

Inhibition of MCF-7 breast cancer cell-induced platelet aggregation using a combination of antiplatelet drugs

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Abstract. Cancer metastasis is a highly coordinated and dynamic multistep process in which cancer cells interact with a variety of host cells. Morphological studies have documented the association of circulating tumor cells with host platelets. Tumor cell-induced platelet aggregation (TCIPA) contributes significantly to hematogenous metastasis; however, the molecular mechanisms involved in breast cancer TCIPA are poorly characterized. In this study, MCF-7 metastatic human breast cancer cells induced dose-dependent aggregation of washed platelets. Four major platelet activation pathways, glycoprotein (GP)-Ib-IX, GPIIb/IIIa, thromboxane (TX)-A₂ and adenosine diphosphate (ADP) were activated during TCIPA and were inhibited by their respective inhibitors, 7E3, SZ-1, aspirin and apyrase. Pretreatment of platelets with 7E3, SZ-1 or apyrase significantly inhibited TCIPA, while pretreatment with aspirin had no effect. Moreover, combined pretreatment of platelets with 7E3, SZ-1 and apyrase significantly inhibited TCIPA, compared to single inhibitors. Combinations of antiplatelet drugs may represent a promising strategy to prevent cancer metastasis.

Introduction

Breast cancer is the most frequent cause of mortality in females in the developed world. Although early detection,

precise resection using wide margins and systematic adjuvant therapy have improved survival, distant metastasis remains the leading cause of breast cancer-related mortality (1). Circulating tumor cells (CTCs) are isolated tumor cells that disseminate from the sites of metastatic and/or primary cancer, including breast cancer, and are identified and measured in peripheral blood (2). In a previous study, we observed that the rate of detection and number of CTCs correlated with the disease stage in breast cancer patients. Moreover, the assessment of CTCs in metastatic breast cancer patients predicts the efficacy of chemotherapy (1).

It is generally accepted that tumor cells become damaged during circulatory transport. This circulatory trauma may be due to humoral factors, including macrophages, natural killer cells and antibody-mediated complement lysis, as well as physical factors, including shear forces and mechanical trauma due to passage through the microvasculature. During hematogenous dissemination, CTCs undergo an extensive variety of interactions with host cells before they establish a secondary metastatic colony (3). The involvement of platelets in hematogenous metastasis has long been recognized. A correlation between venous thromboembolism and cancer was first observed by Trousseau in 1865 (4), while more recently, a study identified that the risk of cancer diagnosis is elevated following primary deep vein thrombosis (DVT) or pulmonary embolism (PE) (5). The ability of malignant tumor cells to aggregate platelets via tumor cell-induced platelet aggregation (TCIPA) (6,7) confers a number of advantages for the successful metastasis of cancer cells. When covered with a coat of platelets, a tumor cell acquires the ability to evade the body's immune system. Indeed, platelets protect tumors from tumor necrosis factor α -mediated cytotoxicity (8). TCIPA also enables embolization of the large tumor-platelet aggregates at new extravasation sites within the microvasculature (9). Additionally, platelets facilitate the adhesion of tumor cells to the vascular endothelium (10) and release a number of growth factors which stimulate tumor cell growth (11). Furthermore, platelets contribute to tumor-induced angiogenesis by releasing

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angiogenic growth factors, including vascular endothelial growth factor (VEGF) (12).

In the initiation phase of primary hemostasis, interaction of the glycoprotein (GP) Ib/V/IX receptor complex with von Willebrand factor (vWF) on the surface of platelets mediates the adhesion of platelets. Subsequently, several platelet-activating factors, including thromboxane A₂ (TXA₂) and adenosine diphosphate (ADP), are secreted in an autocrine/paracrine fashion and activate or prime approaching platelets. The release of ADP and TXA₂ also leads to the conversion of the GPIIb/IIIa receptor into an active form, which mediates platelet aggregation. Tumor cells may use a variety of mechanisms to interact with platelets and induce platelet aggregation. Tumor cells express cell surface molecules, which induce tumor cell-platelet interactions and subsequent platelet aggregation (13), or induce platelet aggregation by generating typical platelet agonists, including thrombin (14) or ADP (15), which are considered as 'soluble stimulators' of platelet aggregation.

