

Didymin reverses phthalate ester-associated breast cancer aggravation in the breast cancer tumor microenvironment

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Abstract. The present study demonstrated two novel findings. To the best of our knowledge, it is the first study to demonstrate that regulated upon activation, normal T-cell expressed and secreted (RANTES), produced by breast tumor-associated monocyte-derived dendritic cells (TADCs) following breast cancer cell exposure to phthalate esters, may contribute to the progression of cancer via enhancement of cancer cell proliferation, migration and invasion. Furthermore, the present study revealed that didymin, a dietary flavonoid glycoside present in citrus fruits, was able to reverse phthalate ester-mediated breast cancer aggravation. MDA-MB-231 cells were treated with butyl benzyl phthalate (BBP), di-*n*-butyl phthalate (DBP) or di-2-ethylhexyl phthalate (DEHP). Subsequently, the conditioned medium (CM) was harvested and cultured with monocyte-derived dendritic cells (mdDCs). Cultures of MDA-MB-231 cells with the conditioned medium of BBP-, DBP- or DEHP-MDA-MB-231 tumor-associated mdDCs (BBP-, DBP- or DEHP-MDA-TADC-CM) demonstrated enhanced proliferation, migration and invasion. Exposure of the MDA-MB-231 cells to DBP induced the MDA-TADCs to produce the inflammatory cytokine RANTES, which subsequently induced MDA-MB-231 cell proliferation, migration and invasion. Depleting RANTES reversed the effects of DBP-MDA-TADC-mediated MDA-MB-231 cell proliferation,

migration and invasion. In addition, didymin was observed to suppress phthalate-mediated breast cancer cell proliferation, migration and invasion. The present study suggested that didymin was capable of preventing phthalate ester-associated cancer aggravation.

Introduction

Phthalates, including butyl benzyl phthalate (BBP), di-*n*-butyl phthalate (DBP) and di-2-ethylhexyl phthalate (DEHP), are utilized as softeners and plasticizers (1,2). Epidemiological studies have observed that phthalate exposure may increase the risk of breast cancer (3-6). However, the effects of phthalate esters in the breast cancer tumor microenvironment remain to be elucidated.

The tumor microenvironment is known to have a significant role in the progression of tumors and the development of chemoresistance to anticancer drugs (7). The tumor microenvironment is comprised of stromal cells, immune cells [lymphocytes, macrophages and dendritic cells (DCs)], growth factors, extracellular matrix constituents, metabolites and cytokines/chemokines (8). As antigen-presenting cells, DCs have been observed to exhibit significant roles in the initiation and regulation of the immune response to cancer (9). Tumor-associated DCs (TADCs) have been observed to contribute to the metastasis of tumors in various cancers (10,11). Regulated upon activation, normal T-cell expressed and secreted (RANTES), also known as C-C chemokine ligand 5, is a cytokine consistently observed in increased levels in breast cancer subtypes (12), and has been observed to be associated with the progression of breast cancer and the promotion of metastasis (13-16).

Epidemiological studies have provided evidence that a high dietary intake of flavonoids via fruits and vegetables may be associated with reduced cancer rates in humans (17-20). Flavonoids are a class of phenolic compounds that are widely distributed throughout the plant kingdom; they display diverse biological activities, including the inhibition of tumor progression and the prevention of cancer initiation (21,22). Didymin, a dietary flavonoid glycoside present in citrus fruits, demonstrates antioxidant and anticancer properties (23-28).

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The present study evaluated the effects of phthalate esters in the breast cancer tumor microenvironment and investigated didymin, a dietary flavonoid glycoside present in citrus fruits, as a possible antidote for phthalate ester-associated cancer aggravation.

Materials and methods

Chemicals. Didymin was obtained from Extrasynthese (Genay, France), and was dissolved in dimethyl sulfoxide (DMSO; Sigma-Aldrich, St. Louis, MO, USA) and stored at -20°C. Control cultures received the carrier solvent (0.1% DMSO). All other chemicals utilized were in the purest form available commercially.

Cell culture and conditioned medium. Human breast adenocarcinoma MDA-MB-231 cells (American Type Culture Collection, Manassas, VA, USA) were cultured in α -minimum essential medium (α -MEM; Thermo Fisher Scientific, Inc., Waltham, MA, USA) supplemented with non-essential amino acids, 0.1 mmol/l sodium pyruvate, 1% antibiotic/anti-mycotic solution and 10% fetal bovine serum (FBS) (all Thermo Fisher Scientific, Inc.). In order to obtain the various conditioned media (CM), the MDA-MB-231 cells (2×10^6 /100-mm dish) were treated with or without BBP, DBP or DEHP (all Sigma-Aldrich) at identical concentrations of 1 μ M for 6 h. Following washing and culturing for 24 h, the CM of phthalate ester-treated MDA-MB-231 cells (BBP-, DBP- or DEHP-MDA-CM) were harvested (Fig. 1A).

