

# MiR-593 mediates curcumin-induced radiosensitization of nasopharyngeal carcinoma cells via MDR1

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**Abstract.** Curcumin (Cur) exhibits radiosensitization effects to a variety of malignant tumors. The present study investigates the radiosensitizing effect of Cur on nasopharyngeal carcinoma (NPC) cells and whether its mechanism is associated with microRNA-593 (miR-593) and multidrug resistance gene 1 (MDR1). A clonogenic assay was performed to measure the radiosensitizing effect. The expression of miR-593 and MDR1 was analyzed by quantitative polymerase chain reaction (qPCR) or western blot assay. A transplanted tumor model was established to identify the radiosensitizing effect *in vivo*. A luciferase-based reporter was constructed to evaluate the effect of direct binding of miR-593 to the putative target site on the 3' UTR of MDR1. The clonogenic assay showed that Cur enhanced the radiosensitivity of cells. Cur (100 mg/kg) combined with 4 Gy irradiation inhibited the growth of a transplanted tumor model *in vivo*, resulting in the higher inhibition ratio compared with the radiotherapy-alone group. These results demonstrated that Cur had a radiosensitizing effect on NPC cells *in vivo* and *in vitro*; Cur-mediated upregulation of miR-593 resulted in reduced MDR1 expression, which may promote radiosensitivity of NPC cells.

## Introduction

Nasopharyngeal carcinoma (NPC), a type of malignant head-and-neck cancer, is a geographical predilection in South-east Asia, particularly in the Southern provinces of China with

an estimated incidence rate of ~20-50/100,000 (1,2). Despite the specificity of the pathological location and the anatomical structure, the majority of patients are unfortunately diagnosed at advanced stage (3). Currently, radiotherapy is the mainstay of treatment for NPC. However, a high proportion of NPC patients, particularly patients with advanced NPC, exhibit radioresistance and exhibit poor outcomes (4,5). Therefore, exploring the molecular mechanisms underlying sensitivity to radiation or resistance as well as the combination of chemo- and radiotherapy is of crucial significance for NPC therapy.

Curcumin (diferuloylmethane, Cur) is a non-flavonoid polyphenol derived from turmeric plants that demonstrates great potential in tumorigenesis and tumor progression (6,7). Accumulating evidence has indicated that Cur can sensitize numerous radioresistant tumor cells, including NPC (8-10). Our previous work also demonstrated that Cur could enhance the radiosensitivity in the CNE2 cell line at an appropriate concentration (11,12). Although recent discoveries have revealed the underlying mechanisms to a certain extent, the relevance of the association between Cur and its radiosensitivity to NPC and the function of related miRNAs are not fully recognized.

MicroRNAs (miRNAs) are a class of small endogenous non-coding RNA that can regulate gene expression at the post-transcriptional level by inhibiting translation of messenger RNA and by inducing mRNA degradation (13). Recent studies indicate that various biological and pathological processes, including cellular proliferation, differentiation, and apoptosis, have been caused by the deregulation of mRNAs (14-16). Among the mRNAs, miR-593 has been reported to be down-exposed in esophageal and gastric cancer (17). The aim of the present study was to investigate whether miR-593 could radiosensitize the NPC cell line CNE2 *in vitro* and *in vivo* by regulating multidrug resistance gene 1 (MDR1), and to identify whether MDR1 is a direct and functional target of miR-593.

## Materials and methods

**Cell culture and reagents.** The human NPC cell line CNE2 was obtained from Sun Yat-sen University (Guangzhou, China) and cultured in RMPI-1640 medium (Invitrogen; Thermo Fisher Scientific, Inc., Waltham, MA, USA) supplemented with 10% fetal bovine serum (Gibco; Thermo Fisher

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Scientific, Inc.) in a humid atmosphere containing 5% CO<sub>2</sub> at 37°C. Cur (Sigma-Aldrich, St. Louis, MO, USA) was dissolved in 0.5% DMSO (Sigma-Aldrich) and diluted with RPMI-1640 medium to the desired concentrations.

