

Downregulation of long non-coding RNA ANRIL promotes proliferation and migration in hypoxic human pulmonary artery smooth muscle cells

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Abstract. Pulmonary arterial hypertension (PAH) is a progressive syndrome. When PAH occurs, the circulatory resistance of the pulmonary vasculature will gradually increase, which may lead to right heart failure and death. Pathological features of PAH include abnormal proliferation of pulmonary vascular smooth muscle cells and pulmonary vascular remodeling. Hypoxia is the main cause of PAH, which directly induces the contraction and proliferation of pulmonary artery smooth muscle cells (PASMCs), and eventually leads to pulmonary vascular remodeling. Recent studies have shown that long non-coding RNAs (lncRNAs) play key roles in numerous biological processes, including cell proliferation and the occurrence and development of cardiovascular diseases. Studies have also shown that lncRNA antisense noncoding RNA in the INK4 locus (ANRIL) can promote the proliferation of vascular smooth muscle cells. Therefore, the hypothesis of the present study was that ANRIL may be expressed in PASMCs and play a regulatory role. In this study, the expression of ANRIL was analyzed by quantitative PCR. The effects of ANRIL on human pulmonary artery smooth muscle cells (HPASMCs) were assessed by MTT assay, flow cytometry, bromodeoxyuridine incorporation assay, Transwell assay, scratch-wound assay, immunofluorescence assay and western blotting. These experiments revealed that the expression of ANRIL was significantly downregulated in HPASMCs induced by hypoxia. The downregulation of ANRIL affected the cell cycle, making more HPASMCs move from the G0/G1 phase to the G2/M+S phase and strengthening the cell proliferation. Moreover, downregulated ANRIL increased the migration of HPASMCs under hypoxia. This study identified ANRIL as a critical regulator in HPASMCs

induced by hypoxia and demonstrated the potential of gene therapy and drug development for treating PAH.

Introduction

Pulmonary arterial hypertension (PAH) is an intractable cardiovascular disease, a key feature of which is pulmonary vascular remodeling (PVR) (1). Hypoxia is considered to be the predominant factor in the pathogenesis of PAH (2,3). During the hypoxic exposure, the dysfunction of pulmonary artery smooth muscle cells (PASMCs), including both proliferation and migration, is the major reason for medial hypertrophy in PVR (4). To limit morbidity and mortality, attention has been focused on identifying the cellular and molecular mechanisms underlying aberrant proliferation and migration in human (H) PASMCs. However, the mechanism of PASMCs proliferation and migration at the molecular level is still not very clear, and further research is needed.

Long non-coding RNAs (lncRNAs), with transcripts >200 nucleotides in length, were once considered as irrelevant transcriptional 'noise' without biological function. Previous studies, however, have shown that lncRNAs play a significant role in diverse biological and disease processes (5). lncRNAs can be used as a 'scaffold' to bind a variety of proteins to exert biological functions (6). Also, lncRNAs can form an anchor point to regulate different bioactivities by collecting or sequestering certain protein factors and participating in the synthesis and reconstruction of nucleic acid sequences (7).

On chromosome 9p21, there is a lncRNA named antisense noncoding RNA in the INK4 locus (ANRIL) with ~126,000 bps (8). lncRNA ANRIL was first identified and named in the investigation of a melanoma-neural system tumor syndrome family (9). A previous study identified that ANRIL is associated with ~40% of all tumor types (10). Moreover, ANRIL also plays an important role in the development and progression of various diseases to a certain degree, including coronary heart disease (11,12). Knocking down the expression of ANRIL in human aortic vascular smooth muscle cells, the results obtained suggested that ANRIL splicing variants played a role in coordinating tissue remodeling (13). Based on the above evidence, it is possible to hypothesize that ANRIL may regulate the process of PAH and that this deserves further investigation. Based on the potential roles of ANRIL in maintaining cellular functions, the present study speculated

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that ANRIL participates in hypoxia-induced proliferation and migration of HPASMCs in PAH. To this end, the expression of ANRIL was studied in HPASMCs where PAH had been induced by hypoxia.

Materials and methods

Materials. Antibodies against proliferating cell nuclear antigen (PCNA), Cyclin A, Cyclin D and Cyclin E were purchased from Boster Biological Technology. Antibody against Ki67 was purchased from Abcam. Bromodeoxyuridine (BrdU) proliferation assay kit was purchased from EMD Millipore. DMEM was purchased from Hyclone; GE Healthcare Life Sciences. The remaining chemical reagents were domestic analytical pure and biochemical reagents.

