

Testis-expressed protein 33 is not essential for spermiogenesis and fertility in mice

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Abstract. Gene expression analyses have revealed that there are >2,300 testis-enriched genes in mice, and gene knockout models have shown that a number of them are responsible for male fertility. However, the functions of numerous genes have yet to be clarified. The aim of the present study was to identify the expression pattern of testis-expressed protein 33 (TEX33) in mice and explore the role of TEX33 in male reproduction. Reverse transcription-polymerase chain reaction and western blot assays were used to investigate the mRNA and protein levels of TEX33 in mouse testes during the first wave of spermatogenesis. Immunofluorescence analysis was also performed to identify the cellular and structural localization of TEX33 protein in the testes. *Tex33* knockout mice were generated by CRISPR/Cas9 gene-editing. Histological analysis with hematoxylin and eosin or periodic acid-Schiff (PAS) staining, computer-assisted sperm analysis (CASA) and fertility testing, were also carried out to evaluate the effect of TEX33 on mouse spermiogenesis and male reproduction. The results showed that *Tex33* mRNA and protein were exclusively expressed in mouse testes and were first detected on postnatal days 21-28 (spermiogenesis phase); their expression then remained into adulthood. Immunofluorescence analysis revealed that TEX33 protein was located in the spermatids and sperm within the seminiferous tubules of the mouse testes, and exhibited specific localization to the acrosome, flagellum and manchette during spermiogenesis. These results suggested that TEX33 may play a role in mouse spermiogen-

esis. However, *Tex33* knockout mice presented no detectable difference in testis-to-body weight ratios when compared with wild-type mice. PAS staining and CASA revealed that spermatogenesis and sperm quality were normal in mice lacking *Tex33*. In addition, fertility testing suggested that the *Tex33* knockout mice had normal reproductive functions. In summary, the findings of the present study indicate that TEX33 is associated with spermiogenesis but is not essential for sperm development and male fertility.

Introduction

Male fertility relies on the continuous production of sperm through a complex developmental process termed spermatogenesis. This process involves the differentiation of spermatogonia into spermatocytes, the development of spermatocytes into round spermatids through two successive meiotic divisions and the final elongation of spermatids (1,2). The process of spermatid elongation is also known as spermiogenesis and involves the differentiation of round spermatids into sperm via marked morphological changes. Spermiogenesis includes chromatin condensation, elongation of the nucleus, biogenesis of the acrosome, extension of the flagellum and removal of excess cytoplasm (3-5).

The acrosome is a unique cap-like organelle that is derived from the Golgi apparatus and gradually expands to cover the apical region of the sperm nucleus (6). Formation of the acrosome is one of the most readily detectable changes in the seminiferous tubules (7), and the process responsible for acrosomal development has been well characterized. First, proacrosomal granules originating from the trans-Golgi stacks assemble in the concave region of the nuclear surface and fuse with each other to form a single acrosomal granule. This subsequently enlarges and spreads to form a cap over the nucleus. Finally, the acrosome migrates over the surface of the sperm nucleus and forms its characteristic shape (8-10).

Microtubule structure plays an important role in spermiogenesis. For example, the manchette, a temporary microtubular structure, first appears in step-8 spermatids but subsequently disappears as spermiogenesis enters steps 13 and 14 (2). During the elongation of spermatids, the manchette shapes the distal half of the head and facilitates the intracellular transport of proteins to the flagellum (11,12).

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A previous study reported that >2,300 genes are expressed predominantly in mouse testes (13), and a number of them are identified as serving important roles in numerous events during spermiogenesis through gene-editing technology, such as the biogenesis of the acrosome, the morphology of the manchette and the assembly of the flagellum. For example, mice lacking autophagy related 7, sirtuin 1, serine-rich single-pass membrane protein 1, vacuolar protein sorting (*Vps*)13b, *Vps54* and sperm acrosome associated 1 exhibit abnormal round-headed sperm as a result of aberrant acrosome formation (8,9,14-16). Leucine-rich repeats and guanylate kinase domain containing isoform 1, serine/threonine kinase 22, katanin-like 2, sperm flagellar 2, hook microtubule tethering protein 1 and calcium and integrin binding family member 4 are essential for the correct formation of the manchette; deficiency in any of these genes can lead to abnormal elongation of the manchette and thus, a malformed head or flagellum (11,12,17-20). In addition, the deletion of sperm associated antigen 16, kinesin light chain 3, kinase suppressor of ras 2, testis-specific serine kinase 4 or genes of the septin (*Sep*) family (*Sept2*, *Sept4* and *Sept12*) causes alterations in the intra-flagellum structure and reduces sperm motility (21-25). Despite the crucial function of these genes in spermiogenesis, the localization and functional requirement of other genes requires further characterization.

Testis-expressed protein 33 (TEX33), also known as Ean57, 1700061J05Rik, cE81G9.2 and C22orf33, is evolutionarily conserved. Orthologs of this protein have been identified in a number of different species, including humans, mice, Norwegian rats, zebrafish and Rhesus monkeys. A previous bioinformatics analysis revealed that TEX33 was expressed abundantly in the testis and associated with the formation of cilia (26). Therefore, we hypothesized that TEX33 may be a testis-enriched or specific protein that localizes to the sperm flagellum and is potentially associated with spermiogenesis.

The aim of the present study was to investigate the function of TEX33 in mouse spermiogenesis and fertility using CRISPR/Cas9 gene-editing. It is hoped that the results will enrich our understanding of the molecular networks underlying spermiogenesis and provide new insight into the genetics of male fertility.

Materials and methods

Animals. A total of three male C57BL/6N mice aged 10 weeks and two female C57BL/6N mice aged 8 weeks were purchased from Cyagen Biosciences, Inc. The CRISPR/Cas9 system was used to generate *Tex33* knockout mice. Two guide RNAs (gRNAs) were designed using the E-CRISPR database (<http://www.e-crisp.org/E-CRISPR/designcrispr.html>) to target exons 2-4 of *Tex33*: gRNA1, TACCAGAATCATCTAGTCCCTGG (matching the forward strand of the gene) and gRNA2, GCTAGCCAAGGCCAACACCTGGG (matching the reverse strand of the gene). The two gRNAs and Cas9 NLS endonuclease (cat. no. M0646M; New England BioLabs, Inc.) were micro-injected into the cytoplasm of fertilized eggs from C57BL/6N mice. These eggs were then implanted into the oviducts of pseudo-pregnant female mice. Founder mice were then crossed with C57BL/6N mice to generate *Tex33*^{+/-} offspring. Finally, *Tex33*^{-/-} mice were obtained by breeding from the *Tex33*^{+/-} mice.

All mice were maintained under specific pathogen-free conditions with free access to standard mouse lab pellet food and water, and housed at 20-22°C and 50-70% humidity with a 12-h light/dark cycle. All animal experiments were approved by the Animal Ethics Committee of Yangzhou University.