Characterization of the interactions between platelets and metastasizing tumor cells could potentially be used to develop therapies to disrupt this correlation and prevent cancer metastasis (16). Therefore, in the present study, we investigated the activation of the GPIb-IX, TXA₂, ADP and GPIIb/IIIa pathways in platelets during TCIPA and determined the effect of inhibiting these pathways on TCIPA.

Materials and methods

Cell culture. MCF-7 human breast cancer cells (American Type Culture Collection, Manassas, VA, USA) were maintained in RPMI-1640 (Gibco, Grand Island, NY, USA) supplemented with 10% fetal calf serum (FCS; Hyclone Laboratories Inc., Logan, UT, USA), 100 U/ml penicillin and 100 mg/ml streptomycin at 37°C in a humidified atmosphere with 5% CO₂ and passaged every 2-3 days to maintain exponential growth. The study was approved by the Ethics Committee of the First Affiliated Hospital of Soochow University, Suzhou, China.

Reagents. Aspirin, apyrase and the anti-GPIb-IX complex monoclonal antibody, 7E3, were obtained from Sigma (St. Louis, MO, USA). The anti-GPIb-IX complex monoclonal antibody, SZ-1, was prepared according to our previously described methods (17). Fluorescein-isothiocyanate (FITC)-conjugated monoclonal antibody against high-affinity GPIIb/IIIa (PAC-1-FITC) was purchased from Becton Dickinson Biosciences (Mississauga, ON, Canada). Recombinant-phycoerythrin (PE)-conjugated monoclonal antibody against human platelet GPIb (CD42b-PE) was purchased from Dako Diagnostics (Glostrup, Denmark).

Preparation of washed platelets. Fresh blood obtained from healthy volunteers was anticoagulated with a 1/7 volume of acid-citrate dextrose (ACD; 85 mM trisodium citrate, 110 mM dextrose, 78 mM citric acid) as previously described (18), washed twice with CGS buffer (0.12 M sodium chloride, 0.0129 M trisodium citrate and 0.03 M D-glucose; pH 6.5), resuspended in freshly prepared Tyrode's buffer (18) and allowed to rest for at least 1 h at 37°C before use.

Platelet aggregation. The interactions between platelets and tumor cells were measured by light aggregometry using a whole-blood ionized calcium lumi-aggregometer (Chrono-Log, Havertown, PA, USA). Briefly, 200 μ l platelets (200x10⁶ cells/ml) were placed in the aggregometer and incubated for 2 min at 37°C with stirring at 900 rpm, prior to the addition of cancer cells. TCIPA was initiated by the addition of 50 μ l tumor cells (0.05-50x10⁶ cells/ml) and the reactions were monitored and analyzed using the Aggro-link data processing system (Chrono-Log) for up to 15 min. Platelet aggregation was expressed as a percentage of the maximum aggregation rate. The structure of the platelet-tumor cell aggregates were also observed using an Olympus CKX41 phase-contrast microscope (Olympus, Melville, NY, USA).

Flow cytometry analysis. The abundance of GPIb-IX and GPIIb/IIIa on the surface of the platelets during TCIPA was measured by flow cytometry. When TCIPA reached 50% maximal light transmission, the reaction was terminated by 10-fold dilution with physiological saline. The samples were then incubated with saturating concentrations (10 μ g/ml) of PE-anti GPIb-IX (CD42b-PE) or FITC-anti GPIIb/IIIa (CD41-FITC) in the dark for 5 min at room temperature and analyzed using a FC500 dual-laser five-color flow cytometer (Beckman Coulter, Fullerton, CA, USA). The mean fluorescence intensity was determined following correction for cell autofluorescence.

Enzyme-linked immunosorbent assay (ELISA). As TXA₂ quickly transforms into TXB₂ in aqueous solution, the concentration of TXB₂ was measured using an enzyme immunoassay kit (Amersham Pharmacia Biotech, Buckinghamshire, UK) (18). When TCIPA reached 50% maximal light transmission, the reaction was terminated by 10-fold dilution with physiological saline, centrifuged and the supernatants were assayed for the generation of TXB₂ using ELISA.

ADP assays. ADP secreted from dense granules in stimulated platelets was measured using a whole blood ionized calcium lumi-aggregometer as previously described (19). Briefly, platelets were incubated with luciferin-luciferase reagent (440 U/ml luciferase and 4 μ g/ml luciferin) for 2 min at 37°C to convert ADP to adenosine triphosphate (ATP) and to generate chemiluminescence. Following incubation, the agonist was added and luminescence was monitored. To quantify the generation of ATP by platelets, standard curves were constructed using standard ATP.