Cell culture. HPASMCs were obtained from the Center Laboratory of Harbin Medical University Daqing Campus. The cells were secondary cultured in 20% Clark serum (CLARK Bioscience)-DMEM at 37°C in a 5% CO₂ humidified incubator. HPASMCs were cultured with a gas mixture containing 92% N₂, 5% CO₂ and 3% O₂ to create hypoxic conditions.

Small interfering (si)RNA and transfection. The sequence of siRNA against lncRNA ANRIL and scrambled negative control siRNA (si-NC) were synthesized by Shanghai GenePharma Co., Ltd. The target sequences of these siRNAs were follows: Si-lncRNA ANRIL, 5'-GCCCCAAGCAUUAUGAUAATT UUGAUCUAUAUGCUUGGGCTT-3' and si-NC, 5'-UUC UCCGAACGUGUCACGUTTACGUGACACGUUCGGAGA ATT-3'. HPASMCs cultured on six-well plates were transfected with the si-lncRNA ANRIL or si-NC (3.75 µl per 1.1 ml DMEM) using X-tremeGENE (Sigma-Aldrich; Merck KGaA) (14). A total of 24 h post transfection, cells were harvested for quantitative (q)PCR and western blotting analyses.

Revers transcription-qPCR. Total RNA was extracted and isolated from HPASMCs using TRIzol reagent (Invitrogen; Thermo Fisher Scientific, Inc.). For the qPCR assay, the total RNA was reverse transcribed (15 min at 37°C, 5 min at 50°C, 5 min at 98°C and holding at 4°C) into cDNA using One-step cDNA First Strand Reverse Transcription kit (HaiGene) according to the manufacturer's protocol. The expression levels were normalized against the expression of GAPDH. The PCR primers were as follows: ANRIL forward, 5'-TCC TGCCACTTCCTCTACTGA-3' and reverse, 5'-TGGTAT GGAAGGTGCTATGGA-3'; GAPDH forward, 5'-CAATGA CCCCTTCATTGACC-3' and reverse, 5'-TGGAAGATGGTG ATGGGATT-3'. The amplification reaction using SYBR® Green Realtime PCR Master Mix was performed for 1 min at 95°C, 15 min at 95°C, followed by 40 cycles at 95°C for 15 sec and 52°C for 15 sec. The relative quantification of target genes expression was relative to the GAPDH levels. The 2^{-ΔΔCq} method was used to calculate the expression of lncRNA ANRIL (15). All qPCR reactions were performed in duplicate.

Western blotting. Proteins were solubilized and extracted with lysis buffer (Tris 50 mM, pH 7.4, NaCl 150 mM, Triton X-100 1%, EDTA 1 mM, and PMSF 2 mM) and incubated for 30 min on ice. The lysates were sonicated for 30 sec at 25 kHz in an

ice bath and centrifuged at 15,000 x g for 15 min at 4°C and the insoluble fractions were discarded. Protein concentrations were determined by the Bradford assay using bovine serum albumin (Sigma-Aldrich; Merck KGaA) as standard. Cell samples containing 20 µg of protein were subjected to electrophoresis on a 10% SDS-polyacrylamide gel. Following electrophoresis, proteins were transferred to nitrocellulose membranes. These membranes were blocked at room temperature for 1 h in blocking buffer (Tris 20 mM, pH 7.6, NaCl 150 mM and Tween 20 0.1%) containing 5% nonfat dry milk and incubated with PCNA (Boster Biological Technology; cat. no. BM3888; anti-PCNA antibody; 1:400), cyclin A (Boster Biological Technology; cat. no. BM4673; anti-Cyclin A2 antibody; 1:200), cyclin D (Boster Biological Technology; cat. no. BM4272; anti-Cyclin D1 antibody; 1:200), cyclin E (Boster Biological Technology; cat. no. BM4658; anti-Cyclin E2 antibody; 1:100), β-actin (Boster Biological Technology; cat. no. BM0627; anti-β-actin antibody; 1:2,000) at 4°C overnight. Secondary antibodies [Beyotime institute of Biotechnology; cat. no. A0208; horseradish peroxidase (HRP)-labeled Goat Anti-Rabbit IgG(H+L); 1:10,000; cat. no. A0216; HRP-labeled Goat Anti-Mouse IgG(H+L); 1:12,000] were prepared according to the corresponding proportion and species, and incubated at room temperature for 1 h. The proteins were visualized with enhanced chemiluminescence reagents (Super Signal; Pierce; Thermo Fisher Scientific, Inc.). Grayscale values were calculated using Quantity One v4.6.2 (Bio-Rad Laboratories, Inc.) and statistical data were obtained using Microsoft Excel and Origin 7.5 (OriginLab).

