

# Lysyl oxidase-like-1 enhances lung metastasis when lactate accumulation and monocarboxylate transporter expression are involved

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**Abstract.** The role that lysyl oxidase-like-1 (LOXL-1) may play in cancer metastasis due to its specific collagen accumulation characteristics has not been investigated extensively. This study was performed to examine the role of LOXL-1 in cancer metastasis. *In vitro* and *in vivo* cancer metastasis experiments were performed with B16F10 cells. Using the immunoblotting technique, the expression of LOXL-1, monocarboxylate transporter (MCT)1/2 and matrix metalloproteinase (MMP)2/9 was examined in a cell culture model and in primary and metastatic site samples from non-small cell lung carcinoma patients. Immunohistochemistry was also performed. According to immunohistochemical analysis of the non-small cell lung carcinoma patient samples, LOXL-1, MCT1/2 and MMP2/9 were expressed more highly in metastatic sites compared to primary sites. In *in vivo* studies, LOXL-1-overexpressing B16F10 cells yielded higher numbers of cancer nodules following their injection into mouse tail veins. Transfection of LOXL-1 siRNA into the cells prior to injection blocked lung metastasis. *In vitro*, the overexpression of LOXL-1 increased cell mobility and invasiveness, with increased extracellular accumulation of lactate at a low pH. The lactate transporter, MCT1/2, was highly expressed in LOXL-1-overexpressing cells. LOXL-1 knockdown through siRNA inhibited cell motility and invasiveness, showing relatively lower lactate accumulation and expression of MCT1/2 than under control conditions. This study elucidates

extracellular pH-associated matrix degradation as a potential mechanism for LOXL-1-induced cancer metastasis.

## Introduction

Metastasis, the spread of cancer cells from the primary tumor to distant organs, remains a major clinical challenge. The transition from a localized tumor to an invasive/metastatic tumor is a landmark in the development of malignant disease, since it is associated with a markedly worse prognosis (1,2). Most tumors/cancers develop a pathophysiological micro-environment characterized by hypoxic conditions, including low oxygen tension and elevated interstitial fluid pressure (3). The hypoxic condition stimulates anaerobic glycolysis, instead of oxygen-induced mitochondrial respiration. The alterations in glucose metabolism regulate cancer metastasis by causing high glucose uptake, high lactate formation and reduction of intra- and extra-tumoral pH (4-7). The acidic environment is produced by the activity of ion channels and transporters including Na<sup>+</sup>/H<sup>+</sup>, Cl<sup>-</sup>/HCO<sub>3</sub><sup>-</sup> exchangers, or the lactate transporter MCT (8,9). High lactic acid production is a common feature of numerous solid-tumor cells (10-12). MCT isoform 1 (MCT1) and MCT4 are highly activated in human cancer cells (13), suggesting that the level and/or affinity of proton/lactate export transporters were increased in cancer cells and that these transporters acted to prevent a lethal intracellular pH decrease, as tumors grow and become more acidic. To maintain the intracellular pH in cancer cells, extracellular pH becomes acidic through transporters such as MCT (14-16). In the acidic surroundings, the activities of extracellular matrix-degrading enzymes such as metalloproteinases, including MMP2/9, are also highly activated, revealing the correlation of acidic pH with MMP activities (3,17).

Lysyl oxidase-like protein-1 (LOXL-1), one of the lysyl oxidase (LOX) family proteins, was originally described as a copper-dependent amine oxidase responsible for catalyzing the cross-linking of collagens and elastins within the extracellular matrix (18). Previously, it was observed that, in epithelial tumors, the transition from a localized to an invasive/metastatic tumor is associated in many cases with the formation of

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fibrotic foci and desmoplasia (the presence of unusually dense collagenous stroma) within the primary tumor (19,20). These observations may represent apparent paradoxes as invasiveness has long been associated with the destruction of the extracellular matrix (ECM) by the ECM-degrading enzymes or metalloproteases (21). However, the deposition of excess ECM may in turn stimulate the expression of matrix-degrading enzymes that contribute to tumor invasion. Evidence suggests that an increase in ECM deposition affects the production of ECM-degrading enzymes (22). Although LOX and LOXL-1 were found to be expressed in areas of fibrogenesis in non-invasive *in situ* ductal breast carcinomas (23), metastatic cancer cell lines express LOX or LOXL-1, whereas non-metastatic cancer-derived cell lines do not (24). Furthermore, LOX-expressing cells exhibit increased invasiveness in *in vitro* invasiveness assays (24). In addition, LOXL-1 is highly expressed in metastatic breast cancer-derived cell lines (18). However, the relationship between LOX or LOXL-1 and cancer metastasis has not been clarified.