Statistical analysis. Each experiment was performed in triplicate, at least. Results are expressed as mean \pm standard deviation (SD). Statistical analysis was performed using unpaired Student's t-tests. P<0.05 was considered to indicate a statistically significant difference.

Results

MCF-7 cells induce platelet aggregation. The TCIPA effect of MCF-7 cells was observed using phase-contrast microscopy and quantified by light aggregometry. MCF-7 cells induced platelet aggregation in a similar manner to collagen, a classic

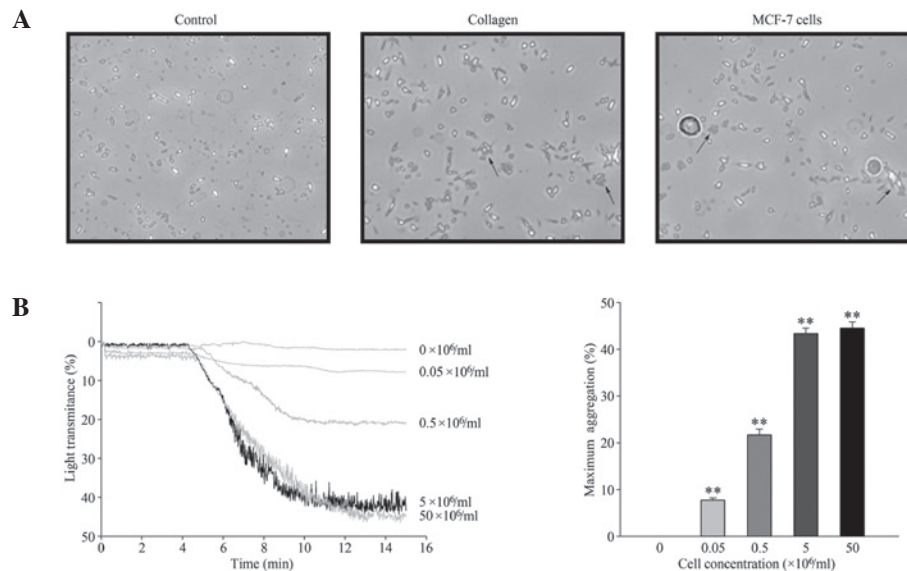


Figure 1. (A) Photomicrographs of MCF-7 cell-induced platelet aggregation. Untreated platelets were used as controls and collagen was used as a positive control to induce platelet aggregation. Platelet aggregation was triggered by MCF-7 cells at a concentration of 5×10^6 cells/ml. (B) Quantitative measurement of MCF-7 cell-induced platelet aggregation. MCF-7 cells induced platelet aggregation in a concentration-dependent manner. ** $P < 0.01$ vs. the control group.