Reverse transcription-polymerase chain reaction (RT-PCR). For RNA extraction, heart, brain, spleen, liver, lung, kidney and trachea were collected from adult wild-type mice (n=3) and testes were harvested from wild-type mice at different spermatogenesis stages (postnatal days 7, 14, 21, 28, 35 and 56; n=3 each). Total RNAs were isolated on ice using TRIzol® reagent (Invitrogen; Thermo Fisher Scientific, Inc.) and then dissolved in RNase-free distilled water. The purity of the total RNAs was determined using ultraviolet spectrophotometry (Eppendorf) and agarose gel electrophoresis. Only RNA samples with a ratio of absorbance at 260 and 280 nm of ≥1.8 were used for RT-PCR. For reverse transcription, 500 ng total RNA in a final volume of 10 μl was mixed with 5X PrimeScript RT Master Mix (Takara Bio, Inc.) and RNase-free distilled water. For cDNA synthesis, the mixture was incubated at 37°C for 15 min, and then at 85°C for 15 sec. *Tex33* transcript variants were then amplified by PCR using the following primers: *Tex33* transcript variant V1: 5'-GGTTCATTTGCCTTCTCC-3' (forward), 5'-TCCTTGTCGTTCCGGGTGTG-3' (reverse); *Tex33* transcript variant V2: 5'-ACCTGGGGAGAGCTGCTGCTCG-3' (forward), 5'-GTTGGTGGTAGAGGTGGAG-3' (reverse); *Tex33* transcript variant V3: 5'-TGGGTCA GGAAGGAGTGTG-3' (forward), 5'-GTTGGTGGTAGAGGTGGAG-3' (reverse). Glyceraldehyde 3-phosphate dehydrogenase (*GAPDH*) was used as a positive control using the following primer sequences: 5'-GACGGCCGCATCTTCTTGT-3' (forward) and 5'-ACACCGACCTTCACCATTTTGT-3' (reverse). PCR was performed using a Taq DNA polymerase kit (cat. no. M1661S; Promega Corporation). Amplification conditions for *Tex33* transcript variant V1, V2 and V3 were as follows: V1, initial denaturation step at 95°C for 5 min followed by 35 cycles of at 95°C for 30 sec, an annealing step at 60°C for 45 sec, an extension step at 72°C for 45 sec, and a final extension at 72°C for 10 min; V2, initial denaturation at 95°C for 5 min, followed by 35 cycles at 95°C for 30 sec, 58°C for 30 sec, 72°C for 45 sec, and elongation at 72°C for 10 min; V3, an initial denaturation step of 5 min at 95°C, followed by 35 cycles 95°C for 30 sec, 52°C for 30 sec, 72°C for 45 sec, ended by a final elongation at 72°C for 10 min. PCR protocol for *GAPDH* included a 5 min denaturation step at 95°C followed by 35 cycles of 50 sec at 95°C, 1 min annealing step at 55°C and 1 min extension at 72°C and a final elongation at 72°C for 10 min. PCR products were electrophoresed on 1.2% agarose gels using 1X Tris/Borate/EDTA buffer (TBE). Gels were stained with SYBR Safe™ (Thermo Fisher Scientific, Inc.) and the bands visualized using a gel imaging system (Tanon Science and Technology Co., Ltd.).

Synthesis of an anti-TEX33 antibody. PCR was used to amplify the open reading fragment (ORF) of *Tex33* transcript variant V2 (encoding 266 amino acids), which represents the predominant transcript according to the GenBank database and shares the same ORF as V1. The following primers were used: 5'-GGGAATTCATGGAGCTGAGCCACCGAC-3' (forward)

and 5'-GGAAGCTTGCTTGGCCTTGGATCTGCTC-3' (reverse). The PCR product was purified with a DNA gel extraction kit (Corning, Inc.), sequenced by next generation sequencing (BGI Group) and finally subcloned into a pET-30a (+) vector (Novagen; Merck KGaA). After induction with isopropyl β -D-thiogalactoside for 12-14 h at 37°C, the prokaryotic protein samples were analyzed by electrophoresis on 10% sodium dodecyl sulfate (SDS) polyacrylamide gels using Coomassie Brilliant Blue solution. The TEX33 prokaryotic protein was successfully expressed in *Escherichia coli* BL21 (DE3) plysS (Novagen; Merck KGaA). The resulting His-tag fusion protein was purified using Ni-NTA affinity resin (RuiXiang Bio Inc.) in accordance with the manufacturer's instructions. The concentration of the purified protein was determined using a bicinchoninic acid (BCA) kit (CWBio).

A total of 5 male New Zealand white rabbits (~2 kg body weight) aged 10 weeks were used for animal immunization. They were purchased from Laboratory Animal Center of Shanghai Branch of Chinese Academy of Science and maintained in individual cages at 22°C, 42% humidity, with a 12-h light/dark cycle and free access to food and water. Purified protein (0.75 mg/ml) was mixed with an equal volume of Freund's adjuvant (Sigma-Aldrich; Merck KGaA) and 500 μ l purified protein/adjuvant mixture was subsequently injected into the backs of adult male rabbits at 0, 2, 4, 6 and 8 weeks. After the last immunization, blood was harvested from each rabbit and the serum was separated. The antibody titer from each rabbit was determined using an enzyme-linked immunosorbent assay (ELISA; Fig. S1A).

For ELISA, 5 μ g/ml purified TEX33 prokaryotic protein was added to each well of a 96-well plate (Corning Inc.) and incubated at 4°C overnight. After washing with 0.05% Tween-20 solution in phosphate-buffered saline (PBS-T), each well was blocked with 1% bovine serum albumin (BSA; Beijing Solarbio Science & Technology Co., Ltd.) for 1 h at room temperature. After washing, 1:10, 1:100, 1:1,000, 1:10,000, 1:100,000, and 1:1,000,000 dilutions of serum were added and the plate was incubated at 37°C for 1 h. The wells were then washed thrice with PBS-T and incubated with goat-anti-rabbit IgG-HRP secondary antibody (cat. no. ZB-2301; 1:2,500 dilution; OriGene Technologies, Inc.) at 37°C for 1 h. TMB Two-Component Substrate solution (Beijing Solarbio Science & Technology Co., Ltd.) was then added and allowed to react for 10 min at room temperature in the dark. The reaction was stopped with 1 mM hydrochloric acid and the absorbance was measured at 450 nm using a Multiskan FC microplate reader (Thermo Fisher Scientific, Inc.). The specificity of the antibody was confirmed by western blot (Fig. S1B) and immunofluorescence analyses (Fig. S1C-F).

