

# Upregulation of miR-34a by diallyl disulfide suppresses invasion and induces apoptosis in SGC-7901 cells through inhibition of the PI3K/Akt signaling pathway

GUOJUN WANG, GUANGHUI LIU, YANWEI YE, YANG FU and XIEFU ZHANG

Department of Gastrointestinal Surgery, The First Affiliated Hospital of Zhengzhou University,  
Zhengzhou, Henan 450052, P.R. China

Received January 15, 2015; Accepted January 21, 2016

DOI: 10.3892/ol.2016.4266

**Abstract.** Diallyl disulfide (DADS) exerts anticarcinogenic activity in various types of cancer. However, the mechanism underlying its anticarcinogenic activity remains to be elucidated. The aim of the present study was to explore the mechanism of the anticarcinogenic activity of DADS in gastric cancer (GC). The expression levels of microRNA (miR)-34a in GC and normal tissues were measured using reverse transcription-quantitative polymerase chain reaction (RT-qPCR). The expression of miR-34a was also measured using RT-qPCR in SGC-7901 cells following treatment with DADS. In addition, the effect of DADS on the invasion capability of SGC-7901 cells was observed in the presence of miR-34a or anti-miR-34a using a Matrigel invasion assay. Furthermore, in identical conditions, the apoptosis of SGC-7901 cells was observed using flow cytometry. Finally, the present study investigated the effects of DADS and miR-34a on the phosphoinositide 3-kinase (PI3K)/Akt signaling pathway *in vitro*. The level of miR-34a in GC tissues was reduced compared with that in adjacent normal tissues ( $P<0.05$ ). Treatment with DADS upregulated miR-34a expression in SGC-7901 cells ( $P<0.05$ ). In the Matrigel invasion assay, DADS inhibited the invasive capability of SGC-7901 cells ( $P<0.05$  vs. control), which was improved by overexpression of miR-34a ( $P<0.01$  vs. control) but reduced by downregulation of miR-34a ( $P<0.05$  vs. DADS treatment group). Furthermore, DADS induced apoptosis of SGC-7901 cells ( $P<0.05$  vs. control); and DADS and miR-34a synergistically enhanced apoptosis of SGC-7901 cells ( $P<0.01$  vs. control). In addition, DADS and miR-34a inhibited the expression levels of phosphorylated (p)-PI3K and p-Akt

( $P<0.05$  vs. control). By contrast, downregulation of miR-34a alleviated the decrease in p-PI3K and p-Akt expression induced by DADS ( $P<0.05$  vs. DADS treatment group). Cell viability was reduced with increasing concentrations of DADS, however, DADS did not affect cell viability following inhibition of the PI3K/Akt signaling pathway. In conclusion, DADS suppresses invasion and induces apoptosis of SGC-7901 cells by upregulation of miR-34a, via inhibition of the PI3K-Akt signaling pathway.

## Introduction

Gastric cancer (GC), a type of malignant tumor that is prone to invasion and metastasis (1), is the fourth most prevalent cancer (2) and the second most frequent cause of cancer-related mortality worldwide, with an estimated 1,000,000 new cases per year (3). In China, GC remains the second leading cause of cancer-related mortality (4). At present, ~80% of patients diagnosed with GC are at an advanced stage with limited surgical options (5). Accumulating evidence has demonstrated that the pathogenesis of GC is associated with the activity of oncogenes, tumor suppressors and precancerous lesions, including chronic atrophic gastritis and intestinal metaplasia (6,7). However, the underlying mechanisms remain to be elucidated.

Diallyl disulfide (DADS) is a major component of cooked garlic and a natural organosulfur compound in processed garlic that accounts for 40-60% of the total oil-soluble sulfides in garlic oil (5,8). Similar to the majority of plant products, DADS exerts anticarcinogenic activity in certain types of cancer, including human breast (9), colon (10,11) and prostate cancer (8,12), as well as hepatoma cells (13). It has been observed that DADS, as an effective modulator of protein kinases/phosphatases, inhibits the initiation and promotion phases of cancer (5).

MicroRNAs (miRs), which are expressed by all multicellular eukaryotes, are a class of small non-coding RNAs 18-25 nucleotide long that negatively regulate gene expression at the post-transcriptional level. This is achieved by miRs binding to the 3'-untranslated region (3'-UTR) of target messenger RNAs (14,15). miRs have been shown to have significant roles in a wide variety of biological processes, including development, the cell cycle, stress responses,

**Correspondence to:** Mr. Guojun Wang, Department of Gastrointestinal Surgery, The First Affiliated Hospital of Zhengzhou University, 1 Jianshedong Road, Zhengzhou, Henan 450052, P.R. China  
E-mail: wwanguojun@126.com

**Key words:** gastric cancer, diallyl disulfide, microRNA-34a, invasion, apoptosis, phosphoinositide 3-kinase/Akt signaling pathway

immunity, proliferation, differentiation and apoptosis (15,16). Notably, the dysfunction of miRs has been associated with certain human pathologies, including cancer (17,18).

Previous studies have demonstrated that dysregulation of miRs occurs in GC (19,20). Using reverse transcription-quantitative polymerase chain reaction (RT-qPCR) analysis, Liu *et al* (21) identified a profile of serum miRs that were biomarkers for GC detection; the results revealed that the expression levels of miR-1, miR-20a, miR-27a, miR-34 and miR-423-5p were correlated with tumor stage. However, the mechanism of miR regulation in GC remains to be elucidated. In the present study, it was observed that miR-34a was upregulated in the SGC-7901 GC cell line following treatment with DADS. In addition, the regulation of miR-34a by DADS, as well as the association between miR-34a and the phosphoinositide 3-kinase (PI3K)-Akt signaling pathway, were investigated in SGC-7901 cells.