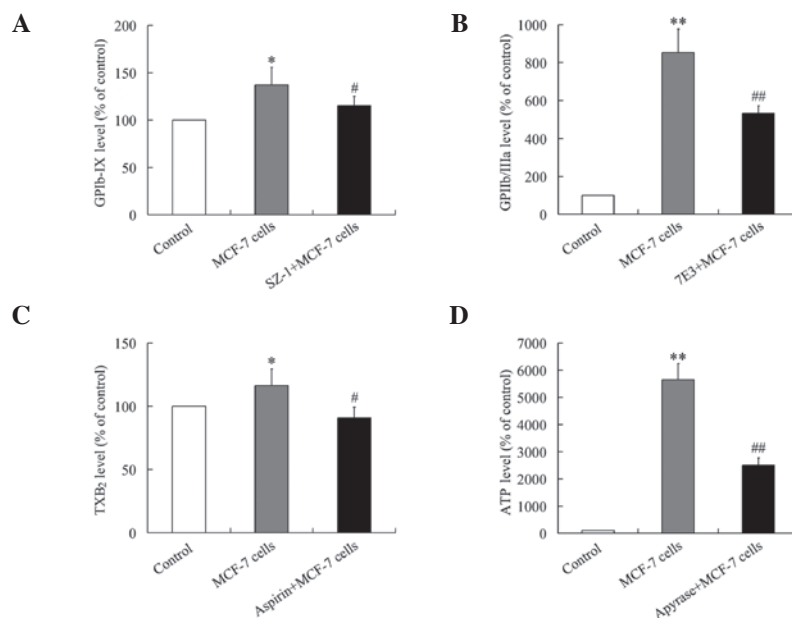


Figure 2. Activation of the GPIb-IX, TXA₂, ADP and GPIIb/IIIa pathways during MCF-7 cell-induced TCIPA. GPIb-IX (A) and GPIIb/IIIa (B) were upregulated on the surface of platelets during TCIPA. These effects were attenuated by pretreatment of the platelets with SZ-1 or 7E3, respectively. (C) The stable metabolite of TXA₂, TXB₂, was upregulated during TCIPA. This effect was attenuated by pretreatment of the platelets with aspirin. (D) ADP release increased during TCIPA. This effect was attenuated by pretreatment of the platelets with apyrase. * $P < 0.05$ and ** $P < 0.01$ vs. the control group; # $P < 0.05$ and ## $P < 0.01$ vs. the MCF-7 cell-treated group. GP, glycoprotein; TX, thromboxane; ADP, adenosine diphosphate; TCIPA, tumor cell-induced platelet aggregation.

inducer of platelet aggregation (Fig. 1A). MCF-7 cells induced platelet aggregation in a concentration-dependent manner (Fig. 1B), up to a maximal concentration of 5×10^6 cells/ml. Therefore, 5×10^6 cells/ml was selected as the standard cell concentration for all further experiments.

Activation of the GPIb-IX, TXA₂, ADP and GPIIb/IIIa pathways during TCIPA. Activation of GPIb-IX and GPIIb/IIIa were evaluated by quantifying the abundance of GPIb-IX and GPIIb/IIIa on the surface of platelets using flow cytometry.

As shown in Fig. 2A and B, GPIb-IX and GPIIb/IIIa were upregulated on the surface of platelets during TCIPA. These effects were repressed by pretreatment of the platelets with SZ-1 (10 μ g/ml) or 7E3 (20 μ g/ml), respectively.

TXA₂ release was measured as the level of the stable TXA₂ metabolite, TXB₂, using ELISA (19). As shown in Fig. 2C, the increased levels of TXB₂ observed during TCIPA were attenuated by pretreatment of the platelets with 50 μ g/ml aspirin.

ADP was measured as the level of ATP generated (20). As presented in Fig. 2D, the increased quantity of ADP released

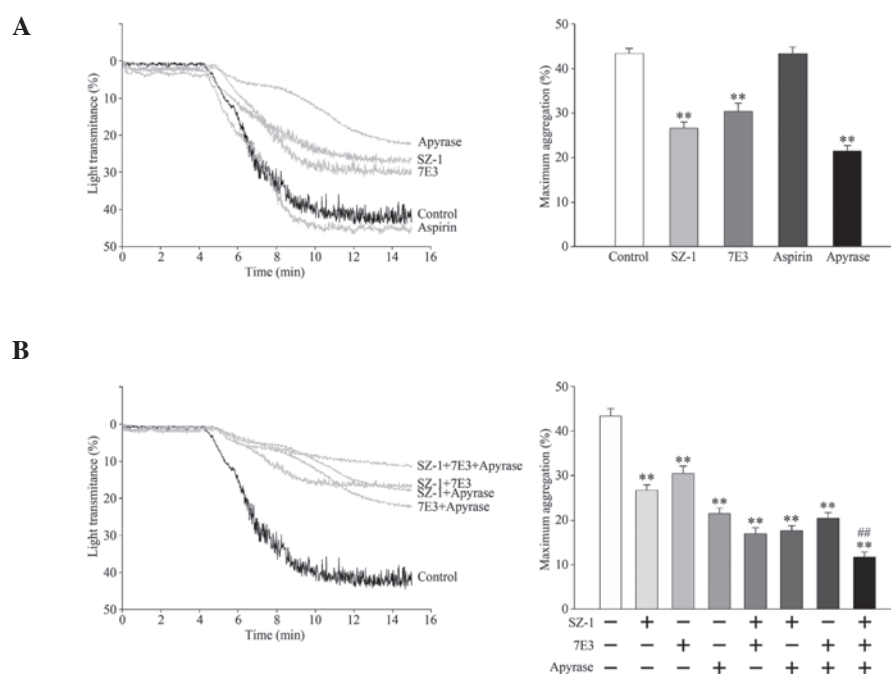


Figure 3. Effect of GPIb-IX, GPIIb/IIIa, TXA2 and/or ADP pathway inhibitors on MCF-7-induced platelet aggregation. (A) Platelets were pre-incubated for 5 min with SZ-1 (10 μ g/ml), 7E3 (20 μ g/ml), aspirin (50 μ g/ml) or apyrase (250 μ g/ml) before use in the MCF-7-induced platelet aggregation assays. (B) Inhibition of TCIPA using combinations of pathway inhibitors. ** P <0.01 vs. the control group; ## P <0.01 vs. all other groups. GP, glycoprotein; TX, thromboxane; ADP, adenosine diphosphate; TCIPA, tumor cell-induced platelet aggregation.

during TCIPA was inhibited by pretreatment of the platelets with 250 μ g/ml apyrase.

