

Lycium barbarum polysaccharide protects ARPE-19 cells against H₂O₂-induced oxidative stress via the Nrf2/HO-1 pathway

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Abstract. Age-related macular degeneration (AMD) is a global health problem. Lycium barbarum polysaccharide (LBP), a traditional Chinese herbal medicine, has been proven to be effective against several eye diseases. However, only a few studies have investigated the effectiveness of LBP for AMD. In the present study, the human retinal epithelial cell line, ARPE-19, was pretreated with LBP for 24 h before exposure to H₂O₂ (500 μ M). Cell viability was assessed, and a series of oxidative and antioxidant indicators were evaluated to determine the influence of LBP on H₂O₂-triggered oxidative stress. The present study also determined the apoptosis status, as well as the expression levels of apoptotic proteins and nuclear factor erythroid 2-related factor 2 (Nrf2)/heme oxygenase-1 (HO-1) pathway proteins. The present study aimed to determine the protective role for LBP pretreatment and its underlying molecular mechanism. The results of the present study suggest that pretreatment of ARPE-19 cells with LBP exhibit high efficacy at reducing oxidative damage and inhibiting cell apoptosis. Furthermore, LBP may modulate the expression of proteins involved in the apoptotic pathway and activate the Nrf2 signaling pathway.

Introduction

Patients presenting with age-related macular degeneration (AMD) suffer from progressive visual degeneration due to a damaged macular area (1). It is estimated that ~300 million

people will be diagnosed with AMD by 2040 (1). There are two prominent pathological features associated with AMD, the formation and accumulation of drusen, and damage to retinal pigment epithelial (RPE) cells (2). Furthermore, the decrease of neuronal nitric oxide synthase in RPE cell nuclei may be associated with the redox status of the RPE in patients with AMD (3). Therefore, protecting these cells from injury is important to prevent AMD pathology.

As a critical part of the blood-retinal barrier, the RPE plays an essential role in supporting the neural retina and visual cycle, by protecting the fundus tissue from oxidation (4). The RPE cell layer is easily damaged by reactive oxygen species (ROS) compared with other cells, due to the high oxygen consumption of the retina (5,6). In addition, ROS-induced damage to RPE cells is an irreversible process and is an early sign of AMD (7). Thus, therapies against oxidative stress (OxS) should be effective in protecting the RPE and may help prevent the development of AMD.

The antioxidant, nuclear factor erythroid 2-related factor 2 (Nrf2) plays an essential role in the immune defense system (8). For example, in the cytoplasm, Nrf2 combines with the inhibitor epichlorohydrin-related protein 1 (Keap1) like Kelch (9) under physiological conditions. However, when the cell is damaged, Nrf2 disassociates from Keap1 and translocates to the nucleus, triggering the downstream gene expression of heme oxygenase-1 (HO-1) (10). Recent studies have reported that Nrf2 and HO-1 participate in the etiology of AMD (11), and that they are involved in maintaining the dynamic balance of the retina under stress or trauma (12). Therefore, the activation of Nrf2 may represent a potentially useful therapy for the treatment of AMD.

Lycium barbarum polysaccharide (LBP) has been reported to exhibit several biological functions, including immunomodulation, neuroprotection, anti-aging and antioxidative capabilities (13). Furthermore, LBP has been reported to reduce the levels of ROS and the extent of apoptosis in human lens epithelial cells (14). In addition, ischemia-induced retinal damage in diabetic rats is ameliorated by LBP (15). However, the protective effect of LBP on AMD has not yet been studied; thus, the present study aimed to investigate the inhibitory effect of LBP on H₂O₂-induced OxS and apoptosis in RPE cells, as well as to investigate its effect on the Nrf2/HO-1 pathway, and employed an *in vitro* AMD model by exposing human retinal epithelial cell lines, ARPE-19, to H₂O₂, which may help to provide an alternative potential neoteric strategy for AMD therapy.

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Abbreviations: AMD, age-related macular degeneration; LBP, Lycium barbarum polysaccharide; RPE, retinal pigment epithelium; OxS, oxidative stress; ROS, reactive oxygen species

Key words: age-related macular degeneration, oxidative stress, Lycium barbarum polysaccharide, retinal pigment epithelium, nuclear factor erythroid 2-related factor 2/heme oxygenase-1 pathway

Materials and methods

Materials and chemicals. LBP (purity >90%; cat. no. SP9311), RIPA lysis buffer (cat. no. R0010) and Annexin V-FITC/PI double staining kit (cat. no. CA1020) were purchased from Beijing Solarbio Science & Technology Co., Ltd. DMEM/F12 medium (cat. no. PM150312) and fetal bovine serum (FBS, cat. no. 164210) were purchased from Procell Life Science & Technology Co., Ltd. Primary antibodies against histone H3 (cat. no. ab1791), Bcl2 (cat. no. ab32124), Caspase-3 (cat. no. ab13585), Cleaved caspase-3 (cat. no. ab214430), Bax (cat. no. ab3191) and β -actin (cat. no. ab6276) were purchased from Abcam (dilutions 1:500 or 1:1,000). Antibodies against Nrf2 (cat. no. 12721) and HO-1 (cat. no. 5853) were purchased from Cell Signaling Technology, Inc. (dilution 1:500). Commercial kits for the detection of 2,7-dichlorodihydrofluorescein diacetate (DCFH-DA, cat. no. S0033), malondialdehyde (MDA, cat. no. A003-1-2), superoxide dismutase (SOD, cat. no. A001-1-2), GSH-peroxidase (GSH-Px, cat. no. A005-1-2) and catalase (CAT, cat. no. A007-1-1) were purchased from Nanjing Jiancheng Bioengineering Institute. Horseradish peroxidase secondary antibody (1:500, cat. no. A0216), PBS (cat. no. ST447-5L) and enhanced chemiluminescence (ECL, cat. no. P0018S) reagent were purchased from Beyotime Institute of Biotechnology, and all other chemicals were purchased from Sigma-Aldrich; Merck KGaA.