Western blot analysis. For protein extraction, heart, brain, spleen, liver, lung, kidney and trachea were collected from multiple tissues of adult wild-type mice (n=3) and testes were harvested from wild-type mice at different spermatogenesis stages (postnatal days 7, 14, 21, 28, 35 and 56; n=3 each). Tissues were lysed in RIPA buffer (CWBio) on ice, followed by sonication on ice for 10 min. After centrifugation at 14,000 x g for 10 min at 4°C, the supernatants were collected. The protein concentration was determined using a BCA kit. Protein samples (40 μ g) were then separated on

10% SDS-polyacrylamide gels and transferred onto polyvinylidene fluoride membranes (Millipore; Merck KGaA). Non-specific antibody binding was blocked using 5% non-fat milk in Tris-buffered saline (TBS) for 1 h at room temperature. Membranes were then incubated at 4°C overnight with the following primary antibodies: TEX33 antibody (1:1,000 dilution; generated in the present study), GAPDH antibody (cat. no. CW0100M; 1:2,500 dilution; CWBio) and β -actin antibody (cat. no. ET1701-80; 1:2,500 dilution; HuaBio). After washing with 0.05% TBS-T, membranes were incubated with goat-anti-rabbit IgG-HRP secondary antibody (cat. no. ZB-2301; 1:2,500 dilution; OriGene Technologies, Inc.) for 1 h at room temperature. After washing three times with TBS-T, protein bands were visualized with enhanced chemiluminescence reagent (New Cell and Molecular Biotech Co., Ltd.) using an automatic chemiluminescence image analysis system (Tanon Science and Technology Co., Ltd.).

Immunofluorescence analysis. In order to analyze tissue sections by immunofluorescence, testicular samples were collected from the mice and fixed in 4% paraformaldehyde solution (pH 7.0) for 24-48 h at 4°C. The testicular samples were dehydrated in an ascending alcohol gradient, then cleared in chloroform overnight. The cleared samples were infiltrated in xylene, embedded in paraffin and cut into 4- μ m sections. The sections were rehydrated in a descending alcohol gradient followed by antigen retrieval in 10 mM sodium citrate (pH 6.0) at 100°C for 10 min. After blocking with 1% BSA for 1 h at room temperature, the sections were incubated with primary antibodies at 4°C overnight. The following primary antibodies were used: TEX33 antibody (1:200 dilution; generated in this study) and mouse α -tubulin antibody (cat. no. AC012; 1:100 dilution; ABclonal Biotech Co., Ltd.). After washing with PBS, sections were further incubated with Alexa Fluor 488-conjugated donkey-anti-mouse secondary antibody (cat. no. A32766TR; 1:1,000 dilution; Thermo Fisher Scientific, Inc.) and Alexa Fluor 555-conjugated donkey-anti-rabbit secondary antibody (cat. no. A-31572; 1:1,000 dilution; Thermo Fisher Scientific, Inc.) for 1 h at room temperature, and then counterstained with 4',6-diamidino-2-phenylindole (DAPI) dye (cat. no. C0060; 1:1,000 dilution; Beijing Solarbio Science & Technology Co., Ltd.) for 10 min at room temperature. Fluorescence intensity was finally detected by confocal laser-scanning microscopy (Carl Zeiss AG).

In order to investigate sperm using immunocytochemistry, the cauda epididymis of each mouse was cut into small pieces to allow motile sperm to swim out. After filtration and centrifugation at 800 x g for 15 min at room temperature, the sperm sample was rinsed with PBS, incubated in PBS at 37°C for 15 min, smeared on slides and fixed with 4% paraformaldehyde solution for 30 min at room temperature. The slides were blocked with donkey serum (cat. no. SL050; 1:100 dilution; Beijing Solarbio Science & Technology Co., Ltd.) at room temperature for 1 h, and then incubated with anti-TEX33 primary antibody (1:100 dilution; generated in the present study) at 4°C overnight. This was followed by incubation with FITC-conjugated peanut agglutinin (PNA-FITC; cat. no. L7381; 1:50 dilution; Sigma-Aldrich; Merck KGaA) and Alexa Fluor 555 donkey-anti-rabbit secondary antibody (cat. no. A-31572; 1:1,000 dilution). After washing three times

with PBS, the nuclei were stained with DAPI for 10 min at room temperature. Images were captured by confocal laser-scanning microscopy.

Mouse genotyping. Mouse tail samples (0.2–0.3 cm) were obtained and lysed at 55°C overnight in lysis buffer containing 1 mM Tris-HCl (pH 8.0), 1 mM NaCl, 1 mM EDTA (pH 8.0), 0.5% SDS and 10 µg/µl proteinase K (Sigma-Aldrich; Merck KGaA). Genomic DNA was extracted from the lysate using phenol and chloroform (Beijing Solarbio Science & Technology Co., Ltd.), precipitated with isopropanol (Sinopharm Chemical Reagent Co., Ltd.) and dissolved in distilled water at 60°C. Subsequent PCR was performed using a Taq DNA polymerase kit (cat. no. M1661S; Promega Corporation) according to the manufacturer's instruction. Primers for genotyping were designed: 5'-CAGCCGATGCTTCTATGACAAC-3' (forward), 5'-TCCCACATTTTCCGCAGGTG-3' (reverse 1), 5'-TCTGGGCTCCGTAATGTCTG-3' (reverse 2). The PCR conditions were as follows: Denaturation at 94°C for 5 min, followed by 35 cycles at 94°C for 50 sec, 55°C for 1 min, 65°C for 50 sec, and a final extension at 65°C for 10 min. PCR products were separated on 1.2% agarose gels by electrophoresis in 1X TBE buffer. The products were visualized using a gel imaging system (Tanon Science and Technology Co., Ltd.) after staining with SYBR Safe™ (Thermo Fisher Scientific, Inc.).

Histological analysis. Paraffin sections (~4 µm) fixed in Bouin's solution (including testes, cauda epididymides and ovaries) were used for both histological and morphological analyses. The sections were dewaxed with xylene, and then rehydrated with 100, 95, 90, 80 and 70% alcohol for 2 min each. After washing with RNase-free distilled water, the sections were used for hematoxylin and eosin (H&E) or periodic acid-schiff (PAS) staining.

For H&E staining, the sections were first stained with hematoxylin (Beyotime Institute of Biotechnology) for 2 min at room temperature, followed by 6 dips in 1% hydrochloric alcohol and rinsing in RNase-free distilled water. The sections were then stained with eosin (Beyotime Institute of Biotechnology) for 3 sec at room temperature, followed by dehydration in 70, 90 and 100% alcohol for 1 min each. Finally, the sections were immersed in xylene for 5 min. Images were visualized by light microscopy (Nikon-70i; Nikon Corporation).

For PAS staining, a Schiff's kit (cat. no. G1360; Beijing Solarbio Science & Technology Co., Ltd.) was used in accordance with the manufacturer's protocols. In brief, the sections were pretreated with periodic acid for 5 min at room temperature, slowly rinsed in RNase-free distilled water and then stained with Schiff's solution for 10 min at room temperature in the dark. The nuclei were stained with hematoxylin for 5 min at room temperature, followed by 6 dips in 1% hydrochloric alcohol. After dehydration with 70, 90 and 100% graded alcohol, the sections were immersed twice in xylene for 10 min each. Images were captured using light microscopy (Nikon-70i).

Analysis of sperm motility and morphology. Cauda epididymides were collected and rinsed with RNase-free distilled water, followed by the incubation of swim-out sperm in 1 ml

warm human tubal fluid (Wisent Biotechnology) at 37°C. The total sperm count was determined using a hemocytometer and sperm motility was determined with a computer-assisted sperm analysis system (CASA; Sperm Class Analyzer v. 4.0.0; Microptic S.L., Barcelona, Spain). The sperm deformity rate was studied using sperm smears and microscopy counting methods. Sperm smears were stained with H&E and images were captured by light microscopy (Nikon-70i). More than 1,000 sperm were observed and counted from *Tex33^{+/+}* and *Tex33^{-/-}* males (n=3 mice/group). The numbers of sperm showing normal and abnormal morphology were counted and analyzed.