**Clonogenic survival assay.** Cells were divided into three groups: control group (CN), IR (irradiation) group (CX), and IR+Cur group (JX). Cells were seeded in 6-well plates and routinely cultured for 24 h. The CX and JX groups were irradiated with X-rays at 6 MV using 600C/D linear accelerator (Varian Medical Systems, Inc., Palo Alto, CA, USA) to deliver the indicated doses (2, 4, 6 and 8 Gy), and all cells were further cultured for another 12 days. The cells were fixed and stained with methanol containing 1% crystal violet (Beijing Dingguo Changsheng Biotech Co., Ltd., Beijing, China). Colonies ( $\geq 50$  cells) were counted under the microscope (U-LH100L-3; Olympus Corporation, Tokyo, Japan) by using the following formula: Plate clone formation efficiency = number of colonies/number of cells inoculated (18).

**Animal studies.** The study was conducted in accordance with the guideline for the Administration of Affairs Concerning Experimental Animals, and all procedures involving animals were approved by Animal Care and Ethics Committee of Southern Medical University. Balb/c nude mice (Experimental Animal Center of Southern Medical University, Weifang, China) for tumor implantation were 4-6 weeks old with a body mass of 18-22 g. The mice were housed under controlled laboratory conditions at an ambient temperature of 23 $\pm$ 2°C for 2 weeks.

For the xenograft tumor assay, 1 $\times 10^6$  cells in 200  $\mu$ l of RPMI-1640 were injected subcutaneously into the right flank of nude mice. The mice were divided into 5 groups of 6 mice when the tumor volume reached 61 mm<sup>3</sup>. Cur was intragastrically administered at 3 different dosages [50, 100 (which was determined to be the optimal concentration of Cur, according to the results of a preliminary test) and 150 mg/kg] once a day for 7 days. Saline was injected as a control. After 7 days, all groups were irradiated with 4 Gy IR every other day, 3 times. Following the final treatment, mice were euthanized, and the tumors were dissected and weighed.

**Reverse transcription-quantitative polymerase chain reaction (RT-qPCR) analysis.** Total RNA from cells and nude mice was extracted using TRIzol reagent (Invitrogen; Thermo Fisher Scientific, Inc.), following the manufacturer's instructions. Quantification was performed using the Quantitect SYBR Green PCR Kit (Stratagene, San Diego, CA, USA) with an MX3005P multiplex quantitative qPCR system (Stratagene), according to the manufacturer's instructions. GAPDH and U6 were used as the internal control for detecting mRNA and miRNA, respectively. The comparative  $\Delta\Delta C_q$  method was employed as previously described (19). The fold changes were calculated according to 2 <sup>$-\Delta\Delta C_q$</sup>  equation. All of the primers used are listed in Table I.

**Western blot analysis.** The portion of xenograft tumors were extracted by RIPA buffer with protease inhibitors (Cell Biolabs Inc., San Diego, CA, USA) and quantified by the BCA method (Thermo Fisher Scientific, Inc.). Equal amounts of total protein

Table I. Primers used in this study.

Primer name	Primer sequence
GAPDH	F: ATCATCAGCAATGCCTCCTG R: ATGGACTGTGGTCATGAGTC
MDR1	F: TCATTCGAGTAGCGGCTCTT R: CTTCTTTGCTCCTCCATTGC
U6	F: CTCGCTTCGGCAGCACA R: AACGCTTCACGAATTTGCGT
miR-593	F: TGTCTCTGCTGGGGTTTCT R: GTGCAGGGTCCGAGGTATT
pGL3-MDR1	F: gattatagaACTCTGACTGTATGAGATGT R: gattatagaTCACATGAAAGTTTAGT
si-MDR1	S: CAGAAAGCUUAGUACCAAdTdT
Si-NC	S: UAACGACGCGACGACGUAAAdTdT