## Materials and methods

**Tissue specimens and cell line.** A total of 25 pairs of human GC and adjacent normal tissues were surgically collected between May 2012 and August 2013 from the Department of Gastrointestinal Surgery at The First Affiliated Hospital of Zhengzhou University (Zhengzhou, China). The matched adjacent normal tissues were obtained a distance of 5 cm away from the tumor margin. Tissue specimens were immediately frozen in liquid nitrogen following collection and were stored at -80°C until use. Informed consent was obtained from all patients and donors, and the study was approved by the Ethics Committee of Zhengzhou University (Zhengzhou, China).

The SGC-7901 human GC line was purchased from the Shanghai Institute of Cell Biology (Shanghai, China). SGC-7901 cells were maintained in RPMI 1640 medium (Gibco; Thermo Fisher Scientific, Inc., Waltham, MA, USA) supplemented with 10% fetal bovine serum (Thermo Fisher Scientific, Inc.), 100 U/ml penicillin (Sigma-Aldrich, St. Louis, MO, USA) and 100 µg/ml streptomycin (Sigma-Aldrich) at 37°C in a humidified incubator at an atmosphere of 5% CO<sub>2</sub> (4). A volume of 400 µM DADS (Sigma-Aldrich) was used to investigate the effects on SGC-7901 cells, and cells were incubated at 37°C in a humidified atmosphere with 5% CO<sub>2</sub> for 24 h.

**RNA extraction and RT-qPCR.** Total RNA was extracted from the tissue specimens and cell line using TRIzol® reagent (Thermo Fisher Scientific, Inc.), according to the manufacturer's protocol. In total, 1 µl MultiScribe™ RT was used on the RNA (50 U/µl; Thermo Fisher Scientific, Inc.) RT and PCR primers for miR-34a were purchased from Guangzhou RiboBio Co., Ltd. (Guangzhou, China). Primer sequences were as follows: miR-34a forward, 5'-CCCGTTGGCAGTGTCTTAGCT-3' and reverse, 5'-GTGCAGGGTCCGAGGT-3'; and glyceraldehyde-3-phosphate dehydrogenase (GAPDH) forward, 5'-AACGGCAGTCAAGGCTGA-3' and reverse, 5'-ACGCCAGTA GACTCCACGACAT-3'. Total RNA was reverse-transcribed using the TaqMan® Reverse Transcription kit (Thermo Fisher Scientific, Inc.), according to the manufacturer's protocol. First strand complementary (c)DNA was synthesized using oligo (dT) primers and Superscript™ II reverse transcriptase

(both Invitrogen; Thermo Fisher Scientific, Inc.). RT-PCR was performed with a PCR mixture containing 1 µmol/l of each primer and SYBR-Green Master Mix (Applied Biosystems; Thermo Fisher Scientific, Inc.), using a StepOnePlus™ Real-Time PCR System (Thermo Fisher Scientific, Inc.). The PCR cycling conditions were as follows: 95°C for 10 min, and then 40 cycles of 95°C for 10 sec and 60°C for 60 sec. GAPDH expression was determined and used as an endogenous control. The relative miR expression levels were calculated and normalized to GAPDH using the 2<sup>-ΔΔCq</sup> method (22). All the experiments were performed in triplicate. A no template control was performed (negative control), which used water instead of the cDNA template.

**Western blot analysis.** For protein extraction, cells were lysed on ice in radioimmunoprecipitation assay buffer (Sigma-Aldrich) with an added cocktail of protease inhibitors (Sigma-Aldrich). Briefly, 50 µg total protein samples were resolved on 10-15% sodium dodecyl sulfate-polyacrylamide gel electrophoresis gels (Sigma-Aldrich) and transferred to polyvinylidene difluoride membranes (EMD Millipore, Billerica, MA, USA). The membranes were incubated overnight at 4°C with primary antibodies against phosphorylated (p)-PI3K (rabbit anti-p-PI3K polyclonal; catalog no., 4228; Cell Signaling Technology, Inc., Danvers, MA, USA; dilution, 1:1,000), p-Akt (polyclonal rabbit anti-human anti-p-Akt; catalog no., sc-33437; Santa Cruz Biotechnology, Inc., Dallas, TX, USA; dilution, 1:500), cleaved caspase-3 (polyclonal rabbit anti-human anti-cleaved caspase-3; Cell Signaling Technology, Inc.; dilution, 1:1,000) and rabbit anti-GAPDH monoclonal antibody (catalog no., 2118; Cell Signaling Technology, Inc.; dilution, 1:1,000). The secondary antibody used was alkaline phosphatase peroxidase-conjugated goat anti-rabbit immunoglobulin G (catalog no., ab6722; dilution, 1:4,000; Abcam, Cambridge, MA, USA) and were incubated for 16 h at 4°C. ImageQuant™ LAS 4000 software (GE Healthcare Life Sciences, Chalfont, UK) was used to quantify the bands and the levels of protein expression were normalized to GAPDH.

**Oligonucleotide transfection.** SGC-7901 cells were transfected with miR-34a mimics (Thermo Fisher Scientific, Inc.), control non-specific miR precursor [negative control (NC)], scrambled mimics and anti-miR-34a (Thermo Fisher Scientific, Inc.) using Lipofectamine® 2000 (Invitrogen; Thermo Fisher Scientific, Inc.), according to the manufacturer's protocol.