Isolation of cluster of differentiation (CD)14⁺ monocytes and differentiation of monocyte-derived dendritic cells (mdDCs). Monocytes were purified from peripheral blood mononuclear cells obtained from healthy consenting donors. Mononuclear cells were isolated from the blood by Ficoll-Hypaque gradient (GE Healthcare Life Sciences, Chalfont, UK). CD14⁺ monocytes were purified with CD14⁺ monoclonal antibody-conjugated magnetic beads (MACS MicroBeads; Miltenyi Biotec GmbH, Bergisch Gladbach, Germany), according to the manufacturer's protocols. mdDCs were generated by culturing CD14⁺ monocytes in α -MEM containing FBS and 20 ng/ml granulocyte-macrophage colony-stimulating factor (GM-CSF) and 10 ng/ml interleukin (IL)4 (R&D Systems, Inc., Minneapolis, MN, USA) for 5 days. The medium was replaced with fresh medium containing GM-CSF and IL4 on day 3. For the maturation of DCs, immature mdDCs were stimulated with lipopolysaccharide (100 ng/ml; Sigma-Aldrich) following priming with interferon- γ (EMD Millipore, Billerica, MA, USA) for 3 h. MDA-MB-231 tumor-associated mdDCs (MDA-TADCs), BBP-MDA-TADCs, DBP-MDA-TADCs or DEHP-MDA-TADCs were generated by culturing CD14⁺ monocytes in α -MEM medium containing FBS, IL4 and GM-CSF in 20% MDA-CM, BBP-MDA-CM, DBP-MDA-CM or DEHP-MDA-CM, and subsequently stimulated as aforementioned. Following washing, the supernatants were collected and identified as MDA-MB-231-TADC-CM, BBP-MDA-TADC-CM, DBP-MDA-TADC-CM or DEHP-MDA-TADC-CM (Fig. 1B). The Institutional Review Board (IRB) of Kaohsiung Medical University Hospital (Kaohsiung, Taiwan) approved the present study protocol

and all participants provided written informed consent in accordance with the Declaration of Helsinki (IRB numbers: KMH-IRB-990174 and KMH-IRB-20120362).

Reverse transcription-quantitative polymerase chain reaction (RT-qPCR). TRIzol reagent (Invitrogen; Thermo Fisher Scientific, Inc.) was utilized for RNA isolation, while complementary (c)DNA was prepared using an oligo(dT) primer and reverse transcriptase (Takara Bio, Inc., Otsu, Japan) following standard protocols (29). RT-qPCR was performed using SYBR Green on the ABI 7500 Real-Time PCR System (Applied Biosystems; Thermo Fisher Scientific, Inc.). Each PCR mixture contained 200 nM of each primer, 10 μ l of 2X SYBR Green PCR Master Mix (Applied Biosystems; Thermo Fisher Scientific, Inc.), and 5 μ l of cDNA and RNase-free water, in a total volume of 20 μ l. The RT-qPCR was performed with a denaturation step at 95°C for 10 min, then for 40 cycles at 95°C for 15 sec and 60°C for 1 min. All PCRs were performed in triplicate and normalized to internal control glyceraldehyde-3-phosphate dehydrogenase mRNA. The relative expression level was presented using the $2^{-\Delta\Delta C_q}$ method. The primer sequences of target genes in the present study were as follows: RANTES, F 5'-cgctgtcat cctcattgcta-3' and R 5'-acacacttgccggttcttctc-3'; and GAPDH F 5'-GAGTCAACGGATTTGGTCGT-3' and R 5'-TTGATT TTGGAGGGATCTCG-3'.

Enzyme-linked immunosorbent assay (ELISA). RANTES levels were determined using an ELISA-based kit (R&D Systems Europe Ltd., Abingdon, UK). ELISA was performed according to the manufacturer's protocols. Depletion of RANTES from various CMs was performed using a mouse monoclonal anti-RANTES antibodies (2 μ g/ml; Abcam, Cambridge, UK) and Sepharose™ Protein A/G beads (PAG50-00-0002, Rockland Immunochemicals Inc., Gilbertsville, PA, USA) following standard immunoprecipitation techniques (30). Cytokine depletion was additionally confirmed using an ELISA-based kit.

Cell proliferation. The cells were plated in 96-well culture plates. Following 24 h of incubation, the cells were treated with vehicle mdDC-CM or specific CM for 72 h. At the conclusion of the assay period, cell proliferation was measured using a water-soluble tetrazolium salts (WST)-1 assay. Cell proliferation was determined using Premixed WST-1 Cell Proliferation Reagent (Clontech Laboratories, Inc., Mountainview, CA, USA) in accordance with the manufacturer's protocols.