Immunofluorescence assay. The pre-configured primary antibody against Ki67 (Santa Cruz Biotechnology, Inc.; cat. no. sc-23900, Ki-67 Antibody; 1:50) was added to the cells of different components and incubated at 4°C overnight. Then the cells were incubated with the secondary antibody [Invitrogen; Thermo Fisher Scientific, Inc.; cat. no. R-6393; Goat anti-Mouse IgG (H+L)] (1:100) conjugated with rhodamine for 1 h in the dark at 37°C and washed with PBS. The nuclei were stained with 4,6-diamidino-2-phenylindole (DAPI) at room temperature for 5 min. The slides were examined using NA1.4 inverted fluorescence microscope (Leica DMI6000; Leica Microsystems, Inc.), images were visualized by Hamamatsu ORCA-R2 camera (Hamamatsu Photonics K.K.) and recorded by LAS AF 2.6.0 software (Leica Microsystems, Inc.). The experiments were conducted in triplicate.

Scratch-wound assay. The confluent HPASMCs cultured in 6-well plates were wounded by pipette tips and the detached cells were washed out by PBS (16). After that, the cells were treated with vehicle or the chemicals of interest in 5% FBS-DMEM. The cells in all study groups were imaged in the same area of the culture plate immediately after 24 h after the wound was inflicted.

Transwell assay. Cell migration was measured using a Matrigel®-coated modified Boyden chamber with a polycarbonate filter with a pore size of 8 µm. A total of 5x10⁴ HPASMCs was added to each upper chamber in a serum-free medium. The lower chambers contained 20% Clark serum (CLARK Bioscience)-DMEM. After 24 h of incubation at 37°C

in a 5% CO₂ humidified incubator, the nonmigrating cells in the upper chamber removed. The cells on the underside of the membrane were incubated in 4% formaldehyde solution for 10 min at room temperature, followed by incubation in 0.4% Crystal Violet for 5 min at room temperature. The number of migrated cells was measured by counting the number of stained nuclei per high power field using a light microscope (Olympus Corporation). Each sample was counted randomly at nine separate locations in the center of the membrane and HPASMCs migration activity was reported as the number of cells migrating per field of view. The experiments were performed at least three times in quadruplicate.

BrdU incorporation. HPASMCs were plated in 96-well plates at a density of 1×10^4 cells/well and then transfected by siRNA after growth arrest for 24 h. BrdU was added to a final concentration of 30 μ M and incubated for 30-60 min. BrdU was removed and washed 3 times with PBS. 4% paraformaldehyde was used to fix cells at room temperature for 30 min. Then cells were washed three times with PBS. The following steps were done according to the manufacturer's protocol. Finally, the plate was read using a spectrophotometer microplate reader at dual wavelengths of 450/550 nm.

3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay. After the model was established, the cells were drained of the original culture solution and 90 μ M culture solution and 10 μ M MTT (5 mg·ml⁻¹) were added to each well (17). The cultures were incubated at 37°C for 4 h, then drained of liquid and 150 μ l DMSO was added to each well. The absorbance was read at 490 nm using a spectrophotometer, to determine the cell viability.

Cell cycle analysis. The proportions of cells in the G0/G1, S and G2/M phases were detected using a flow cytometer. The cell cycle analysis kit (Beyotime institute of Biotechnology) was used. The cells were digested by trypsin then centrifuged at 1,000 x g for 5 min at room temperature to remove the supernatant. A total of 1 ml ice bath precooled PBS buffer was added to suspend the cells and centrifuged again (1 000 x g, 5 min, room temperature). A total of 1 ml 70% ethanol precooled by ice bath was added and mixed. Cells were fixed at 4°C for 2 h or longer then centrifuged at 1,000 x g for 3-5 min at room temperature. A total of 1 ml ice bath precooled PBS buffer was added then the mixture was centrifuged again (1,000 x g, 5 min, room temperature) and discard the supernatant. The prepared staining solution was added according to the protocol of the kit and the cell precipitation was resuspended. The cells were then placed in a temperature bath at 37°C for half an hour. Finally, the cells were filtered once through 400-mesh sieves and detected by flow cytometry (CytoFLEX, Beckman Coulter, Inc.). The results were analyzed by CytExpert 2.0 (Beckman Coulter, Inc.).

