

MicroRNA-214 promotes the EMT process in melanoma by downregulating CADM1 expression

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Abstract. Melanoma is a malignant skin cancer type associated with a high mortality rate, but its treatment is currently not ideal. Both microRNA (miR)-214 and cell adhesion molecule 1 (CADM1) are differentially expressed in melanoma, but their role in this cancer type remains unknown. Therefore, the aim of the present study was to investigate the role of CADM1 and miR-214 in melanoma to identify novel targets for its treatment. The expression levels of CADM1 and miR-214 in cells were detected by reverse transcription-quantitative PCR (RT-qPCR). Moreover, cell viability, migration and invasion were measured by MTT, wound healing and Transwell assays, respectively. In addition, the relative expression levels of epithelial-mesenchymal transition (EMT)-related proteins in cells were detected by RT-qPCR and western blotting. It was found that the expression of CADM1 was inhibited in melanoma cells, while miR-214 expression was increased during melanoma tumorigenesis. Furthermore, miR-214 mimics promoted the viability, migration and invasion of melanoma cells. It was also demonstrated that the downregulation of CADM1 reversed the inhibitory effect of the miR-214 inhibitor in melanoma. Moreover, overexpression of CADM1 inhibited the EMT process in melanoma, while the miR-214 inhibitor suppressed the EMT process. The results also indicated that miR-214 promoted the EMT process by downregulating CADM1, which may represent a novel mechanism for the progression of melanoma.

Introduction

Melanoma is the second most dangerous malignant tumor type and ranks second only to pancreatic cancer (1). Moreover, melanoma is characterized by early metastasis

and high mortality rates. In recent years, the incidence of numerous malignant tumors has decreased from 40 to 20% worldwide, but the incidence of malignant melanoma remains high (2). Epithelial-mesenchymal transition (EMT) has a critical role in the process of cell remodeling during tumorigenesis (3). Furthermore, previous studies have reported that EMT plays a key role in the metastasis of malignant tumors (4-6). However, as the metastasis of melanoma has not been effectively controlled (7), it is important to identify the mechanism via which the EMT process is promoted.

Cell adhesion molecule 1 (CADM1), which is regarded as a suppressor gene in the tumorigenesis of non-small cell lung cancer, has been identified as a diagnostic marker for adult T-cell leukemia and lymphoma (8). In addition, it has been previously reported that CADM1 plays an inhibitory role in the progression, migration and invasion of cancer, such as cervical cancer and liver cancer (9-11), indicating that CADM1 is an inhibitor of tumor cell malignant features. You *et al* (12) showed that CADM1 was differentially expressed in melanoma, but its function in melanoma remains unknown. Moreover, microRNAs (miRNAs) act as imperfect sequence guides to recruit a ribonucleoprotein complex to the complementary RNA (13). Furthermore, miRNA expression profiles differ between different stages of cancer and healthy tissues, and previous studies have used miRNAs as diagnostic markers, either alone or in combination with other known biomarkers (14-16). Preliminary studies examining miRNA expression also used tissues to determine the functional and diagnostic roles of miRNAs (13,17,18), which have been reported to regulate the development and progression of cancer (19,20). It has also been revealed that miRNAs (miRs) can promote or inhibit the EMT process in cancer types (21-23). For instance, miR-214 is a member of the miRNA family, and it can induce immune suppression and promote the occurrence of cancer (24), thus demonstrating that miR-214 plays a key role in tumorigenesis. In addition, Mirzaei *et al* (25) revealed that miR-214 is differentially expressed in melanoma. However, the relationship between CADM1 and miR-214 is not fully understood.

Based on these previous findings, the aims of the present study were to investigate the mechanism via which miR-214 regulates the EMT process in melanoma, and to identify the relationship between CADM1 and miR-214. Thus, the

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present results may provide a potential novel target for treating melanoma.

Materials and methods

Cell lines and cell culture. The PIG-1, WM-266-4, A2058 and A375 cell lines were purchased from the Cell Bank of Chinese Academy of Science, and cultured in 90% RPMI-1640 (Thermo Fisher Scientific, Inc.) with 10% FBS (Thermo Fisher Scientific, Inc.) and maintained at 37°C in a humidified incubator containing 5% CO₂. The BR-V-108 plasmid vector (Shanghai Biosciences Co., Ltd.) and competent TOP10 *E. coli* cells (Shanghai Biosciences Co., Ltd.) were used.

Vector construction and cell transfection. RNA interference target sequences were designed using the *CADM1* gene as a template to construct a target gene RNA interference 5'-CCA CAGGACAATGCTGAACCTT-3' lentiviral vector (Shanghai Biosciences Co., Ltd.). A single-stranded DNA oligo containing the interference sequence was synthesized, and the obtained lyophilized powder was dissolved in an annealing buffer (final concentration, 100 M; Beyotime Institute of Biotechnology) and incubated in a water bath at 90°C for 15 min. After naturally cooling to room temperature, a double-stranded oligo with overhang ends was formed, which was then directly ligated into the digested lentiviral vector through restriction sites at both ends. A 50 µl reaction system was prepared according to the NEB (New England Biolabs, Inc.) instructions, the BR-V-108 vector was linearized by double digestion and the ligation product was introduced into prepared TOP10 *E. coli* (Tiangen Biotech Co., Ltd.) competent cells. The sequencing results were compared with those of the correct clones and the plasmid was isolated.

