

# The emerging role of CXCL10 in cancer (Review)

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**Abstract.** The chemokine interferon- $\gamma$  inducible protein 10 kDa (CXCL10) is a member of the CXC chemokine family which binds to the CXCR3 receptor to exert its biological effects. CXCL10 is involved in chemotaxis, induction of apoptosis, regulation of cell growth and mediation of angiostatic effects. CXCL10 is associated with a variety of human diseases including infectious diseases, chronic inflammation, immune dysfunction, tumor development, metastasis and dissemination. More importantly, CXCL10 has been identified as a major biological marker mediating disease severity and may be utilized as a prognostic indicator for various diseases. In this review, we focus on current research elucidating the emerging role of CXCL10 in the pathogenesis of cancer. Understanding the role of CXCL10 in disease initiation and progression may provide the basis for developing CXCL10 as a potential biomarker and therapeutic target for related human malignancies.

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## 1. Introduction

Chemokines are small, structurally related proteins which play a significant role in leukocyte trafficking (1) by producing chemotactic activity in cells expressing corresponding chemokine receptors. Based on the position of the first two conserved cysteine residues within the N-terminal, the chemokines are divided into two major (CX3C and CXC) and two minor (CC and C) subfamilies (2-4). The CX3C subfamily has three

intervening residues separating the two N-terminal cysteines, whereas the CXC subfamily only has one non-conserved amino acid residue separating the N-terminal cysteines. CC chemokines are those in which two cysteines are adjacent to each other, and a single known C chemokine lacks the first cysteine of the N-terminal pair. CXCL10 is a member of the CXC subfamily. Target cells of chemokines express corresponding receptors to which chemokines bind and mediate function (5). Therefore, the receptors of CC and CXC chemokine are referred to as CCRs and CXCRs, respectively. CC chemokines bind to CC chemokine receptors, and CXC chemokines bind to CXC chemokine receptors. Most receptors usually bind to more than one chemokine, and most chemokines usually bind to more than one receptor. CXCL10 specifically activates a receptor, CXCR3, which is a seven trans-membrane-spanning G protein-coupled receptor (6) predominantly expressed on activated T lymphocytes (Th1) (7), natural killer (NK) cells, inflammatory dendritic cells, macrophages and B cells (8,9). The interferon-induced angiostatic CXC chemokines, monokine induced by interferon (Mig/CXCL9) and interferon-inducible T-cell chemoattractant (I-TAC/CXCL11), also activate CXCR3. These CXC chemokines are preferentially expressed on Th1 lymphocytes (6,10,11).

Under proinflammatory conditions CXCL10 is secreted from a variety of cells, such as leukocytes, activated neutrophils, eosinophils (12), monocytes, epithelial cells, endothelial cells, stromal cells (fibroblasts) and keratinocytes in response to IFN- $\gamma$  (13,14). This crucial regulator of the interferon response, preferentially attracts activated Th1 lymphocytes to the area of inflammation and its expression is associated with Th1 immune responses (15-17). CXCL10 is also a chemoattractant for monocytes, T cells and NK cells.

CXCL10 is highly expressed in a diverse range of human diseases. It has been shown to be involved in the pathological processes of three main human disorders, infectious diseases, inflammatory (18-20) and autoimmune diseases (2), and cancer. Since CXCL10 plays a significant role in leukocyte homing to inflamed tissues, it exacerbates inflammation and causes significant tissue damage (2). Additionally, the CXC chemokines are a unique family of cytokines that either stimulate or inhibit angiogenesis depending on the presence of the structural domain of Glu-Leu-Arg; an ELR motif. CXCL10 is an ELR-negative CXC chemokine that attenuates angiogenesis and has anti-tumor actions (21-23). However, an increased expression of CXCL10 and its corresponding receptor CXCR3 have also been associated with advanced human cancers, including