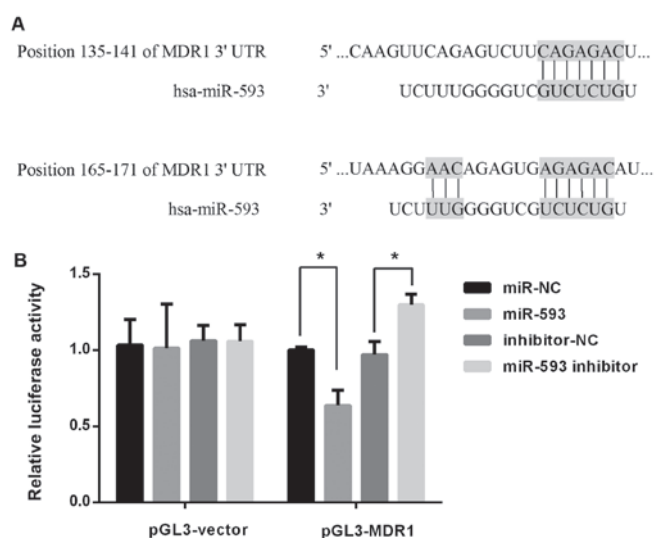


Figure 1. Effect of the putative miR-593 binding site derived from the MDR1 3'UTR on luciferase expression. (A) Schematic of the potential miR-593 binding sites containing MDR1 3'UTR. (B) Luciferase activity in HEK293 cells transfected with miR-NC, miR-593 mimics, inhibitor-NC, and miR-593 inhibitor with pGL3-vector or pGL3-MDR1 36 h post-transfection. Data represent mean  $\pm$  standard deviation of 3 independent experiments, and each experiment was performed in triplicate. \*P<0.05. MDR1, multidrug resistance gene 1; 3'UTR, 3'-untranslated region; miR, micro RNA.

(30-50  $\mu$ g) were resolved by 10% SDS-PAGE (Bio-Rad Laboratories, Inc., Hercules, CA, USA) and transferred onto the PVDF membrane (Thermo Fisher Scientific, Inc.), which was blocked in 5% non-fat milk at room temperature for 1 h. Thereafter, they were incubated with rabbit monoclonal anti-MDR1 antibody (1:2,000; catalogue no. ab170904; Abcam, Cambridge, UK) overnight at 4°C. The membranes were blotted for 1 h at room temperature with goat anti-rabbit horseradish peroxidase-conjugated immunoglobulin G secondary antibody (1:1,000; catalogue no. 7074; Cell Signaling Technology, Inc., Danvers, MA, USA). The bands were developed using ECL Kit (Cell Signaling Technology, Inc.) and quantified on Gel Logic 2200 PRO Imaging System (Kodak, Rochester, NY, USA).