Statistical analysis. Each experiment was repeated ≥ 3 times. The composite data were expressed as the mean \pm standard error of the mean. Statistical analysis was performed with analysis of variance followed by Dunnett's test or Student's t-test or Pearson correlation test. Quantity One-4.6.2 (Bio-Rad Laboratories, Inc.), Microsoft Office Excel 2007 (Microsoft

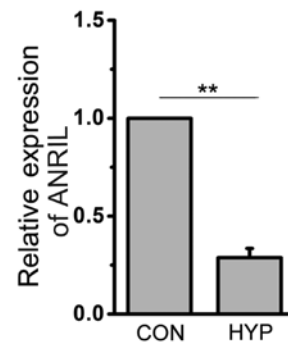


Figure 1. Quantitative PCR analysis of ANRIL expression in HPASMCs. **P<0.01 (n=6). CON, normoxia control; HYP, hypoxia; HPASMCs, human pulmonary artery smooth muscle cells; ANRIL, antisense noncoding RNA in the INK4 locus.

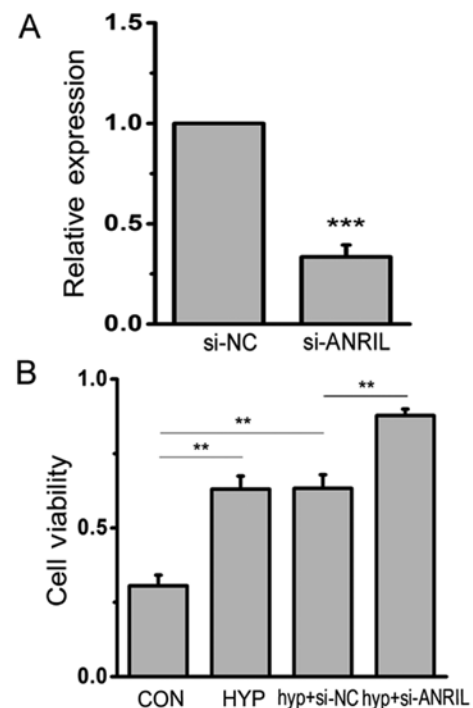


Figure 2. Downregulated ANRIL promotes the cell viability of HPASMCs *in vitro*. (A) ANRIL expression in HPASMCs following treatment with si-NC or si-ANRIL (n=5). The experimental groups were established as follows: si-NC, cells treated with negative control; si-ANRIL, cells treated with si-ANRIL. (B) The MTT assay was used to detect the cell viability (n=3). **P<0.01 and ***P<0.001. CON, normoxia control; HYP, hypoxia; hyp+si-NC, cells treated with the negative control under hypoxic conditions; hyp+si-ANRIL, cells treated with si-ANRIL under hypoxic conditions; HPASMCs, human pulmonary artery smooth muscle cells; si-ANRIL, small interfering RNA against antisense noncoding RNA in the INK4 locus.

Corporation), Origin 7.5 (OriginLab) and IPP 6.0 (Media Cybernetics, Inc.) software were used to analyze data. P<0.05 was considered to indicate a statistically significant difference.

Results

ANRIL expression is downregulated in hypoxic HPASMCs. In order to prove that lncRNA ANRIL was related to hypoxic PAH, the expression of ANRIL in HPASMCs exposed to both normal and hypoxic conditions was detected by qPCR. The

