# A chronic myeloid leukemia case with a unique variant Philadelphia translocation: t(9;22;21)(q34;q11;p12)

WALID AL-ACHKAR<sup>1</sup>, ABDULSAMAD WAFA<sup>1</sup>, FATEN MOASSASS<sup>1</sup> and THOMAS LIEHR<sup>2</sup>

<sup>1</sup>Molecular Biology and Biotechnology Department, Human Genetics Division, Atomic Energy Commission, Damascus, Syria; <sup>2</sup>Jena University Hospital, Institute of Human Genetics, Jena, Germany

Received May 26, 2011; Accepted July 25, 2011

## DOI: 10.3892/ol.2012.623

Abstract. The so-called Philadelphia (Ph) chromosome is present in more than 90% of chronic myeloid leukemia (CML) patients. Approximately, 5-10% of these patients show complex translocations involving a third chromosome in addition to chromosomes 9 and 22. Since at present the majority of CML cases are treated with imatinib, variant rearrangements do not exhibit specific prognostic significance. However, events of therapy resistance remain to be studied. In this study, we report a unique case of CML exhibiting an uncommon t(21;22)(p12;q11). This translocation has been characterized by fluorescence in situ hybridization (FISH) and array-proven multicolor banding (aMCB). Using specific probes for the BCR and ABL genes, results of FISH showed a three-way variant Philadelphia translocation (9;22;21)(q34;q11;p12) with a BCR/ABL fusion residing on the der(22) and the 3'BCR region translocated on the short arm of the derivative chromosome 21. In addition, the aMCB technique is significant in the detection of the breakpoints of genetic changes. The underlying mechanisms and prognostic significance of these changes are discussed.

## Introduction

Chronic myeloid leukemia (CML) is a myeloproliferative disease that originates in an abnormal pluripotent bone marrow stem cell and is consistently associated with the Philadelphia (Ph) chromosome, usually leading to a BCR/ABL gene fusion. The Ph chromosome produced as a result of t(9;22)(q34;q11) is observed in over 90% of cases, whereas variant Ph translocations are observed in 5-10% of cases (1). By standard cytogenetics, variant translocations have been

classified as simple when they involve the distal section of chromosome 22 and another chromosome distinct from chromosome 9, and as complex when chromosomes 9, 22 and at least one or more other chromosomes are involved (1). The BCR-ABL fusion gene is formed by the transposing of the 3' portion of the ABL oncogene from 9q34 to the 5' portion of the BCR gene on chromosome 22, and this fusion gene encodes a constitutively active tyrosine kinase (2). Imatinib mesylate (Glivec, formerly STI571) was designed specifically to inhibit the tyrosine kinase activity of the BCR/ABL protein and other tyrosine kinases, such as cABL, c-KIT and platelet-derived growth factor receptor (PDGF). By binding to an active site of the tyrosine kinase, Glivec switches off downstream signaling, cells are prevented from proliferating and apoptosis ensues (3). Various studies showed that a high efficacy of imatinib therapy achieves a complete or major cytogenetic response, i.e., a reduction to 0-34% Ph-positive cells. This positive effect is achieved in cases with a simple t(9;22) combined with complex translocations, resulting in BCR/ABL gene fusion, as well as in cases with clonal evolution (4,5).

In this case report, we present a unique translocation, t(21;22), which was further characterized by fluorescence *in situ* hybridization (FISH) and array-proven high-resolution multicolor banding (aMCB) as t(9;22;21)(q34;q11;p12) with a BCR/ABL fusion residing on the der(22) and the 3'BCR region translocated on the short arm of derivative chromosome 21, nonetheless successfully treatable with imatinib.