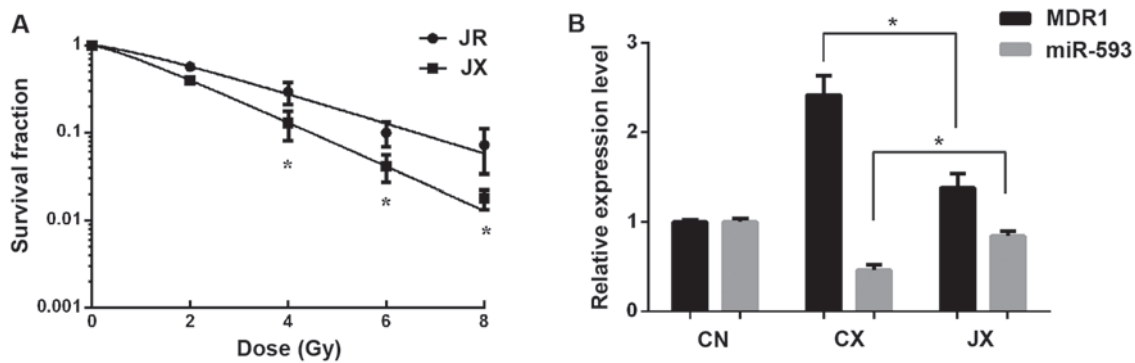


Figure 2. Cur sensitizes nasopharyngeal carcinoma cells to irradiation treatment through upregulating mir-593 to reverse IR induced MDR1 expression. (A) CNE2 cells were treated with 0, 2, 4, 6, or 8 Gy of IR with Cur pretreatment. The cell survival fraction was calculated by clonogenic assay. (B) Relative miR-593 and MDR1 expression were determined by quantitative polymerase chain reaction. IR-induced miR-593 downregulation and MDR1 upregulation were reversed during Cur-enhanced radiosensitization. Data are presented as mean  $\pm$  standard deviation from 3 replicate experiments. \* $P < 0.05$ . CN, control group; CX, irradiated group; JX, irradiation + Cur group; Cur, curcumin; MDR1, multidrug resistance gene 1; IR, irradiation; miR, micro RNA.

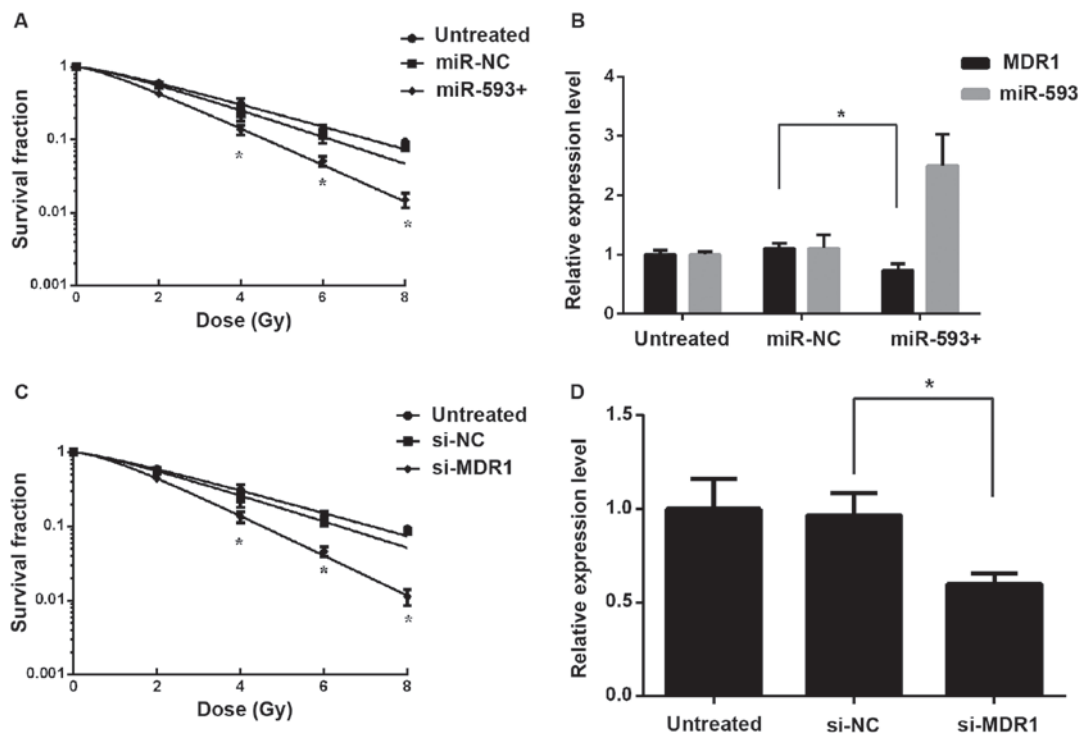


Figure 3. Effect of MDR1 knockdown on radiosensitivity of nasopharyngeal carcinoma cells *in vitro*. (A) Survival fraction was calculated by clonogenic assay after treatment with miR-593 mimics (miR-593+) and (B) relative miR-593 and MDR1 expression were determined by qPCR. (C) Survival fraction after treatment with si-MDR1 and (D) relative miR-593 and MDR1 expression were determined by qPCR. All data are presented as mean  $\pm$  standard deviation from 3 replicate experiments. \* $P < 0.05$ . Cur, curcumin; MDR1, multidrug resistance gene 1; si, short interfering; qPCR, quantitative polymerase chain reaction; miR, micro RNA.