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malignant melanoma (24), ovarian carcinoma (25), multiple myeloma (26), B-cell lymphoma (27) and basal cell carcinoma (14). In the central nervous system (CNS), microglia, astrocytes and even neurons express and secrete soluble CXCL10 (28,29). CXCL10 chemoattracts microglia cells into the circulation of CNS. CXCL10 and its receptor CXCR3 play a role in both peripheral and various CNS pathologies since the interference of CXCL10/CXCR3 signaling alters the initiation and progression in various CNS disease models (29,30). The increased production of CXCL10 in CNS has been associated with cerebral ischemia, epilepsy, brain inflammation and a number of neurodegenerative diseases such as multiple sclerosis (MS), Alzheimer's disease, amyotrophic lateral sclerosis, and human immunodeficiency virus encephalitis (29). The aim of this review is to focus mainly on the current understanding of the role of CXCL10 in cancer.

## 2. CXCL10 gene structure, function and signaling pathways

The human CXCL10 gene, was initially isolated in 1985 by Luster *et al* (31) while treating a lymphoma cell line (U937) with recombinant IFN- $\gamma$ . CXCL10 cDNA has an open reading frame of 1,173 bp containing 4 exons and encoding a protein of 98 amino acids with a molecular mass of 10,000 dalton. The primary translation product of CXCL10 is a 12-kDa protein and constitutes two internal disulfide cross bridges (13). The predicted signal peptidase cleavage generates a 10-kDa secreted polypeptide with four conserved cysteine residues in the N-terminal (13). The CXCL10 gene localizes on chromosome 4 at band q21, a locus associated with an acute monocytic/B-lymphocyte lineage leukemia exhibiting translocation of t (4; 11) (q21; q23). The CXCL10 protein shows significant homology in sequence with a family of proteins having chemotactic (platelet factor 4,  $\beta$ -thromboglobulin) and mitogenic (connective tissue-activating peptide HI) activities, which are associated with inflammation and cell proliferation (13,32). Human CXCL10 has a 63% homology in cDNA sequence with mouse CXCL10.

As with other chemokines, CXCL10 is a structurally specific protein, in which the potency and biological activities vary as a result of structural differences. The monomeric structure of CXCL10 exhibits a typical chemokine fold consisting of a three-stranded  $\beta$  sheet overlaid by an  $\alpha$  helix with a number of the receptor binding residues located in the associated loops stabilized by the disulfide bonds (3). A monomeric variant structure of CXCL10 was found by nuclear magnetic resonance spectroscopy. This new variant showed that the regions of the N-terminal and 40s loops within the CXCL10 molecule interplay with each other and form a hydrophobic cleft. This unusual structural characteristic of CXCL10 provides an explanation for the ability of CXCL10 to bind to both CXCR3 and CCR3 receptors (33). Another stereotypical oligomerization of CXCL10 provides an additional source of structural diversity (3) which is essential for CXCL10 to recruit activated T cells that bind to endothelial cells and subsequently trans-migrate *in vivo* (34). This hypothesis was supported by Campanella *et al* (34), who determined that CXCL10 knockout mice lose their ability to recruit activated CD8 $^{+}$  T cells into their airways due to the presence of the N-methyl group disrupting the interaction of hydrogen bonds between the main chains,

preventing the formation of dimers and oligomerization. *In vitro*, the N-methylated Leu27 monomeric mutants were capable of inducing CXCR3 internalization and the chemotaxis of CD8 $^{+}$  T cells expressing CXCR3, but this induction required at least ten times higher concentrations than wild-type CXCL10; heparin and CXCR3 binding were noted, but at greatly reduced efficacy. These results indicate that, *in vitro*, a considerably higher concentration of monomer mutant CXCL10 ligand is required to bind receptor CXCR3 and heparin (34). In an experiment designed by Swaminathan *et al* (3), CXCL10 molecules were found to exist in three different crystal forms: monomer, dimer and tetramer. In free solution, CXCL10 exists in monomer-dimer equilibrium, and tetrameric structures may represent species promoted by the binding of glycosaminoglycans (GAG). Findings of these authors suggest that only oligomeric forms of CXCL10 bind to endothelial and epithelial cells in a GAG-dependent manner (3). In agreement with the results of Swaminathan *et al*, a novel tetramer in the mouse CXCL10 structure has been discovered where two typical CXC chemokine dimers bind to their N-terminal regions to form a tetrameric assembly (35). Furthermore, the free N-terminal areas of two molecules at each terminal of the tetramer, enhance the probability of further attachment of molecules to generate higher order oligomers that may have functional relevance (35). The study by Jabeen *et al* (35) greatly contributes to the theory that the existence of CXCL10 in different oligomeric forms is crucial for its *in vivo* activity.