Cell culture and treatments. ARPE-19 cells (Procell Life Science & Technology Co., Ltd., certified by STR) were maintained in DMEM/F-12 supplemented with 10% FBS, streptomycin (100 mg/ml) and penicillin (100 U/ml), at 37°C with 5% CO₂. All treatments were performed when the cells reached ~80% confluence.

Cell viability assay. ARPE-19 cells were seeded into 96-well plates at a density of 1x10⁴ cells/well, with six replicates for each group. Following incubation overnight at 37°C, the cells were incubated with different concentrations of H₂O₂ (0, 125, 250, 500 and 1,000 μ M) for 2 h at 37°C to determine the optimal concentration. ARPE-19 cells were also pretreated with different concentrations of LBP (0, 0.25, 0.5, 1 and 2 mg/ml) for 24 h at 37°C to optimize the dose of LBP to be used in the present study. Pretreatment with LBP was preceded by co-incubation with H₂O₂ (500 μ M) for 2 h at 37°C to assess the protective effect of LBP on H₂O₂-triggered cell death.

Cell viability was assessed via the Cell Counting Kit-8 (CCK-8, cat. no. HY-K0301; MedChemExpress) assay. Briefly, CCK-8 solution was added into each well and incubated for 2 h at 37°C, in the dark. Cell viability was determined using a microplate reader (BioTek Instruments, Inc.) and calculated as follows: Cell viability (%)=[(absorbance of the test sample-absorbance of the control sample)/mean absorbance of the control] x100.

Measurement of intracellular ROS. The DCFH-DA method was used to measure ROS levels. Briefly, ARPE-19 cells (1x10⁶ cells/well) were cultured in the presence or absence of different concentrations of LBP in 6-well plates for 24 h at 37°C, prior to treatment with 500 μ M H₂O₂. Cells were subsequently cultured in the presence of DCFH-DA (10 mM)

at room temperature in the dark for 20 min. Cells were washed three times with cold PBS and the fluorescence intensity of the harvested cells was determined using a FACSCalibur flow cytometer (Beckman Coulter, Inc.). All experimental results are presented as percentages relative to that of the control sample.

Measurement of MDA, SOD, CAT and GSH-Px. Following the different treatments, ARPE-19 cells (1x10⁶ cells/well) in 1.5 ml Eppendorf tubes (Thermo Fisher Scientific, Inc.) were co-incubated with 100 μ l RIPA lysis buffer and 10% protease inhibitor (cat. no. HY-K0010; MedChemExpress) at 4°C for 30 min. Following lysis and centrifugation at 12,000 x g for 15 min at 4°C, the proteins in the lysate were quantified using the BCA kit (cat. no. P0009; Beyotime Institute of Biotechnology). The intracellular activities of MDA and SOD, and levels of CAT and GSH-Px were determined spectrophotometrically using the relevant commercial kits. SOD, CAT and GPX-Px activities are presented as units/mg protein, while MDA levels are presented as nmol/g of protein. All experimental results are presented as percentages of the control value.

Quantification of apoptosis. After collecting cells from different treatment groups, the centrifuged ARPE-19 cells were resuspended in 100 μ l binding buffer at a density of 1x10⁶ cells/ml. Subsequently, 5 μ l Annexin V-FITC and 5 μ l of PI were added and gently mixed into the cell suspension. Following incubation at room temperature for 15 min in the dark, FACSVerse flow cytometer (BD Biosciences) and FACSuite software (version 1.0.4.2650; BD Biosciences) were used for the quantitation of apoptotic cells. The non-apoptotic, early and late apoptotic cells are presented as Annexin⁻/PI⁻, Annexin V-FITC⁺/PI⁻ and Annexin⁺/PI⁺ cell populations, respectively.

Western blotting. Following the different treatments, ARPE-19 cells (1x10⁶ cells/well) in 1.5 ml Eppendorf tubes were co-incubated with 100 μ l RIPA lysis buffer and 10% protease inhibitor at 4°C for 30 min. Following lysis and centrifugation at 12,000 x g for 15 min at 4°C, the proteins in the lysate were quantified using the BCA kit. Following protein quantitation, 10 or 12% SDS-PAGE was used to resolve the proteins (30 μ g/lane), which were transferred onto PVDF membranes (MilliporeSigma) and subsequently blocked with 5% skimmed milk for 2 h at room temperature. The membranes were incubated with primary antibodies overnight at 4°C. After washing three times with PBS, the membranes were incubated with secondary antibodies for 2 h at room temperature. Protein bands were visualized using ECL reagent and analyzed using Image Lab software (version 4.0, Bio-Rad Laboratories, Inc.). β -actin and histone H3 were used as the internal controls.