**Luciferase assay.** The entire 380-base pair fragment of MDR1 3' UTR that contains the putative binding site of miR-593 was amplified by PCR and cloned downstream of the luciferase gene at the XbaI sites in the pGL3-3 plasmid (Promega Corporation, Madison, WI, USA). The construct was designated as pGL3-MDR1 (primers are listed in Table I). HEK293 cells were co-transfected with 30 pmol of either miR-593 mimics or miR-593 NC and pGL3-MDR1 using Lipofectamine 2000 (Invitrogen; Thermo Fisher Scientific, Inc.). Luciferase activity was measured using the Promega dual-luciferase assay kit and normalized by  $\beta$ -galactosidase activity. Relative protein levels were expressed as Renilla/firefly luciferase ratios. Each experiment was repeated twice (11).

**RNA interference.** CNE2 cells at 20-30% confluence were transfected with 50 nM of siRNAs using Lipofectamine 2000 following the manufacturer's protocol. Small interfering RNA (siRNA) and scrambled negative control siRNA (siRNA-NC) were obtained from Invitrogen (Thermo Fisher Scientific, Inc.). The target sequence of MDR1 is listed in Table I. A total of 36 h after transfection, cells were harvested for RT-qPCR.

**Statistics.** SPSS software, version 13.0 (SPSS Inc., Chicago, IL, USA) was used for all the statistical analyses in the present study. Quantitative data are presented as mean  $\pm$  standard deviation.  $\chi^2$  test and Student's t-test were appropriately applied

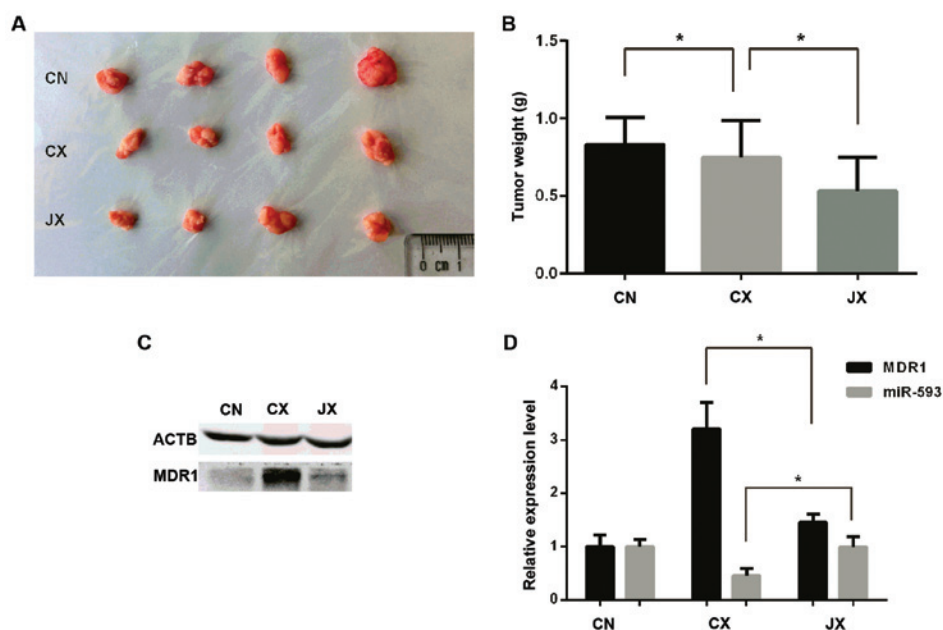


Figure 4. Radiosensitization of Cur in xenograft tumors *in vivo*. (A) Image of tumors in each group. (B) Tumor weight of each group. (C) Relative mir-593 and MDR1 expression were determined by (C) western blot assay or (D) qPCR. All data are presented as mean  $\pm$  standard deviation, \* $P < 0.05$ . Cur, curcumin; MDR1, multidrug resistance gene 1; qPCR, quantitative polymerase chain reaction; miR, micro RNA; CN, control group; CX, irradiated group; JX, irradiation + Cur group.

for different types of data.  $P < 0.05$  was considered to indicate a statistically significant difference.