Fertility testing. *Tex33^{+/+}* and *Tex33^{-/-}* male mice (10–12 weeks old) were continuously mated with *Tex33^{+/+}* female mice (8–10 weeks old) with demonstrable fertilizing ability (at a ratio of 1:2) for 1 month. The *Tex33^{+/+}* female mice were changed once a week. Plugs and the numbers of pups born were recorded.

Statistical analysis. Sperm quality and average litter sizes are expressed as mean ± standard error of mean and the Student's t-test was used to identify significant differences. Plugs and pregnancies were compared between *Tex33^{+/+}* and *Tex33^{-/-}* mice using the Chi-squared test. P<0.05 was considered to indicate a statistically significant result.

Results

Expression of *Tex33* during spermatogenesis in mice. The GenBank database revealed that the mouse *Tex33* gene is located in the E1 region of chromosome 15 and contains seven exons (Fig. 1A). There are three transcript variants: V1, V2 and V3 (Fig. 1B). V1 is the longest *Tex33* transcript variant; it is 1,147 bp in size and lacks exon 3 as the result of alternative splicing events (Fig. 1B). V2 is the predominant transcript variant of the *Tex33* gene according to GeneBank, and exhibits alternative splice sites in exon 1 and exon 3, resulting in an additional exon and the deletion of exon 3 (Fig. 1B). Another transcript variant, V3, is the shortest variant and exhibits alternative splice sites in exons 1 and 2 (Fig. 1B). The translation of V1 and V2 begins from the same site in exon 2 and these two variants share one stop codon in exon 7; thus, they both encode the same isoform (isoform 1; Fig. 1C). However, V3, encoding isoform 2, translates from exon 4 and lacks a portion of the 5' coding region; this leads to a shorter N-terminus compared with isoform 1 (Fig. 1C).

To examine the mRNA levels of *Tex33* *in vivo*, specific primers were designed for the three transcript variants and the expression profiles of the three transcript variants were then determined by RT-PCR using cDNA samples synthesized from various adult mouse organs, including the heart, brain, spleen, liver, lung, kidney, trachea and testis. The results showed that all transcript variants were only detected in the testes (Fig. 1D). To investigate the levels of TEX33 protein in these tissues, a TEX33 antibody with high efficiency was generated as described in our previous study (27). Western blot analysis performed using the TEX33 antibody revealed that TEX33 was expressed exclusively in the testes, which is

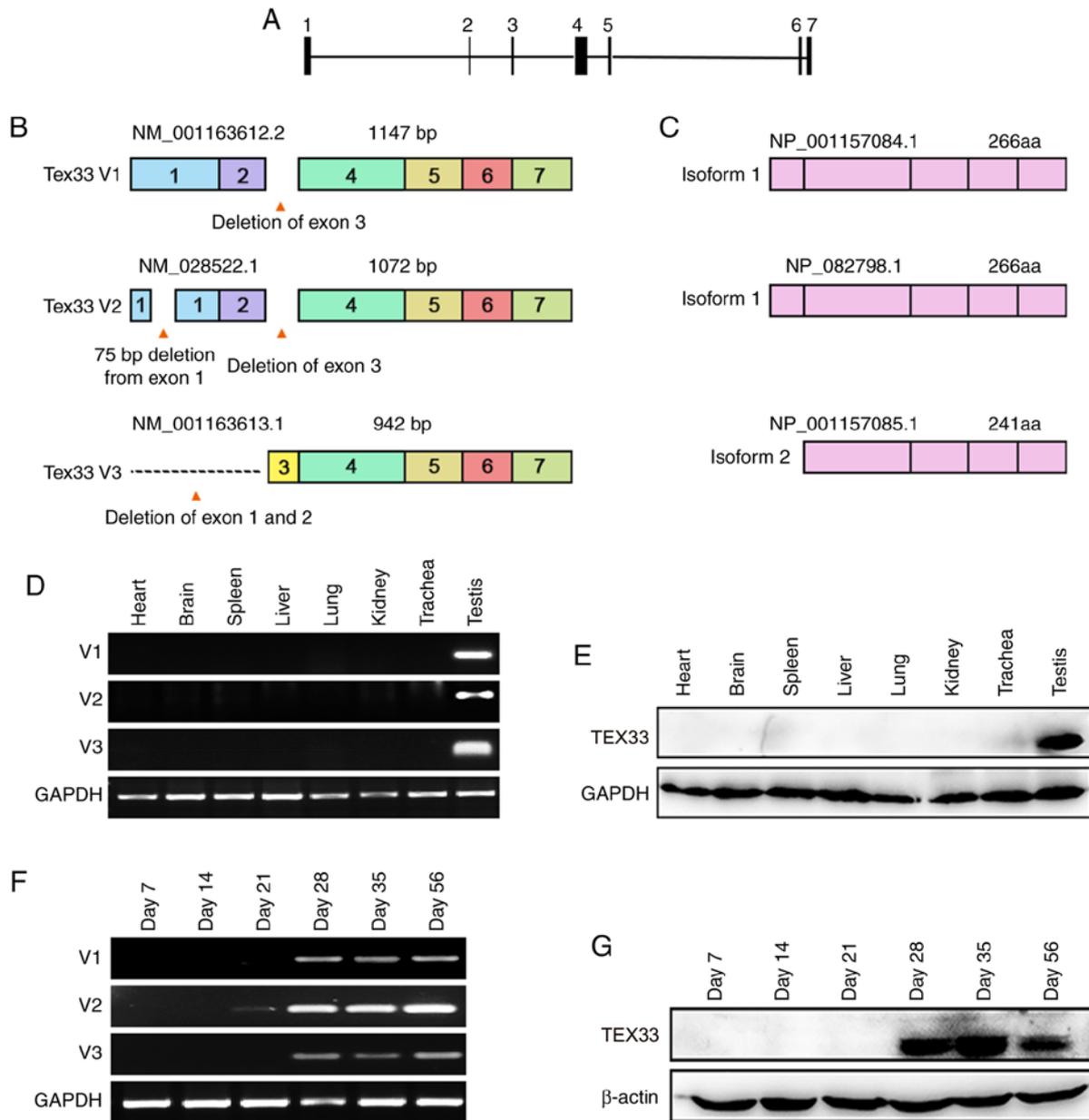


Figure 1. Characterization of mouse *Tex33*. (A) Analysis of the *Tex33* gene in mice. The *Tex33* gene consists of seven exons. (B) *Tex33* transcript variants in mice. Deleted regions reveal alternate splicing events and different combinations of exons. Transcript variant V1 contains six exons, transcript variant V2 has seven exons and transcript variant V3 has five exons. (C) Two different isoforms are encoded by transcript variants V1, V2 and V3. (D) mRNA levels of *Tex33* transcript variants in various mouse tissues were analyzed by RT-PCR. Transcript variant V1, 220 bp; transcript variant V2, 348 bp; transcript variant V3, 261 bp. (E) Expression of TEX33 protein in various mouse tissues, as determined by western blot analysis. (F) Expression of *Tex33* transcript variants during the first wave of spermatogenesis, as determined by RT-PCR. (G) Expression of TEX33 protein during the first wave of spermatogenesis, as determined by western blot analysis. TEX33, testis-expressed protein 33; RT-PCR, reverse transcription-polymerase chain reaction.

consistent with the expression profiles of the transcript variants (Fig. 1E).