Small interfering (si)RNA. ARPE-19 cells (1x10⁵ cells/well) were transfected with 100 μ M negative control (NC, 5'-CAC ACTGGATGGCTAGGAGGATAT-3') siRNA or 100 μ M siRNA Nrf2 (5'-CACACTGGATCAGACAGGAGGATAT-3') (Shanghai GenePharma Co., Ltd.), using Lipofectamine[®] 2000 (Invitrogen; Thermo Fisher Scientific, Inc.). After 12 h of transfection at 37°C, ARPE-19 cells were pretreated with LBP for 24 h and then exposed to 500 μ M H₂O₂ for 2 h. Next,

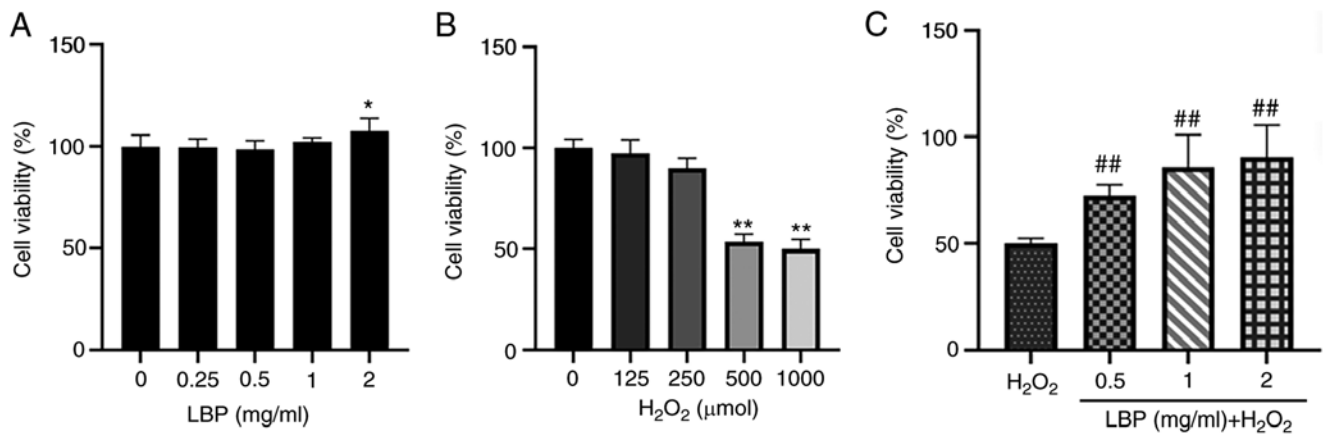


Figure 1. LBP reduces H_2O_2 -induced cytotoxicity. (A) Effect of LBP on the viability of ARPE-19 cells treated with different concentrations of LBP (0, 0.25, 0.5, 1 and 2 mg/ml) for 24 h. (B) Effect of H_2O_2 on the viability of ARPE-19 cells. ARPE-19 cells were treated with different concentrations of H_2O_2 (0, 125, 250, 500 and 1,000 μ M) for 2 h. (C) Effect of LBP on H_2O_2 -induced cytotoxicity in ARPE-19 cells. ARPE-19 cells were pretreated with different concentrations of LBP (0.5, 1 and 2 mg/ml) for 24 h followed by 500 μ M H_2O_2 for 2 h. Data are presented as the mean \pm SEM (n=3). *P<0.05, **P<0.01 vs. control; ##P<0.001 vs. H_2O_2 -treated cells with no LBP pretreatment. LBP, Lycium barbarum polysaccharide.

western blotting and the CCK-8 assay were performed to assess the effect of LBP on the protection of ARPE-19 cells, and determine its underlying molecular mechanism.

Statistical analysis. Statistical analysis was performed using GraphPad Prism version 8.0 (GraphPad Software, Inc.). All experiments were performed in triplicate and data are presented as the mean \pm SEM. Unpaired Student's t-test was used to compare differences between two groups, while one-way ANOVA and Tukey's post hoc test was used to compare differences between multiple groups. P<0.05 was considered to indicate a statistically significant difference.

Results

LBP reduces H_2O_2 -induced cell damage. The toxicity of LBP against ARPE-19 cells was assessed. Following 24 h of pretreatment with LBP (0, 0.25, 0.5, 1 or 2 mg/ml), cell viability was assessed via the CCK-8 assay. The results demonstrated that cell viability was retained before and after 24 h of pretreatment, suggesting that the assessed concentrations of LBP were safe and did not affect the cells (Fig. 1A). To evaluate the potential impact of H_2O_2 , ARPE-19 cells were incubated with 0, 125, 250, 500 and 1,000 μ M H_2O_2 for 2 h, and the resulting cell toxicity was assessed. As expected, cell viability significantly decreased following treatment with H_2O_2 compared with the control group, in a dose-dependent manner (Fig. 1B). Notably, cell viability significantly decreased by 53.6% (P<0.01) at a concentration of 500 μ M H_2O_2 . Thus, this concentration was selected for subsequent experimentation. The antioxidant effect of LBP pretreatment was subsequently evaluated. ARPE-19 cells were treated with different concentrations of LBP (0.5, 1 or 2 mg/ml) and 500 μ M H_2O_2 , and cell viability was assessed via the CCK-8 assay. The results demonstrated that ARPE-19 cell viability increased up to 90.33% following pretreatment with 2 mg/ml LBP (P<0.001; Fig. 1C). Taken together, these results suggested that 24 h of pretreatment with LBP (0.5–2 mg/ml) effectively reduced the H_2O_2 -induced damage in these cells.