## Materials and methods

*Case report*. A 36-year-old male was diagnosed as suffering from CML in the chronic phase (CP). In August 2007, the white blood cell count (WBC) of the patient was 11.8x10<sup>9</sup>/l, constituting 53% neutrophils, 21% lymphocytes, 4% monocytes, 4% eosinophiles, 16% basophiles and 2% blasts. The platelet count was 118x10<sup>9</sup>/l and the hemoglobin level was 12.9 g/dl. A previous physical examination revealed splenomegaly. The patient was treated with imatinib mesylate at 400 mg/day for eight months in total, and the previous relevant symptoms appeared to have improved. The serum lactate dehydrogenase (LDH) level was 301 U/l (normal level up to 414 U/l) and serum alkaline phosphatase level was 94 U/l (normal level up to 90 U/l). In February 2008, the patient presented for the second time with a WBC of 54.5x10<sup>9</sup>/l consisting

*Correspondence to:* Dr Walid Al-Achkar, Molecular Biology and Biotechnology Department, Human Genetics Division, Atomic Energy Commission of Syria, P.O. Box 6091, Damascus, Syria E-mail: ascientific@aec.org.sy

*Key words:* chronic myeloid leukemia, variant Philadelphia chromosome, fluorescence *in situ* hybridization, high-resolution array-proven multicolor banding, imatinib mesylate

of 44% neutrophils, 11% lymphocytes, 1% monocytes, 29% basophiles and 15% blasts. The platelet count was 303x10<sup>9</sup>/l and the hemoglobin level was 13.5 g/dl. The serum LDH level was 403 U/l and the serum alkaline phosphatase level was 104 U/l. The patient was treated again with imatinib mesylate at 400 mg/day for 14 months in total. The patient was then lost during follow-up.

*Cytogenetic analysis.* Chromosome analysis using GTGbanding was performed according to standard procedures (6). A total of 20 metaphase cells derived from the unstimulated bone marrow of the patient were analyzed. Karyotypes were described according to the international system for human cytogenetic nomenclature (7).

*Molecular cytogenetics*. FISH using a LSI BCR/ABL dual color dual fusion translocation probe (Abbott Molecular/ Vysis, Des Plaines, IL, USA) was applied according to the manufacturer's instructions (6). aMCB sets based on microdissection-derived region-specific libraries for chromosome 9, 21 and 22 were applied as previously described (8,9). A total of 20 metaphase spreads were analyzed, using a fluorescence microscope (Axio Imager Z1 mot, Zeiss, Hertfordshire, UK) equipped with appropriate filter sets to discriminate between a maximum of five fluorochromes and the counterstain DAPI. Image capturing and processing were carried out using an ISIS imaging system (MetaSystems, Altlussheim, Germany) for the MCB evaluation.

## Results

Karyotyping was performed following the initiation of chemotherapy treatment, showing the following karyotypic changes.



Figure 1. GTG-banding revealed a complex karyotype involving one further chromosome besides chromosomes 9 and 22. Derivative chromosomes are shown by the arrowheads.

A complex karyotype 47,XY,t(9;22),der(21;22),+der(22) [3]\46,XY,t(9;22),der(21;22)[10]\46,XY,t(9;22)[7] was determined by GTG-banding (Fig. 1) and was further specified by molecular cytogenetic studies (Fig. 2). A dual-color-FISH using a probe specific for BCR and ABL revealed that a typical Ph chromosome with a BCR/ABL fusion gene was present. However, sections of chromosome 22 were present on a der(21) (Fig. 2A). Thus, aMCB using probes for the corresponding chromosomes was performed as previously reported (9). A complex translocation among the three chromosomes was detected (Fig. 2 B-D) and the final karyotypes obtained were: 47,XY,t(9;22)(q34;q11),der(21;22)



Figure 2. (A) Fluorescence *in situ* hybridization (FISH) using probes for BCR (green) and ABL (red) confirmed an involvement of chromosome 21 in the rearrangement, presence of the BCR/ABL translocation and Philadelphia (Ph) chromosome in this case. (B-D) Array-proven multicolor banding (aMCB) was applied to determine the involved breakpoints in this complex rearrangement. Each lane shows the results of aMCB analysis using probe sets for chromosomes 9, 21 and 22. The normal chromosomes are shown in the first column and the derivatives of the three chromosomes in the subsequent ones. The aMCB probe's unstained regions on the derivative chromosomes are shown in gray. *#*, chromosome; der, derivative chromosome; Ph, Philadelphia chromosome.