In this study, the role of LOXL-1 in cancer metastasis was examined. It was hypothesized that this role may be related to acidic extracellular pH, probably through extracellular lactate accumulation.

## Materials and methods

**Materials.** Monoclonal anti-LOXL-1 antibody was obtained from Professor Y. Kim (Wonkwang University, Iksan, Korea). The antibody against hemagglutinin antigen (HA) was purchased from Cell Signaling Technologies (Beverly, MA, USA). Dulbecco's modified Eagle's medium (DMEM), fetal bovine serum (FBS), trypsin and other tissue culture reagents were supplied by Life Technologies, Inc. (Grand Island, NY, USA). Bicinchoninic acid (BCA) protein assay reagents were obtained from Pierce Biotechnology (Rockford, IL, USA). All other chemicals were at least of analytical grade and were purchased from Sigma Chemical Company (St. Louis, MO, USA).

**Tumor metastasis model.** In the experimental lung metastasis model, B16F10 cells ( $5 \times 10^4$ ), which were transfected with a pcDNA3 vector, LOXL-1, or non-specific or LOXL-specific siRNA, were injected into the tail veins of C57BL/6 mice on day 0. Eighteen days after cell inoculation the mice were bled, sacrificed, and the lungs were removed, weighed, and fixed in formalin solution. Tumor nodules on the five lobes of the lungs were then counted macroscopically.

**Immunoblotting assays.** Equal amounts of the protein that was extracted from cells with RIPA buffer was fractionated by 10% SDS-PAGE. Blots were stripped, probed with antibodies including LOXL-1 antibody and reprobed with a polyclonal antibody against  $\beta$ -actin (Santa Cruz Biotechnology, Santa Cruz, CA, USA) to confirm equal protein loading and transfer.

***In vitro* invasion assay.** An *in vitro* invasion assay was performed using a 24-well transwell unit with polycarbonate filters (Corning Costar, Cambridge, MA, USA) as previously described (17). The lower side of the filter was coated with type I collagen and the upper side was coated with matrigel

(Collaborative Research, Lexington, KY, USA). The lower compartment was filled with serum-free medium containing 0.1% FBS. Cells were placed in the upper section of the transwell plate, incubated for 17 h, then fixed and stained with crystal violet dye for 10 min. Invasive phenotypes were determined by counting the cells that migrated to the lower side of the filter with microscopy at  $\times 400$ . Thirteen fields were counted for each filter and each sample was assayed in triplicate.

**Wound migration assay.** An injury line was made with a 2 mm wide tip on the cells plated in culture dishes at 90% confluence. After being rinsed with PBS, cells were allowed to migrate in complete medium and images were captured ( $\times 40$ ) at the indicated time points.

**Gelatin zymography.** Cells were cultured in serum-free DMEM for 48 h. Conditioned media were collected and centrifuged at 3,000 rpm for 10 min to remove cell debris. The protein concentration was measured using BCA protein assay reagents (Pierce, Rockford, IL, USA). Equal amounts of protein from the conditioned media were mixed with 2X Laemmli non-reducing sample buffer, incubated for 15 min at room temperature and then electrophoresed on 10% SDS-PAGE gels containing 1 mg/ml gelatin. Following electrophoresis, the gels were washed with 2.5% Triton X-100 three times for 30 min, rinsed for 15 min with 50 mM Tris-HCl buffer (pH 7.6) containing 5 mM  $\text{CaCl}_2$ , 0.02% Brij-35 and 0.2% sodium azide and incubated overnight at 37°C. The gels were stained with 0.5% Coomassie Brilliant Blue R-250 solution containing 10% acetic acid and 20% methanol for 30 min and de-stained with 10% acetic acid solution. Areas of gelatinase activity were detected as clear bands against the blue-stained gelatin background.

**Patients.** A total of 71 non-small cell lung carcinoma patients, including patients with tumors in 55 primary regions and 16 metastatic regions, were selected for this study. Metastatic region tissues were obtained from lymph nodes. The patients underwent surgical resection at the Chonbuk National University Hospital. Clinical information was obtained through a computerized tumor registry database. Samples derived from the National Biobank of Korea were obtained with informed consent, and approval was obtained from the institutional review board.