Functionally, CXCL10 exerts its biological effects by binding to CXCR3, and by inducing signaling effects in a paracrine or autocrine manner (14). CXCL10 induction depends predominantly on the carboxyl-terminal region of CXCR3, which is essential for CXCR3 internalization, chemotaxis and calcium mobilization induced by the CXCL10 ligand (8,36).

**Regulation of CXCR3 $^{+}$  cell chemotaxis.** CXCL10 performs 'homing' functions to chemoattract CXCR3-positive cells, including macrophages [microglia cell in CNS (37-39)], dendritic cells, NK cells and activated T lymphocytes (CD4 $^{+}$  T cells and CD8 $^{+}$  T cells) towards inflammatory, infectious and neoplastic regions. Consequently, CXCL10 is involved in modulating both innate and adaptive immunity, inducing tissue damage and contributing to tumorigenesis (37-39).

**Induction and variation in the conditions for CXCL10 induced apoptosis.** Using an *in vitro* model of cultured cortical neurons, neuronal CXCL10 expression recruits glial cells during embryogenesis, indicating that CXCL10 may be involved in apoptosis during the development of the nervous system (29,40). Alternatively, CXCL10 has been shown to facilitate cellular clearance of myeloid cells and strengthen the interaction between glial cells and neurons, which is a crucial step for synaptogenesis in the later stages of development of the nervous system (29). CXCL10 also significantly increased the apoptotic rate of cancer cells in cervical carcinoma (41).

**Promotion of cell growth and proliferation (14).** CXCL10, along with other CXC chemokines, binds to G-protein coupled receptors and induces a wide spectrum of biological and physiological activities. One of these activities involves the increase of cell growth and proliferation. CXCL10 colocalizes

lizes with the cell proliferation marker, cytokeratin 17 (K17) in tumor cells (14), whose proliferating actions are cell cycle dependent (42).

CXCL10 appears to have dual effects on cell growth. The proliferative or anti-proliferative action of CXCL10 appears to be cell-type-dependent; in other words, it may depend on the subtype of its receptor CXCR3. There are three CXCR3 splice variants: CXCR3-A, CXCR3-B and CXCR3-alt. Different cell types demonstrate various expression patterns. Additionally, various isoforms of CXCR3 induce the opposing actions of CXCL10 on proliferation. The main isoform, known as CXCR3-A, found in most cell types, codes for a protein of 368 amino acids (42) and couples with G $\alpha$ i to activate the ERK1/2, p38/MAPK, JNK and PI3-kinase/Akt signaling pathways, thereby inducing intracellular calcium influx, DNA synthesis and cell proliferation or chemotaxis (8,42-44). These types of cells include normal human bronchial epithelial cells (42), astrocytes, glioma cells (44), microglia cells (45), MDA-MB-231 breast cancer cells (46) and basal cell carcinoma (14). CXCR3-alt, which is known to co-express with CXCR3-A at a very low level (14,42), has not been found to be involved in cell growth.