Cell migration and invasion assay. Cell migration and invasion assays were performed using QCM™ 24-well Cell Migration and Invasion Assay kits (EMD Millipore). Briefly, the cells were seeded into the migration chamber and mdDC-CM or various CM were added to the lower wells for 24 h as a chemoattractant. At the conclusion of the treatment, the cells were stained using CyQuant GR dye (cat no. 90131) in cell lysis buffer for 15 min at room temperature. The fluorescence of the migrated and invaded cells was subsequently measured using a fluorescence plate reader at excitation/emission wavelengths of 485/520 nm.

Statistical analysis. Data are expressed as the mean \pm standard deviation. Statistical comparisons of the results were

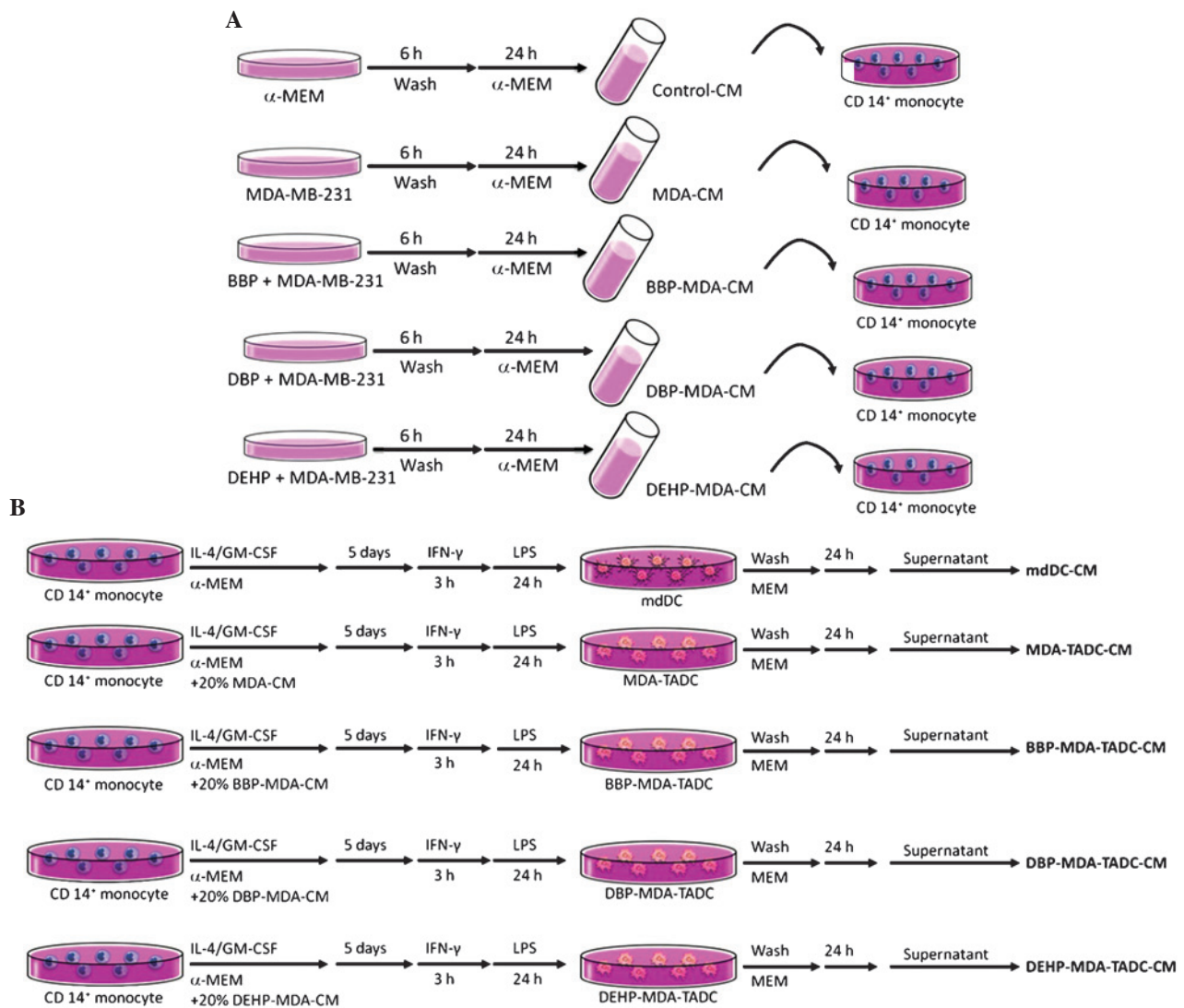


Figure 1. Flow chart of the production of various CM. (A) Flow chart of the production of control-CM, MDA-CM, BBP-MDA-CM, DBP-MDA-CM and DEHP-MDA-CM. (B) Flow chart of the production of mdDC-CM, MDA-TADC-CM, BBP-MDA-TADC-CM, DBP-MDA-TADC-CM and DEHP-MDA-TADC-CM. CM, conditioned media; MDA, MDA-MB-231 cells; BBP, butyl benzyl phthalate; DBP, di-*n*-butyl phthalate; DEHP, di-2-ethylhexyl phthalate; mdDC, monocyte-derived dendritic cells; TADC, tumor-associated mdDC; MEM, minimum essential medium; IL, interleukin; GM-CSF, granulocyte-macrophage colony-stimulating factor; IFN, interferon; LPS, lipopolysaccharide; CD, cluster of differentiation.

performed using an analysis of variance. Significant differences between the means of the test groups were analyzed using Student's *t*-test. $P < 0.05$ was considered to indicate a statistically significant difference.