**Tissue microarray construction.** One representative formalin-fixed, paraffin-embedded block-containing tumor was selected for each of the 71 cases. The arrays were constructed using a 3 mm punch (Microm International, Korea). All 71 cases were reviewed using conventional hematoxylin and eosin-stained slides and the areas of interest were marked on each slide. The corresponding regions on the donor paraffin blocks were arrayed on recipient blank blocks. Histological typing was performed according to the histological classification of lung cancer by the World Health Organization.

**Immunohistochemistry.** Sections (5  $\mu\text{m}$ ) were cut from a tissue microarray block, de-paraffinized in xylene and rehydrated

in graded alcohols. Antigen retrieval was achieved by incubation of tissue sections in boiling 10  $\mu$ mol/l citrate buffer (pH 6.0) for 10 min in a microwave oven and immunostaining was undertaken using the avidin-biotinylated enzyme complex method (Dako, Denmark) with LOXL-1 antibody overnight at 4°C. Following incubation with the appropriate biotin-conjugated secondary antibody and subsequently with streptavidin solution, color development was performed using 3-amino-9-ethylcarbazole (AEC) (Lab Vision, CA, USA) as a chromogen. The sections were counterstained with hematoxylin. Appropriate negative controls, consisting of tissue sections of each case, processed without the addition of primary antibody, were prepared along with positive control sections. Tumors were considered to be positive for LOXL-1 when cytoplasmic reactivity was observed in >25% of the tumor cells at any intensity.

**Statistical analysis.** Associations between LOXL-1 expression and clinicopathological parameters were analyzed using the SPSS software package, version 11.5 (Seoul, Korea). P-values were determined via Chi-square tests.

## Results

**LOXL-1 is involved in lung cancer metastasis.** Table I shows immunohistochemistry results for LOXL-1. A total of 71 cases of lung carcinoma were available for analysis. Fifty-five tissue samples were obtained from primary sites and 16 tissues were obtained from metastatic lymph nodes. LOXL-1 expression was more frequently observed in the metastatic sites (81%) than in the primary sites (40%,  $p=0.004$ ). In primary regions, LOXL-1 expression was associated with histological type ( $p=0.014$ ); 14 of 23 squamous cell carcinomas (61%) and 6 of 24 adenocarcinomas (25%) were positive for LOXL-1. No correlations were found between LOXL-1 expression and tumor size or histological grade.

**LOXL-1 induces lung metastasis in vivo.** An *in vivo* study was used to investigate the involvement of LOXL-1 in cancer metastasis. In one set of experiments, lung metastasis was examined using a tail vein injection model. For this study, transfected stable B16F10 cells that highly express LOXL-1 (LOXL cells) were used. The expression of LOXL-1 is confirmed in the upper panel of Fig. 1A. The LOXL-1-expressing tumor cells produced a greater number of metastatic lung colonies than neomycin-resistant control vector-transfected tumor cells (Neo cells; Fig. 1A and B), but without altering tumor colony size (data not shown). The endogenous role of LOXL-1 was also investigated. The same *in vivo* experiments were performed with non-specific or LOXL-1 siRNA-transfected B16F10 cells. Decreased expression of LOXL-1 in LOXL-1 siRNA-transfected cells was confirmed (upper panel, Fig. 1C). The injection of the LOXL-1 knock-down cells significantly reduced the number of tumor nodules compared with the non-specific siRNA-transfected cell-injected model (Fig. 1C). The quantified number of nodules is shown in Fig. 1D.

**LOXL-1 induces cancer metastasis in vitro.** *In vitro* analysis of tumor cell migration was conducted using infiltration

Table I. Correlation between the expression of LOXL-1 and clinicopathological parameters in non-small cell lung carcinoma.

	No. of cases	LOXL-1 expression		p-value
		Negative	Positive	
Tumor size				0.435
≤3.0 cm	27	17	10	
>3.0 cm	28	16	12	
Histological type				0.019
Squamous cell carcinoma	23	9	14	
Adenocarcinoma	24	18	6	
Other	8	6	2	
Histological grade				0.301
Well	10	8	2	
Moderate	28	16	12	
Poor	16	8	8	
Lymph node metastasis				0.177
Negative	40	26	14	
Positive	15	7	8	
Tumor site				0.004
Primary site	55	33	22	
Metastatic site	16	3	16	

and wound healing assays. In the infiltration assay, Neo and LOXL cells were added to the upper surface of a matrigel-coated chamber in a medium containing 0.1% serum (Corning Costar, Cambridge, MA, USA) and were allowed to adhere as previously described (17). Invasion was measured over 17 h by placing the lower chamber in contact with medium containing 0.1% FBS. Cells were stained with Coomassie blue and the number of adherent cells was counted. In the infiltration assay, the quantification data clearly showed a significant increase in infiltrated cells in the LOXL cell chambers (Fig. 2A, lower panel). A complementary wound healing assay of *in vitro* cell migration was performed in which a 300  $\mu$ m-wide linear strip of cells was scraped from confluent monolayers using a pipette tip. In agreement with the infiltration assay, the wound healing assay revealed significantly increased wound closure in the LOXL cells, compared with the Neo cells at 3, 6, 9 and 12 h following scratching (Fig. 2B).