 $(p12;q11),+der(22)[3]\46,XY,t(9;22)(q34;q11),der(21;22)$  $(p12;q11)[10]\46,XY,t(9;22)(q34;q11)[7].$ 

#### Discussion

According to the literature, a number of other CML cases with t(9;22;21)(q34;q11;q22) (10-15), one case with t(9;22;21) (q34;q11;q21) (16) and one with t(9;22;21)(q34;q11;q11.2) (17) have been reported, respectively. To the best of our knowledge, only one case of Ph chromosome-positive CML with a unique translocation of three chromosomes t(9;22;21)(q34;q11;p12) was detected, and this translocation has yet to be observed at 21p12 in CML (18).

Chromosomes are known to be involved in variant rearrangements in CML (19). However, it has been suggested that the distribution of the break-points is non-random with the chromosomal bands most susceptible to breakage being: 1p36, 3p21, 5q31, 6p21, 9q22, 10q22, 11q13, 12p13, 17p13, 17q21, 17q25, 19q13, 21q22, 22q12 and 22q13 (19). However, the fusion gene remained on chromosome 22.

The review by Johansson *et al* (19) observed that a major breakpoint in chromosome 21 is q22, and q11 is very rare. The translocation with 21q22 is also common in other hematologic malignancies, whereas 21q11 has been reported in only a few cases of myelodysplastic syndrome (MDS), chronic lymphocytic leukemia (CLL) and acute myelogenous leukemia (AML) (20). However, CML with variant chromosomal abnormalities generally has a similar prognosis to that of cases with the typical t(9;22)(q34;q11) translocation (1). Further patient studies with involvement of 21p12 are required in order to establish prognosis in such cases.

In conclusion, we reported a unique case of a Ph chromosome-positive CML in the CP with a new variant Ph translocation involving three chromosomal aberrations 9q34, 21p12 and 22q11, and the 3'BCR region translocated on the short arm of derivative chromosome 21, which has not previously been described. Of note is that the patient had a favorable response to imatinib.

#### Acknowledgements

We thank Professor I. Othman, the Director General of Atomic Energy Commission of Syria (AECS) and Dr N. Mirali, Head of the Molecular Biology and Biotechnology Department for their support. This study was supported by the AECS and in parts by the Stefan-Morsch-Stiftung, Monika-Kutzner-Stiftung and the DAAD (D/07/09624).

#### References

- O'Brien S, Thall PF and Siciliiano MJ: Cytogenetics of chronic myelogeneous leukemia. Baillie'res Clin Hematol 10: 259-276, 1997.
- ShtiveIman E, Lifshitz B, Gale RP and Canaani E: Fused transcript of abl and bcr genes in chronic myelogenous leukemia. Nature 315: 550-554, 1985.
- Griffen J: The biology of signal transduction inhibition: basic science to novel therapies. Semin Oncol 28: 3-8, 2001.
- Kantarjian H, Sawyers C, Hochhaus A, Guilhot F, Schiffer C, Gambacorti-Passerini C, Niederwieser D, Resta D, Capdeville R, Zoellner U, Talpaz M, *et al*: International STI571 CML Study Group: Hematologic and cytogenetic responses to imatinib mesylate in chronic myelogenous leukemia. N Engl J Med 346: 645-652, 2002.