Results

Breast cancer cells, following exposure to phthalate esters, affect mdDCs and contribute to breast cancer progression by enhancing cancer cell proliferation, migration and invasion. In order to understand whether phthalate esters exacerbate cancer progression in the breast cancer tumor microenvironment, the effects of BBP-MDA-TADC-CM, DBP-MDA-TADC-CM and DEHP-MDA-TADC-CM on breast cancer cell proliferation, migration and invasion were investigated. MDA-TADC-CM (20%) increased breast cancer cell proliferation ($P = 0.01$), and this stimulatory effect was additionally enhanced when MDA-MB-231 cells were pretreated with BBP, DBP or DEHP (Fig. 2A) ($P = 0.001, 0.0001$ and 0.0007 for BBP, DBP and DEHP,

respectively). In addition, MDA-TADC-CM (20%) induced breast cancer cell migration and invasion ($P = 0.002$ and 0.008 for migration and invasion analysis), and this reinforceable effect was worsened when breast cancer cells were cultured with BBP-MDA-TADC-CM (20%), DBP-MDA-TADC-CM (20%) or DEHP-MDA-TADC-CM (20%) (Fig. 2B and C).

RANTES has a significant role in TADC-mediated cancer progression. In order to determine the primary factors contributing to MDA-TADC, BBP-MDA-TADC, DBP-MDA-TADC and DEHP-MDA-TADC-mediated breast cancer progression, RT-qPCR analysis revealed that RANTES mRNA levels were increased by 6-, 12-, 17- and 11-fold in MDA-TADCs, BBP-MDA-TADCs, DBP-MDA-TADCs and DEHP-MDA-TADCs, respectively (Fig. 3A). As demonstrated by ELISA, the protein levels of RANTES were enhanced in MDA-TADC-CM ($P = 0.01$), BBP-MDA-TADC-CM ($P = 0.005$), DBP-MDA-TADC-CM ($P = 0.002$) and DEHP-MDA-TADC-CM (Fig. 3B)

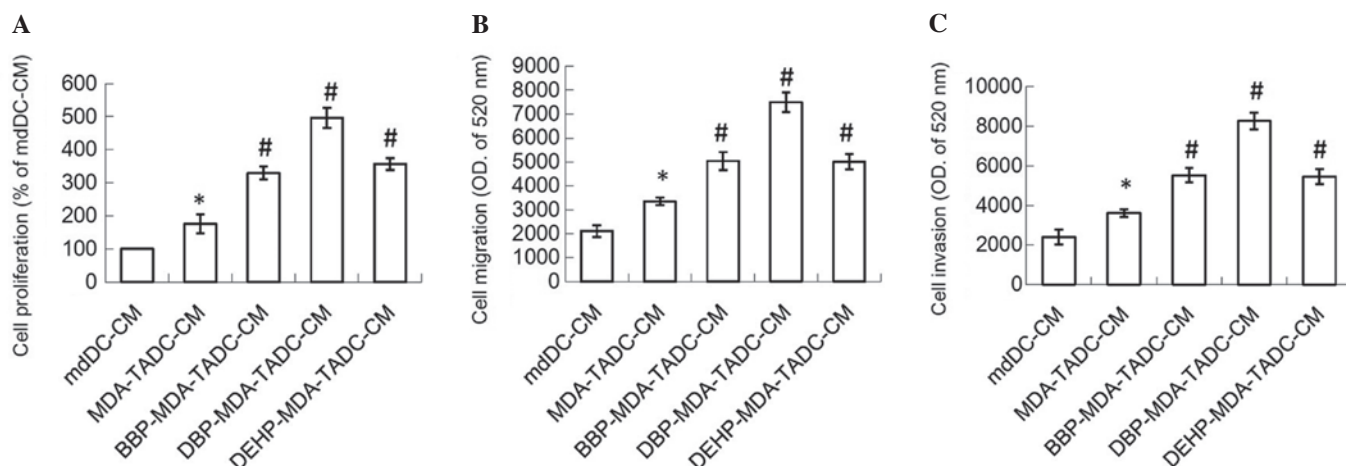


Figure 2. CM of phthalate ester-treated MDA-MB-231 cells cause mDCs to increase breast cancer cell proliferation, migration and invasion. BBP-, DBP- and DEHP-TADC-CM increased breast cancer cell (A) proliferation, (B) migration and (C) invasion. Each value is presented as the mean \pm standard deviation of three independent experiments. * $P < 0.05$ vs. mDC-CM treatment. # $P < 0.05$ vs. MDA-TADC-CM treatment. CM, conditioned media; mDCs, monocyte-derived dendritic cells; BBP, butyl benzyl phthalate; DBP, di-*n*-butyl phthalate; DEHP, di-2-ethylhexyl phthalate; TADC, tumor-associated mDC; MEM, minimum essential medium; OD, optical density.