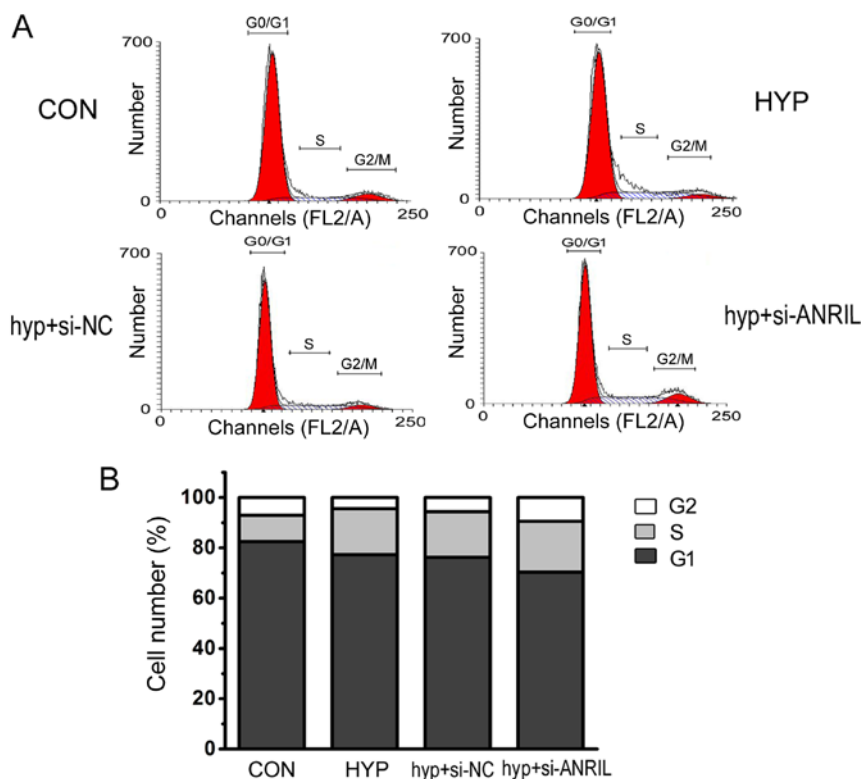


Figure 3. si-ANRIL induces cell cycle progression. ANRIL inhibited HPASMCs cycle progression and decreased the percentage of cells in the G2/M+S phase. (A) Hypoxia increased the cell number in the G2/M+S phases compared with normoxia, which was strengthened by si-ANRIL, as determined by flow cytometry analysis. (B) The percentage of cells in the S and G2/M phases in HPASMCs is shown. CON, normoxia control; HYP, hypoxia; hyp+si-NC, cells treated with negative control under hypoxic conditions; hyp+si-ANRIL, cells treated with si-ANRIL under hypoxic conditions (n=3); HPASMCs, human pulmonary artery smooth muscle cells; ANRIL, antisense noncoding RNA in the INK4 locus; si, small interfering.

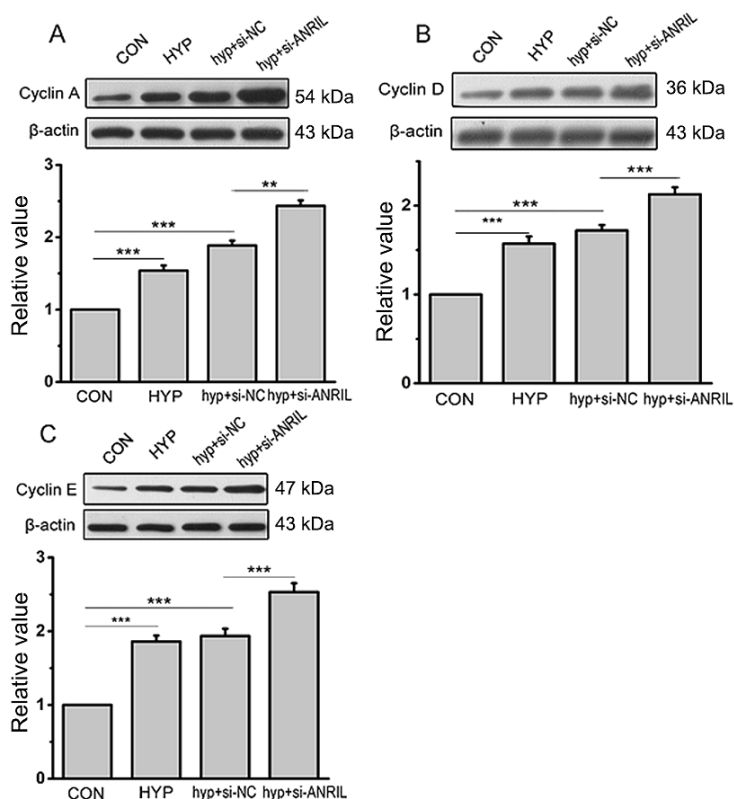


Figure 4. ANRIL regulates the expression levels of Cyclin A, Cyclin D and Cyclin E in HPASMCs under hypoxic conditions. (A) Cyclin A, (B) Cyclin D and (C) Cyclin E were analyzed by western blotting in different groups. * $P < 0.01$ and *** $P < 0.001$ (n=3). CON, normoxia control; HYP, hypoxia; hyp+si-NC, cells treated with negative control under hypoxic conditions; hyp+si-ANRIL, cells treated with si-ANRIL under hypoxic conditions; HPASMCs, human pulmonary artery smooth muscle cells.