Increased glycolysis is a phenotypic trait almost invariably observed in human cancers with an acidic environment that is toxic to competitors but relatively harmless to the tumors themselves. Therefore, we compared the extracellular lactate accumulation between Neo and LOXL cells. The amount of lactate was time-dependently increased in the extracellular medium (Fig. 2C). Particularly in LOXL cells, the lactate increase pattern was significantly clearer than in the Neo cells. The extracellular pH value was also consistently lower in LOXL cells than in Neo cells following the culturing period of 36 h (Fig. 2D). The expression of

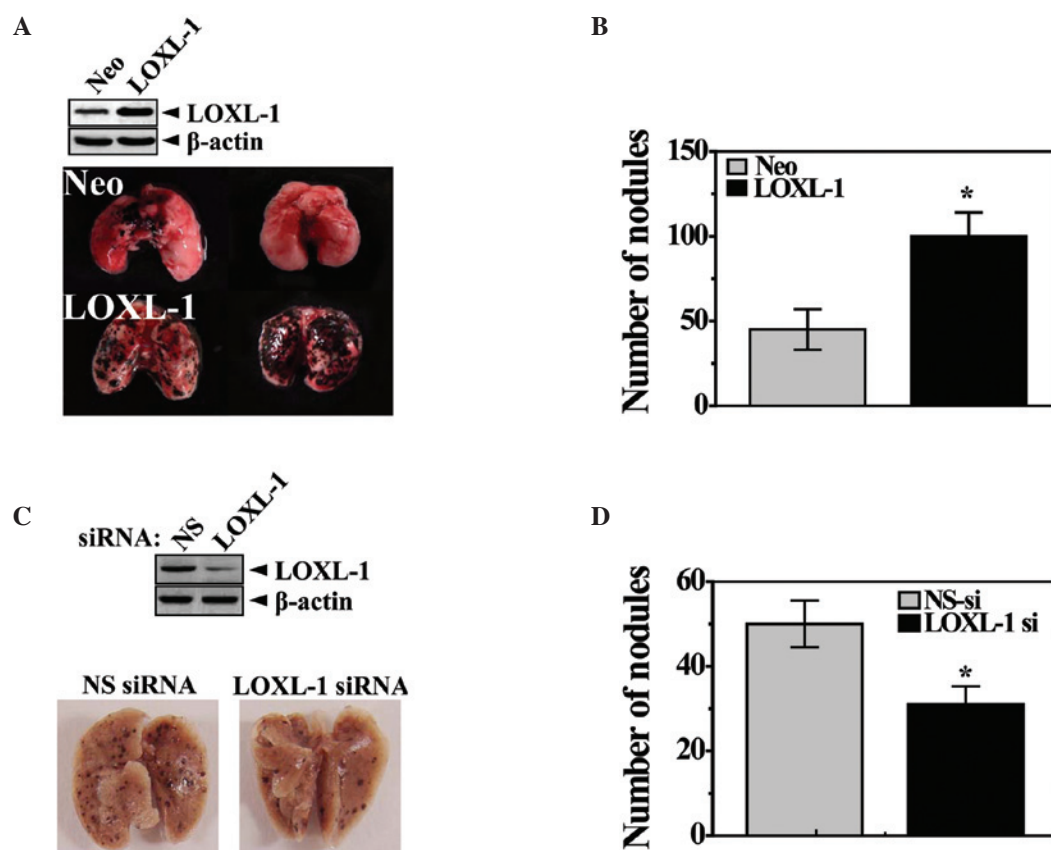
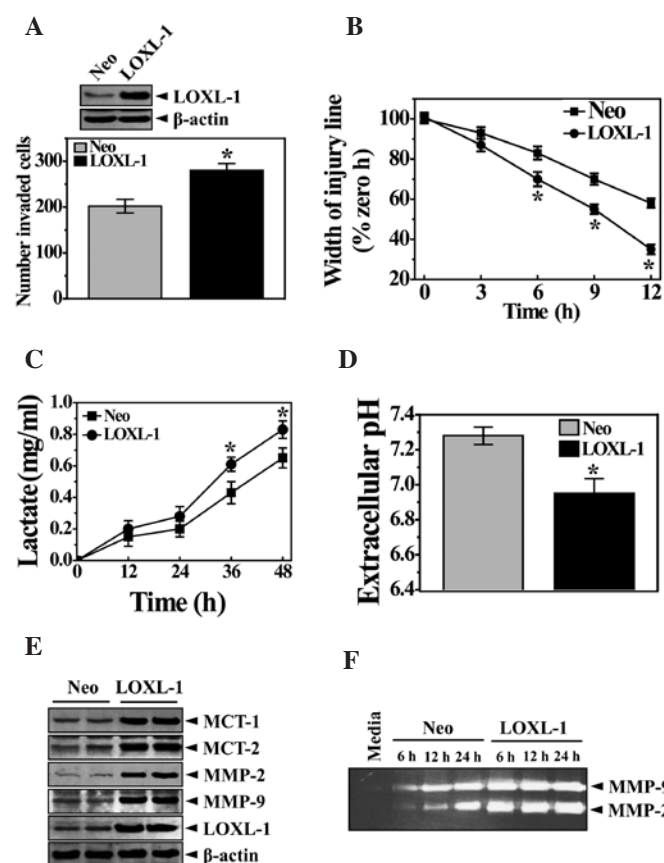