**Matrigel invasion assay.** SGC-7901 cells were seeded at a density of 5x10<sup>3</sup> cells/well into 96-well culture plates (Nunc A/S Plastfabrikation, Roskilde, Denmark) and transfected with miR-34a mimic, NC, scrambled mimics and anti-miR-34a according to the Lipofectamine 2000 transfection manual. A Matrigel invasion assay was used to assay the capability of cell invasion. Briefly, 1x10<sup>4</sup> cells/well were plated with serum-free medium in the upper chamber (Corning, Inc., NY, USA), which was filter-coated with Matrigel (Sigma-Aldrich). The lower chamber was filled with 10% fetal bovine serum. Following 24 h of incubation at 37°C in an atmosphere of 5% CO<sub>2</sub>, the cells that migrated to the bottom of the membrane were fixed with 4% formaldehyde (Sigma-Aldrich) in phosphate-buffered saline (PBS; Sigma-Aldrich), stained with hematoxylin (Wako

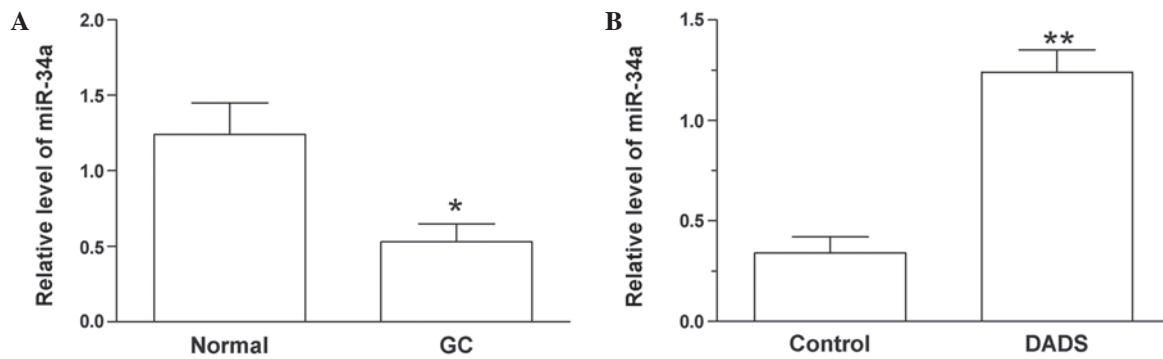


Figure 1. DADS upregulates the levels of miR-34a in GC. (A) The expression of miR-34a was decreased in 25 human GC tissues compared with 25 adjacent normal tissues. \* $P < 0.05$  vs. the normal tissues. (B) DADS increased the expression of miR-34a in SGC-7901 cells compared with untreated cells.  $n = 3$ . \*\* $P < 0.01$  vs. the control cells. Data are presented as the mean  $\pm$  standard error of the mean. Student's  $t$  test was used for analysis. miR, microRNA; GC, gastric cancer; DADS, diallyl disulfide.

Pure Chemical Industries, Ltd., Osaka, Japan) and quantified using an inverted microscope (IX71; Olympus Corp., Tokyo, Japan) at  $\times 200$  magnification.

**Flow cytometry assay.** Annexin V-fluorescein isothiocyanate (FITC)/propidium iodide (PI) staining was performed with an Annexin V-FITC Apoptosis Detection kit (Sigma-Aldrich), according to the manufacturer's protocol. Apoptosis was analyzed using flow cytometry. Briefly, SGC-7901 cells were trypsinized (Sigma-Aldrich), washed with PBS and resuspended in binding buffer (Sigma-Aldrich) 24 h after transient transfection. Cells were incubated with 5  $\mu$ l Annexin V-FITC and 10  $\mu$ l PI for 10 min in the dark at room temperature, prior to suspension in 300  $\mu$ l binding buffer. The cells were immediately analyzed using a FACScan flow cytometer (BD Biosciences, Franklin Lakes, NJ, USA) for relative quantitative apoptosis.

**3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay.** SGC-7901 cells were seeded into 96-well culture plates ( $1 \times 10^5$ /well). Following SGC-7901 cell treatment with PI3K inhibitor LY294002 (10 mM; Cell Signaling Technology, Inc.) for 4 h, DADS (0, 100, 200 and 400  $\mu$ M) was added. Cells were incubated at  $37^\circ\text{C}$  in a humidified atmosphere with 5%  $\text{CO}_2$  for 24 h. Subsequently, culture medium was replaced with fresh medium containing 500  $\mu$ g/ml MTT (Nanjing KeyGen Biotech Co., Ltd., Nanjing, China) and cells were incubated for an additional 4 h. Finally, dimethylsulfoxide (150  $\mu$ l/well; Nanjing KeyGen Biotech Co., Ltd.) was added and optical density was measured using a spectrophotometer (Novaspek III Visible Spectrophotometer; GE Healthcare Life Sciences, Chalfont, UK) at a wavelength of 490 nm.

**Statistical analysis.** SPSS version 13.0 (SPSS, Inc., Chicago, IL, USA) was used to analyze the experimental data. Data was checked for normality. Differences between groups were calculated by one-way analysis of variance and Student's  $t$  test. Data are presented as the mean  $\pm$  standard error of the mean.  $P < 0.05$  was considered to indicate a statistically significant difference.

## Results

**DADS treatment increases the expression levels of miR-34a in SGC-7901 cells.** The expression miR-34a was significantly

decreased in GC tissues compared with adjacent normal tissues ( $P = 0.014$ ; Fig. 1A). In SGC-7901 cells, miR-34a was upregulated following treatment with DADS for 48 h ( $P = 0.008$ ; Fig. 1B). This result indicates that DADS may influence the level of miR-34a, which may have a significant role in GC.