miR-214 mimics (upregulation of miR-214, a chemically modified single-stranded RNA that can mimic and enhance the function of endogenous miR-214) and miR-214 inhibitor (down-regulation of miR-214, a chemically modified single-stranded RNA that can specifically target miR-214 in melanoma cells), as well as their corresponding negative controls (NCs), were designed by Shanghai GenePharma Co., Ltd. For cell transfection, melanoma cells (1.5x10⁵/well) in a 6-well plate were transfected with 200 pmol *CADM1* short hairpin RNA (shRNA), or 100 pmol miR-214 mimics, inhibitor or their NCs using Lipofectamine® 2000 reagent (Thermo Fisher Scientific, Inc.) for 48 h according to the manufacturer's instructions. Transfected cells were incubated for an additional 24 h at 4°C and then used to test the cell viability. The sequences were as follows: miR-214 mimics, 5'-UGCCUGUCUACACUUGCUGUGC-3'; miR-214 inhibitor, 5'-ACUGCCUGUCUGGCCUGCUGU-3'; *CADM1* siRNA sense, 5'-CACCGCAGATGACTTATCCTCTAC AATCAAGAGATTGTAGAGGATAAGTCATCTGTTTTT TG-3' and anti-sense, 5'-GATCCAAAAACAGATGACTT ATCCTCTACAATCTCTTGAATTGTAGAGGATAAGTC ATCTGC-3'.

Reverse transcription-quantitative PCR (RT-qPCR). Total RNA was extracted from melanoma cell lines using TRIzol® reagent (Thermo Fisher Scientific, Inc.) according to the manufacturer's protocol. First-strand cDNA was synthesized using the PrimeScript RT reagent kit (Takara Bio, Inc.) according

to the manufacturer's protocol. RT-qPCR was performed in an ABI7500 system (Thermo Fisher Scientific, Inc.) using SYBR-Green (Takara Bio, Inc.) methods. RT-qPCR was performed in triplicate using the following protocol: Initial denaturation for 2 min at 94°C, followed by 35 cycles for 30 sec at 94°C and 45 sec at 55°C. The primers for all the genes were obtained from Shanghai GenePharma Co., Ltd. Relative gene expression was quantified using the 2^{-ΔΔC_q} method (26). The primer sequences are listed in Table I.

Mature miRNAs were quantified using miRNA assays. The protocol was as following: cDNA was synthesized from 5-10 µg of total RNA using the microRNA cDNA Synthesis kit (Takara Bio, Inc.). Then, PCR assays using were performed using SYBR premix Ex Taq II kit (Takara Bio, Inc.). RT-qPCR conditions used were as follows: Initial denaturation for 10 min at 95°C, followed by 40 cycles of 15 sec at 95°C and 1 min at 60°C. Relative miRNA and mRNA expression levels were measured and normalized to U6 and GAPDH expression, respectively.

Western blot analysis. To investigate EMT-related protein expression levels, A375 or A2058 cells were collected and lysed using RIPA lysis buffer (Cell Signaling Technology, Inc.) on ice according to the manufacturer's instructions. Total cellular proteins (40 µg/lane) were quantified by bicinchoninic protein assay kit (Beyotime Institute of Biotechnology), and then subjected to 10% SDS-PAGE for western blotting. Proteins were transferred to PVDF membranes, the blots were incubated at room temperature with 5% BSA (Thermo Fisher Scientific, Inc.) in TBST (0.5% Tween-20) for 60 min and then incubated overnight at 4°C on a rocker with the following primary antibodies: Anti-CADM1 (1:1,000; cat. no. ab3910; Abcam), anti-E-cadherin (1:3,000; cat. no. ab194982; Abcam), anti-vimentin (1:3,000; cat. no. ab92547; Abcam), anti-Slug (1:1,000; cat. no. ab51772; Abcam) and anti-GAPDH (1:3,000; cat. no. ab181602; Bioworld Technology, Inc.). After washing three times with TBST for 5 min, the membranes were incubated with horseradish peroxidase-conjugated goat anti-rabbit IgG polyclonal secondary antibody (1:3,000; ab191866; Beyotime Institute of Biotechnology) at room temperature for 1 h. An ECL⁺ western blotting system kit (GE Healthcare Life Science) was used for color development. The density of the bands was measured using ImageJ software (version 6.0; National Institutes of Health). Signals were detected with enhanced chemiluminescence using GAPDH as the internal standard (Kodak).

Dual-luciferase reporter assay. The downstream target of *CADM1* was predicted using Starbase 2.0 (Sun Yat-sen University; <http://starbase.sysu.edu.cn/>). Both wild-type and mutant constructs of *CADM1* were cloned into the pmirGLO Dual-Luciferase miRNA target expression vector (Promega Corporation). A375 or A2058 cells (5x10³ cells per well) were seeded in a 24-well plate and co-transfected with wild-type or mutant *CADM1* 3'untranslated region and mimic NC or miR-214 mimics using Lipofectamine® 3000 (Thermo Fisher Scientific, Inc.). Transfection and harvest efficiencies were controlled using the pmirGLO reporter as an internal control. Cells were collected 48 h post-transfection and analyzed using the dual-luciferase reporter assay system (Promega

Table I. Primer sequences used in the present study.