To determine the expression profile of *Tex33* during the first wave of spermatogenesis, mouse testes were obtained at postnatal day 7 (containing only spermatogonia), day 14 (containing spermatogonia and spermatocytes), day 21 (in which round spermatids begin to appear), day 28 (in which spermatids begin to elongate), day 35 (around which time the first wave of spermatogenesis is completed) and day 56 (3,28). The results of RT-PCR analysis suggested that V2 was expressed weakly from day 21 in the mouse testes, peaked at day 28 and was expressed into adult-

hood, while V1 and V3 were expressed from day 28 and maintained peak expression levels into adulthood (Fig. 1F). Western blotting showed that TEX33 protein first presented on day 28 and maintained high expression levels into adulthood (Fig. 1G). The appearance of *Tex33* transcript variants and proteins coincides with the spermiogenesis phase, during which time the appearance of round spermatids begins and spermiogenesis is initiated (28).

Immunofluorescence analysis demonstrated that TEX33 protein was localized in spermatids and testicular sperm within the seminiferous tubules (Fig. 2A). In spermatids, TEX33 was detected in Golgi-derived pro-acrosomal granules,

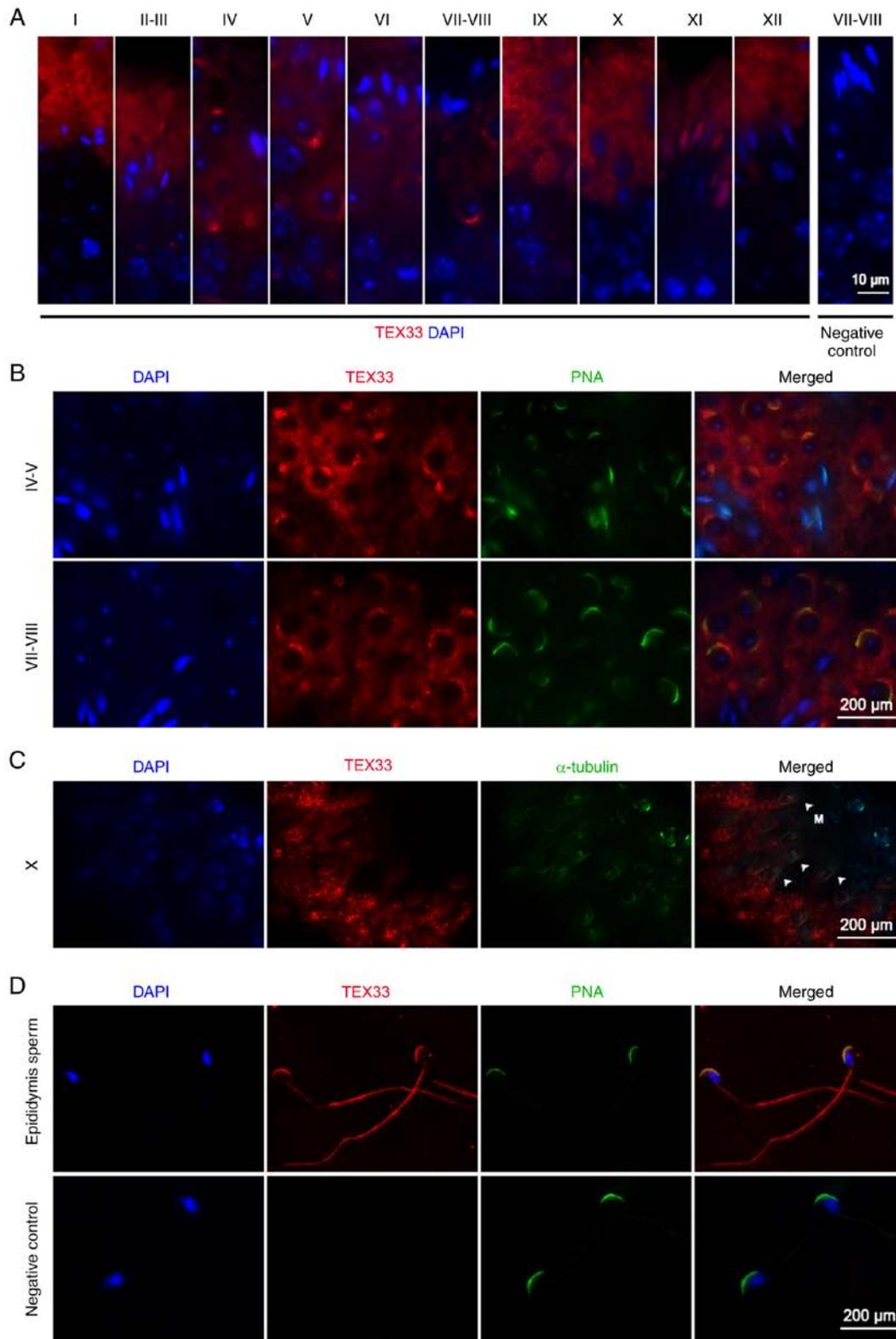


Figure 2. Localization of TEX33 during spermatogenesis. (A) Immunofluorescence staining of TEX33 in the seminiferous tubules during different stages of spermatogenic development (stages I-XII). (B) Co-immunofluorescence staining of TEX33 with PNA-FITC in the seminiferous tubules during different stages of spermatogenic development. (C) Co-immunofluorescence staining of TEX33 and α -tubulin in elongating spermatids during spermatogenic stage X. (D) Co-immunofluorescence staining of TEX33 with PNA-FITC in sperm. TEX33, testis-expressed protein 33; PNA-FITC, FITC-conjugated peanut agglutinin; DAPI, 4',6-diamidino-2-phenylindole; M, manchette (indicated by arrowheads).

the cap-like acrosome and the fully formed acrosome from spermatogenic stages I to XII (Fig. 2A). It has been reported that the acrosome could bind many lectins in a variety of

mammalian species (29,30), such as peanut agglutinin (PNA), which reacts specifically with acrosomal sugar components and is widely used to visualize acrosomal formation (31-35).