**miR-34a enhances the anti-invasion effect of DADS in SGC-7901 cells.** The number of cells that migrate in a Matrigel assay reflect the capacity for invasion. As demonstrated in Fig. 2A-F, DADS inhibited the cell invasion capacity of SGC-7901 cells ( $P = 0.023$  vs. control; Fig. 2C). In addition, upregulation of miR-34a enhanced the anti-invasion effect of DADS ( $P = 0.009$  vs. control; Fig. 2D). Notably, downregulation of miR-34a suppressed DADS-induced anti-invasion capacity ( $P = 0.036$  vs. DADS; Fig. 2E), suggesting that DADS inhibited the invasion of SGC-7901 cells by upregulation of miR-34a.

**miR-34a enhances DADS-induced apoptosis.** As demonstrated in Fig. 3A and B, DADS induced the apoptosis of SGC-7901 cells ( $P = 0.019$  vs. control), which was enhanced by upregulating miR-34a ( $P = 0.008$  vs. control). However, downregulation of miR-34a antagonized the role of DADS in SGC-7901 cells ( $P = 0.011$  vs. DADS). The expression level of cleaved caspase-3 in SGC-7901 cells following treatment with DADS and DADS plus miR-34a or anti-miR-34a similarly indicated that miR-34a enhanced DADS-induced apoptosis (DADS,  $P = 0.031$  vs. control; DADS + miR-34a,  $P = 0.003$  vs. control; DADS + anti-miR-24a,  $P = 0.019$  vs. DADS; Fig. 3C and D).

**DADS suppresses invasion capacity and induces apoptosis through inactivating the PI3K/Akt signaling pathway by upregulation of miR-34a.** To investigate the association between DADS and the PI3K/Akt signaling pathway, DADS, miR-34a mimics and DADS plus miR-34 mimics or anti-miR-34a were transfected into SGC-7901 cells, and a western blot analysis was performed. As demonstrated in Fig. 4A and B, DADS, miR-34a mimics and DADS plus miR-34 mimics decreased the expression levels of p-PI3K (DADS,  $P = 0.033$ ; miR-34a mimics,  $P = 0.041$ ; DADS + miR-34a,  $P = 0.007$ ) and p-Akt (DADS,  $P = 0.025$ ; miR-34a mimics,  $P = 0.012$ ; DADS + miR-34a,  $P = 0.009$ ) compared with the control group. However, treatment with DADS plus anti-miR-34a increased the expression levels of p-PI3K ( $P = 0.039$ ) and p-Akt ( $P = 0.040$ )



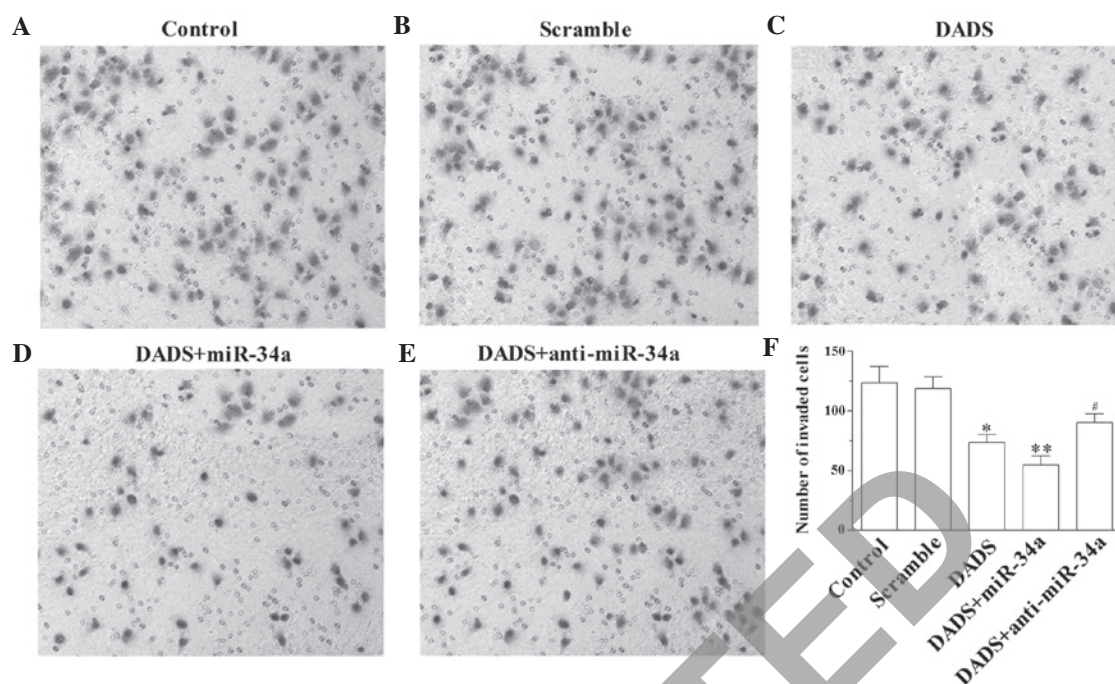


Figure 2. DADS inhibits the invasion of SGC-7901 cells by upregulation of miR-34a. The SGC-7901 cells were treated with (A) control non-specific miR precursor (control), (B) scrambled mimics (scramble), (C) DADS, (D) DADS plus miR-34a mimics (DADS + miR-34a) and (E) DADS plus anti-miR-34a mimics (DADS + anti-miR-34a). (A-E) The invaded cells were observed using an inverted microscope (magnification, x200; hematoxylin staining). (F) DADS inhibited the invasion capability of SGC-7901 cells and miR-34a enhanced the anti-invasion effect of DADS in SGC-7901 cells. \* $P < 0.05$  and \*\* $P < 0.01$  vs. the control cells; # $P < 0.05$  vs. the DADS treatment group. Data are presented as the mean  $\pm$  standard error of the mean ( $n=3$ ). One-way analysis of variance was used for analysis. DADS, diallyl disulfide; miR, microRNA.