*Inhibition of cell growth and proliferation.* The anti-proliferative action of CXCL10 is regulated by a variant isoform, CXCR3-B. CXCR3-B codes for a larger protein of 416 amino acids, couples with G $\alpha$ s to activate adenylyl cyclase and causes the inhibition of endothelial cell proliferation and migration (43,47,48). This appears to be the key mechanism by which CXCL10 exerts its antiproliferative activity. This receptor subtype does not induce chemotaxis (43,47). These types of cells are included in uterine endometrial cancer (46,49), glioblastoma (37), CCL-51 mammary tumor (50,51) and colorectal cancer (53). CXCR3+ T-cell migration into inflammatory and neoplastic regions attracted by CXCL10 along with CXCL9 and CXCL11 also contributes to anti-tumor progression and anti-metastasis (37). The variant CXCR3-B as a common receptor for all four angiostatic chemokines (CXCL4, CXCL9, CXCL10 and CXCL11) has enabled a better understanding of the role of CXC chemokines in the sequential participation of inflammatory cells and in the regulation of the inflammatory reaction resulting in angiostasis, and the inhibition of endothelial cell proliferation (53).

*Regulating angiostatic action.* CXC chemokines have dual effects on angiogenesis, depending on the presence of the Glu-Leu-Arg (ELR) motif. This well-established anti-proliferative (angiostatic) function, particularly on endothelial cells by CXCL10, has been shown to be regulated by the ELR motif. ELR-negative CXCL10 is an angiostatic chemokine that inhibits angiogenesis and is associated with its anti-tumor activities (21-23).

CXCL10 has cross talk with various typical signaling pathways. In breast cancer, Ras-induced CXCL10 overexpression is mediated through the Raf and PI3K signaling pathways, which may contribute to the development of breast tumors through cancer cell proliferation (46). In microglia cells, elevated CXCL10 expression occurs through p38/MAPK, JNK/MAPK and NF- $\kappa$ B cascades (45). In human airway

epithelial cells, p38/MAPK and PI3K signaling play a significant role in CXCL10/CXCR3 chemokine receptor-induced chemotaxis (54). In murine macrophage-like cells, activation of JAK1, JAK2/STAT1, but not the p38 pathway, up-regulates the expression of CXCL10, which is a strong inflammatory factor (55). The inhibition of CXCL10 expression in the cells by targeting the JAK/STAT1 signaling pathway may exert anti-inflammatory effects by attenuating the formation of chemokine CXCL10. Rabies virus (RV) stimulates CXCL10 expression in macrophages by activating extracellular signal-regulated kinases 1 and 2 (ERK1/2) (56). The RV-induced expression of CXCL10 in microglia in CNS was achieved by the activation of p38 and NF- $\kappa$ B pathways (57).

### 3. CXCL10 gene and cancer types

Interactions between chemokines and chemokine receptors were recently proposed to be of importance in the initiation and progression of cancer. CXCL10 has dual actions on tumorigenesis depending on the spliced variant of the corresponding CXCR3 receptor. CXCR3-B possesses growth-inhibitory properties, whereas CXCR3-A promotes cell proliferation (46).

*Anti-tumor effects through angiostatic action.* Various model systems have shown ELR-negative CXC chemokines to inhibit angiogenesis. In xenograft models of lymphoma, squamous cell carcinoma and adenocarcinoma of lung, the production of CXCL10 was inversely correlated with tumor growth, resulting in a marked reduction in tumor-associated angiogenesis. CXCL10 mediates its effects in T cell, macrophages- or NK-independent manner (22,23). CXCL10 may effect the suppression of angiogenesis associated with fibroblast growth factor (bFGF) in advanced uterine endometrial cancers (49). CXCL10 inhibits the growth of cervical carcinoma by down-regulating the formation of microvessels, the expression of proliferating cell nuclear antigens and the expression of human papillomavirus oncoproteins E6 and E7 through an increase in the apoptotic rate (41). In estrogen receptor-positive (ER $^{+}$ ) mammary tumors, CXCL10 inhibits vascular endothelial growth factor levels to reduce tumor burden (50).