LBP ameliorates H_2O_2 -induced OxS. To determine the mechanism by which LBP exerts its cellular protection, indicators of intracellular OxS levels were used along with the evaluation of antioxidant enzyme levels. The DCFH-DA assay was performed to measure ROS levels, as well as the ability of LBP to scavenge H_2O_2 -induced ROS. As presented in Fig. 2A–C, in comparison with the controls, both ROS and MDA levels in ARPE-19 cells significantly increased following treatment with H_2O_2 (P<0.001). However, pretreatment with LBP significantly decreased both ROS and MDA levels (P<0.01 or P<0.001). These results suggest the critical influence of LBP on the inhibition of H_2O_2 -induced OxS in ARPE-19 cells. In addition, antioxidant stress markers (SOD, CAT and GSH-Px) were monitored both in the presence and absence of LBP and H_2O_2 . As presented in Fig. 2D and E, H_2O_2 significantly decreased the activities of these antioxidant enzymes (P<0.001), while LBP pretreatment effectively restored the activities of SOD and CAT to normal levels (P<0.01 or P<0.001). Similarly, the GSH-Px ratio significantly enhanced following pretreatment with LBP compared with cells undergoing H_2O_2 treatment alone (P<0.01 or P<0.001; Fig. 2F), suggesting that LBP pretreatment is a potent inhibitor of OxS damage.

LBP prevents H_2O_2 -induced apoptosis. Flow cytometry was performed to determine the extent of apoptosis occurring in ARPE-19 cells under different experimental conditions. As presented in Fig. 3A and B, the extent of apoptosis in the H_2O_2 group was markedly higher compared with the control group. Notably, H_2O_2 -induced apoptosis in ARPE-19 cells was inhibited following treatment with different concentrations of LBP (Fig. 3C–E). The percentage of apoptotic cells of each experimental group are shown in Fig. 3F, differences were significant (P<0.001). To identify the cause of this anti-apoptotic effect at the protein level, the present study detected the expression levels of apoptosis-related proteins, including pro-apoptotic Bax and cleaved caspase-3 and the anti-apoptotic protein Bcl-2 via western blotting (Fig. 3G). Compared with the controls, cells treated with 500 μ M H_2O_2 exhibited higher levels of Bax and cleaved caspase-3 expression (P<0.01 or P<0.001),