Gene	Sequence
miR-214	F: 5'-AGCACAGCAGGCACAGACA-3' R: 5'-CAGTGCCTGTCGTGGAGT-3'
U6	F: 5'-CGCTTCGGCAGCACACATATAC-3' R: 5'-CAGGGGCCATGCTAATCTT-3'
CADM1	F: 5'-GGTGATGGGCAGAATCTGTT-3' R: 5'-AGGCCTGAAGTCCCTGAAAT-3'
E-cadherin	F: 5'-TCACATCCTACACTGCCAG-3' R: 5'-AGTGTCCCTGTTCCAGTAGC-3'
Vimentin	F: 5'-GGACCAGCTAACCAACGACA-3' R: 5'-AAGGTCAAGACGTGCCAGAG-3'
Slug	F: 5'-ACGCCTCCAAAAAGCCAAAC-3' R: 5'-ACAGTGATGGGGCTGTATGC-3'
GAPDH	F: 5'-CCACCCATGGCAAATTCCATGGCA-3' R: 5'-TCTAGACGGCAGGTCAGGTCCAC-3'

F, forward; R, reverse; miR, microRNA; CADM1, cell adhesion molecule 1.

Corporation). The data were quantified via normalizing to *Renilla* luciferase activity.

MTT assay. An MTT assay was performed to investigate the effect of miR-214 and CADM1 on melanoma cell viability. A375 and A2058 cells (5×10^3 cells per well) were seeded on 96-well plates and incubated at 37°C for 24, 48 and 72 h. Then, 20 μ l MTT solution (Sigma-Aldrich; Merck KGaA) was added to each well and incubated at room temperature for another 4 h. Then, the culture medium with MTT was discarded, 150 μ l DMSO was added and cells were agitated for 10 min at room temperature. The absorbance was measured at 490 nm with a standard microplate reader, and cell viability was analyzed.

Wound healing assay. A375 and A2058 cells (5×10^4 cells per well) were plated in a 6-well plate for culture. When cells were at 90% confluency, the medium was replaced with RPMI-1640 (Thermo Fisher Scientific, Inc.) for the scratch test, and the slide was pushed up to form a scratch. Then, 0.5% FBS was added and cells were imaged using a light microscope (magnification, x100). Cells were incubated at 37°C in a 5% CO₂ incubator for 48 h and then assessed using ImageJ software (version 6.0; National Institutes of Health). The cell migration rate of each group was calculated based on the images acquired after the scratch.

Transwell assay. For the invasion assay, the upper chamber (polycarbonic membrane; diameter 6.5 mm; pore size, 8 μ m; Corning, Inc.) was pretreated with 100 μ l Matrigel (BD Biosciences) at 37°C for 30 min, and melanoma cells (1×10^5 cells) in FBS-free medium (RPMI-1640) were seeded into the upper chamber. The lower chamber contained RPMI-1640 supplemented with 10% FBS. Then, cells were incubated at 37°C for 24 h. The cells that attached to the

underside of the membrane were fixed using 5% glutaraldehyde at 4°C for 30 min and stained with a 0.5% crystal violet solution at room temperature for 30 min. Subsequently, images were captured and the number of invading cells was counted under a light microscope (magnification, x100).

Statistical analysis. All assays were performed in ≥ 3 independent experiments, and data are presented as the mean \pm SD. Comparison between two groups was analyzed using unpaired Student's t-test. Comparisons among multiple groups were performed with one-way ANOVA followed by Tukey's test, using GraphPad Prism 7 (version 7; GraphPad Software, Inc.). $P < 0.05$ was considered to indicate a statistically significant difference.

Results

CADM1 has an inhibitory effect in melanoma. To investigate the role of CADM1 in melanoma cells, the expression of CADM1 was detected using RT-qPCR. It was found that the expression of CADM1 was significantly decreased in melanoma cells (WM-266-4, A2058 and A375) compared with normal melanocytes cells (PIG1; Fig. 1A). Western blotting was then used to assess this result, and it was identified that the protein expression of CADM1 was significantly inhibited in the melanoma cells compared with PIG1 cells (Fig. 1B). Therefore, these results indicated that CADM1 had an inhibitory effect in melanoma *in vitro*.

Next, the efficiency of cell transfection was detected by RT-qPCR. The results suggested that the expression of CADM1 in the melanoma cells was significantly upregulated by overexpression of CADM1, but decreased in the presence of short hairpin (sh)-CADM1 (Fig. 1C). Then, to examine the effect of CADM1 on the EMT process of melanoma, RT-qPCR and western blotting were performed. The results demonstrated that the relative expression levels of vimentin and Slug were significantly inhibited by overexpression of CADM1, but increased by CADM1 knockdown. However, the expression of E-cadherin in melanoma cells was significantly increased after CADM1 overexpression, while knockdown of CADM1 significantly decreased the relative expression of E-cadherin (Fig. 1D and E). Collectively, these results demonstrated that overexpression of CADM1 significantly downregulated the EMT process in melanoma cells, while reducing the expression of CADM1 had the opposite effect.

miR-214 significantly upregulates the progression of melanoma in vitro. RT-qPCR was used to investigate the role of miR-214 in melanoma, and it was identified that the expression of miR-214 was significantly increased in melanoma cells compared with normal cells (Fig. 2A). In addition, miR-214 expression was increased to the highest level in A375 cells. Next, RT-qPCR was used to assess the cell transfection and the results indicated that miR-214 was significantly upregulated in melanoma cells in the presence of miR-214 mimics. However, the expression of miR-214 in melanoma cells was significantly decreased by miR-214 inhibitor (Fig. 2B).