*Antitumor effects through immunogenic action.* In the mouse glioblastoma model, CXCL10-mediated immunostimulation is likely to be responsible for the therapeutic efficacy rather than inhibiting vascularization (37). Immune modulation of CXCL10 has been widely used to modify dendritic cells to increase vaccine potency. Numerous investigators have confirmed that the CXCL10 gene has significant synergistic effects against tumors through its immunomodulatory properties by recruiting immature antigen-presenting, dendritic or early activated T cells into the tumor in murine glioma and the melanoma model (58-60).

Okada (61) utilized CXCL10 as a homing factor for cytotoxic T lymphocytes with a type 1 phenotype (Tc1) to attract cytotoxic T cells into CNS tumors, where cells durably exert antitumor effects in the CNS tumor. This author used type 1 polarizing DCs loaded with glioma-associated antigen peptides in combination with polyinosinic-polycytidylic acid stabilized by lysine and carboxymethylcellulose (poly-ICLC)

Table I. Comparative analysis of CXCL10 expression in various types of human cancers (Oncomine data).

	Fold change	P-value	No. of samples		References
			Normal	Cancer	
Bladder cancer	3.039	6.79E-11	48	81	Sanchez-Carbayo <i>et al</i> , (69)
Brain and CNS cancer	2.237	5.40E-12	23	81	Sun <i>et al</i> , (70)
	4.971	2.67E-4	3	84	Lee <i>et al</i> , (71)
Breast cancer	19.021	8.00E-12	7	40	Richardson <i>et al</i> , (72)
	5.072	1.71E-4	15	7	Karnoub <i>et al</i> , (73)
Cervical cancer	4.009	2.32E-5	22	20	Pyeon <i>et al</i> , (74)
Colorectal cancer	2.547	2.13E-8	41	50	Ki <i>et al</i> , (75)
Head and neck cancer	6.410	1.15E-11	13	41	Ginos <i>et al</i> , (76)
	14.448	1.49E-4	22	6	Pyeon <i>et al</i> , (74)
	3.761	5.41E-4	22	15	Pyeon <i>et al</i> , (74)
	3.073	1.28E-6	28	31	Talbot <i>et al</i> , (77)
Kidney cancer	12.873	3.10E-12	5	26	Yusenko <i>et al</i> , (78)
	5.447	5.90E-8	10	10	Gumz <i>et al</i> , (79)
	11.612	9.94E-11	11	32	Beroukheim <i>et al</i> , (91)
	5.897	4.41E-7	11	27	Beroukheim <i>et al</i> , (91)
Leukemia	3.596	2.40E-5	6	22	Andersson <i>et al</i> , (80)
	2.053	4.10E-5	6	84	Andersson <i>et al</i> , (80)
	-2.354	1.21E-4	14	39	Rosenwald <i>et al</i> , (85)
	-2.274	1.22E-4	6	11	Andersson <i>et al</i> , (83)
Liver cancer	17.693	1.42E-6	10	13	Wurmbach <i>et al</i> , (81)
	6.620	9.20E-5	10	17	Wurmbach <i>et al</i> , (81)
	5.928	1.78E-4	10	35	Wurmbach <i>et al</i> , (81)
Lymphoma	5.390	8.61E-9	14	38	Rosenwald <i>et al</i> , (82)
	5.656	6.66E-8	7	260	Rosenwald <i>et al</i> , (83)
	5.045	4.51E-4	6	5	Storz <i>et al</i> , (84)
	43.703	1.40E-17	25	28	Basso <i>et al</i> , (85)
	8.586	9.08E-7	25	32	Basso <i>et al</i> , (85)
	9.124	3.89E-5	25	17	Basso <i>et al</i> , (85)
Melanoma	5.651	2.52E-4	3	6	Haqq <i>et al</i> , (86)
<i>Other cancer</i>					
Testicular seminoma	2.614	5.49E-4	3	3	Skotheim <i>et al</i> , (87)
Parathyroid adenoma	2.434	1.73E-4	5	35	Morrison <i>et al</i> , (88)
Sarcoma	8.098	1.12E-4	15	9	Detwiller <i>et al</i> , (89)
Embryonal carcinoma	7.103	1.13E-7	6	15	Korkola <i>et al</i> , (90)
Seminoma	6.118	1.71E-6	6	12	Korkola <i>et al</i> , (90)
Mixed germ cell tumor	2.406	1.67E-6	6	41	Korkola <i>et al</i> , (90)