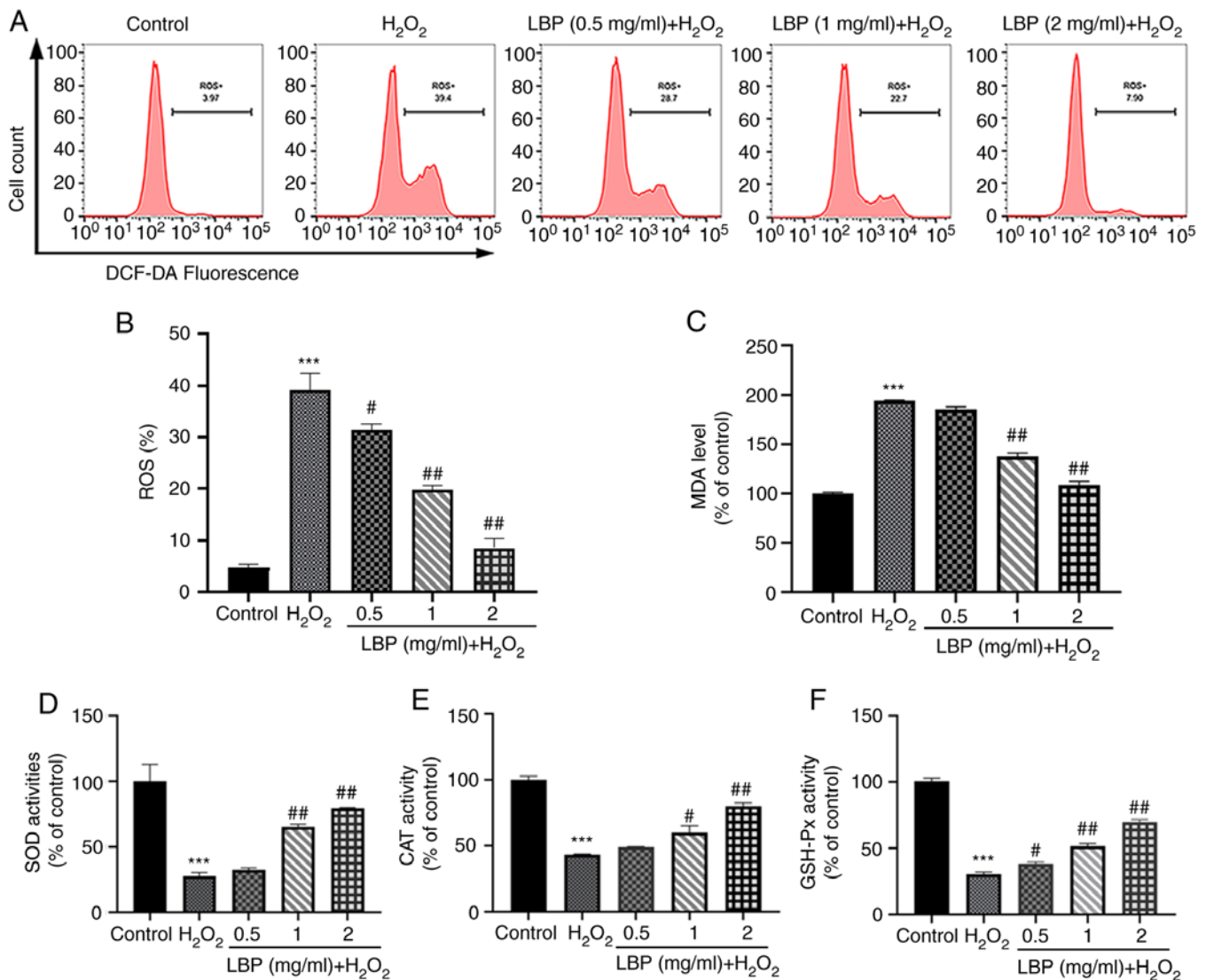


Figure 2. LBP ameliorates H₂O₂-induced oxidative stress. (A and B) Effect of LBP on ROS levels in H₂O₂-treated ARPE-19 cells. (C) Effect of LBP on MDA levels in H₂O₂-treated ARPE-19 cells. (D) Effect of LBP on SOD activity in H₂O₂-treated ARPE-19 cells. (E) Effect of LBP on CAT activity in H₂O₂-treated ARPE-19 cells. (F) Effect of LBP on GSH-Px activity in H₂O₂-treated ARPE-19 cells. Data are presented as the mean \pm SEM (n=3). ***P<0.001 vs. control; #P<0.01, ##P<0.001 vs. H₂O₂-treated cells with no LBP pretreatment. LBP, Lycium barbarum polysaccharide; ROS, reactive oxygen species; MDA, malondialdehyde; SOD, superoxide dismutase; CAT, catalase; GSH-Px, GSH-peroxidase.

which is consistent with the results obtained from flow cytometry. Furthermore, an increased expression of Bcl-2 with a concomitant decrease in Bax and cleaved caspase-3 ($P<0.01$ or $P<0.001$) was observed 24 h after pretreatment with LBP, suggesting that LBP exhibits a significant dose-dependent reversal of H₂O₂-induced apoptosis (Fig. 3G). In addition, the Bcl-2/Bax ratio markedly increased in the LBP pretreatment group ($P<0.001$), the effects of which were reversed following treatment with H₂O₂, suggesting its effective protection against H₂O₂-induced apoptosis in ARPE-19 cells.

LBP alleviates H₂O₂-induced cell damage via the Nrf2/HO-1 pathway. To determine the molecular mechanism involved in this protection against H₂O₂-induced oxidative damage and apoptosis, a potential signaling effect induced by LBP upon Nrf2/HO-1 was investigated. Western blot analysis demonstrated that treatment with H₂O₂ increased the nuclear transcriptional expression of Nrf2 protein ($P<0.05$; Fig. 4A),

the main regulator of the cellular antioxidant response (8). Compared with the H₂O₂ group, pretreatment with LBP increased this nuclear transcriptional expression of Nrf2 protein, in a dose-dependent manner ($P<0.01$ or $P<0.001$; Fig. 4A). In addition, the downstream gene, HO-1 also exhibited a similar trend in expression to that of Nrf2 ($P<0.01$ or $P<0.001$; Fig. 4B). To further investigate the molecular mechanism of LBP on H₂O₂-induced ARPE-19 cell damage, treatment with LBP exhibited a negligible influence on the expression of nuclear Nrf2 and HO-1 (Fig. 4C). However, a statistically significant increase was observed in nuclear Nrf2 and HO-1 upon induction of damage by H₂O₂ ($P<0.01$; Fig. 4C), suggesting that the combination of LBP and Oxs contributed to increasing the nuclear translocation of the Nrf2 protein in a synergistic manner.