($P = 0.003$). The effect of DBP-MDA-TADC-CM on the induction of breast cancer cell proliferation, migration and invasion, and the induction of RANTES was greater than that of BBP-MDA-TADC-CM or DEHP-MDA-TADC-CM (Figs. 2 and 3). As DBP demonstrated the greatest effect in these circumstances, it was selected as the model for investigation of the detailed effects of phthalate ester-associated cancer aggravation in the breast cancer tumor microenvironment. In order to understand its role, RANTES was depleted in MDA-TADC-CM and DBP-MDA-TADC-CM. The effect of MDA-TADC-CM and DBP-MDA-TADC-CM on cancer progression was subsequently assessed. The successful depletion of RANTES from MDA-TADC-CM and DBP-MDA-TADC-CM was confirmed using RANTES ELISA kits (data not shown). As demonstrated in Fig. 4, RANTES depletion inhibited the stimulatory effects of MDA-TADC-CM on breast cancer cell proliferation, migration and invasion ($P = 0.00002$ for cell proliferation, $P < 0.0001$ for migration and invasion), as well as blocking the intensified stimulatory effects of DBP-MDA-TADC-CM on breast cancer cell proliferation, migration and invasion.

Didymin suppresses DBP-MDA-TADC-mediated breast cancer aggravation. The tumor microenvironment has a significant role in the development of chemoresistance to anticancer drugs and tumor progression (7). TADC has been demonstrated to promote the progression of cancer by modulating a number of components in the cancer niche, thereby creating a supportive and permissive microenvironment for tumor survival, proliferation and metastasis (11,31). As phthalate esters stimulate the ability of breast cancer cells to affect mDCs and thereby intensified breast cancer cell proliferation, migration and invasion, the search for a possible antidote in the fight against phthalate esters-induced cancer aggravation in the breast cancer tumor microenvironment has become a matter of importance. The present study therefore assessed the effect of didymin, a dietary flavonoid glycoside derived from citrus fruits, on DBP-induced cancer progression.

As shown in Fig. 5A, the MDA-TADC-CM-induced breast cancer cell proliferation effect was inhibited when cells were pretreated with didymin, and DBP-MDA-TADC-CM-induced breast cancer cell proliferation was additionally reversed when the cells were pretreated with didymin. Similarly, enhancement of breast cancer cell migration and invasion triggered by DBP-MDA-TADC-CM was abrogated upon didymin pretreatment (Fig. 5B and C).

Discussion

To the best of our knowledge, the present study is the first to evaluate the interaction between mDCs and breast cancer cells following exposure to phthalate esters. BBP, DBP and DEHP stimulated the breast cancer cells and subsequently caused the mDCs to secrete RANTES, which enhanced the proliferation, migration and invasion of the human breast cancer cells. To the best of our knowledge, the present study is additionally the first to investigate the effects of didymin in interfering with phthalate ester-mediated breast cancer aggravation in the breast cancer tumor microenvironment. The results of the present study suggested that didymin was capable of preventing phthalate ester-associated breast cancer progression (Fig. 6).

Phthalates, including BBP, DBP and DEHP, are widely utilized in food wraps and cosmetic products (1-3). Individuals are exposed to phthalates throughout their entire lives via ingestion, inhalation and dermal exposure (1,2). Several phthalates have been demonstrated to promote breast cancer development by increasing cell proliferation and migration (32-34). In a previous study, we demonstrated that phthalate esters were able to induce breast cancer bone metastasis by targeting parathyroid hormone-related protein (35). The present study demonstrated that phthalate esters BBP, DBP and DEHP were able to induce breast cancer cells to affect mDCs, thereby intensifying breast cancer cell proliferation, migration and invasion. The results of the present study suggested that phthalate esters may increase cancer progression in the breast cancer tumor microenvironment.