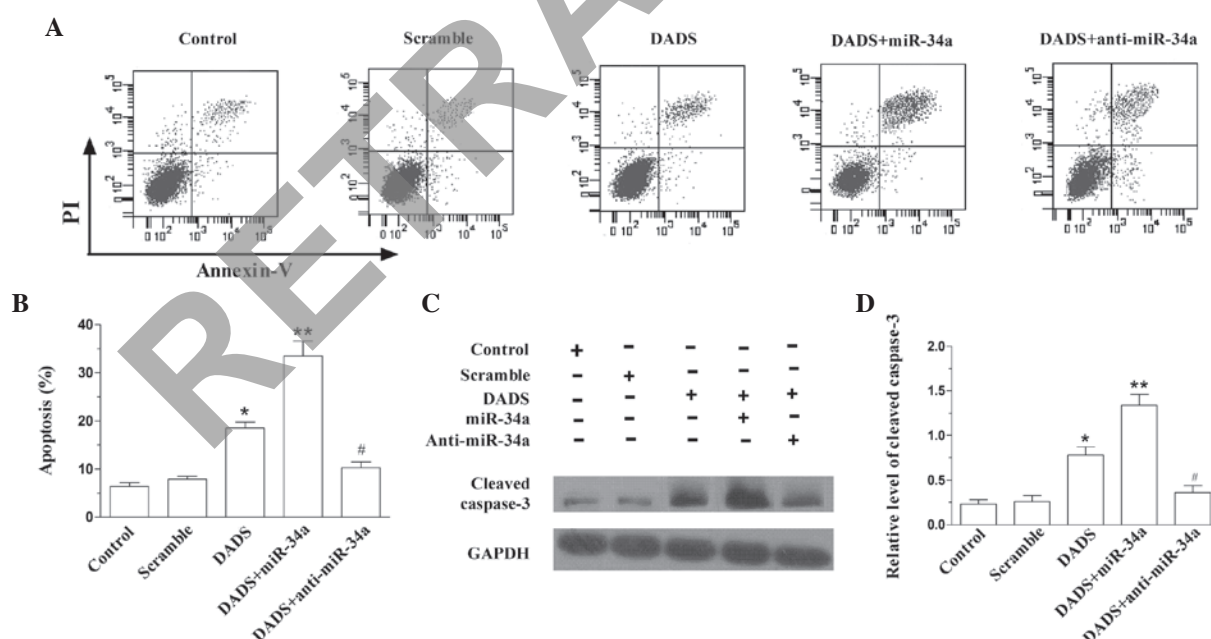


Figure 3. DADS induces the apoptosis of SGC-7901 cells by upregulation of miR-34a. (A and B) The apoptosis rate was measured using a flow cytometer. Upregulation of miR-34a enhanced DADS-induced apoptosis of SGC-7901 cells, while downregulation of miR-34a antagonized the role of DADS in SGC-7901 cells. (C) The expression level of cleaved caspase-3 in SGC-7901 cells following various treatments was determined using western blot analysis. (D) The relative expression level of cleaved caspase-3 were then determined from the western blotting results. \* $P < 0.05$  and \*\* $P < 0.01$  vs. the control; # $P < 0.05$  vs. the DADS treatment group. Data are presented as the mean  $\pm$  standard error of the mean ( $n=3$ ). One-way analysis of variance was used for analysis. DADS, diallyl disulfide; PI, propidium iodide; miR, microRNA; GAPDH, glyceraldehyde-3-phosphate dehydrogenase.

compared with DADS treatment alone. Furthermore, to investigate whether the PI3K/Akt signaling pathway participates in DADS-induced anticancer effects, the cell viability was measured after SGC-7901 cells were exposed to DADS and/or

LY294002. Without LY294002, the cell viability reduced as the concentration of DADS increased (200  $\mu$ M DADS,  $P=0.044$  vs. control; 400  $\mu$ M DADS,  $P=0.031$  vs. control). However, following treatment with LY294002, the effect of