Thus, the GPIb-IX, TXA2, ADP and GPIIb/IIIa pathways were all activated during MCF-7-induced TCIPA and the activation of each pathway during TCIPA was repressed by pretreatment of the platelets with the appropriate inhibitors.

Repression of TCIPA using a combination of GPIb-IX, ADP and/or GPIIb/IIIa pathway inhibitors. To investigate whether activation of the GPIb-IX, TXA2, ADP and GPIIb/IIIa pathways participated in MCF-7-induced TCIPA, platelets were pretreated with the inhibitors of each pathway. As shown in Fig. 3A and B, aspirin did not exert any significant effect on MCF-7-induced TCIPA. However, SZ-1, 7E3 and apyrase inhibited TCIPA, suggesting that MCF-7-induced TCIPA was executed through the GPIb-IX, ADP and GPIIb/IIIa pathways, but not the TXA2 pathway.

The combined effects of these platelet-aggregation inhibitors on MCF-7-induced TCIPA were investigated further. As shown in Fig. 3, paired pretreatment of platelets with SZ-1, 7E3 and/or apyrase lead to a slightly greater inhibition of MCF-7 cell-induced TCIPA and the combination of all three inhibitors significantly inhibited MCF-7-induced TCIPA, compared to the single inhibitors alone.

Discussion

Metastasis is the major cause of mortality in breast cancer patients, yet there is no effective strategy to prevent tumor metastasis. Fewer than 0.01% of the cells that enter the bloodstream result in metastases (21). During hematogenous dissemination, the ability of circulating tumor cells to interact with platelets

via TCIPA is believed to promote tumor cell survival within the circulation (22) and increase the arrest of tumor cell emboli within the microcirculation (21), thereby facilitating metastasis. TCIPA is currently gaining acceptance as a key intermediate step in the process of blood-borne metastasis (23). Pre-clinical animal models have demonstrated that pharmacologically- or genetically-induced thrombocytopenia (6,24) and platelet function defects are associated with reduced metastasis (24-26). These observations have prompted the use of antiplatelet and anticoagulation agents to prevent metastasis in experimental models (7,26) and human cancer patients (27). Thus, in the present study, we investigated the mechanisms involved in MCF-7 breast cancer cell-induced TCIPA and the effect of antiplatelet strategies on MCF-7-induced TCIPA.

In the present study, we observed that the GPIb-IX and GPIIb/IIIa pathways were activated during MCF-7-induced TCIPA and inhibition of the GPIb-IX and GPIIb/IIIa pathways repressed MCF-7-induced TCIPA. GPIb-IX and GPIIb/IIIa are the major platelet surface transmembrane receptors implicated in TCIPA (28). GPIb-IX mainly mediates platelet adhesion (29), while GPIIb/IIIa plays an important role in platelet aggregation (30). A previous study supported the hypothesis that the functions of GPIb-IX in platelets, which support normal hemostasis or pathological thrombosis also contribute to tumor malignancy (25). A functional absence of GPIb-IX correlated with a 15-fold reduction in the number of lung metastatic foci in an animal model using B16F10 melanoma cells, demonstrates that platelet GPIb-IX contributes to experimental metastasis (25). Competitive inhibition of platelet GPIIb/IIIa, either pharmacologically or by using antibodies against GPIIIa (31,32) and knockout of the GPIIIa subunit in mice also diminished the formation of metastases (33).