- Cortes JE, Talpaz M, Giles F, O'Brien S, Rios MB, Shan J, Garcia-Manero G, Faderl S, Thomas DA, Wierda W, Ferrajoli A, *et al*: Prognostic significance of cytogenetic clonal evolution in patients with chronic myelogenous leukemia on imatinib mesylate therapy. Blood 101: 3794-3800, 2003.
- Al-Achkar W, Wafa A and Nweder MS: A complex translocation t(5;9;22) in Philadelphia cells involving the short arm of chromosome 5 in a case of chronic myelogenous leukemia. J Exp Clin Cancer Res 26: 411-415, 2007.
- Shaffer L, Slovak M, Cambell L (eds.): ISCN (2009): An International System for Human Cytogenetic Nomenclature. S. Karger, Basel, 2009.
- Weise A, Mrasek K, Fickelscher I, Claussen U, Cheung SW, Cai WW, Liehr T and Kosyakova N: Molecular definition of high-resolution multicolor banding probes: first within the human DNA sequence anchored FISH banding probe set. J Histochem Cytochem 56: 487-493, 2008.
- Liehr T, Heller A, Starke H, Rubtsov N, Trifonov V, Mrasek K, Weise A, Kuechler A and Claussen U: Microdissection based high resolution multicolor banding for all 24 human chromosomes. Int J Mol Med 9: 335-339, 2002.
- El-Zimaity MM, Kantarjian H, Talpaz M, O'Brien S, Giles F, Garcia-Manero G, Verstovsek S, Thomas D, Ferrajoli A, Hayes K, *et al*: Results of imatinib mesylate therapy in chronic myelogenous leukaemia with variant Philadelphia chromosome. Br J Haematol 125: 187-195, 2004.
- Bartram CR, Anger B, Carbonell F and Kleihauer E: Involvement of chromosome 9 in variant Ph1 translocation. Leuk Res 9: 1133-1137, 1985.
- Guillaume B, Ameye G, Libouton JM, Dierlamm J, Vaerman JL, Straetmans N, Ferrant A, Verellen-Dumoulin C and Michaux L: Chronic myeloid leukemia with a rare variant Philadelphia translocation: t(9;22;21)(q34;q11;q22). Cancer Genet Cytogenet 116: 166-169, 2000.
- Mancini M, Nanni M, Cedrone M, De Cuia MR, Rondinelli MB, Malagnino F and Alimena G: Application of fluorescence in situ hybridization in defining a complex t(9;21;22) Ph formation. Haematologica 79: 536-539, 1994.
- 14. Vallcorba I, García-Sagredo JM, San Román C, Ferro MT, González A, Cabello P and Villegas A: Translocation (9;22;21) in a chronic myeloid leukemia fluorescence in situ hybridization definition. Cancer Genet Cytogenet 104: 72-73, 1998.
- Zhang J, Meltzer P, Jenkins R, Guan XY and Trent J: Application of chromosome microdissection probes for elucidation of BCR-ABL fusion and variant Philadelphia chromosome translocations in chronic myelogenous leukemia. Blood 81: 3365-3371, 1993.
- 16. Calabrese G, Stuppia L, Franchi PG, et al: Complex translocations of the Ph chromosome and Ph negative CML arise from similar mechanisms, as evidenced by FISH analysis. Cancer Genet Cytogenet 78: 153-159, 1994.
- Genet Cytogenet 78: 153-159, 1994.
  17. Takeuchi M, Katayama Y, Okamura A, Yamamoto K, Shimoyama M and Matsui T: Chronic myeloid leukemia with a rare variant BCR-ABL translocation: t(9;22;21)(q34;q11.2;q11.2). Cancer Genet Cytogenet 179: 85-87, 2007.
- Mitelman F, Johansson B and Mertens F (eds.): Mitelman Database of Chromosome Aberrations in Cancer, 2009. http:// cgap.nci.nih.gov/Chromosomes/Mitelman.
- Johansson B, Fioretos T and Mitelman F. Cytogenetic and molecular genetic evolution of chronic myeloid leukemia. Acta Haematol 107: 76-94, 2002.
- 20. Jeandidier E, Dastugue N, Mugneret F, Lafage-Pochitaloff M, Mozziconacci MJ, Herens C, Michaux L, Verellen-Dumoulin C, Talmant P, Cornillet-Lefebvre P, Luquet I, Charrin C, Barin C, Collonge-Rame MA, Pérot C, Van den Akker J, Grégoire MJ, Jonveaux P, Baranger L, Eclache-Saudreau V, Pagès MP, Cabrol C, Terré C and Berger R; Groupe Français de Cytogénétique Hématologique (GFCH): Abnormalities of the long arm of chromosome 21 in 107 patients with hematopoietic disorders: a collaborative retrospective study of the Groupe Français de Cytogénétique Hématologique. Cancer Genet Cytogenet 166: 1-11, 2006.