to induce IFN- $\alpha$  and CXCL10 in the CNS tumor micro-environment. In this experiment, Okada (61) successfully improved the survival of tumor-bearing mice without the generation of detectable autoimmunity. A phase I/II vaccination study based on Okada's concept is currently under way in patients with recurrent malignant glioma. Another obstacle for tumor antigen-specific T-cell immunity is the rapid down-regulation of chemokines, such as CXCL10,

resulting in a negative feedback mechanism. To solve this issue, Kang *et al* introduced the CXCL10 gene into DC2.4 cells using a retroviral system, resulting in the secretion of functionally chemoattractive CXCL10. Findings by these authors have laid the foundation for a future clinical translation of the chemokine-based genetic modification of DCs to increase their vaccine potency (62). CXCL10 has also been determined to have synergistic effects with a deoxycytidine



analog, gemcitabine, which inhibits the proliferation of endothelial cells, induces tumor cells apoptosis, and recruits lymphocytes to the tumor in murine models. Kang *et al* subsequently established an ideal model for the treatment of cancer by a combination of gene and chemokine therapy (63). A human study shows CXCL10 to be down-regulated in colorectal cancer (CRC) tissues with recurrence, indicating that CXCL10 may be utilized as a predictor of recurrence and as a prognostic indicator for survival in CRC patients (52).

**Tumor-promoting effects.** Contrary to tumor-limiting actions, CXCL10 exhibits tumor-promoting ability. Investigators have proposed that CXC chemokines and their receptors, particularly CXCR3 and its ligands CXCL10, CXCL9 and CXCL11, may be involved in tumor progression and metastasis through the overexpressions of CXCR3 in the tumor cells compared to the infiltrating immunocompetent cells, resulting in over-responsiveness to chemokines expressed either by tumors or inflammatory cells (24,64,65). In human breast cancer cell lines MDA-MB-435 and MCF-7, Ras induces CXCL10 over-expression by way of Raf and PI3 kinase signaling pathways. Overexpressed CXCL10 binds to CXCR3 and down-regulates CXCR3-B, promoting breast cancer growth (46). CXCL10 has also been reported as an autocrine invasion factor in nasal natural killer/T-cell lymphoma (66), which promotes colon cancer metastasis (67), and tumorigenesis in basal cell carcinoma (14) and human glioma (44).

**CXCL10 expression in human cancers from the Oncomine database.** CXCL10 information was summarized using the publicized microarray database Oncomine 4.3 (<https://www.oncomine.org/resource>). The P-value cut-off was 0.001, with a fold change threshold of 4. CXCL10 mRNA is up-regulated in the majority of human cancers, but is down-regulated in a limited number of cancers (68-90) (Table I).

#### 4. Conclusions

Although CXCL10 was originally identified as a proinflammatory chemokine that plays a role in leukocyte trafficking, it has been found not to only activate T lymphocytes (Th1) (7), but also NK cells, inflammatory dendritic cells, most macrophages and B cells. CXCL10 is capable of homing to target/threat regions. CXCL10 has multiple roles, such as modulating innate and adaptive immune response, regulating cell growth and angiostatic effects. CXCL10 induction is associated with numerous human disorders, and contributes to infectious diseases, chronic inflammatory and autoimmune diseases, and tumor formation. The features of CXCL10 make it a potential novel candidate for cancer target therapy. The relationship between the downstream and upstream signaling pathways should be investigated in order to develop CXCL10 as a novel therapeutic target in cancer and other human disorders.

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