TXA2 (34) and ADP (35,36) are considered to be 'soluble stimulators' of platelet aggregation. In this study, we observed that the TXA2 and ADP pathways are activated during MCF-7 cell-induced TCIPA. Several lines of evidence support the hypothesis that TXA2 plays an important role in tumor metastasis. Firstly, TXA2 is a potent stimulator of platelet aggregation (37), which promotes the binding of tumor cell-platelet aggregates to endothelial cells (26). Secondly, several types of tumor cells release large amounts of TXA2, compared to normal tissues, which potentiates tumor growth *in vitro* (38). Thirdly, TXA2 has been demonstrated to increase endothelial cell migration and angiogenesis in several *in vitro* and *in vivo* models (39). Fourthly, TXA2 potentiates tumor cell growth in culture and increases metastasis in animals (34). Finally, the use of TXA2 inhibitors has been shown to reduce metastasis in animals (26,40). However, aspirin, an inhibitor of the TXA2-mediated pathway exerted no significant effect on MCF-7 cell-induced TCIPA, suggesting that the TXA2 aggregation pathway is not required during MCF-7-induced aggregation, in agreement with previous studies (5,24,41).

ADP is a potent pro-aggregatory agent, which is released during TCIPA induced by fibrosarcoma, breast carcinoma and neuroblastoma cells (36,42,43), whereas the ADP scavenger apyrase (35,42), ADP receptor antagonist ticlopidine (36) and ADP receptor inhibitor 2-methylthio-AMP (43) inhibit TCIPA. These findings are in agreement with the results of this study, in which MCF-7-induced TCIPA leads to activation of the ADP pathway and MCF-7-induced TCIPA is inhibited by the ADP scavenger, apyrase. ADP-mediated platelet activation is associated with a net increase in the release of VEGF in healthy individuals, with no effect on the release of endostatin. VEGF release in response to ADP-mediated platelet activation is abolished by selective inhibition of the P2Y₁₂ receptor (44). Moreover, ADP depletion is associated with reduced formation of metastases *in vivo* (45) and improved biochemical control in prostate cancer patients receiving radiotherapy with curative intent (46).

We observed that combination of the GPIb-IX, GPIIb/IIIa and ADP pathway inhibitors exhibited a significant repression of TCIPA when compared with inhibition of a single pathway alone. Further studies are required to investigate the interactions between the GPIb-IX, GPIIb/IIIa and ADP pathways during TCIPA. The findings of the present study may be useful for the development of new clinical strategies to arrest TCIPA and prevent or reduce the formation of metastases.