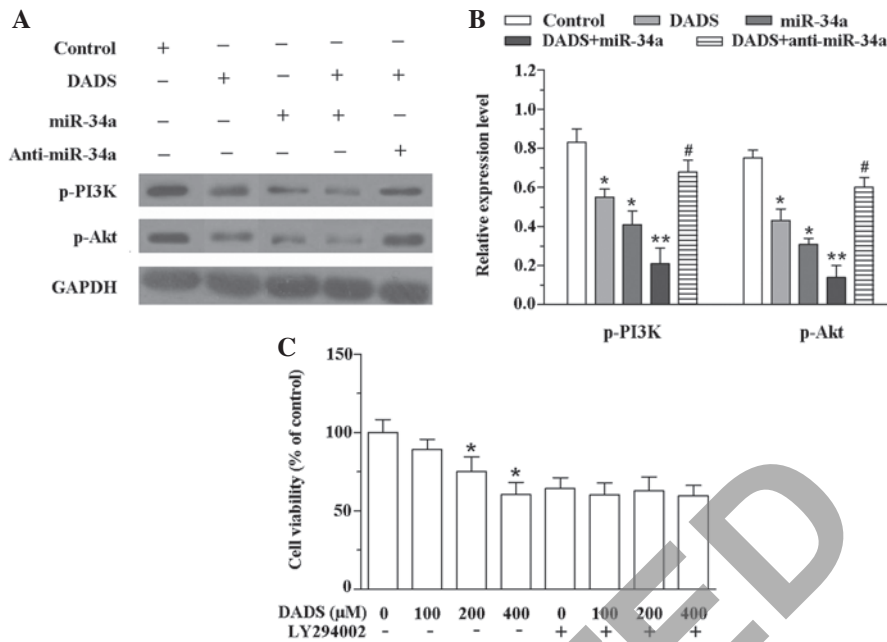


Figure 4. Effect of DADS and miR-34a on the PI3K/Akt signaling pathway. (A) The expression levels of p-PI3K and p-Akt in SGC-7901 cells following various treatments were determined using western blot analysis. (B) The relative levels of p-PI3K and p-Akt were then determined from the western blotting results. DADS, miR-34a mimics and DADS plus miR-34 mimics decreased the expression levels of p-PI3K and p-Akt, while downregulation of miR-34a antagonized the role of DADS in SGC-7901 cells. \* $P < 0.05$  and \*\* $P < 0.01$  vs. the control; # $P < 0.05$  vs. the DADS treatment group. (C) The effect of DADS and LY294002 (a PI3K inhibitor) on cell viability. The viability of SGC-7901 cells was reduced with the increase in the concentration of DADS, but in the presence of LY294002, the cell viability was not obviously reduced even when the concentration of DADS was increased. \* $P < 0.05$  vs. 0  $\mu\text{M}$  DADS. Data are presented as the mean  $\pm$  standard error of the mean ( $n=3$ ). One-way analysis of variance was used for analysis. DADS, diallyl disulfide; miR, microRNA; p-, phosphorylated; PI3K, phosphoinositide 3-kinase; GAPDH, glyceraldehyde-3-phosphate dehydrogenase.

DADS on cell viability was inhibited (Fig. 4C). These results indicate that upregulation of miR-34a by DADS suppresses invasion and induces apoptosis in SGC-7901 cells through inhibition of the PI3K-Akt signaling pathway.

## Discussion

The significant roles of miRs in the regulation of gene expression in the development of GC has been demonstrated by gene expression profiling studies (23,24). However, the expression levels and function of miRs in normal and GC cells remain to be investigated (25). Human miR-223, which is regulated by the transcription factor Twist, was observed to be overexpressed in GC and is associated with poor distal metastasis-free survival. Additional investigation revealed that miR-223 induced the migration and invasion of GC cells *in vitro* and *in vivo* via inhibition of erythrocyte membrane protein band 4.1-like 3 (24). A number of other miRs are also involved in regulation of the biological behavior of GC cells. Overexpression of miR-21 promoted BGC-823 cell growth, invasion and cell migration *in vitro*, while downregulation of miR-21 demonstrated an inhibitory effect on GC cells. Furthermore, inhibition of miR-21 may upregulate the expression levels of phosphatase and tensin homolog (PTEN), indicating PTEN as a target gene for GC initiation and development (26). By contrast, upregulation of miR-34a induced by DADS in GC has been reported (27). However, little is known about the underlying mechanisms. In the present study, it was demonstrated that DADS upregulated miR-34a in SGC-7901 cells. In addition, concurrent miR-34a and DADS treatment suppressed the invasion capacity and induced apoptosis of SGC-7901 cells

via activation of the PI3K-Akt signaling pathway. A number of previous studies have shown that miRs participate in the regulation of GC via certain alternative signaling pathways. For example, miR-200b and miR-22 synergistically inhibited GC growth and enhanced the antitumor effect of DADS *in vitro* and *in vivo* via the Wnt-1 signaling pathway (27). Furthermore, miR-372 regulated cell growth, cell cycle and apoptosis of human GC cells via downregulation of a tumor suppressor gene, large tumor suppressor kinase 2 (28).

PI3K induces the lipid second messenger phosphatidylinositol-3,4,5-triphosphate (PIP3) at the cell membrane. PIP3 in turn enhances the activation of downstream targets involving the serine-threonine protein kinase Akt (29). The PI3K/Akt signaling pathway is able to regulate fundamental cellular functions, including cell growth, survival and motility (30). Aberrant activation of the PI3K-Akt signaling pathway has been implicated in tumor development and in tumor response to cancer treatment (31,32). Therefore, a large number of novel 'targeted agents' have been designed to target the PI3K/Akt signaling pathway (32). Recently, several studies have reported that amplification of PI3K-Akt signaling leads to the development of GC (33-35). Therefore, targeting and downregulating the activation of PI3K/Akt signaling may be a promising approach for GC therapy. Activation of the PI3K/Akt signaling pathway enhances tumor metastasis in GC, followed by the induction of NF- $\kappa$ B and matrix metalloproteinase 9 activity (33). In addition, Shrivastava *et al* (35) observed that P1 Piperlongumine inhibits the growth potential of GC cells by targeting the PI3K/Akt/mammalian target of rapamycin signaling pathway. In agreement with the aforementioned studies, the present study detected the expression levels of p-PI3K, p-Akt and

cleaved caspase-3 in SGC-7901 cells following treatment with DADS, and observed that DADS induced GC cell apoptosis by aberrant activation of the PI3K-Akt signaling pathway. DADS is derived from garlic and is beneficial to human health, particularly due its protective effects against carcinogenesis and chemically-induced toxicity, meaning that DADS may be used safely and is a potential novel candidate for GC therapy (36).