Figure 1. LOXL-1 induces lung metastasis *in vivo*. Neo and LOXL-1-transfected B16F10 cells (expression was confirmed, upper panel) were injected into mouse tail veins. Eighteen days later, (A) images of the lungs were captured and the nodules were quantified. (B) Non-specific or LOXL-1 siRNA-transfected B16F10 cells (expression confirmed, lower panel) were injected into mice tail veins. Eighteen days later, (C) images of the lungs were captured and the (D) nodules were quantified. Arrowheads, metastatic nodules in the lung. \* $P < 0.05$ , significantly different from the lung nodule number in the Neo cell-injected model. Neo, neomycin-resistant vector-transfected; NS-si, non-specific siRNA.



MCT1/2 was then assessed. MCT1/2 regulates the transport of the monoamine compounds including lactate to the extracellular surroundings, which explains how lactate is accumulated in the medium (25). As expected, the expression of MCT1/2 was high, along with MMP2/9, suggesting pH-associated ECM degradation and cancer metastasis (Fig. 2E and F). To show the endogenous role of LOXL-1 in cancer metastasis, *in vitro* analysis of tumor cell invasion was conducted with an infiltration assay using siRNA. The knockdown of LOXL expression is confirmed in Fig. 3A (upper panel). LOXL-1 siRNA-transfected cells revealed a significantly lower infiltration efficiency than non-specific siRNA-transfected cells (Fig. 3A, lower panel). The quantification data clearly showed the significance in the LOXL-1

Figure 2. LOXL-1 accelerates *in vitro* migration and invasion when lactate and MCT expression are involved. (A) Neo and LOXL cells that invaded through the matrigel-coated membrane were stained with 0.1% crystal violet. The amount was then determined by counting the stained cells in the lower well. (B) Through phase-contrast micrographs of scratched monolayers of Neo and LOXL-1-expressing cells at 0 and 15 h, the width of the injury line was quantified. (C) Lactate in the medium was measured when Neo and LOXL-1 cells were cultured for the indicated periods (12, 24, 36 or 48 h) as described in Materials and methods. (D) The extracellular pH of Neo and LOXL-1 cells was measured during the 36 h incubation period. (E) Western blotting was performed with anti-MCT1/2, MMP2/9, LOXL-1 or β-actin antibody. (F) Zymography was performed for MMP2/9 activity. \* $P < 0.05$ , significantly different from Neo cells. Neo, neomycin-resistant vector-transfected.