To further examine the effect of miR-214 on the tumorigenesis of melanoma, an MTT assay was used to measure the viability of melanoma cells. The results suggested that

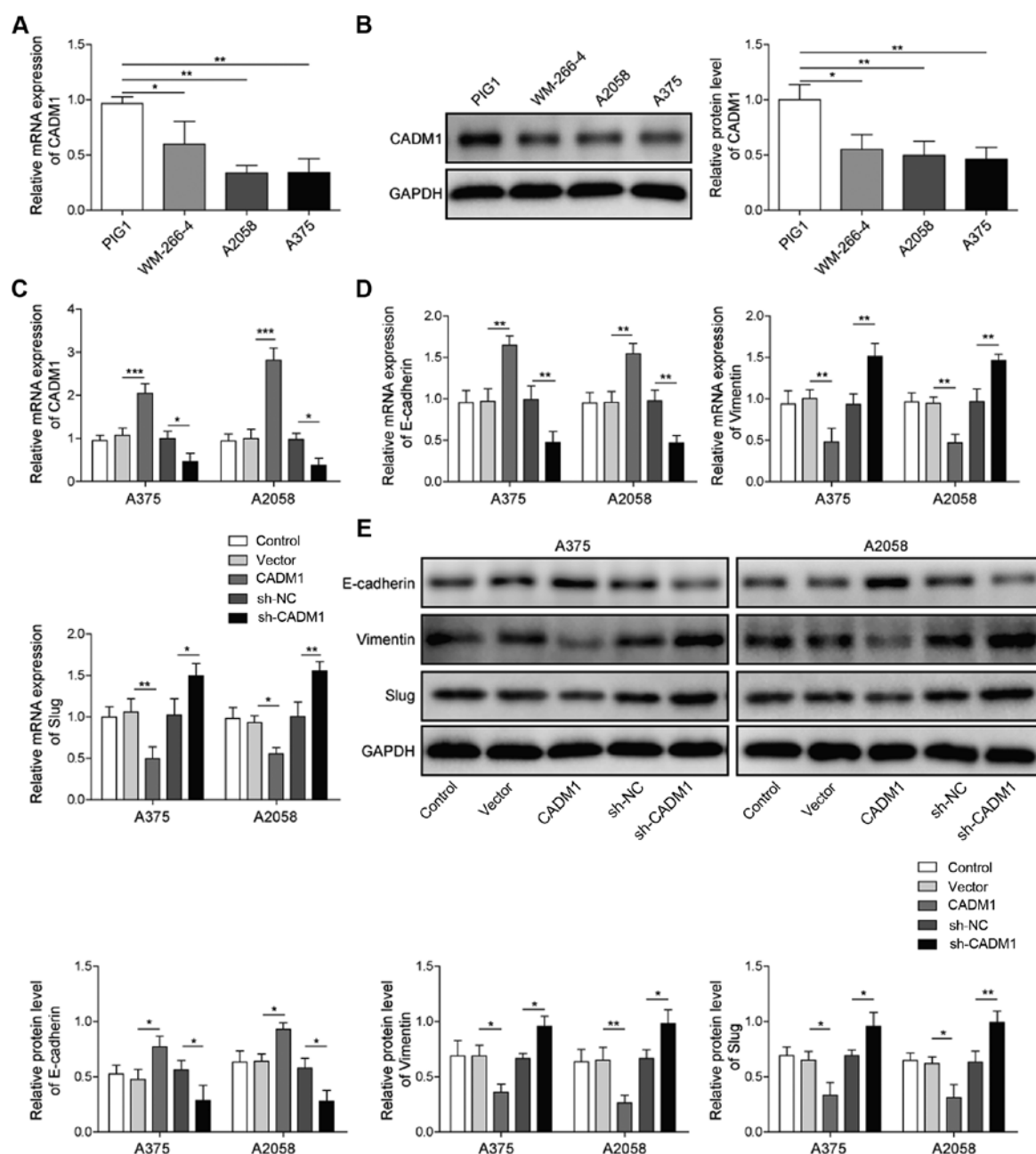


Figure 1. CADM1 downregulates the EMT process in melanoma. Relative expression of CADM1 in PIG1, WM-266-4, A2058 and A375 cells was detected by (A) RT-qPCR and (B) western blotting. (C) A375 or A2058 cells were transfected with overexpression of CADM1, NC or sh-CADM1 for 24 h. Then, cell transfection was assessed by RT-qPCR. Relative expression levels of EMT-related proteins in melanoma cells were detected by (D) RT-qPCR and (E) western blotting. There were ≥ 3 independent experiments performed for each group. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$. sh, short hairpin RNA; NC, negative control; miR, microRNA; CADM1, cell adhesion molecule 1; RT-qPCR, reverse transcription-quantitative PCR; EMT, epithelial-mesenchymal transition.