DADS inhibits the initiation and promotion phases of certain types of cancer in rodent models (37,38). A number of mechanisms have been proposed to explain the anticarcinogenic properties of DADS. DADS inhibited human colon tumor cell proliferation via inhibition of histone deacetylase activity, leading to histone hyperacetylation and an increase in p21<sup>waf1/cip1</sup> expression (38). Treatment with DADS induced G2/M phase arrest of GC cells by activation of p38 mitogen-activated protein kinase pathways (5). DADS also suppressed the growth of human prostate cancer xenografts *in vivo* via induction of Bcl-2-associated X protein and Bcl-2 homologous antagonist killer (8). In the present study, DADS suppressed invasion and induced apoptosis in human GC cells via inhibition of the PI3K-Akt signaling pathway by upregulation of miR-34a.

In conclusion, the present study demonstrated that DADS treatment upregulates the miR-34a expression levels in SGC-7901 cells. Furthermore, miR-34a synergistically enhanced the anti-invasion effect and induced apoptosis following treatment with DADS in SGC-7901 cells. DADS appeared to suppress cell invasion capacity and induce apoptosis via inactivating the PI3K/Akt signaling pathway. While DADS is an effective anticancer agent, the molecular mechanisms involved in the regulation of cell cycle control remain to be elucidated for the development of novel anticancer strategies.

## References

- Cao W, Fan R, Wang L, Cheng S, Li H, Jiang J, Geng M, Jin Y and Wu Y: Expression and regulatory function of miRNA-34a in targeting survivin in gastric cancer cells. *Tumor Biol* 34: 963-971, 2013.
- Xie B, Zhou J, Shu G, Liu DC, Zhou J, Chen J and Yuan L: Restoration of klotho gene expression induces apoptosis and autophagy in gastric cancer cells: Tumor suppressive role of klotho in gastric cancer. *Cancer Cell Int* 13: 18, 2013.
- Hartgrink HH, Jansen EP, van Grieken NC and van de Velde CJ: Gastric cancer. *Lancet* 374: 477-490, 2009.
- Li Z, Wang Y, Dong S, Ge C, Xiao Y, Li R, Ma X, Xue Y, Zhang Q, Lv J, *et al*: Association of CXCR1 and 2 expressions with gastric cancer metastasis *in vivo* and tumor cell invasion *in vitro*. *Cytokine* 69: 6-13, 2014.
- Yuan JP, Wang GH, Ling H, Su Q, Yang YH, Song Y, Tang RJ, Liu Y and Huang C: Diallyl disulfide-induced G2/M arrest of human gastric cancer MGC803 cells involves activation of p38 MAP kinase pathways. *World J Gastroenterol* 10: 2731-2734, 2004.
- Zhang XD, Shu YQ, Liang J, Zhang FC, Ma XZ, Huang JJ, Chen L, Shi GM, Cao WG, Guo CY, *et al*: Combination chemotherapy with paclitaxel, cisplatin and fluorouracil for patients with advanced and metastatic gastric or esophagogastric junction adenocarcinoma: A multicenter prospective study. *Chin J Cancer Res* 24: 291-298, 2012.
- Liu X, Liu Q, Fan Y, Wang S, Liu X, Zhu L, Liu M and Tang H: Downregulation of PPP2R5E expression by miR-23a suppresses apoptosis to facilitate the growth of gastric cancer cells. *FEBS Lett* 588: 3160-3169, 2014.
- Xiao D, Lew KL, Kim YA, Zeng Y, Hahm ER, Dhir R and Singh SV: Diallyl trisulfide suppresses growth of PC-3 human prostate cancer xenograft *in vivo* in association with Bax and Bak induction. *Clin Cancer Res* 12: 6836-6843, 2006.
- Lei XY, Yao SQ, Zu XY, Huang ZX, Liu LJ, Zhong M, Zhu BY, Tang SS and Liao DF: Apoptosis induced by diallyl disulfide in human breast cancer cell line MCF-7. *Acta Pharmacol Sin* 29: 1233-1239, 2008.
- Robert V, Mouillé B, Mayeur C, Michaud M and Blachier F: Effects of the garlic compound diallyl disulfide on the metabolism, adherence and cell cycle of HT-29 colon carcinoma cells: Evidence of sensitive and resistant sub-populations. *Carcinogenesis* 22: 1155-1161, 2001.
- Knowles LM and Milner JA: Diallyl disulfide induces ERK phosphorylation and alters gene expression profiles in human colon tumor cells. *J Nutr* 133: 2901-2906, 2003.
- Gunadharini DN, Arunkumar A, Krishnamoorthy G, Muthuvel R, Vijayababu MR, Kanagaraj P, Srinivasan N, Aruldas MM and Arunakaran J: Antiproliferative effect of diallyl disulfide (DADS) on prostate cancer cell line LNCaP. *Cell Biochem Funct* 24: 407-412, 2006.
- Wen J, Zhang Y, Chen X, Shen L, Li GC and Xu M: Enhancement of diallyl disulfide-induced apoptosis by inhibitors of MAPKs in human HepG2 hepatoma cells. *Biochem Pharmacol* 68: 323-331, 2004.
- Tu F, Pan ZX, Yao Y, Liu HL, Liu SR, Xie Z and Li QF: miR-34a targets the inhibin  $\beta$  B gene, promoting granulosa cell apoptosis in the porcine ovary. *Genet Mol Res* 13: 2504-2512, 2014.
- Forte E, Salinas RE, Chang C, Zhou T, Linnstaedt SD, Gottwein E, Jacobs C, Jima D, Li QJ, Dave SS and Luftig MA: The Epstein-Barr virus (EBV)-induced tumor suppressor microRNA MiR-34a is growth promoting in EBV-infected B cells. *J Virol* 86: 6889-6898, 2012.
- Hwang HW and Mendell JT: MicroRNAs in cell proliferation, cell death, and tumorigenesis. *Br J Cancer* 94: 776-780, 2006.
- Garzon R, Calin GA and Croce CM: MicroRNAs in cancer. *Annu Rev Med* 60: 167-179, 2009.
- Di Leva G, Garofalo M and Croce CM: MicroRNAs in cancer. *Annu Rev Pathol* 9: 287-314, 2014.
- Ueda T, Volinia S, Okumura H, Shimizu M, Taccioli C, Rossi S, Alder H, Liu CG, Oue N, Yasui W, *et al*: Relation between microRNA expression and progression and prognosis of gastric cancer: A microRNA expression analysis. *Lancet Oncol* 11: 136-146, 2010.
- Guo J, Miao Y, Xiao B, Huan R, Jiang Z, Meng D and Wang Y: Differential expression of microRNA species in human gastric cancer versus non-tumorous tissues. *J Gastroenterol Hepatol* 24: 652-657, 2009.
- Liu R, Zhang C, Hu Z, Li G, Wang C, Yang C, Huang D, Chen X, Zhang H, Zhuang R, *et al*: A five-microRNA signature identified from genome-wide serum microRNA expression profiling serves as a fingerprint for gastric cancer diagnosis. *Eur J Cancer* 47: 784-791, 2011.
- Livak and Schmittgen: Analysis of relative gene expression data using real-time quantitative PCR and the 2- $\Delta\Delta C_t$  method. *Methods* 25: 402-408, 2001.
- McManus MT: MicroRNAs and cancer. *Semin Cancer Biol* 13: 253-258, 2003.
- Li X, Zhang Y, Zhang H, Liu X, Gong T, Li M, Sun L, Ji G, Shi Y, Han Z, *et al*: miRNA-223 promotes gastric cancer invasion and metastasis by targeting tumor suppressor EPB41L3. *Mol Cancer Res* 9: 824-833, 2011.
- Ambros V: MicroRNA pathways in flies and worms: Growth, death, fat, stress, and timing. *Cell* 113: 673-676, 2003.
- Zhang BG, Li JF, Yu BQ, Zhu ZG, Liu BY and Yan M: microRNA-21 promotes tumor proliferation and invasion in gastric cancer by targeting PTEN. *Oncol Rep* 27: 1019-1026, 2012.
- Tang H, Kong Y, Guo J, Tang Y, Xie X, Yang L, Su Q and Xie X: Diallyl disulfide suppresses proliferation and induces apoptosis in human gastric cancer through Wnt-1 signaling pathway by up-regulation of miR-200b and miR-22. *Cancer Lett* 340: 72-81, 2013.
- Cho WJ, Shin JM, Kim JS, Lee MR, Hong KS, Lee JH, Koo KH, Park JW and Kim KS: miR-372 regulates cell cycle and apoptosis of ags human gastric cancer cell line through direct regulation of LATS2. *Mol Cells* 28: 521-527, 2009.
- Luo J, Manning BD and Cantley LC: Targeting the PI3K-Akt pathway in human cancer: Rationale and promise. *Cancer Cell* 4: 257-262, 2003.
- West KA, Castillo SS and Dennis PA: Activation of the PI3K/Akt pathway and chemotherapeutic resistance. *Drug Resist Updat* 5: 234-248, 2002.