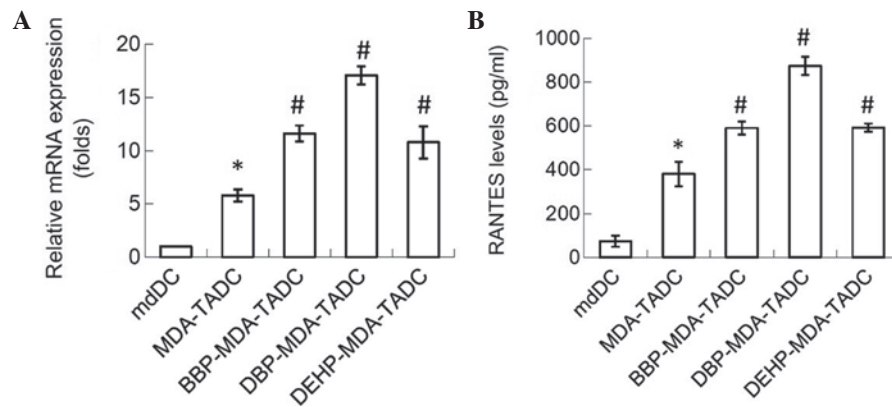


Figure 3. MDA-CM, BBP-MDA-CM, DBP-MDA-CM and DEHP-MDA-CM increase the expression of RANTES in mdDCs. (A) MDA-CM, BBP-MDA-CM, DBP-MDA-CM and DEHP-MDA-CM increased the levels of RANTES, as assessed by reverse transcription-polymerase chain reaction. (B) MDA-CM, BBP-MDA-CM, DBP-MDA-CM and DEHP-MDA-CM increased the RANTES protein levels, as detected by enzyme-linked immunosorbent assay. Each value is presented as the mean \pm standard deviation of three independent experiments. * $P < 0.05$ vs. mdDC treatment. # $P < 0.05$ vs. MDA-TADC treatment. MDA, MDA-MB-231 cells; CM, conditioned media; BBP, butyl benzyl phthalate; DBP, di-*n*-butyl phthalate; DEHP, di-2-ethylhexyl phthalate; RANTES; regulated upon activation, normal T-cell expressed, and secreted; mdDCs, monocyte-derived dendritic cells.