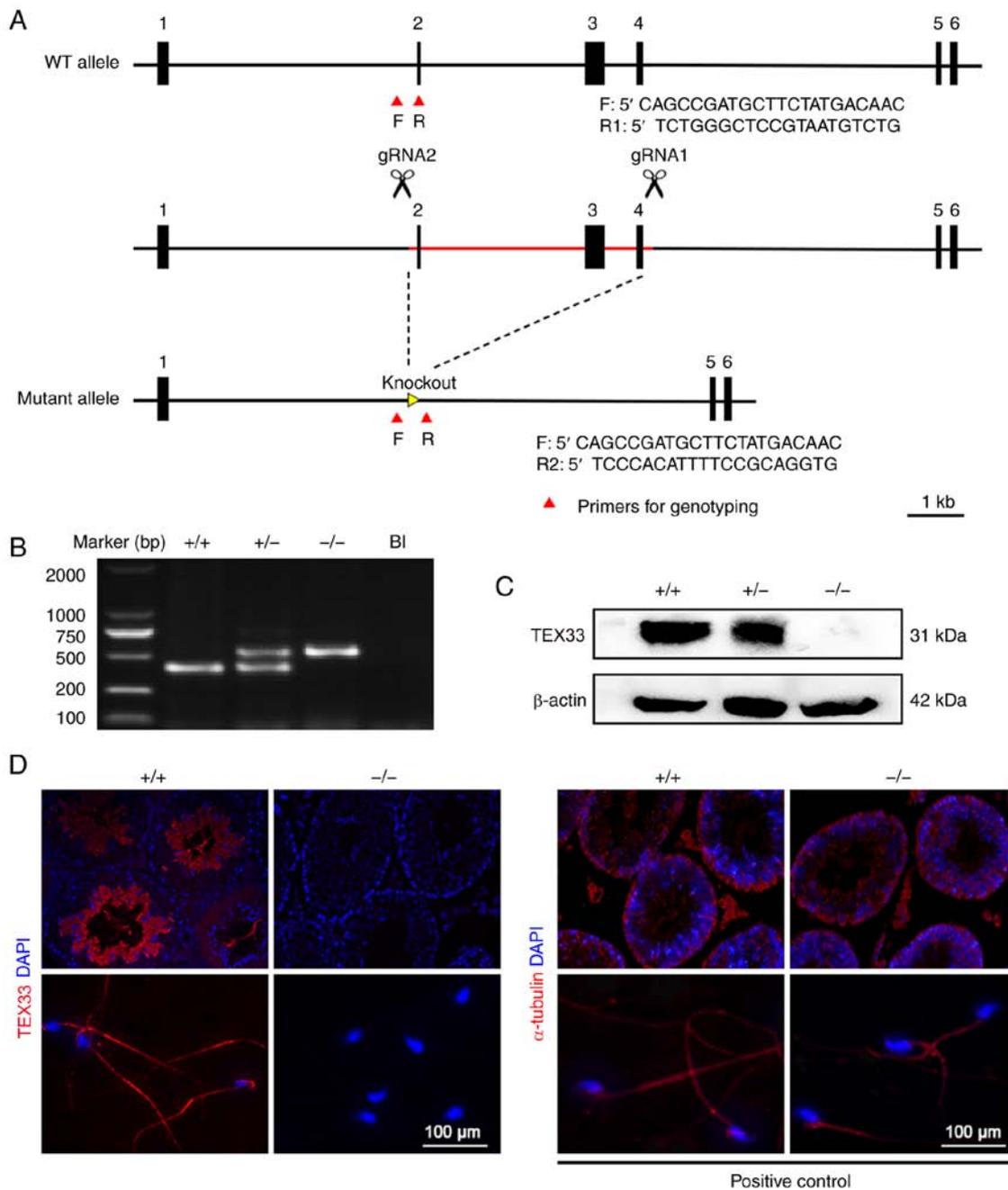


Figure 3. Generation of *Tex33* knockout mice. (A) Schematic representation of the deletion of *Tex33* exons 2-4 via Cas9 microinjection. Primers were designed for genotyping. F targets intron 1, R1 targets exon 2 and R2 targets intron 4. (B) Genotyping of *Tex33* offspring by PCR assay. (C) Comparison of TEX33 knockout efficiency between *Tex33*^{+/+}, *Tex33*^{+/-} and *Tex33*^{-/-} mice by western blot analysis. (D) Detection of TEX33 expression in testes (upper panels) and sperm (lower panels) by immunofluorescence analysis. WT, wild-type; F, forward; R, reverse; gRNA, guide RNA; TEX33, testis-expressed protein 33; +/+, *Tex33*^{+/+} (wild-type) mice; +/-, *Tex33*^{+/-} (heterozygous) mice; -/-, *Tex33*^{-/-} (homozygote) mice; Bl, blank; DAPI, 4',6'-diamidino-2-phenylindole.

In the present study, TEX33 was co-localized with PNA on the acrosomes during different stages of spermatid development (Fig. 2B) and on mature acrosomes (Fig. 2D). TEX33 was also detected on the manchette of elongating spermatids (spermatogenic stage X) via α -tubulin co-labeling (Fig. 2C), as well as the sperm flagellum (Fig. 2D). These findings indicate that TEX33 may be involved in the formation of the acrosome, manchette and flagellum during spermiogenesis.

Deletion of Tex33 in mice. The initial data indicate that TEX33 is expressed at high levels in mouse testes. On the basis of

this, the biological function of TEX33 during spermatogenesis was investigated. CRISPR/Cas9-mediated genome engineering was used to create *Tex33* knockout mice. Exons 2, 3 and 4 were selected as the target sites. Cas9 and gRNA were co-injected into fertilized eggs in order to delete exons 2-4 (Fig. 3A). Mice were then genotyped by PCR using specific primers (Fig. 3A and B). Western blot analysis was performed on testis samples to validate the efficiency of *Tex33* removal. As shown in Fig. 3C, TEX33 protein was present in the *Tex33*^{+/+} and *Tex33*^{+/-} mice, but absent from the *Tex33*^{-/-} mice. Furthermore, immunofluorescence analysis of the testis and