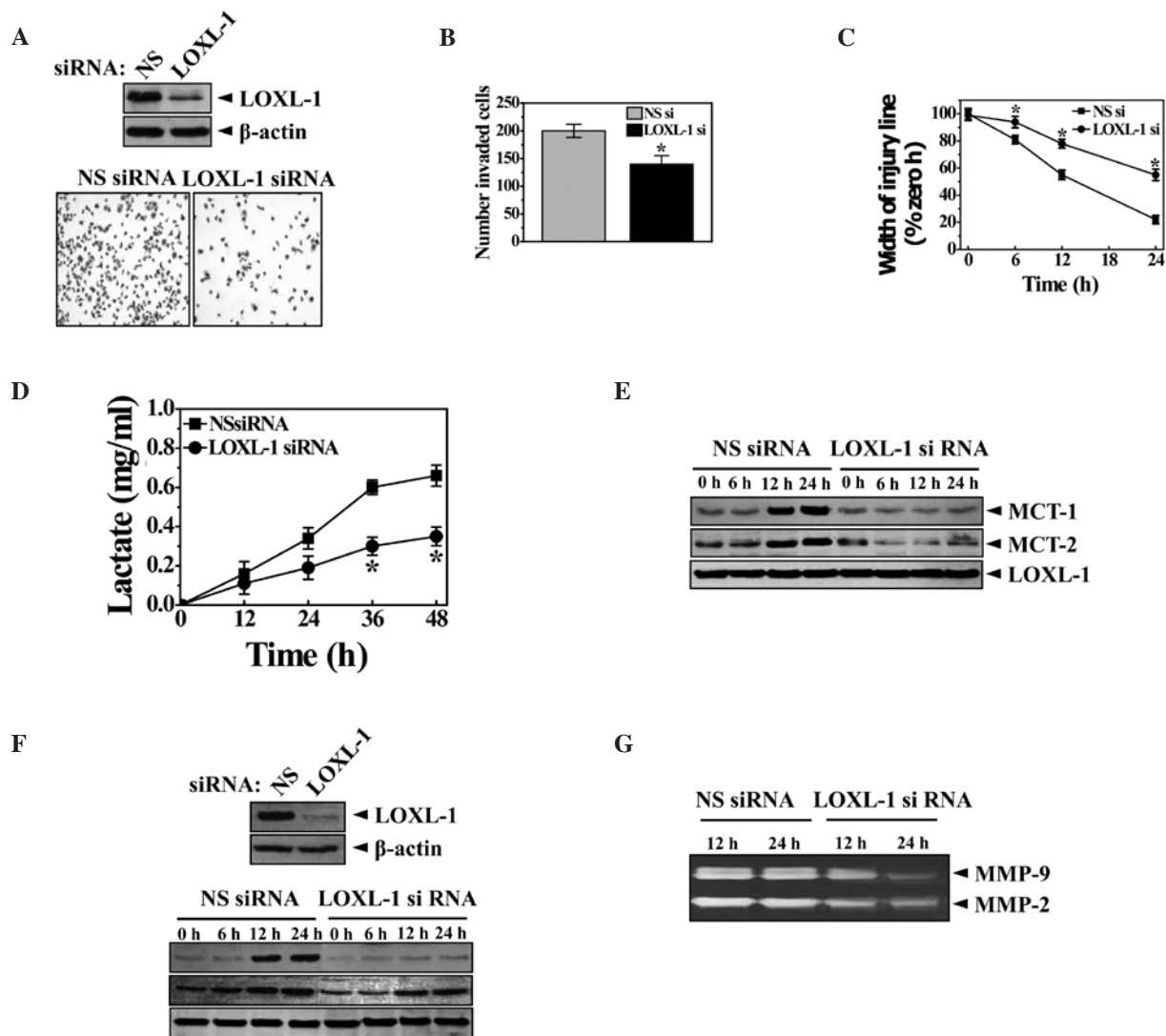


Figure 3. Endogenous LOXL-1 plays a significant role in the extracellular accumulation of lactate and the expression of its transporter, MCT1/2, in *in vitro* cancer metastasis. (A) HT1080 cells were transfected with non-specific or LOXL-1-specific siRNA and the expression of LOXL-1 was confirmed (upper panel). The invasion analysis was performed as in Fig. 2A. The 0.1% crystal violet staining cells in the lower chamber are shown (A, lower panel). The number was determined by counting cells in the lower wells. (B) The width of the injury line was quantified at the indicated culturing periods, 0, 6, 12, 18 or 24 h. (C) Lactate in the medium was measured when non-specific and LOXL-1 siRNA-transfected cells were cultured for the indicated periods (12, 24, 36 or 48 h) (D) Western blotting was performed with anti-MCT1/2, MMP2/9, LOXL-1 or  $\beta$ -actin antibodies. (E) Zymography was performed for MMP2/9 activity. (F) \* $P < 0.05$ , significantly different from the non-specific siRNA-transfected condition. NSsi, non-specific siRNA.