31. Vivanco I and Sawyers CL: The phosphatidylinositol 3-kinase AKT pathway in human cancer. *Nat Rev Cancer* 2: 489-501, 2002.
32. Fresno Vara JA, Casado E, de Castro J, Cejas P, Belda-Iniesta C and González-Barón M: PI3K/Akt signalling pathway and cancer. *Cancer Treat Rev* 30: 193-204, 2004.
33. Kang MH, Oh SC, Lee HJ, Kang HN, Kim JL, Kim JS and Yoo YA: Metastatic function of BMP-2 in gastric cancer cells: The role of PI3K/AKT, MAPK, the NF- $\kappa$ B pathway and MMP-9 expression. *Exp Cell Res* 317: 1746-1762, 2011.
34. Yun SM: Abstract 4304: PPP1R1B-STARD3 fusion gene promotes gastric tumorigenesis through activation of PI3K/Akt signaling. *Cancer Res* 73: 4304, 2013.
35. Shrivastava S, Jeengar MK, Thummuri D and Naidu VGM: P1 Piperlongumine inhibits growth potential of gastric cancer cells by targeting PI3K/Akt/mTOR signaling pathway. *Eur J Cancer* 50: S9, 2014.
36. Lee IC, Kim SH, Baek HS, Beak HS, Moon C, Kim SH, Kim YB, Yun WK, Kim HC and Kim JC: Protective effects of diallyl disulfide on carbon tetrachloride-induced hepatotoxicity through activation of Nrf2. *Environ Toxicol* 30: 538-548, 2015.
37. Takahashi S, Hakoi K, Yada H, Hirose M, Ito N and Fukushima S: Enhancing effects of diallyl sulfide on hepatocarcinogenesis and inhibitory actions of the related diallyl disulfide on colon and renal carcinogenesis in rats. *Carcinogenesis* 13: 1513-1518, 1992.
38. Druesne N, Pagniez A, Mayeur C, Thomas M, Cherbuy C, Duée PH, Martel P and Chaumontet C: Diallyl disulfide (DADS) increases histone acetylation and p21(waf1/cip1) expression in human colon tumor cell lines. *Carcinogenesis* 25: 1227-1236, 2004.

RETRACTED