## Results

**MDR1 is a direct target of miR-593.** A bioinformatics analysis was performed using TargetScan version 7.0 software (<http://www.targetscan.org/>), which predicted that miR-593 may target the MDR1 3'UTR region (Fig. 1A). A luciferase-based reporter was constructed to evaluate the effect of miR-593 direct binding to the putative target site on the 3'UTR of MDR1. To substantiate the assumption that miR-593 can directly repress MDR1, the reporter-construct pGL3-vector and pGL3-MDR1 were co-transfected with miR-593 mimic, miR-NC, and miR-593 inhibitor or inhibitor-NC to HEK293 cells. Luciferase activity was then assayed. As shown in Fig. 1B, for pGL3-MDR1 construct, miR-593 mimic significantly lowered luciferase activity compared with miR-NC ( $P < 0.05$ ). By contrast, miR-593 inhibitor increased luciferase activity in HEK293 cells compared with inhibitor-NC ( $P < 0.05$ ). These findings support the hypothesis that miR-593 directly targets MDR1 expression.

**Cur sensitizes CNE2 cells to IR through upregulating mir-593 to reverse IR-induced MDR1 expression.** The colony survival assay is considered a canonical standard to determine radiosensitivity (20). The results confirmed that cells pretreated with Cur were much more sensitive to IR than their untreated counterparts. The  $\alpha$  and  $\beta$  components were 0.2476/Gy and 0.01650/Gy<sup>2</sup> for the cells treated with radiation alone, and 0.4201/Gy and 0.02029/Gy<sup>2</sup> for the cells treated with the combination treatment (Fig. 2A), respectively, leading to significantly different ( $P < 0.001$ ) survival fractions, as tested with the linear regression analysis. The data were further analyzed according to the multitarget single-hit model. When pretreated with 10  $\mu$ M Cur ( $IC_{20}$ ), the sensitization enhancement ratio of

CNE-2 reached 1.44. These data indicate that Cur has an effective radiosensitization effect on the CNE-2 cell line *in vitro*. In addition, miR-593 and MDR1 expression were detected by RT-qPCR (Fig. 2B). The result demonstrated that IR-induced miR-593 downregulation was reversed during Cur-enhanced radiosensitization ( $P < 0.05$ ).

**The effect of MDR1 knockdown on the radiosensitivity of NPC cells in vitro.** Cur enhances the radiosensitivity involving the reversal of differentially expressed miR-593 and MDR1. To elucidate whether the effect of Cur on radiosensitivity was mediated by repression of MDR1, miR-593 mimics or si-MDR1 were transfected into CNE2 cells. A clonogenic assay suggested that the ectopic expression of MDR1 significantly reduced miR-593-induced radiosensitivity (Fig. 3A and C), which was consistent with the results of RT-qPCR (Fig. 3B and D). These observations indicated that Cur may have sensitized cells to IR by stimulating miR-593 to downregulate the expression of MDR1.

**Radiosensitization of Cur in xenograft tumors in vivo.** The tumor weight of each mouse in each treatment group was measured (Fig. 4A). A single dose of 4 Gy irradiation (CX group) exhibited the expected effect of tumor growth inhibition compared with the untreated CN group ( $P < 0.05$ , Fig. 4B). However, the JX group (Cur 100 mg/kg and 4 Gy irradiation) had the highest inhibition ratio (62.18%) and expressed a significant tumor growth inhibition effect compared with the CX group ( $P < 0.05$ , Fig. 4B). Mice body weights were monitored to assess the tolerability of systemic Cur. Body weight changes over the course of the experiment were minimal in all treatment groups, suggesting that Cur is well tolerated (data not shown).