knock-down cells (Fig. 3B), suggesting that the endogenous expression of LOXL-1 is related to tumor cell invasion. A complementary wound healing assay of *in vitro* cell migration was also performed. In agreement with the infiltration assay (Fig. 3A and B), the wound healing assay revealed significantly reduced wound closure in LOXL-1 siRNA-transfected cells, compared with non-specific siRNA-transfected cells at 6, 12, 18 and 24 h after scratching (Fig. 3C). Lactate accumulation was also compared between non-specific and LOXL-1-specific siRNA-transfected cells. Consistent with data regarding LOXL-1 overexpressing cells (Fig. 2C and D), less lactate was accumulated in LOXL-1 than in non-specific siRNA-transfected cells (Fig. 3D). The expression of MCT1/2 was also negatively regulated in LOXL-1-specific siRNA-transfected cells (Fig. 3E). When non-specific and LOXL-1 siRNA-transfected cells were cultured, MMP2/9 expression was measured from a point (12 h after the transfection of siRNAs) that was marked at 0 h.

As expected, the MMP expressions were highly expressed in a time-dependent manner in non-specific siRNA-transfected cells than in LOXL-1 siRNA-transfected cells (Fig. 3F). The MMP2/9 activity was also measured using medium from the 12- and 24 h-culturing period, which was also counted from the 0 h point (12 h after the transfection of siRNAs). MMP2/9 was also more activated in non-specific siRNA-transfected cells than in LOXL-1 siRNA-transfected cells (Fig. 3G), demonstrating a consistent role of LOXL-1 on MMP2/9 expression and activity.

*LOXL-1 is highly expressed with MCT1/2 and MMP2/9.* Consistent with Table I, Fig. 4A shows the higher expression of LOXL-1 in metastatic cancer samples than in primary site samples. Consistent with *in vitro* data, MCT1/2 was more highly expressed in metastatic cancer samples compared with primary cancer samples. In cancer patients, the invasive phenotype of cancer cells is often associated with an

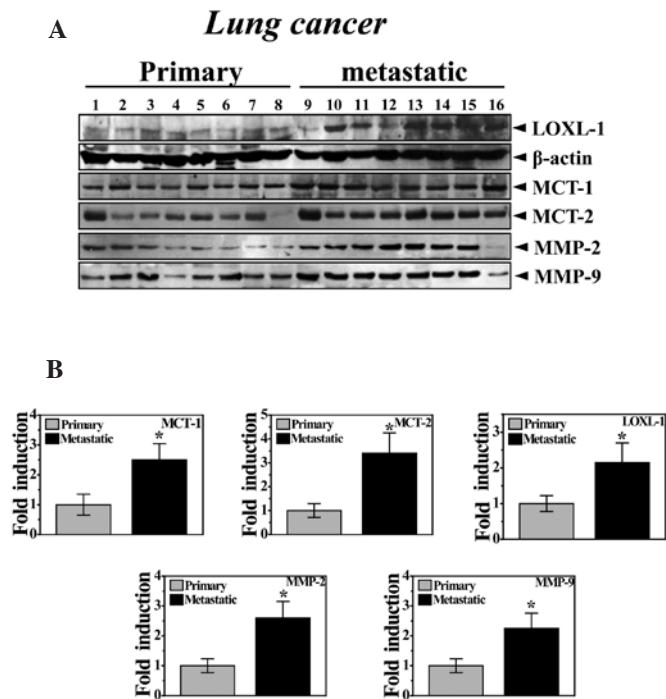


Figure 4. LOXL-1 is highly expressed in metastatic site samples and a correlation with MCT1/2 expression is confirmed. (A) Western blotting was performed with primary and secondary (metastatic) tumor samples from lung cancer patients using anti-LOXL-1, MCT1/2 or MMP2/9 antibodies. (B) The quantification for LOXL-1, MCT1/2 or MMP 2/9 was analyzed. \* $P < 0.05$ , significantly different from primary site samples.

increased expression of MMP-2 and/or MMP-9, which may degrade major structural collagen (26). As expected, MMP-2/9 was highly expressed in the samples from metastatic sites. Fig. 4B clearly shows the quantification of the expression of these proteins.

## Discussion

The present study provides evidence indicating that the expression of LOXL-1, one of the amine oxidases, is involved in non-small cell lung cancer metastasis. Results of the *in vitro* and *in vivo* studies also showed LOXL-1-associated cancer metastasis. In this study, MCT1/2 was suggested to be involved in LOXL-1-associated cancer metastasis through lactate accumulation and the consequential acidic extracellular pH where MMP2/9 was highly activated. Furthermore, among a total of 71 cases of lung carcinoma, LOXL-1 expression was more frequently observed in metastatic sites than in primary sites. Throughout this study, the role of LOXL-1 was clearly linked to cancer metastasis.