To determine the molecular mechanisms involved in this process, siRNA transfection was performed to silence Nrf2 expression. The results demonstrated that Nrf2 protein expression

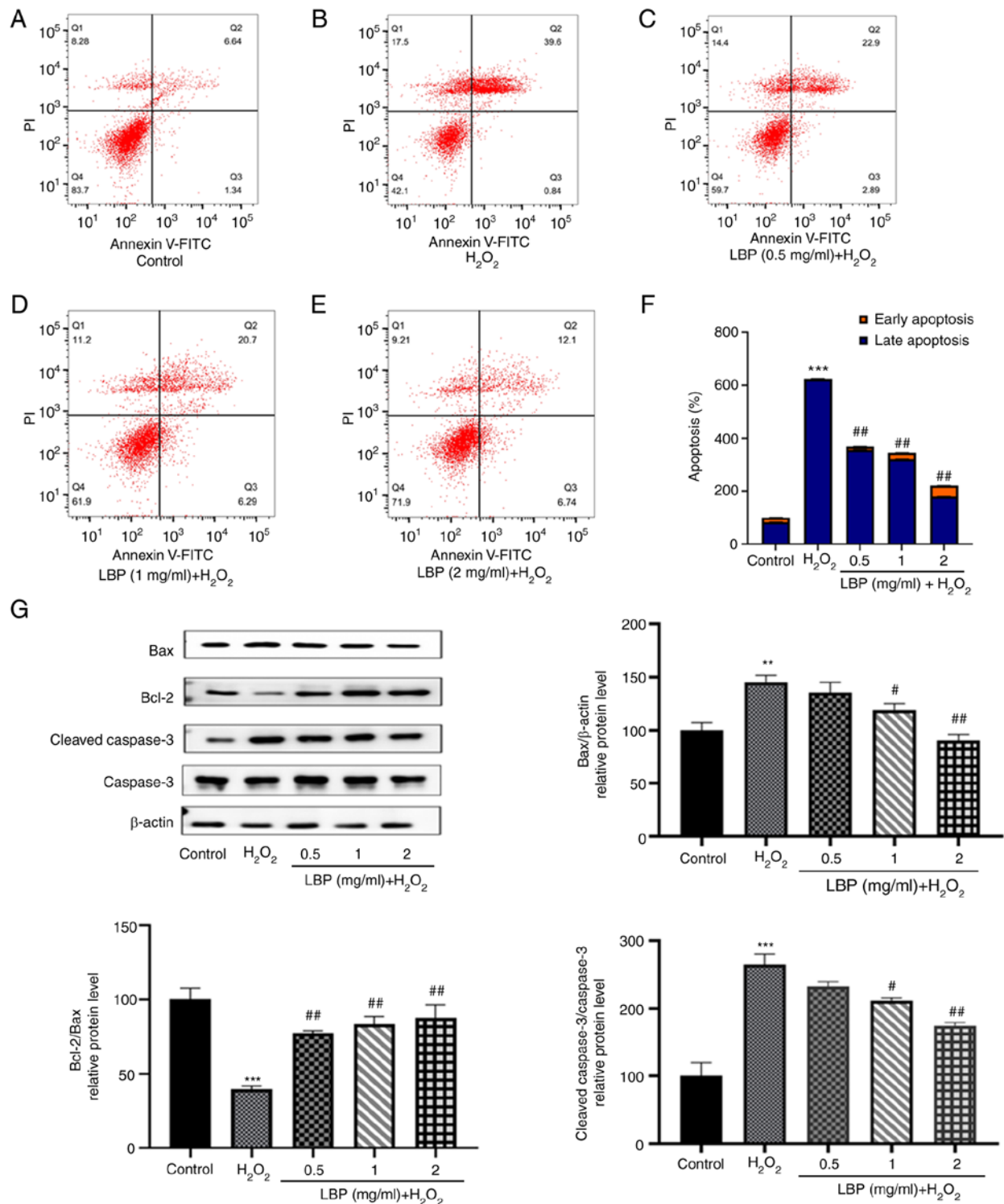


Figure 3. LBP protects cells from H₂O₂-induced apoptosis. (A-E) Apoptosis was detected via staining with Annexin V-FITC and PI. Flow cytometric analysis of ARPE-19 cells in each group. (F) Quantification of the extent of apoptosis in each group. (G) Western blot analysis was performed to detect the levels of apoptosis-related proteins (cleaved caspase-3, Bax and Bcl-2). Data are presented as the mean ± SEM (n=3). **P<0.01, ***P<0.001 vs. control; #P<0.01, ##P<0.001 vs. H₂O₂-treated cells with no LBP pretreatment. LBP, Lycium barbarum polysaccharide; PI, propidium iodide.

significantly decreased in ARPE-19 cells (P<0.01; Fig. 4D) and LBP-mediated expression of HO-1 was almost eliminated (P<0.001; Fig. 4D). In addition, transfection with Nrf2-siRNA enhanced H₂O₂-induced cell death, thereby offsetting the protection by LBP (P<0.05; Fig. 4E). Taken together, these results suggest that LBP activates the Nrf2/HO-1 pathway, and thus protects ARPE-19 cells from H₂O₂-induced cell damage (Fig. 5).