the viability of A375 cells was significantly increased in the presence of miR-214 mimic, which was reversed by miR-214 inhibitor (Fig. 2C). Moreover, to investigate the role of miR-214 in the progression of melanoma *in vitro*, wound healing and Transwell assays were performed to detect the migration and invasion of melanoma cells, respectively. It was found that the miR-214 mimic significantly increased the migration of melanoma cells, while the miR-214 inhibitor significantly inhibited the migration of A375 and A2058 cells (Fig. 2D). Furthermore, during the invasion assay, A375 cells treated with miR-214 mimic exhibited a significant invasive ability. However, the miR-214 inhibitor significantly inhibited the invasive ability of melanoma cells (Fig. 2E). Therefore, the

results demonstrated that miR-214 overexpression significantly promoted the tumorigenesis and progression of melanoma *in vitro*, while downregulation of miR-214 exhibited an inhibitory effect.

miR-214 overexpression significantly downregulates the expression of CADM1. To investigate the relationship between miR-214 and CADM1, the binding site of CADM1 and miR-214 was analyzed by bioinformatics prediction software, and a dual-luciferase reporter assay was performed to verify the specific binding of miR-214 and CADM1. Luciferase reporter analysis results identified that miR-214 mimic significantly downregulated the activity of the reporter containing wild-type

