immunoreactivity in BSCC in case 1 was also observed in the membrane of squamous components. We therefore examined whether SOX2 overexpression affected the expression of claudin-4 and occludin, but it had little effect. This implies that claudin-4 and occludin overexpression may occur independently of SOX2. Finally, the effect of SOX2 overexpression on cell proliferation was determined, involving PCNA expression. No significant effect was observed implying that other factors, such as c-Myc, may be associated with cell proliferation of oral cancer.

Human papillomavirus (HPV) affects the progression, recurrence and radiotherapy sensitivity of tonsillar and posterior tongue SCC (34,35). In addition, the protein expression of angiogenesis-related proteins and TNF-receptors differ between HPV-positive and -negative tissues (34). Although our BSCC sample from case 2 was from the tonsil, the p16 immunoreactivity was negative. It is possible that the observed immunoreactivities of case 2 may not be due to HPV infection. Our tongue SCC cases were from a side margin lesion; however, future studies needs to clarify whether the HPV status affects progression and immunoreactivities.

Poorly differentiated SCC has no squamous components, thus it is extremely difficult to distinguish poorly differentiated SCC from BSCC without squamous components. In this study, it was demonstrated that the staining patterns of SOX2 and PCNA immunoreactivities in BSCC and SCC were different. SOX2 and PCNA presented as dot-like staining, appearing as partially marginal staining in cancer cells of SCC, whereas they were diffuse staining in almost all cancer cells of BSCC. On the other hand, there were no differences in claudin-4 and occludin immunoreactivities. These diffuse staining patterns of SOX2 and PCNA suggest that BSCC has more aggressive and proliferative potential than SCC. It has been reported that the diffuse staining pattern of NOTCH1 is correlated with mutation status and poor prognosis in adenoid cystic carcinoma (36). Therefore, diffuse staining patterns of SOX2 and PCNA may correlate with BSCC mutation and progression.

This observation may be useful for additional diagnosis. In conclusion, claudin-4 and occludin immunoreactivities and staining patterns of SOX2 and PCNA may be utilized to potentially carry out a differential diagnosis between BSCC and SCC.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors’ contributions

FS and UKB performed experiments, carried out the pathological diagnosis and completed the draft. FS, SIM and YM performed the pathological diagnosis. IT and SF performed the clinical diagnosis and submitted clinical images and materials. UKB, IT, SF, SIM and YM revised manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate

This study was approved by the Wakayama Medical University Research Ethics Committee (Protocol no. 1715) and historical specimens were retrieved from our hospital archives. Informed consent was provided by each patient for the use of the clinical images.

Patient consent for publication

The patient provided written informed consent for the publication of clinical images.

Competing interests

The authors declare that they have no competing interests.

References