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References

1. Tao M, Ma D, Li Y, Zhou C, Zhang Y, Duan W, Xu X, Wang R, Wu L and Liu H: Clinical significance of circulating tumor cells in breast cancer patients. *Breast Cancer Res Treat* 129: 247-254, 2011.
2. Swaby RF and Cristofanilli M: Circulating tumor cells in breast cancer: a tool whose time has come of age. *BMC Med* 9: 43, 2011.
3. Honn KV, Tang DG and Crissman JD: Platelets and cancer metastasis: a causal relationship? *Cancer Metastasis Rev* 11: 325-351, 1992.
4. Sørensen HT, Mellemkjaer L, Steffensen FH, Olsen JH and Nielsen GL: The risk of a diagnosis of cancer after primary deep venous thrombosis or pulmonary embolism. *N Engl J Med* 338: 1169-1173, 1998.
5. Bazou D, Santos-Martinez MJ, Medina C and Radomski MW: Elucidation of flow-mediated tumour cell-induced platelet aggregation using an ultrasound standing wave trap. *Br J Pharmacol* 162: 1577-1589, 2011.
6. Gasic GJ, Gasic TB and Stewart CC: Antimetastatic effects associated with platelet reduction. *Proc Natl Acad Sci USA* 61: 46-52, 1968.
7. Gasic GJ, Gasic TB, Galanti N, Johnson T and Murphy S: Platelet-tumor-cell interactions in mice. The role of platelets in the spread of malignant disease. *Int J Cancer* 11: 704-718, 1973.
8. Philippe C, Philippe B, Fouqueray B, Perez J, Lebreton M and Baud L: Protection from tumor necrosis factor-mediated cytotoxicity by platelets. *Am J Pathol* 143: 1713-1723, 1993.
9. Malik AB: Pulmonary microembolism. *Physiol Rev* 63: 1114-1207, 1983.
10. Mehta P: Potential role of platelets in the pathogenesis of tumor metastasis. *Blood* 63: 55-63, 1984.
11. Honn KV, Tang DG and Chen YQ: Platelets and cancer metastasis: more than an epiphenomenon. *Semin Thromb Hemost* 18: 392-415, 1992.
12. Verheul HM, Hoekman K, Lupu F, Broxterman HJ, van der Valk P, Kakkar AK and Pinedo HM: Platelet and coagulation activation with vascular endothelial growth factor generation in soft tissue sarcomas. *Clin Cancer Res* 6: 166-171, 2000.
13. Katagiri Y, Hayashi Y, Baba I, Suzuki H, Tanoue K and Yamazaki H: Characterization of platelet aggregation induced by the human melanoma cell line HMV-I: roles of heparin, plasma adhesive proteins, and tumor cell membrane proteins. *Cancer Res* 51: 1286-1293, 1991.
14. Tohgo A, Tanaka NG and Ogawa H: Platelet-aggregating activities of metastasizing tumor cells. III. Platelet aggregation as resulting from thrombin generation by tumor cells. *Invasion Metastasis* 5: 96-105, 1985.
15. Ugen KE, Mahalingam M, Klein PA and Kao KJ: Inhibition of tumor cell-induced platelet aggregation and experimental tumor metastasis by the synthetic Gly-Arg-Gly-Asp-Ser peptide. *J Natl Cancer Inst* 80: 1461-1466, 1988.
16. Nash GF, Turner LF, Scully MF and Kakkar AK: Platelets and cancer. *Lancet Oncol* 3: 425-430, 2002.
17. Du X, Beutler L, Ruan C, Castaldi PA and Berndt MC: Glycoprotein Ib and glycoprotein IX are fully complexed in the intact platelet membrane. *Blood* 69: 1524-1527, 1987.
18. Zhang G, Han J, Welch EJ, Ye RD, Voyno-Yasenetskaya TA, Malik AB, Du X and Li Z: Lipopolysaccharide stimulates platelet secretion and potentiates platelet aggregation via TLR4/MyD88 and the cGMP-dependent protein kinase pathway. *J Immunol* 182: 7997-8004, 2009.
19. Sakai H, Suzuki T, Murota M, Takahashi Y and Takeguchi N: Nitric oxide-induced Cl⁻ secretion in isolated rat colon is mediated by the release of thromboxane A₂. *J Physiol* 543 (Pt 1): 261-271, 2002.
20. Chung AW, Jurasz P, Hollenberg MD and Radomski MW: Mechanisms of action of proteinase-activated receptor agonists on human platelets. *Br J Pharmacol* 135: 1123-1132, 2002.
21. Tsuruo T and Fujita N: Platelet aggregation in the formation of tumor metastasis. *Proc Jpn Acad Ser B Phys Biol Sci* 84: 189-198, 2008.
22. Egan K, Crowley D, Smyth P, O'Toole S, Spillane C, Martin C, Gallagher M, Canney A, Norris L, Conlon N, *et al*: Platelet adhesion and degranulation induce pro-survival and pro-angiogenic signalling in ovarian cancer cells. *PLoS One* 6: e26125, 2011.
23. Swaim MW, Chiang HS and Huang TF: Characterisation of platelet aggregation induced by PC-3 human prostate adenocarcinoma cells and inhibited by venom peptides, trigramin and rhodostomin. *Eur J Cancer* 32A: 715-721, 1996.
24. Camerer E, Qazi AA, Duong DN, Cornelissen I, Advincula R and Coughlin SR: Platelets, protease-activated receptors, and fibrinogen in hematogenous metastasis. *Blood* 104: 397-401, 2004.
25. Jain S, Zuka M, Liu J, Russell S, Dent J, Guerrero JA, Forsyth J, Maruszak B, Gartner TK, Felding-Habermann B and Ware J: Platelet glycoprotein Ib alpha supports experimental lung metastasis. *Proc Natl Acad Sci USA* 104: 9024-9028, 2007.