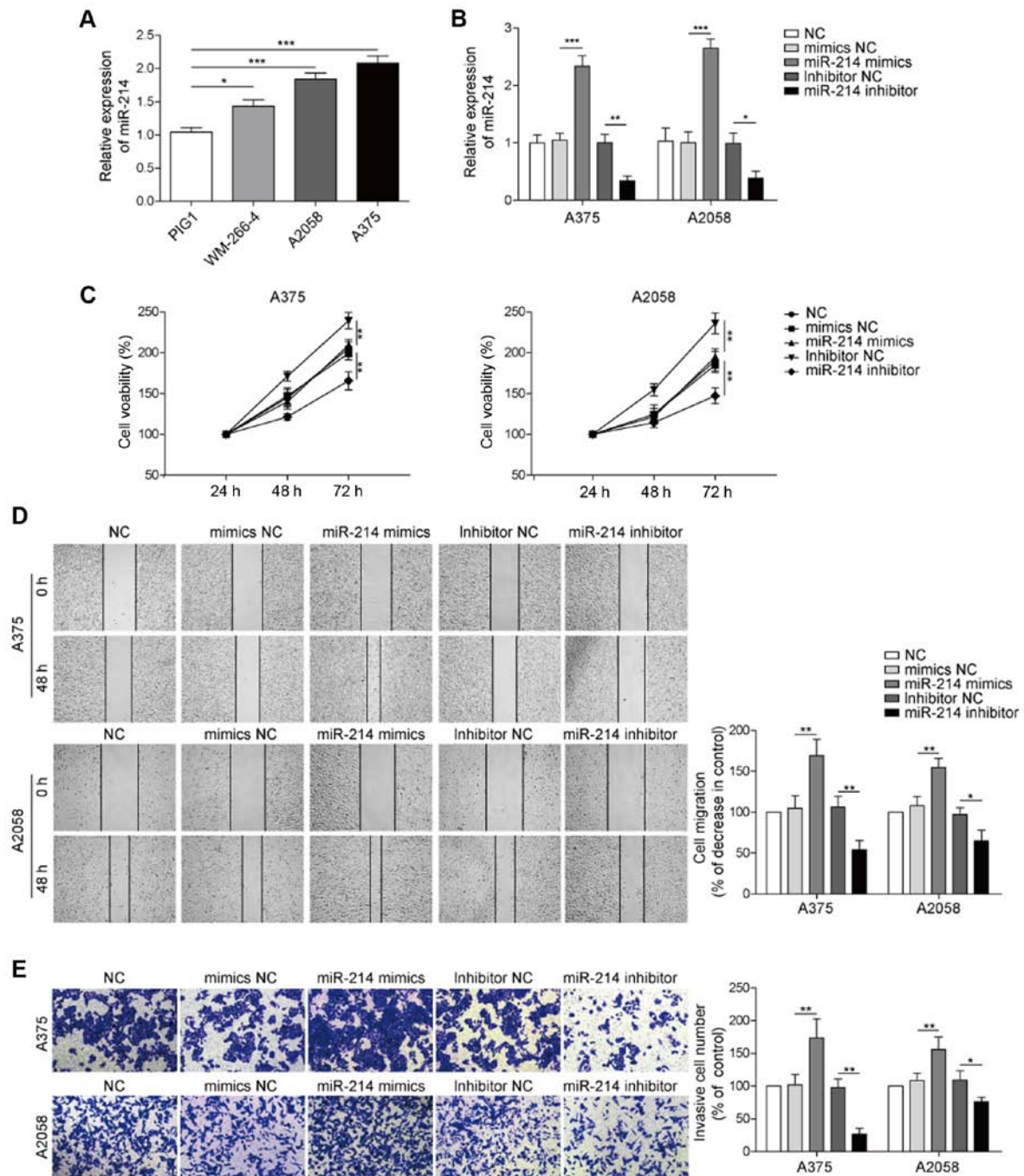


Figure 2. miR-214 upregulates the tumorigenesis and progression of melanoma. (A) Relative expression of miR-214 in PIG1, WM-266-4, A2058 and A375 cells was detected by RT-qPCR. (B) A375 or A2058 cells were transfected with miR-214 mimics, NC or miR-214 inhibitor for 24 h. Then, cell transfection was assessed by RT-qPCR. (C) Cell viability in the NC, mimic NC, inhibitor NC, miR-214 mimic and miR-214 inhibitor groups was measured after 24, 48 and 72 h using MTT assay. (D) Cell migration in the NC, mimic NC, inhibitor NC, miR-214 mimic and miR-214 inhibitor groups was tested by wound healing assay (magnification, $\times 100$). (E) Cell invasion in the NC, mimic NC, inhibitor NC, miR-214 mimic and miR-214 inhibitor groups was tested by Transwell assay (magnification, $\times 100$). NC, negative control. There were ≥ 3 independent experiments performed for each group. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$. NC, negative control; miR, microRNA; RT-qPCR, reverse transcription-quantitative PCR.

CADM1; however, mutating the binding site of miR-214 in CADM1 abolished this suppression (Fig. 3A and B). Thus, the results suggested that CADM1 may be a direct target regulated by miR-214.

In addition, RT-qPCR was used to determine the expression of CADM1 in melanoma cells transfected with miR-214 mimic or inhibitor. It was found that the expression of CADM1 was significantly enhanced by the miR-214 inhibitor, but was reversed by miR-214 mimic (Fig. 3C). Moreover, the results

indicated that the protein expression of CADM1 was significantly inhibited by the presence of miR-214 mimic, which was reversed by miR-214 inhibitor (Fig. 3D). Therefore, it was suggested that the overexpression of miR-214 downregulated the expression of CADM1 in melanoma cells, while down-regulation of miR-214 increased the expression of CADM1.

miR-214 promotes EMT in melanoma cells by downregulating CADM1 expression. To further investigate the mechanism