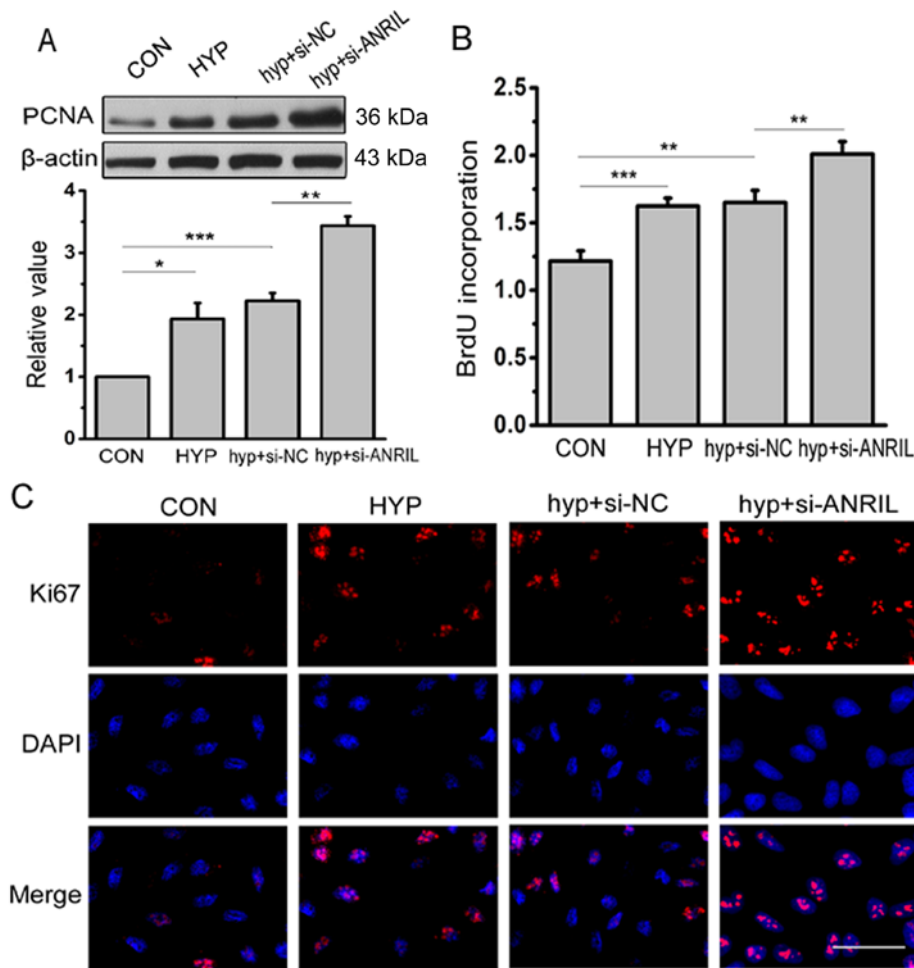


Figure 5. ANRIL induces higher levels of proliferation in HPASMCs. (A) The protein level of PCNA was analyzed by western blotting after transfection with si-ANRIL; densitometric quantification of protein bands showed a significant increase in PCNA following 24 h of transfection with si-ANRIL (n=3). (B) 5-bromodeoxyuridine incorporation assay was performed to determine the proliferation of HPASMCs (n=5). (C) Immunofluorescence assay was used to detect the expression of Ki67. The red color denotes Ki67 staining by rhodamine, whereas the blue color denotes nucleus staining by DAPI (n=3). Scale bar=50 μ m. *P<0.05, **P<0.01 and ***P<0.001. CON, normoxia control; HYP, hypoxia; hyp+si-NC, cells treated with negative control under hypoxic conditions; hyp+si-ANRIL, cells treated with si-ANRIL under hypoxic conditions; HPASMCs, human pulmonary artery smooth muscle cells; PCNA, proliferating cell nuclear antigen; ANRIL, antisense noncoding RNA in the INK4 locus; DAPI, 4,6-diamidino-2-phenylindole.

results revealed that the expression of ANRIL in hypoxic HPASMCs was significantly downregulated compared with that in normoxic cells (P<0.01; Fig. 1).

ANRIL regulates the viability of HPASMCs. To determine the biological function of ANRIL in HPASMCs under hypoxic conditions, siRNA targeting lncRNA ANRIL were designed to inhibit the expression. The efficiency and specificity of siRNA transfection and the downregulation of endogenous lncRNA ANRIL were confirmed by qPCR (Fig. 2A). Meanwhile, to explore the effects of ANRIL on cell survival ability, the viability of HPASMCs was examined by MTT assay. The result revealed the significantly increased viability of hypoxic HPASMCs compared with those under conditions of normoxia, which was strengthened by si-ANRIL under hypoxic conditions (P<0.05; Fig. 2B).

ANRIL regulates the cell cycle progression of HPASMCs. In order to investigate whether hypoxia affected cell cycle progression through the lncRNA ANRIL pathway, flow cytometry was used to detect the number of cells at different

cell cycle phases. The results showed that hypoxia increased the percentage of cells in the G2/M+S phase and the downregulated ANRIL strengthened the increase in the percentage of cells in the G2/M+S phase under hypoxic conditions (Fig. 3).

In addition, cyclins, proteins that regulate the cell cycle and play important roles in both the S phase and G2/M phase of cell cycle, have been generally considered as markers of cellular proliferation. In order to understand the molecular mechanism by which ANRIL regulates the cell cycle, the expression of cell cycle-associated proteins, Cyclin A, Cyclin D and Cyclin E, were detected by western blotting. The results revealed that the expression of cyclins in hypoxic HPASMCs was significantly increased compared with normoxia, which was further promoted by ANRIL siRNA (P<0.001; Fig. 4).