Discussion

RPE cells are critical for maintaining the structural integrity of the retina (16,17) and are particularly susceptible to the negative effects of OxS, and are generally exposed to high levels of ROS (18) due to the high oxygen demands of the retina. Previous studies (7,19,20) have reported the

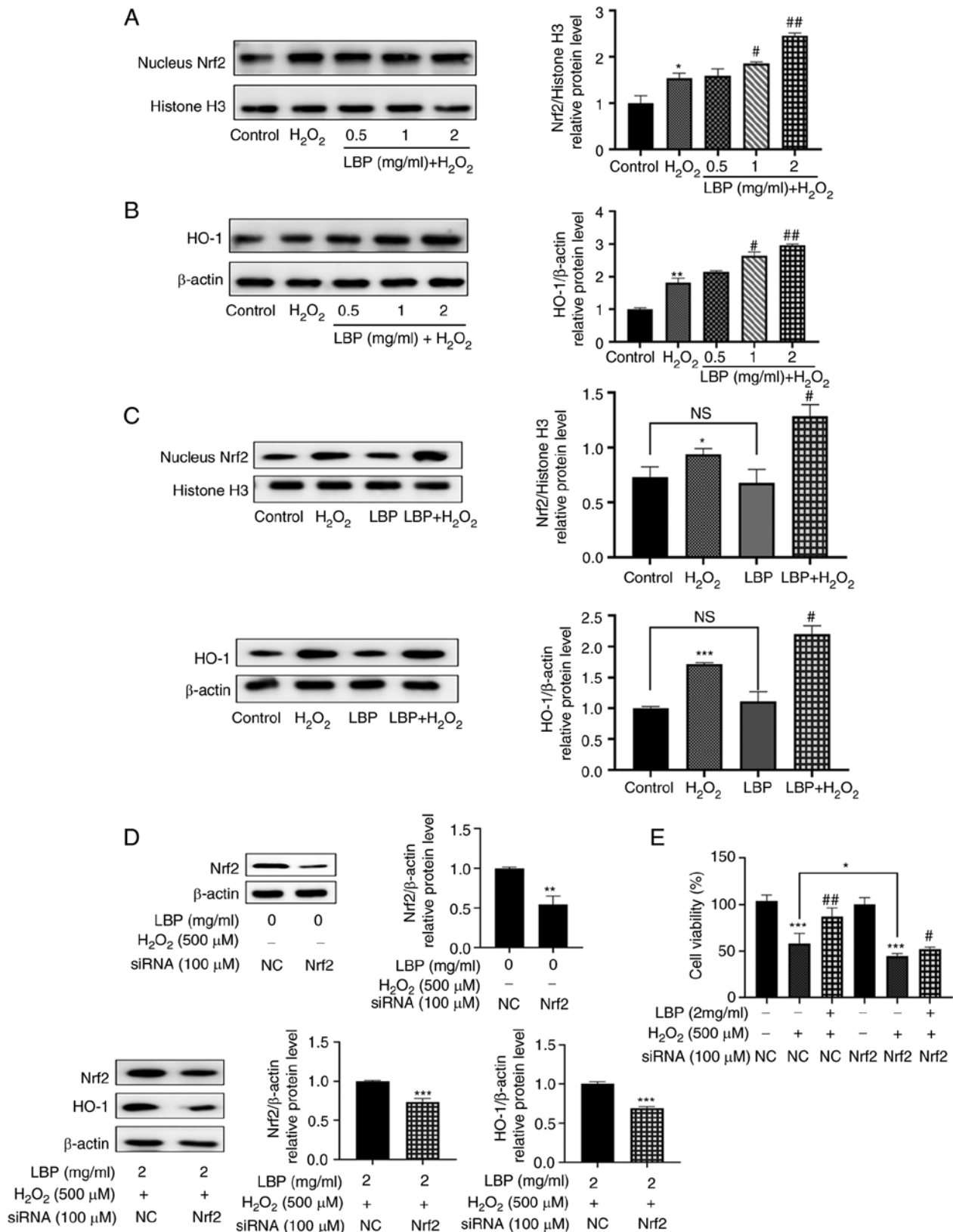


Figure 4. LBP alleviates H₂O₂-induced RPE cell damage via the Nrf2/HO-1 pathway. ARPE-19 cells were incubated in the presence or absence of LBP for 24 h, and subsequently treated with 500 μM H₂O₂ for 2 h. (A) The relative protein expression levels of nuclear Nrf2 were determined via western blotting. (B) The relative protein expression levels of HO-1 were determined via western blotting. (C) Western blot analysis was performed to detect the protein expression levels of nuclear Nrf2 and HO-1. (D) ARPE-19 cells were transfected with siRNA (NC or Nrf2) for 12 h, incubated with LBP for 24 h and subsequently treated with H₂O₂ for 2 h. Protein expression levels of Nrf2 and HO-1 were analyzed via western blotting. (E) ARPE-19 cells were transfected with siRNA (NC or Nrf2) for 12 h, incubated in the presence or absence of LBP for 24 h and subsequently treated with H₂O₂ for 2 h. The cytoprotective effect of LBP was analyzed via the Cell Counting Kit-8 assay. +, presence of H₂O₂ or LBP; -, absence of H₂O₂ or LBP. *P<0.05, **P<0.01, ***P<0.001 vs. control; #P<0.01, ##P<0.001 vs. H₂O₂-treated cells with no LBP pretreatment. LBP, Lycium barbarum polysaccharide; RPE, retinal pigment epithelium; Nrf2, nuclear factor erythroid 2-related factor 2; HO-1, heme oxygenase-1; si, small interfering; NC, negative control.