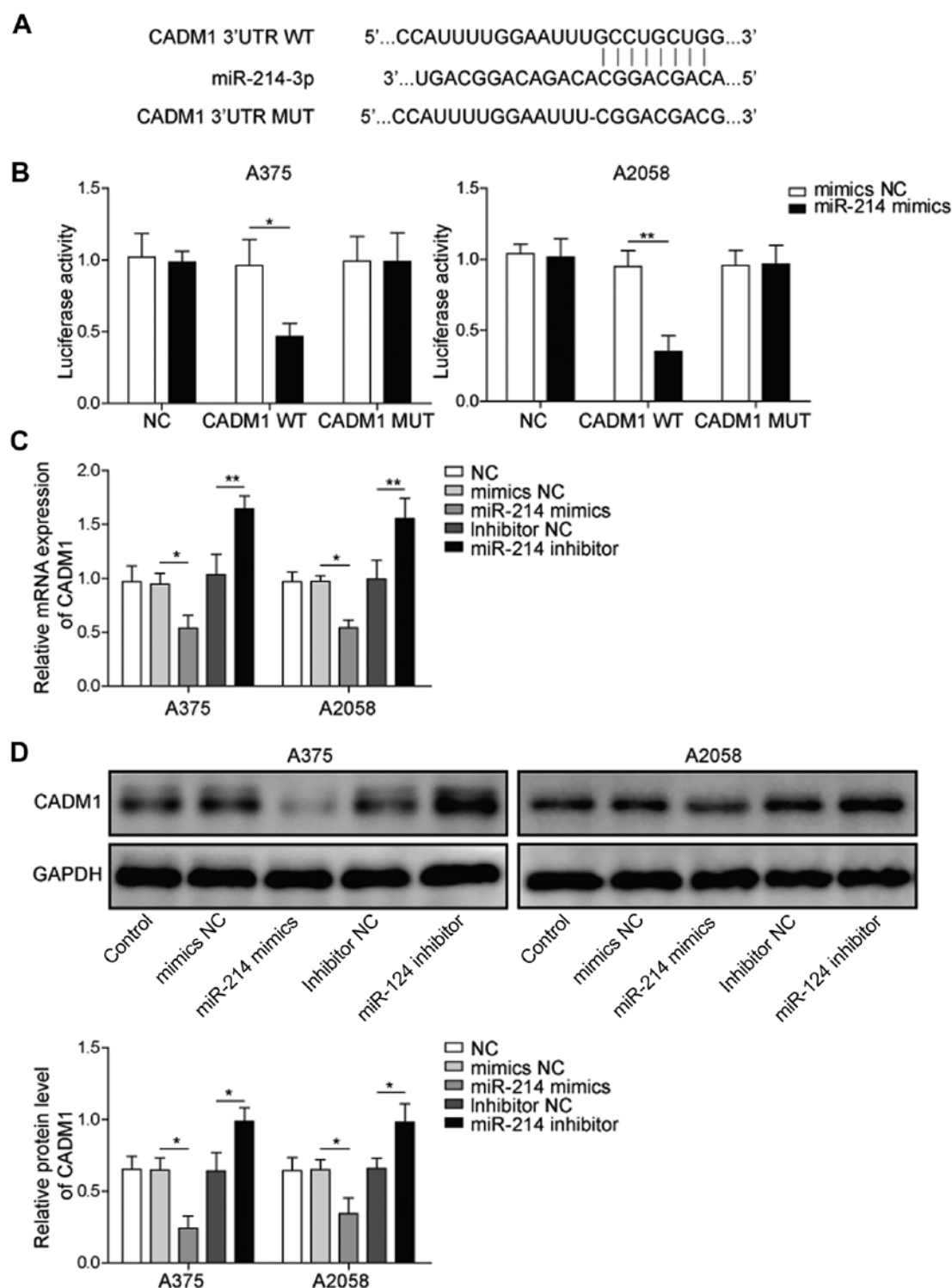


Figure 3. miR-214 regulates CADM1 expression. (A) Binding site of miR-214 to CADM1 was analyzed by bioinformatics prediction software. The position and sequences of the predicted binding sites are shown. (B) A luciferase reporter assay was performed to assess the predicted binding sites with the sequence in miR-214 mimic-transfected melanoma cells. Relative expression of CADM1 in the NC, miR-214 mimic and miR-214 inhibitor groups was detected by (C) reverse transcription-quantitative PCR and (D) western blotting. There were ≥ 3 independent experiments performed for each group. * $P < 0.05$, ** $P < 0.01$. NC, negative control; miR, microRNA; WT, wild-type; MUT, mutant; CADM1, cell adhesion molecule 1.

via which miR-214 regulated the EMT process of melanoma *in vitro*, the expression levels of CADM1 and EMT-related proteins were detected by RT-qPCR and western blotting. It was demonstrated that the expression of CADM1 was significantly upregulated in the presence of miR-214 inhibitor (Fig. 4A and B). Moreover, the miR-214 inhibitor significantly

decreased the relative expression levels of vimentin and Slug, but increased the relative expression of E-cadherin in melanoma cells, which was significantly reversed by the knockdown of CADM1.

Furthermore, to investigate the effect of CADM1 on melanoma cells transfected with miR-214 inhibitor, wound healing