ANRIL affects the proliferation of HPASMCs. Subsequently, the effect of ANRIL on HPASMC proliferation under hypoxic conditions was further investigated. The results of western blotting displayed that the expression of PCNA was significantly elevated under hypoxic conditions contrasted with normoxia and si-ANRIL increased the trend further

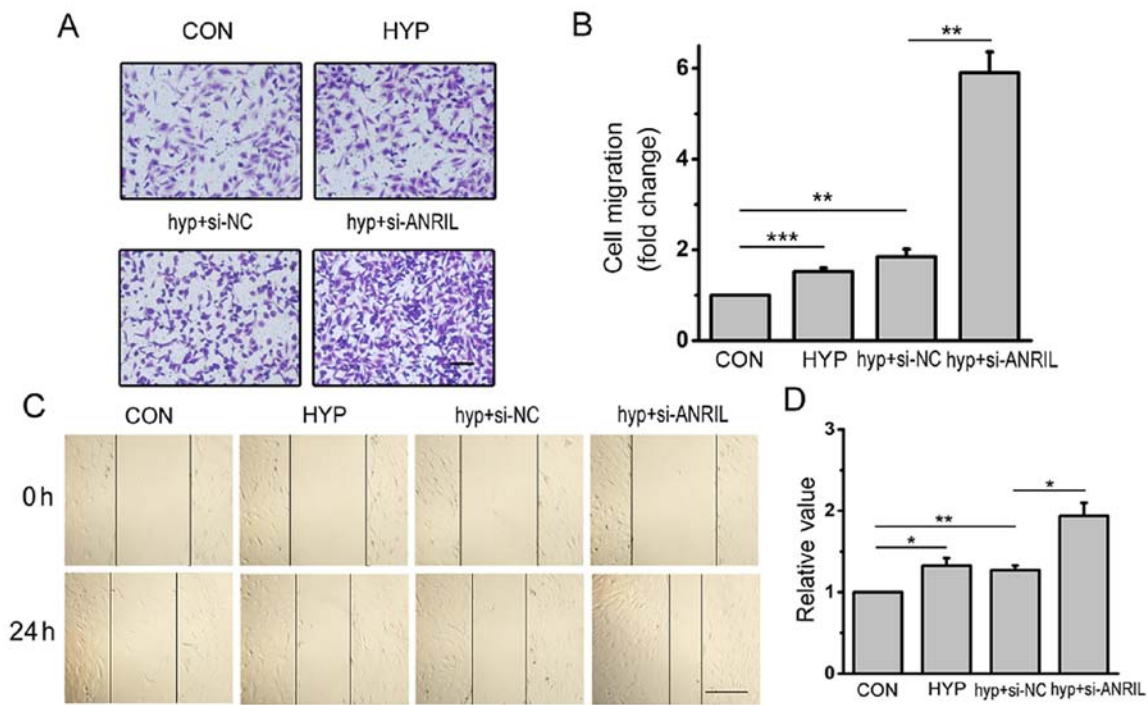


Figure 6. ANRIL induces the migration of HPASMCs under hypoxic conditions. (A) HPASMCs were subjected to a Transwell migration assay. (B) Quantification of the data from the Transwell assay. (C) Scratch-wound assay of HPASMCs after treatment with si-NC or si-lncRNA ANRIL; (D) Quantification of the data from the scratch-wound assay. Scale bar=100 μ m (n=3). * P <0.05, ** P <0.01 and *** P <0.001. CON, normoxia control; HYP, hypoxia; hyp+si-NC, cells treated with negative control under hypoxic conditions; hyp+si-ANRIL, cells treated with si-ANRIL under hypoxic conditions; HPASMCs, human pulmonary artery smooth muscle cells; ANRIL, antisense noncoding RNA in the INK4 locus.

(P <0.05; Fig. 5A). To evaluate the number of cells that were actively synthesizing DNA, the BrdU incorporation assay was carried out. The present results revealed that the ratio of proliferating cells in HPASMCs under hypoxic conditions was significantly increased compared with under normoxic conditions (P <0.001), which was enhanced by si-ANRIL (Fig. 5B). The Ki67 protein is a cell proliferation marker that is closely related to cell proliferation and the cell cycle. The result of immunofluorescence assay showed that the expression of Ki67 increased in hypoxia compared with under normal conditions. When the expression of ANRIL was downregulated by siRNA, Ki67 fluorescence staining was found to be increased (Fig. 5C). Consistently, these results demonstrated that the proportion of proliferating cells with a low ANRIL content was higher.