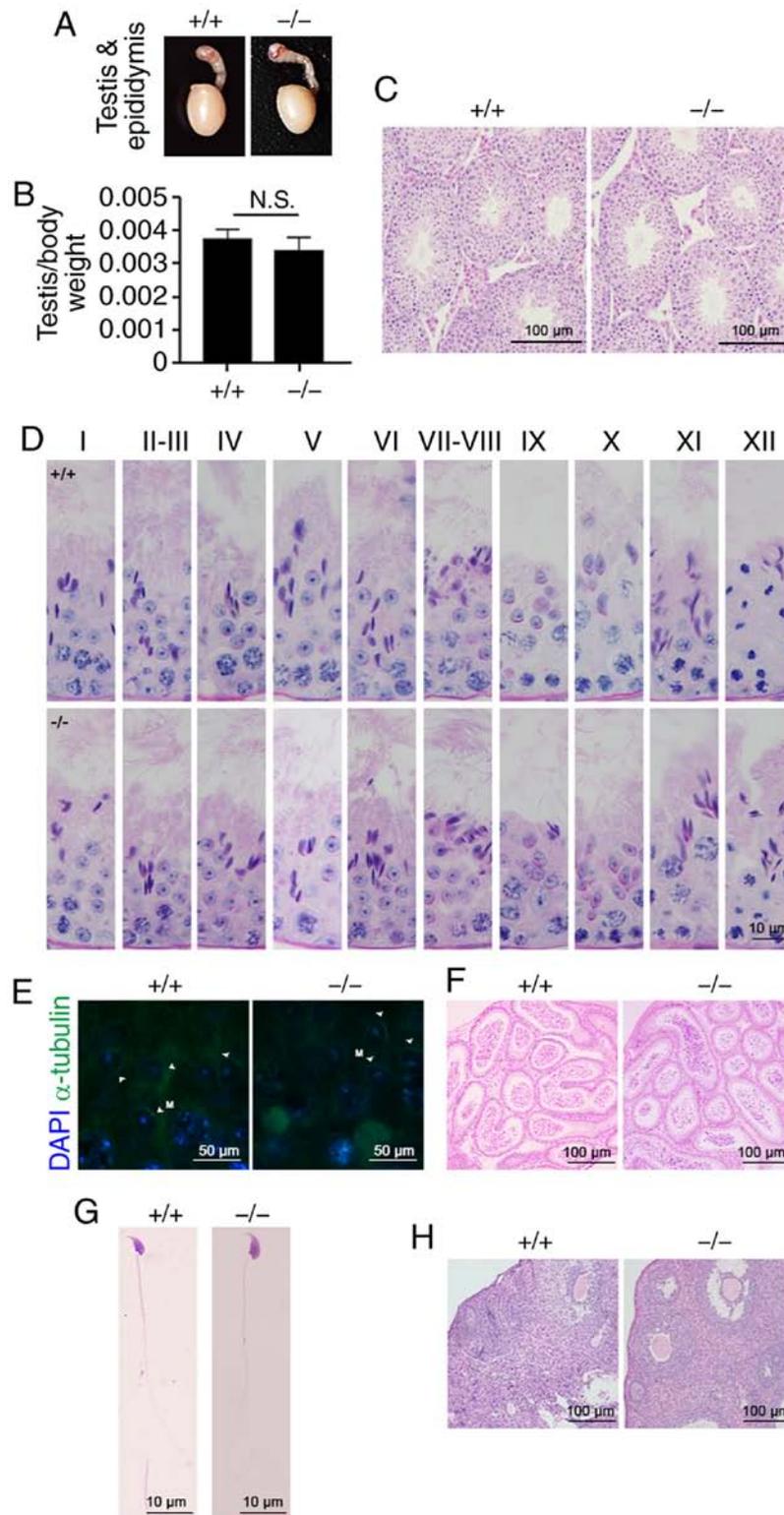


Figure 4. Phenotypic analysis of *Tex33* knockout mice. (A) Testis and epididymis from *Tex33*^{+/+} and *Tex33*^{-/-} males. Tissue samples were collected from 10-week-old male mice (n=5 per genotype). (B) Testis/body weight ratio of *Tex33*^{+/+} and *Tex33*^{-/-} males. Values are expressed as mean \pm standard error of mean. (C) H&E staining showing seminiferous tubules with normal histology in *Tex33*^{+/+} and *Tex33*^{-/-} males. (D) Periodic acid-Schiff staining showing normal acrosome development and spermatogenesis in *Tex33*^{+/+} and *Tex33*^{-/-} males. (E) Co-immunofluorescence staining of TEX33 and α -tubulin in elongating spermatids revealed normal manchette morphology in both *Tex33*^{+/+} and *Tex33*^{-/-} males. H&E staining of *Tex33*^{+/+} and *Tex33*^{-/-} males showing normal (F) cauda epididymis and (G) sperm. (H) H&E staining revealed normal ovarian morphology in *Tex33*^{+/+} and *Tex33*^{-/-} females. H&E, hematoxylin and eosin; Tex33, testis-expressed protein 33; $+/+$, *Tex33*^{+/+} (wild-type) mice; $-/-$, *Tex33*^{-/-} (homozygote) mice; M, manchette (indicated by arrowheads); DAPI, 4',6-diamidino-2-phenylindole; N.S., not significant.

sperm confirmed that TEX33 expression was absent from the *Tex33*^{-/-} mice (Fig. 3D).

Tex33 deletion does not affect spermatogenesis and fertility. To determine whether the deletion of *Tex33* has an effect on

Table I. Analysis of sperm quality in *Tex33*^{+/+} and *Tex33*^{-/-} male mice.

Genotype	Sperm count (x10 ⁶ /ml)	Total sperm motility (%)	Progressive motility (%)	Sperm deformation (%)
+/+ (n=3)	9.02±0.46	80.82±3.69	57.59±5.06	16.31±3.76
-/- (n=3)	8.98±0.35	77.96±4.51	58.26±3.47	18.12±4.08
P-value	0.91	0.47	0.86	0.60

Data are expressed as mean ± standard error of the mean and were analyzed using Student's t-test. *Tex33*, testis-expressed protein 33; +/+, *Tex33*^{+/+} (wild-type) males; -/-, *Tex33*^{-/-} (homozygous) males.

Table II. Fertility analysis of *Tex33*^{+/+} and *Tex33*^{-/-} mice.

Test	Males	Females	Average litter size	P-value
Male fertility	+/+ (n=8)	+/+ (n=64)	8.69±0.37	0.13
	-/- (n=8)	+/+ (n=64)	8.41±0.34	
Female fertility	+/+ (n=5)	+/+ (n=12)	8.67±0.86	0.61
	+/+ (n=5)	-/- (n=12)	8.92±0.80	

Data are expressed as mean ± standard error of the mean and were analyzed using the Student's t-test. *Tex33*, testis-expressed protein 33; +/+, *Tex33*^{+/+} (wild-type) mice; -/-, *Tex33*^{-/-} (homozygous) mice.

Table III. Comparison of pregnancies in *Tex33*^{+/+} and *Tex33*^{-/-} mice.

Test	Males	Females	Pregnancies (%)	P-value
Male fertility	+/+ (n=8)	+/+ (n=64)	90.00	0.23
	-/- (n=8)	+/+ (n=64)	94.64	
Female fertility	+/+ (n=5)	+/+ (n=12)	81.81	0.59
	+/+ (n=5)	-/- (n=12)	90.00	

Data were analyzed using Chi-square tests. *Tex33*, testis-expressed protein 33; +/+, *Tex33*^{+/+} (wild-type) mice; -/-, *Tex33*^{-/-} (homozygous) mice.

fertility, functional analyses were then carried out on *Tex33* knockout mice. As shown in Fig. 4A, the sizes of the testes and epididymis were comparable between adult *Tex33*^{+/+} and *Tex33*^{-/-} males. The testis-to-body weight ratios were not significantly different between the *Tex33*^{+/+} and *Tex33*^{-/-} male mice (Fig. 4B). The process of sperm development in the testis was examined by H&E staining. The results revealed that *Tex33*^{+/+} and *Tex33*^{-/-} mice exhibited seminiferous tubules with normal morphology and spermatogenic cells (Fig. 4C).