The expression level of miR-593 was significantly higher in the JX group compared with the CX group. To investigate the regulation of MDR1 by miR-593, the relative expression of MDR1 was measured by western blot assay (Fig. 4C) and



RT-qPCR (Fig. 4D). In accordance with the altered expression level of miR-593, MDR1 was significantly downregulated in the JX group. These findings also indicate that MDR1 is a target of miR-593.

## Discussion

Radiotherapy is considered one of the most effective treatments for patients with NPC, and radioresistance is the main risk factor that contributes to poor prognosis (21). Radioresistance occurs in primary IR treatment, and the survived cells may be more resistant to the second IR treatment, thereby leading to radiotherapy failure (12,22,23). In this regard, the exact molecules and signaling pathway involved in radiosensitization should be determined to develop target therapy and enhance the efficacy of radiation. In the present study, it was observed that IR-induced downregulation of miR-593 or upregulation of MDR1 expression was almost reversed by Cur.

Cur regulates the gene expression involved in survival, proliferation, angiogenesis, invasion, and metastasis. This phytochemical also modulates various mechanisms that are associated with radioresistance, including the following: downregulating COX-2, MRP, Bcl-2, and survivin expression; inhibiting PI3K/AKT activation; suppressing growth factor signaling pathways; and inhibiting STAT3 activation (24-26). The present study demonstrated that Cur enhanced radiosensitivity in the NPC cell line CNE2 at 10  $\mu\text{mol/l}$  by MTT or clonogenic survival test (27), although Cur exhibited higher anti-proliferative effects when used alone at a concentration of 20 or 40  $\mu\text{mol/l}$ . Considering the cytotoxicity of Cur and IR, a concentration of 10  $\mu\text{mol/l}$  and 24 h pretreatment was more suitable as a radioenhancer (data not shown). In addition, in the animal studies, Cur 100 mg/kg was chosen as optimal concentration after a preliminary test. Therefore, no significant data were obtained by conjoint analysis with other groups although the single Cur group (JN group) was performed (data not shown). In the present study, the radiosensitizing effect of Cur was evaluated *in vitro* and *in vivo*. The data indicated that the radiosensitizing effect of Cur may be associated with miR-593 and MDR1.

P-glycoprotein (P-gp), encoded by MDR1, has attracted great interest because of its role in MDR in a variety of cancers. P-gp is overexpressed in cancer cells that actively extrude chemotherapeutic agents (28). MDR1 mediates not only chemoprotection by drug efflux but has also been found to inhibit apoptosis induced by chemotherapeutics, death receptor ligands, serum starvation, and UV or ionizing irradiation. Therefore, blocked MDR1 may enhance the efficacy of chemotherapy and reverse radioresistance in patients. Maier *et al* (29) confirmed that a protective effect of retroviral overexpression of MDR1 is increasing the radiotolerance of haematopoietic cells and the related apoptosis gene was downregulated. In the present study, it was also observed that MDR1 was a radioresistant factor which could be downregulated by miR-593.

Certain miRNAs were reported to affect the radiosensitivity of cancer cells, such as let-7, miR-21, miR-101, miR-421, and miR-181a (30-34). In esophageal cancer (EC), miR-593 degrades polo-like kinase 1 (PLK1) mRNA by direct binding to the 3'-UTR of PLK1 mRNA and reduced proliferation of

EC cells (17). Bioinformatics analysis was used in the present study to predict that miR-593 may also target the MDR1 3'UTR region. A luciferase-based reporter showed that miR-593 regulated MDR1 expression by directly targeting 3'-UTR, consistently resulting in reduced expression of MDR1. Furthermore, modulated expression tests demonstrated that radiosensitization of Cur was triggered by miR-593 instead of directly by MDR1.

Taken together, the results demonstrated that Cur had a radiosensitization effect on NPC cells *in vivo* and *in vitro*; curcumin-mediated upregulation of miR-593 causes the depression of MDR1 expression, which may promote radiosensitivity of NPC cells.

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