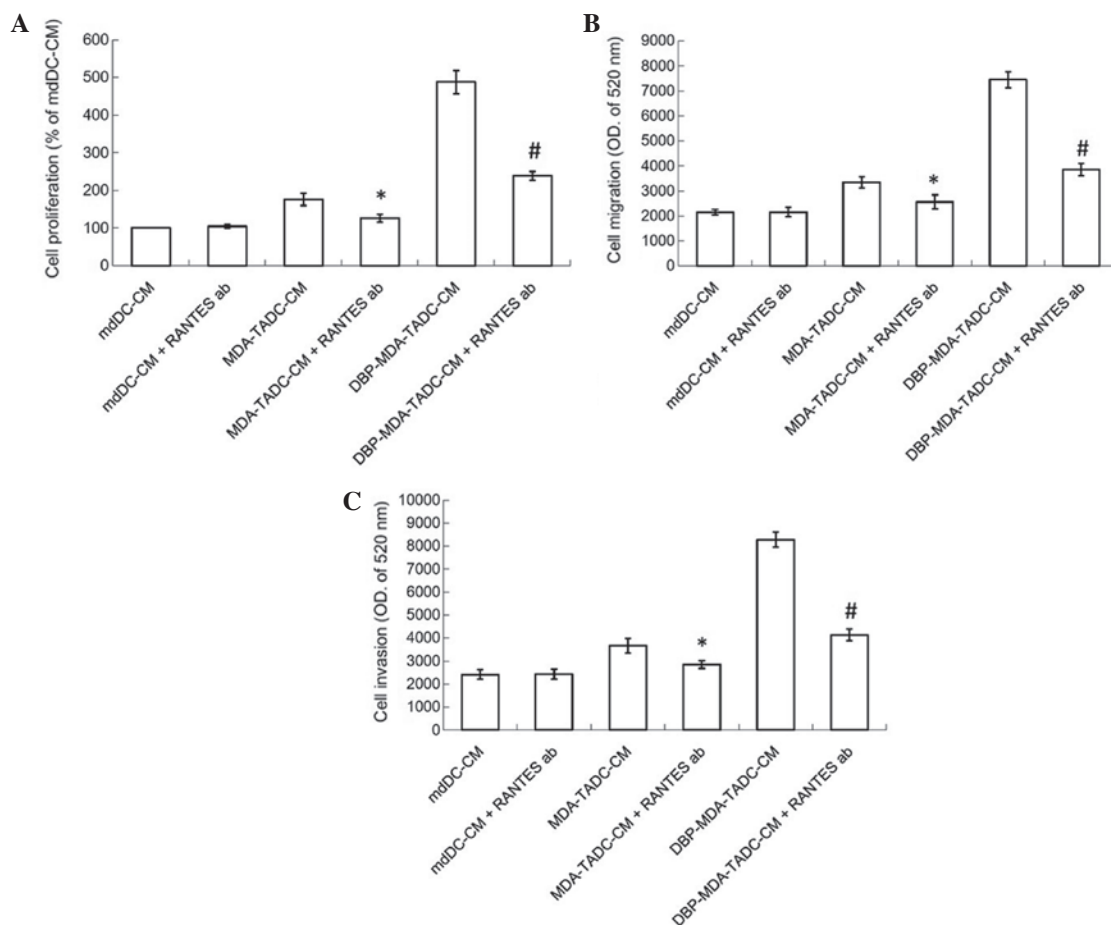


Figure 4. RANTES is a significant factor for MDA-TADC-CM and DBP-MDA-TADC-CM in increasing breast cancer cell proliferation and migration. (A) The proliferation of MDA-MB-231 cells observed in RANTES-depleted MDA-TADC-CM or DBP-MDA-TADC-CM. (B) Migration and (C) invasion of MDA-MB-231 cells present in RANTES-depleted MDA-TADC-CM or DBP-MDA-TADC-CM. Each value is presented as the mean \pm standard deviation of three independent experiments. * $P < 0.05$ vs. MDA-TADC-CM treatment. # $P < 0.05$ vs. DBP-MDA-TADC-CM treatment. RANTES; regulated upon activation, normal T-cell expressed, and secreted; MDA, MDA-MB-231 cells; mdDCs, monocyte-derived dendritic cells; TADC, tumor-associated mdDC; CM, conditioned media; DBP, di-*n*-butyl phthalate; OD, optical density; ab, antibody.

The tumor microenvironment has been observed to affect cancer progression and generation. The cells surrounding a tumor provide essential supportive factors that promote

progression (36-39). RANTES has been observed to be a significant contributor to various chronic inflammatory diseases and malignancies via the recruitment of inflammatory