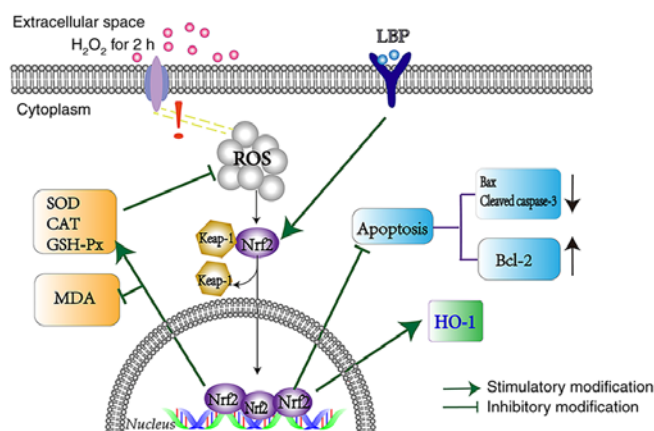


Figure 5. LBP protects ARPE-19 cells against H_2O_2 -induced oxidative stress via the Nrf2/HO-1 pathway. LBP, Lycium barbarum polysaccharide; Nrf2, nuclear factor erythroid 2-related factor 2; HO-1, heme oxygenase-1; SOD, superoxide dismutase; CAT, catalase; GSH-Px, GSH-peroxidase; MDA, malondialdehyde; ROS, reactive oxygen species.

association between ROS-induced damage in RPE cells and AMD, suggesting that early intervention is important for the prevention of OxS-induced damage. The anti-oxidative and anti-apoptotic functions of LBP have been reported in several eye diseases, including glaucoma (21), retinal ischemia-reperfusion injury (22) and diabetic retinopathy (23). Studies on the chemical composition of LBP have revealed that glycopeptides within its structure can alleviate lipid peroxidation (24-26). Thus, the present study aimed to investigate how LBP prevents OxS and apoptosis in ARPE-19 cells and its potential mechanism of action.

The present study used H_2O_2 to mimic the pathogenesis of AMD to determine the influence of LBP on OxS (27-29). The results of the CCK-8 assay demonstrated that treatment with 500 μM H_2O_2 significantly reduced the viability of ARPE-19 cells, the effects of which were reversed following pretreatment with LBP, in a concentration-dependent manner.

It has been reported that H_2O_2 -induced OxS is associated with increased ROS levels, which can be eliminated by enhancing the activity of antioxidant enzymes, thereby reducing the apoptotic state of aging RPE cells (30,31). The present study performed DCFH-DA staining to detect ROS levels, and flow cytometric analysis demonstrated that the fluorescence intensity of ROS in the H_2O_2 group significantly increased. Conversely, pretreatment with LBP reduced H_2O_2 -triggered ROS enhancement. The level of MDA was also consistent with the level of ROS, and the antioxidant levels in ARPE-19 cells, including SOD, CAT and GSH-Px, were maintained at high levels in response to LBP pretreatment.

Previous studies have demonstrated that the activation of apoptosis triggered by ROS represents a contributing factor for AMD pathogenesis (32,33). The effect of H_2O_2 exposure resulted in an increase in pro-apoptotic proteins (Bax and cleaved caspase-3), and a decrease in the anti-apoptotic protein, Bcl-2. However, 24 h of LBP pretreatment before H_2O_2 addition reversed the previously observed phenomenon, as shown by the reduced expression of Bax and cleaved caspase-3 and increased expression of Bcl-2.

Furthermore, Nrf2 is heavily involved in the process of cell redox homeostasis, which serves to reduce OxS by promoting the expression of antioxidant enzymes (34,35). However, few studies have focused on the association between LBP and the Nrf2 pathway during oxidative damage (36,37). It has been reported that once stimulated by OxS, Nrf2 becomes dissociated from Keap1 and translocates to the nucleus where it activates the HO-1 gene (38,39). The results of the present study demonstrated that LBP pretreatment alone did not increase nuclear translocation of Nrf2, and as a result, HO-1 expression was not affected. However, when OxS was induced, LBP increased the nuclear translocation of Nrf2 and HO-1 expression. Thus, H_2O_2 is essential for Nrf2 translocation and the expression of antioxidant proteins; these results are consistent with previous findings (40). The results of the present study demonstrated that transfection with Nrf2 siRNA partially reversed the protective effect of LBP on H_2O_2 -induced cell death. However, only one retinal cell line was assessed in the present study. Thus, other retinal cell lines and *in vivo* studies are required to verify the results presented here.

In conclusion, the results of the present study suggest that LBP exerts a protective effect on ARPE-19 cells, particularly by inhibiting H_2O_2 -induced OxS and apoptosis, enhancing antioxidant enzymes and activating the Nrf2/HO-1 pathway. Thus, LBP, a nutritional supplement (41), has the ability to reduce the risk of AMD and OxS-associated retinal disorders.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author upon reasonable request.

Authors' contributions

QZhuo and RL designed the present study. RL, MG and QZhuo performed the experiments. XH analyzed the data. YW helped perform the analysis with constructive discussions. XH and YW confirm the authenticity of all the raw data. All authors have read and approved the final manuscript.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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