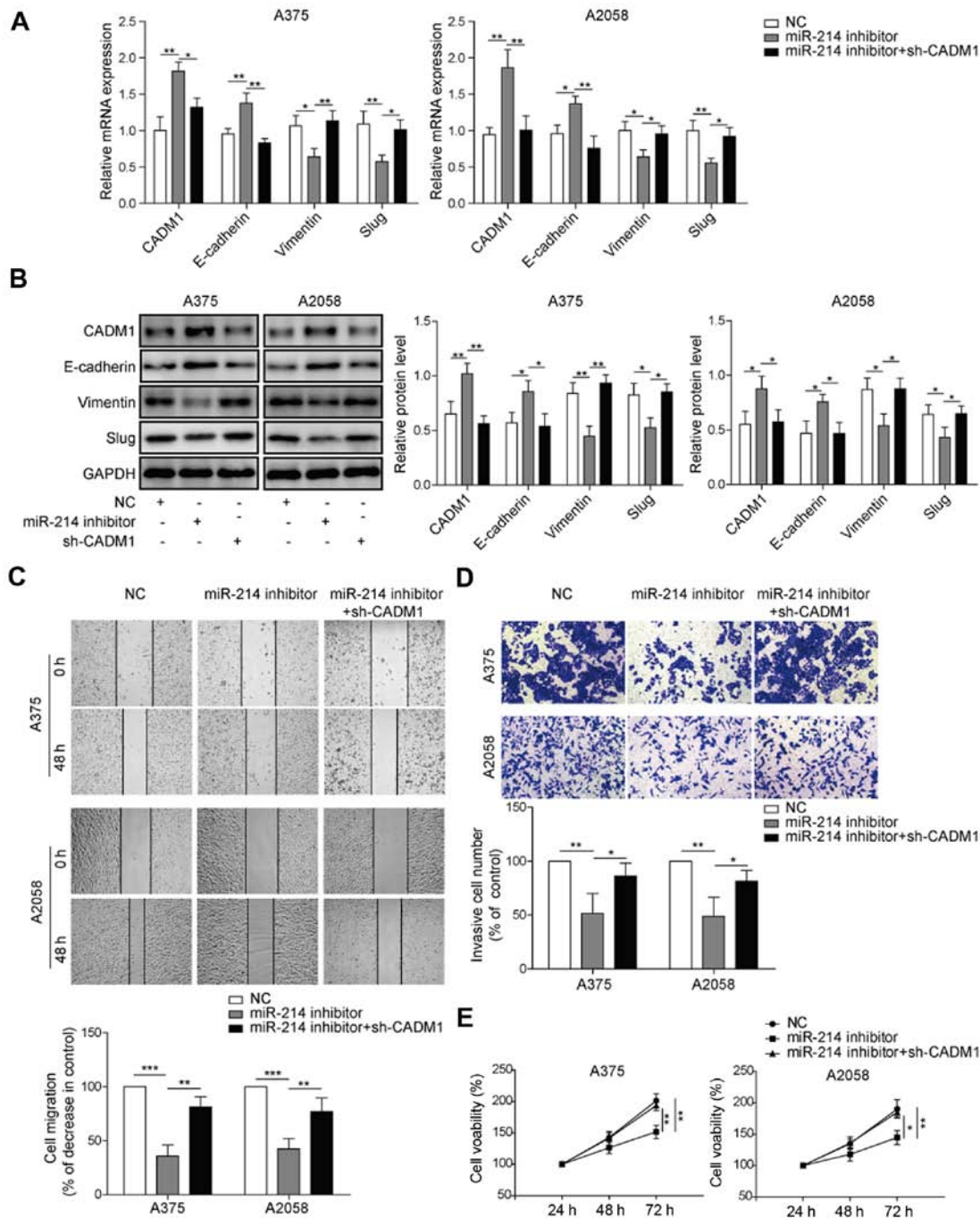


Figure 4. miR-214 downregulates CADM1 and upregulates the EMT process in melanoma cells. (A) Relative mRNA expression levels of E-cadherin, Slug and vimentin in the NC, miR-214 inhibitor and sh-CADM1 + miR-214 inhibitor groups were detected by reverse transcription-quantitative PCR. (B) Relative protein expression levels of EMT-related proteins in the NC, miR-214 inhibitor and sh-CADM1 + miR-214 inhibitor groups were detected by western blotting. (C) Cell migration in the NC, miR-214 inhibitor and sh-CADM1 + miR-214 inhibitor groups was assessed by wound healing assay (magnification, x100). (D) Cell invasion in the NC, miR-214 inhibitor and sh-CADM1 + miR-214 inhibitor groups was measured by Transwell assay (magnification, x100). (E) Cell viability in the NC, miR-214 inhibitor and sh-CADM1 + miR-214 inhibitor groups was examined after 24, 48 and 72 h by MTT assay. There were ≥ 3 independent experiments performed for each group. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$. NC, negative control; miR, microRNA; WT, wild-type; MUT, mutant; CADM1, cell adhesion molecule 1; sh, short hairpin RNA; EMT, epithelial-mesenchymal transition.

and Transwell assays were performed. The results demonstrated that the migration of A375 cells was significantly inhibited by miR-214 inhibitor, which was partially rescued by downregulation of CADM1 (Fig. 4C). In the Transwell assay, the invasion of A375 and A2058 cells was significantly inhibited in the presence of miR-214 inhibitor. However, CADM1 downregulation partially reversed the inhibitory effect of miR-214 inhibitor on cell invasion in melanoma

(Fig. 4D). Moreover, to test the effect of CADM1 on the viability of A375 or A2058 cells treated with miR-214 inhibitor, an MTT assay was conducted. It was found that cell viability was decreased by the miR-214 inhibitor, which was partially reversed by knockdown of CADM1 (Fig. 4E). Collectively, the results indicated that miR-214 affected the EMT process of melanoma by modulating the expression of CADM1.

Discussion

Previous studies have focused on the metastasis of malignant tumors (27-29); however, the understanding of the mechanism of melanoma development remains limited (30). It has been reported that the immune system plays a key role in the treatment of metastasis in advanced cancer (31-36). Moreover, E-cadherin, vimentin and Slug are key regulators during the EMT process of malignant tumors (37,38). Previous studies have also shown that EMT-related proteins are closely associated with tumor metastasis (39,40), which is in line with the present results, thus indicating that EMT is closely related to tumor metastasis. Furthermore, it is possible that metastatic cells after EMT are vulnerable to host immunosurveillance (41). Consistent with these findings, the present results suggested that metastasis in melanoma may be closely associated with the EMT process, and thus EMT may play a critical role in the metastasis of melanoma.

It has been reported that CADM1 plays an inhibitory role in tumorigenesis and progression of cancer. For instance, overexpression of CADM1 inhibits cell-cycle progression and invasion of hepatocellular carcinoma (11). Moreover, the present results indicated that CADM1 acted as a tumor suppressor in melanoma, which was similar to the results of a previous study (9). Furthermore, overexpression of CADM1 significantly inhibited the EMT process of melanoma. Therefore, it was speculated that CADM1 may inhibit the tumorigenesis and progression of melanoma by downregulating the EMT process.

miR-214 functions as a pro-metastatic factor in malignant tumors (42), and plays a critical role in the migration and invasion of cancer cells (43-45). The present study identified a potential link between miR-214 and CADM1 in the coordination of metastasis in melanoma. Furthermore, the present results suggested that CADM1 expression decreased when miR-214 was upregulated in melanoma cell lines. It was also found that miR-214 significantly promoted the EMT process of melanoma, while CADM1 inhibited the EMT process. These results were similar to those of previous studies (46-48), thus indicating that miR-214 increased the invasion and migration of melanoma by downregulating CADM1 to promote the EMT process. However, it has also been revealed that miR-214 promotes cancer progression by activating the NF- κ B signaling pathway (49). Therefore, future studies should focus on the association between melanoma and the NF- κ B signaling pathway. In addition, the present study used 0.5% FBS to test the cell migration in wound healing assay, which may be a limitation. Although 0.5% FBS may enhance the proliferative ability of cells to a certain extent, the effect of proliferation on scratches may be negligible due to the low concentration, however, in order to further eliminate this interference, follow-up studies will use other methods to detect cell migration ability.

In conclusion, the present results suggest that miR-214 promoted the EMT process of melanoma by downregulating CADM1, which may serve as a potential novel target for the treatment of melanoma.

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Availability of data and materials

All data generated or analyzed during this study are included in this published article.

Authors' contributions

SJW, WWL and TLZ conceived and supervised the study. SJW and TLZ designed the study. CJW and YLD performed the experiments and analyzed the data. All authors reviewed the results and approved the final version of the manuscript.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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