PAS staining was then performed to analyze the process of acrosome development during spermatogenesis. No apparent morphological differences were observed between the testes of *Tex33*^{+/+} and *Tex33*^{-/-} mice in spermatogenic stages I-XII (Fig. 4D). The morphology of the manchette was evaluated by α -tubulin immunofluorescence staining and immunostaining was identified in elongating spermatids in the seminiferous tubules of *Tex33*^{+/+} and *Tex33*^{-/-} male mice at spermatogenic stage IX (Fig. 4E). H&E staining revealed the presence of mature sperm in the cauda epididymides of *Tex33*^{+/+} and

Tex33^{-/-} mice (Fig. 4F). In addition, the morphology of sperm exhibited no clear differences between the *Tex33*^{+/+} and *Tex33*^{-/-} mice (Fig. 4G and Table I). Sperm were counted and their motility assessed to reveal the effect of *Tex33* knockout on sperm quality. Sperm counts were determined using a counting board while motility was determined by a CASA analysis that provided total motility and progressive motility data. These investigations revealed no detectable differences in either sperm count or motility between the *Tex33*^{+/+} and *Tex33*^{-/-} mice (Table I).

A breeding assay was performed to evaluate the fertility of the *Tex33* knockout mice. *Tex33*^{+/+} and *Tex33*^{-/-} male mice (10-12 weeks old) were mated separately with female mice (8-10 weeks old) for 1 month. The *Tex33*^{+/+} and *Tex33*^{-/-} male mice exhibited normal fertility (Tables II-IV), and the litter sizes sired by *Tex33*^{-/-} mice were similar to those sired by *Tex33*^{+/+} mice (Table II). These results indicate that the fertilizing ability of *Tex33*^{-/-} male mice was not significantly different from that of *Tex33*^{+/+} mice. No marked differences in female fertility were observed between the two groups of

Table IV. Comparison of plugs in *Tex33*^{+/+} and *Tex33*^{-/-} mice.

Test	Males	Females	Plugs (%)	P-value
Male fertility	+/+ (n=8)	+/+ (n=64)	93.75	0.35
	-/- (n=8)	+/+ (n=64)	87.50	
Female fertility	+/+ (n=5)	+/+ (n=12)	91.67	0.54
	+/+ (n=5)	-/- (n=12)	83.33	

Data were analyzed using Chi-square tests. *Tex33*, testis-expressed protein 33; +/+, *Tex33*^{+/+} (wild-type) mice; -/-, *Tex33*^{-/-} (homozygous) mice.

mice, as the *Tex33*^{-/-} female mice exhibited normal ovarian structures (Fig. 4H), underwent normal pregnancies and produced equivalent litter sizes to the *Tex33*^{+/+} female mice (Tables II and III).

Discussion

Mature sperm is the end product of spermiogenesis and exhibits highly specialized morphology. Spermiogenesis is facilitated by an important microtubule structure referred to as the manchette, which appears in elongating spermatids and helps to create the shape of the sperm head and transport proteins to the flagellum (2,11). Failure in spermiogenesis, commonly results in subfertility or infertility due to sperm malformation and/or dysfunction (36,37). A number of genes have been reported to be responsible for spermiogenesis (38-42); however, the expression profiles of other genes and their respective roles in spermiogenesis remain unknown and require further investigation. In the present study, *Tex33*, a gene previously shown to be associated with cilia formation (26), was demonstrated for the first time, to the best of our knowledge, to be testis-specific and associated with spermiogenesis. Its role in spermatogenesis was investigated further using a *Tex33* knockout mouse model.

In mammals, primary RNA transcripts give rise to different transcript variants through alternative splicing, thereby resulting in different mRNA and protein isoforms that may have distinct functions and localization patterns (43). Analysis of GenBank data revealed that *Tex33* pre-mRNA undergoes alternative splicing to produce three transcripts. Exon 3 is present in V3, but lacking from V1 and V2; this may be due to exon skipping. Similarly, exon skipping may have contributed to the deletion of exons 1 and 2 from V3. In addition, unlike exon 1 of V1, exon 1 of V2 has a 75-bp deletion. The results of RT-PCR analysis revealed that all three transcript variants are testis specific. Furthermore, TEX33 protein was also detected only in the testis, indicating that the *Tex33* gene is testis specific and conserved from transcription to RNA translation. However, the distribution of this gene in the testis has not previously been elucidated. It has been reported that *Tex33* is associated with cilia formation; therefore, we hypothesized that the TEX33 protein may localize on the sperm flagellum. RT-PCR analysis revealed that the expression of *Tex33* transcript variants began during spermiogenesis, between days 21 and 28, when round spermatids begin to appear and undergo morphological changes. Consistent with this finding, the expression of TEX33 protein

was first detected at day 28, and localized on the acrosome at different developmental stages, including at maturity. Furthermore, it was also detected on the manchette of elongating spermatids during spermatogenesis stages IX and X, as well as on the sperm flagellum. These results confirm that *Tex33* is not only associated with cilia, as previously reported, but is also testis-specific and associated with spermiogenesis. Thus, *Tex33* may serve a crucial role in mouse spermiogenesis, particularly in terms of acrosome biogenesis, manchette morphology and flagellum formation.

Considering the enrichment and cellular localization of TEX33 in the testis, *Tex33* knockout mice were generated by application of the CRISPR/Cas9 system, as reported by Miyata *et al.* (44). The association between TEX33 and spermatid elongation was then explored. Exons 2-4 were selected as targets to ensure the non-functionality of *Tex33* after knockout. The deletion was confirmed by genotyping, western blotting and immunofluorescence analysis.

Recent studies have identified that numerous testis-enriched/specific genes are necessary for male fertility, including CFAP97 domain containing 1, kinesin family member 9, intraflagellar transporter protein 140, protease serine 55 and RIKEN cDNA 1700102P08 (39,40,45-47). Mice lacking any of these genes have been shown to be sterile due to a failure to complete spermatogenesis. However, the *Tex33* knockout mice were found to be fertile. Further analyses of testis and cauda epididymis histology, acrosome biogenesis, manchette morphology and sperm quality also indicated that there were no significant differences between *Tex33*^{-/-} mice and the controls. The possible reason for these knockout mice exhibiting normal spermatogenesis and fertility may be functional redundancy (48-50). Paralogous genes may compensate for the loss of function of knockout genes when they contribute to single function (50). Likewise, it is plausible that *Tex33* paralogs, such as *Tex11* and *Tex14*, have compensational functions in the absence of *Tex33*.

The function of *Tex33* during mouse spermatogenesis has been preliminarily clarified using CRISPR/Cas9 system in the present study. However, considering the cellular and subcellular localization of *Tex33* on sperm and the acrosome, further investigations using transmission electron microscopy or *in vitro* fertilization experiments are recommended.

In summary, the present study preliminarily describes the expression profile and role of TEX33 during spermatogenesis in mice. Although *Tex33* was testis-specific and associated with spermiogenesis, deletion of this gene had no effect on sperm development and reproduction in mice. This indicates

that *Tex33* may regulate spermiogenesis in association with cooperating factors; however, it is not required for spermiogenesis and male fertility.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

MX, JX and CN designed and performed the experiments. MX wrote the manuscript. YaZ collected and analyzed the data. MX, TG and YD generated knockout mice and the TEX33 antibody. YiZ designed the study. All authors read and approved the final manuscript.

Ethics approval and consent to participate

All animal experiments were approved by the Animal Ethics Committee of Yangzhou University.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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