LOXL-1, similarly to LOX, plays a critical role in the formation and repair of the ECM by oxidizing lysine residues in elastin and collagen, thereby stabilizing the fibrous proteins (18,27). In addition to its fibrogenic function, LOX expression is associated with tumor suppression and tumor progression, and its role in tumorigenesis appears to be dependent on cellular location, cell type and transformation status (27-32). Microarray studies have shown that the expression of LOX is elevated in hypoxic human tumor cells (33), suggesting that the LOX expression is regulated by hypoxia-

inducible factor (HIF) and is associated with hypoxia in human breast, head and neck tumors (28). Furthermore, LOX secreted by hypoxic primary tumor cells accumulates with fibronectin at sites of future metastasis (34,35), suggesting a crucial role for LOX that is secreted by hypoxic tumor cells and is critically involved in a pre-metastatic state formation resulting in cancer metastasis. Possible mechanisms of LOXL-1-related cancer metastasis may also be linked to a hypoxia-associated mechanism. Although the data do not provide any direct evidence regarding hypoxia, indirect phenomena, including lactate accumulation, were demonstrated (Figs. 2C and 3D). Hypoxic status is observed in the areas around tumors due to high proliferation at the primary site, leading to vessel formation through angiogenesis factor activation and further leading to metastasis (4,9). Cancer cells usually facilitate glucose uptake, which is a diagnostic marker of cancer (5-7,9). The expression of lysyl oxidation enzymes is directly linked to collagen cross-linking at a specific site and step; however, the expression is associated with hypoxic status and hypoxia-associated signaling (28,33-35). Therefore, the expression of LOXL-1 may also be linked to hypoxic cancer status, leading to high glucose uptake and the resultant lactate production.

To assess the mechanism of LOXL-1-associated lactate accumulation, the expression of MCT1/2 was also shown. In LOXL-1 overexpressing cells, MCT1/2 was highly expressed (Fig. 2E). Furthermore, LOXL-1 knockdown inhibited the expression of MCT1/2, indicating the upstream role of LOXL-1 on MCT1/2 expression (Fig. 3E). MCT opening results in the extracellular accumulation of lactate and acidic extracellular surroundings (25). The cancer model showed the extrusion of lactate but not lactate uptake through MCT, which is correlated to lactate accumulation in the extracellular cancer environment (14). Considering that a number of human tumors are hypoxic (3,17), probably due to the compromised micro-circulation within tumors, LOX/LOXL-1 expression in hypoxic tumors may be associated with a more aggressive malignant phenotype and increased risk of metastasis.

In clinical sample analysis, cancer tissues from metastatic sites revealed higher expression of LOXL-1 than those from primary site tissues in non-small cell lung carcinoma patients (Fig. 4A and B). In accordance with LOXL-1, the lactate transporter MCT1/2 was highly expressed in the metastatic cancer samples. In this study, LOXL-1 is suggested as a cancer metastasis-associated protein, likely correlated to cancer hypoxic status-associated metastasis. Similarly, MCT1 is significantly increased in breast carcinomas compared with normal breast tissue and, significantly, MCT1 is associated with poor prognostic variables including high-grade tumors (36). MCT1 expression is also associated with advanced gastric carcinoma and lymph node metastasis (37).

This study also demonstrates a favorable correlation between MMP2/9 and MCT expression in LOXL-1-expressing cells (Fig. 2E and F). The critical pathological turning point in cancer is the initiation of local invasion, leading to the dissemination of tumor cells. Invasion is an active translocation of neoplastic cells across tissue boundaries through host cellular and extracellular matrix barriers (38,39), where MMPs are directly involved. Most invasive colonic, gastric, breast and ovarian carcinomas have high MMP expression, particu-

larly MMP-2 and MMP-9 (40). The evidence for MMPs as active contributors to cancer progression stems from animal studies (41). Furthermore, acidic media (pH 5.4-6.5) increase MMP activity in cell culture (3), in which expression correlates with tumor aggressiveness.

In conclusion, LOXL-1 is closely related to lung cancer metastasis, and MMP2/9 is also highly activated. MCT1/2, a lactate transporter, is involved in LOXL-1-associated cancer metastasis by contributing to an acidic extracellular environment. This study suggests that LOXL-1 is a potential drug target for cancer metastasis.

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