ANRIL promotes migration of hypoxic HPASMCs. The effects of ANRIL on cell migration was examined by Transwell migration assay and scratch-wound assay. As shown in Fig. 6A, the number of migrated cells under hypoxic conditions was more than that under normoxia. Downregulated ANRIL significantly increased the cell migration according to the Transwell migration assay (P <0.01; Fig. 6B). The scratch-wound assay results showed that the migration distance of hypoxic HPASMCs was longer than that in normoxia, which was strengthened by ANRIL siRNA under hypoxic condition in Fig. 6C. Histograms showing the results of the Transwell migration and scratch-wound assays are shown Fig. 6B and D, respectively. Taken together, these data indicated that the downregulation of ANRIL induced by hypoxia enhanced the capacity of migration in HPASMCs.

Discussion

Previous studies have indicated that, in numerous diseases, lncRNA ANRIL regulates cell proliferation, migration and cell cycle progression including cardiovascular diseases, tumors and diabetes (18,19). In this study, it was found that the expression of ANRIL was downregulated in hypoxic HPASMCs. Hypoxia accelerated the cell cycle progression and induced cell proliferation of HPASMCs through the downregulation of the lncRNA ANRIL pathway. Moreover, downregulated ANRIL in the hypoxic HPASMCs promoted cell migration. Therefore, solid evidence demonstrated that ANRIL is a critical regulator in HPASMCs induced by hypoxia.

It is well established that hypoxia is the main cause of PAH. Recent reports have highlighted the relationship between lncRNAs and hypoxia in PAH. The downregulated lncRNA, maternally expressed 3, in hypoxic PSMCs was shown to trigger cell proliferation and migration via the p53 signaling pathway (1). The lncRNA Hoxa cluster antisense RNA 3 (Hoxaas3) was upregulated in both pulmonary vessels in hypoxic mice and PSMCs under hypoxic conditions, while a high expression level of Hoxaas3 was associated with cell proliferation and modulated cell cycle distribution by upregulating Homeobox a3 at the mRNA and protein levels (20). The results from the present study showed that the decrease in the expression level of ANRIL when downregulated in hypoxic HPASMCs was twice that which occurred under normoxic conditions by qPCR, indicating that ANRIL was able to regulate hypoxic PAH.

The proliferation of PSMCs is one of the main causes of medial hypertrophy of PVR in PAH (4). A previous study

showed that in epithelial ovarian cancer, ANRIL promoted proliferation and cell cycle progression (21). The present study demonstrated that si-ANRIL may accelerate cell cycle progression by increasing the percentage of cells in the G2/M+S phase and also elevated the expression level of cyclins under hypoxic conditions. Downregulated ANRIL promoted the expression of PCNA in HPASMCs and increased the expression of Ki-67 under hypoxia. The above results indicated that ANRIL could regulate the cell cycle progression and the proliferation of HPASMCs.

A recently published study reported that the expression of ANRIL in the pancreatic cancer cells was higher than that in normal pancreatic duct epithelial cells and after ANRIL is silenced, the migratory and invasive abilities of the cells were decreased (22). In the present study, Transwell and scratch-wound assays were used to examine the effects of ANRIL on cell migration. The results showed that downregulated ANRIL increased the migration of hypoxic HPASMCs. These data indicated that the downregulation of ANRIL could enhance the capacity of migration in HPASMCs.

At present, the importance of cell biology and the role of ANRIL in the pathogenesis of various diseases have been major topics for study. To the best of our knowledge, this is the first study that has reported on the involvement of ANRIL in PAH, however, there remain several limitations that need to be resolved in future studies. Although the downregulation of ANRIL was detected in HPASMCs, it is still unclear how the decrease of ANRIL is caused by hypoxia. In addition, it is worth noting that one lncRNA may target multiple microRNAs and exert different effects. Whether ANRIL can effectively function as a competing endogenous RNA by binding with microRNAs remains to be further studied. Further experiments will be performed in the future to explore the role of ANRIL in PAH.

In conclusion, the present study has demonstrated that the expression of lncRNA ANRIL in HPASMCs of PAH was reduced. The decreased expression of lncRNA ANRIL contributed to the proliferation and migration of HPASMCs in the pathogenesis of PAH. Therefore, the authors' proposal is that ANRIL-mediated PSMCs are involved in the pathogenesis of vascular remodeling in PAH. These findings provide a novel potential target for the treatment of PAH and may help to improve the therapeutic efforts in the future.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

SW, CZ and XZ made substantial contributions towards the conception and design of the experiments. SW completed all the studies and aggregated the figures and discussed the results. SW and CZ drafted the manuscript and critically revised and added important intellectual content.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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