26. Honn KV: Inhibition of tumor cell metastasis by modulation of the vascular prostacyclin/thromboxane A2 system. *Clin Exp Metastasis* 1: 103-114, 1983.
27. Zacharski LR, Henderson WG, Rickles FR, Forman WB, Cornell CJ Jr, Forcier RJ, Harrower HW and Johnson RO: Rationale and experimental design for the VA Cooperative Study of Anticoagulation (Warfarin) in the Treatment of Cancer. *Cancer* 44: 732-741, 1979.
28. Santos-Martinez MJ, Medina C, Jurasz P and Radomski MW: Role of metalloproteinases in platelet function. *Thromb Res* 121: 535-542, 2008.
29. Andrews RK and Berndt MC: Platelet physiology and thrombosis. *Thromb Res* 114: 447-453, 2004.
30. Shattil SJ, Kashiwagi H and Pampori N: Integrin signaling: the platelet paradigm. *Blood* 91: 2645-2657, 1998.
31. Amirkhosravi A, Mousa SA, Amaya M, Blaydes S, Desai H, Meyer T and Francis JL: Inhibition of tumor cell-induced platelet aggregation and lung metastasis by the oral GPIIb/IIIa antagonist XV454. *Thromb Haemost* 90: 549-554, 2003.
32. Trikha M, Zhou Z, Timar J, Raso E, Kennel M, Emmell E and Nakada MT: Multiple roles for platelet GPIIb/IIIa and α v β 3 integrins in tumor growth, angiogenesis, and metastasis. *Cancer Res* 62: 2824-2833, 2002.
33. Bakewell SJ, Nestor P, Prasad S, Tomasson MH, *et al*: Platelet and osteoclast β 3 integrins are critical for bone metastasis. *Proc Natl Acad Sci USA* 100: 14205-14210, 2003.
34. de Leval X, Benoit V, Delarge J, Julémont F, Masereel B, Pirotte B, Merville MP, David JL and Dogné JM: Pharmacological evaluation of the novel thromboxane modulator BM-567 (II/II). Effects of BM-567 on osteogenic sarcoma-cell-induced platelet aggregation. *Prostaglandins Leukot Essent Fatty Acids* 68: 55-59, 2003.
35. Boukerche H, Berthier-Vergnes O, Penin F, Tabone E, Lizard G, Bailly M and McGregor JL: Human melanoma cell lines differ in their capacity to release ADP and aggregate platelets. *Br J Haematol* 87: 763-772, 1994.
36. Bastida E, Escolar G, Almirall L and Ordinas A: Platelet activation induced by a human neuroblastoma tumor cell line is reduced by prior administration of ticlopidine. *Thromb Haemost* 55: 333-337, 1986.
37. Dogné JM, de Leval X, Delarge J, David JL and Masereel B: New trends in thromboxane and prostacyclin modulators. *Curr Med Chem* 7: 609-628, 2000.
38. Ogletree ML: Overview of physiological and pathophysiological effects of thromboxane A2. *Fed Proc* 46: 133-138, 1987.
39. Nie D, Lamberti M, Zacharek A, Li L, Szekeres K, Tang K, Chen Y and Honn KV: Thromboxane A(2) regulation of endothelial cell migration, angiogenesis, and tumor metastasis. *Biochem Biophys Res Commun* 267: 245-251, 2000.
40. Yokoyama I, Hayashi S, Kobayashi T, Negita M, Yasutomi M, Uchida K and Takagi H: Prevention of experimental hepatic metastasis with thromboxane synthase inhibitor. *Res Exp Med (Berl)* 195: 209-215, 1995.
41. Medina C, Jurasz P, Santos-Martinez MJ, Jeong SS, Mitsky T, Chen R and Radomski MW: Platelet aggregation-induced by caco-2 cells: regulation by matrix metalloproteinase-2 and adenosine diphosphate. *J Pharmacol Exp Ther* 317: 739-745, 2006.
42. Jurasz P, Sawicki G, Duszyk M, Sawicka J, Miranda C, Mayers I and Radomski MW: Matrix metalloproteinase 2 in tumor cell-induced platelet aggregation: regulation by nitric oxide. *Cancer Res* 61: 376-382, 2001.
43. Alonso-Escolano D, Strongin AY, Chung AW, Deryugina EI and Radomski MW: Membrane type-1 matrix metalloproteinase stimulates tumour cell-induced platelet aggregation: role of receptor glycoproteins. *Br J Pharmacol* 141: 241-252, 2004.
44. Bambace NM, Levis JE and Holmes CE: The effect of P2Y-mediated platelet activation on the release of VEGF and endostatin from platelets. *Platelets* 21: 85-93, 2010.
45. Uluckan O, Eagleton MC, Floyd DH, Morgan EA, *et al*: APT102, a novel adpase, cooperates with aspirin to disrupt bone metastasis in mice. *J Cell Biochem* 104: 1311-1323, 2008.
46. Choe KS, Correa D, Jani AB and Liauw SL: The use of anti-coagulants improves biochemical control of localized prostate cancer treated with radiotherapy. *Cancer* 116: 1820-1826, 2010.