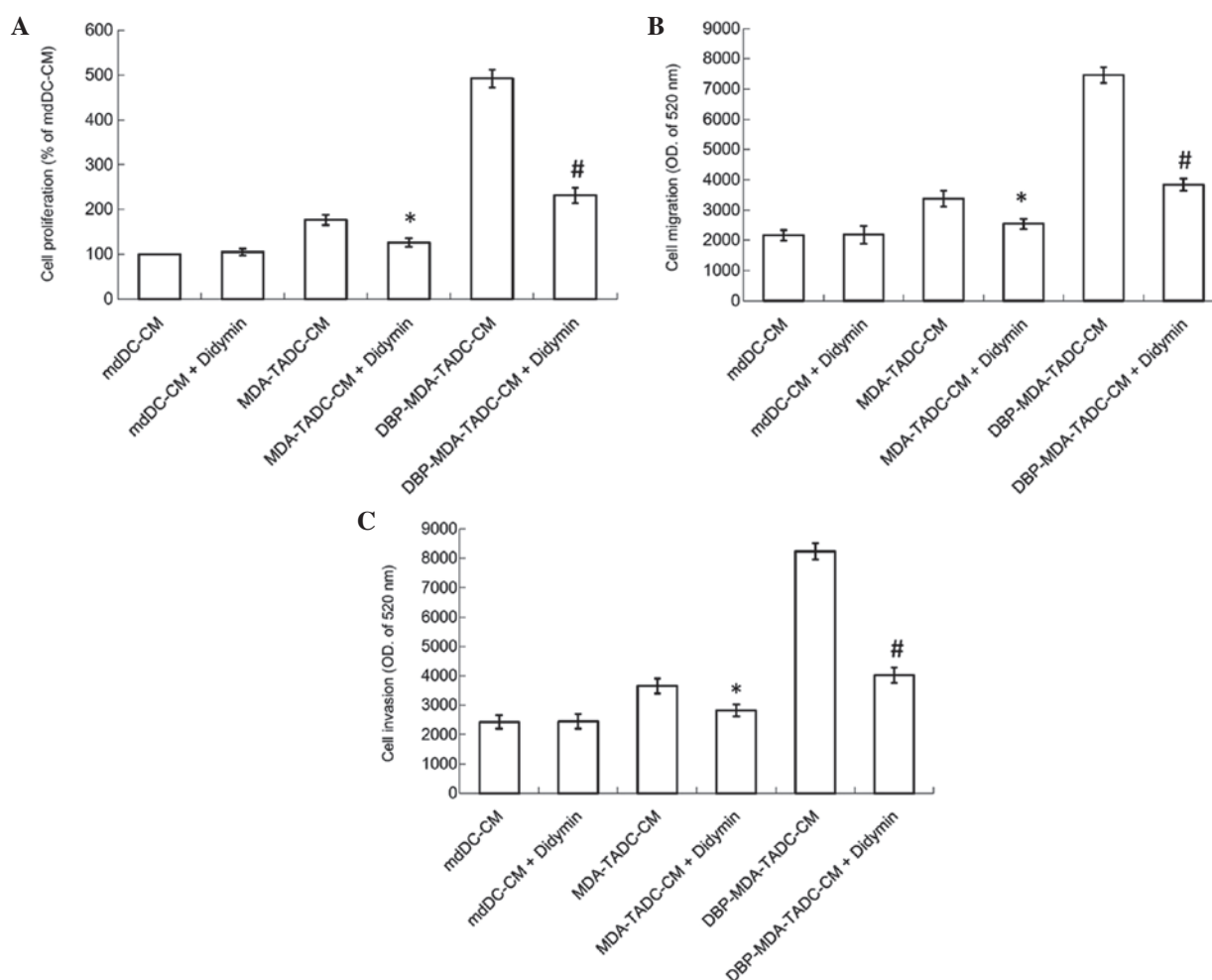


Figure 5. Didymine suppresses DBP-mediated cancer aggravation in the breast cancer tumor microenvironment. Didymine reversed (A) proliferation, (B) migration and (C) invasion of MDA-MB-231 breast cancer cells exposed in culture to MDA-TADC-CM or DBP-MDA-TADC-CM. Each value is presented as the mean \pm standard deviation of three independent experiments. * $P < 0.05$ vs. MDA-TADC-CM treatment. # $P < 0.05$ vs. DBP-MDA-TADC-CM treatment. DBP, di-*n*-butyl phthalate; mdDCs, monocyte-derived dendritic cells; TADC, tumor-associated mdDC; CM, conditioned media; OD, optical density.

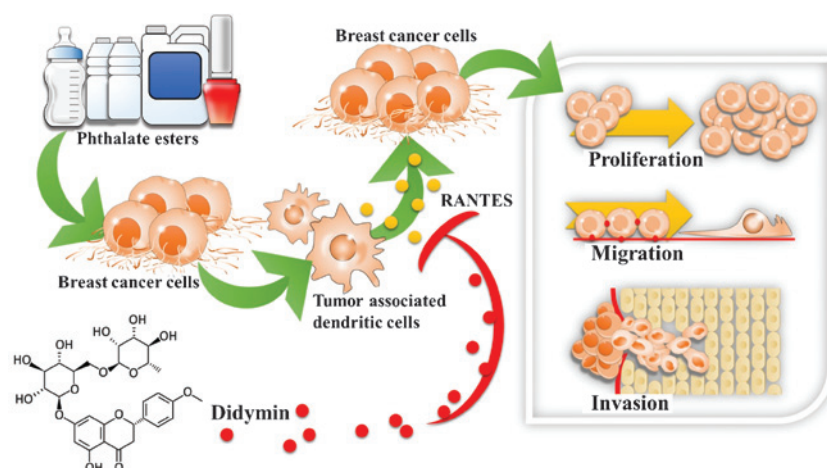


Figure 6. Didymine reverses phthalate ester-associated breast cancer aggravation in the breast cancer tumor microenvironment. The present study demonstrated two novel findings: i) RANTES, produced by breast tumor-associated monocyte-derived dendritic cells following breast cancer cells' exposure to phthalate esters, contributed to cancer progression by enhancing cancer cell proliferation, migration and invasion; and ii) didymine, a dietary flavonoid glycoside present in citrus fruits, reversed phthalate ester-mediated breast cancer aggravation.

cells (15,40,41). RANTES has additionally been reported to be overexpressed in certain cancers and is involved in critical steps of cancer spread, including proliferation, migration, invasion,

angiogenesis and metastatic colonization following activation (42-45). Furthermore, RANTES has been associated with resistance to conventional chemotherapeutic drugs, including

cisplatin and tamoxifen (46). The present study revealed that the phthalate esters BBP, DBP and DEHP were able to cause breast cancer cells to affect mdDCs to produce the inflammatory cytokine RANTES, which subsequently enhanced breast cancer cell proliferation, migration and invasion. Depletion of RANTES reversed the effects of MDA-TADC-CM- and DBP-MDA-TADC-CM-mediated breast cancer cell proliferation, migration and invasion. Elimination of all phthalate exposure may not be possible, as phthalate esters are extensively used in modern life (1,2). It is therefore important that strategies are developed for the prevention of breast cancer progression in the breast cancer tumor microenvironment. The results of the present study revealed that didymine, a dietary flavonoid glycoside obtained from citrus fruits, is able to reverse the negative actions of DBP-stimulated breast cancer cells, which cause mdDCs to enhance the proliferation, migration and invasion of human breast cancer cells.

In conclusion, to the best of our knowledge, the present study is the first to investigate the interaction between mdDCs and breast cancer cells following exposure to phthalate esters. These esters, BBP, DBP and DEHP, were used to stimulate human breast cancer MDA-MB-231 cells to mediate RANTES upregulation in mdDCs, thereby inducing breast cancer cell proliferation, migration and invasion. To the best of our knowledge, this is the first study to provide evidence that didymine, a dietary flavonoid glycoside present in citrus fruits, has potential for the prevention of phthalate ester-associated breast cancer progression in the breast cancer tumor microenvironment. Therefore, the present study suggested that didymine may be capable of preventing phthalate ester-associated cancer aggravation.

Acknowledgements

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