Clinicopathological relevance of kinesin family member 18A expression in invasive breast cancer

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Abstract. Recently, kinesin motor proteins have been focused on as targets for cancer therapy. Kinesins are microtubule-based motor proteins that mediate diverse functions within the cell, including the transport of vesicles, organelles, chromosomes and protein complexes, as well as the movement of microtubules. In the current study, the expression of kinesin family member 18A (KIF18A), a member of kinesin superfamily, was investigated in breast cancer using immunohistochemistry, and its effect on breast cancer prognosis was examined. KIF18A expression level was significantly associated with lymph node metastasis (P=0.047). In patients with high levels of KIF18A expression, survival was significantly poorer compared to patients with low levels of KIF18A expression (disease-free survival, P=0.030). Multivariate analysis revealed that venous invasion (hazard ratio, 9.22; 95% confidence interval, 3.90-23.66; P<0.001) and KIF18A expression (hazard ratio, 3.20; 95% confidence interval, 1.34-6.09; P=0.010) were independent predictive factors for lymph node metastasis. KIF18A may be a useful predictive marker for lymph node metastasis in breast cancer, which could facilitate curative adjuvant treatment.

Introduction

Breast cancer is one of the most prevalent cancers and is a major public health concern among women. The most recent statistics for Japan document >76,000 cases per year (1), with a mortality rate of >13,000 per year (2). Breast cancer care and research has improved early detection and treatment. However, even after apparently successful localized treatments, there are long-term risks of recurrence and metastasis (3).

Breast cancer is currently classified into subtypes based on immunohistochemical (IHC) classification (4,5). Subtypes are defined by clinicopathological biomarkers, including estrogen receptor (ER), progesterone receptors (PgR), human epidermal growth factor receptor 2 (HER2) and Ki-67; these biomarkers can indicate the optimum treatment and play a key role in breast cancer treatment to improve prognosis (6,7). To facilitate the treatment of cancer, it is important to exploit new biomarkers that can improve the reliability of prognosis prediction, and to develop targeted therapies for breast cancer patients.

Recently, specific kinesin motor proteins have been studied as key proteins that regulate mitotic events and potential targets of therapy (8-11). KIF18A is a member of the kinesin protein superfamily, which is associated with the molecular motor proteins that use ATP hydrolysis to produce force and movement along microtubules (12-17). The basic mechanism for these activities is not well understood. However, recent studies have demonstrated that KIF18A regulates chromosome congression (18) and suppresses kinetochore movements to control mitotic chromosome alignment (19). Chromosome congression relies on the presence of KIF18A, indicating that this motile microtubule depolymerase has a key role in the dynamics of kinetochore microtubules driving chromosome alignment in the pre-anaphase state of the human cell cycle (18-20).

It has been reported that KIF18A is involved in breast, colorectal and hepatocellular cancers, and cholangiocarcinoma (21-24). However, there is essentially no information regarding the clinical relevance of KIF18A in breast cancer patients. Therefore, the present study investigated the clinicopathological significance of KIF18A expression in human breast cancer.

Materials and methods

Patients and specimen collection. Primary breast cancer and paired normal tissues were obtained from the operative specimens of 144 patients, who underwent radical surgery at the Medical Hospital of the Tokyo Medical and Dental University (Tokyo, Japan) between January 2004 and December 2006. Normal tissue was collected from at least 1 cm away from the primary breast cancer site and was histopathologically identified as normal by a pathologist at the Tokyo Medical and Dental University. Written informed consent was obtained...
from all patients according to the guidelines approved by the
Institutional Research Board. Patients ranged in age from 26 to
91 years, with a mean age of 54 years. Patients were excluded
if they received anti-hormone therapy, chemotherapy or radio-
therapy prior to surgery. Patients with non-invasive breast cancer
were also not included in the study. All patients were closely
followed up after surgery at regular 3- to 6-month intervals, and
the total follow-up periods ranged from 3 months to 7.6 years,
with a median of 5.9 years. Following surgery, all patients were
clearly classified into a category of breast cancer based on the
clinicopathological criteria described by the Japanese Society
for Breast Cancer (25). All data, including age, tumor size,
lymphatic invasion, vascular invasion, nuclear grade, lymph
node metastasis, ER status, PgR status, HER2 score, recurrence,
pathological histology and clinical stage, were obtained from
the clinical and pathological records. HER2 status was scored
using the HER2 expression criteria (26). For primary tumors
with a HER2 score of 2+, the IHC results were additionally
validated with fluorescence in situ hybridization. All patients
were treated with anti-hormonal therapy, chemotherapy and/or
radiotherapy subsequent to surgery, according to breast cancer
treatment guidelines in Japan, which were based on St. Gallen
International Breast Cancer Conference and National Compre-
hensive Cancer Network recommendations (27).

**Immunohistochemistry.** IHC studies for KIF18A expression
were performed on formalin-fixed, paraffin-embedded breast
cancer tissues. After tissue sections (4 µm) were deparaffinized
over 5 x 10 min incubations in xylene, the sections were rehy-
drated, and antigen retrieval was performed by incubation in
antigen activation liquid (pH 9.0) in a microwave processor at
98°C for 30 min. Endogenous peroxidase activity was blocked
using a solution of 3% hydrogen peroxide in absolute methanol
for 15 min. A polyclonal rabbit anti-KIF18A antibody (catalog
no., A301-080A; Bethyl Labotatories, Montgomery, TX, USA)
was applied at a dilution of 1:150 and incubated overnight at
4°C. The Histofine Simple Stain MAX PO kit (Nichirei Corpo-
ration, Tokyo, Japan) was used according to the manufacturer’s
instructions to block non-specific binding and to detect bound
primary antibody. The color was developed by diaminobenzi-
dine (Nichirei Corporation) for 10 min at room temperature.
The sections were then counterstained with Mayer's hemato-
ylin. Negative control staining was conducted by substituting
the primary antibody. The color was developed by diaminobenzi-
dine for 10 min at room temperature. The sections were then counterstained with Mayer's hematoxylin. Negative control staining was conducted by substituting non-immune rabbit serum and phosphate-buffered saline for primary antibodies. An expert pathologist selected and evaluatedfive representative fields at 400x magnification from each slide to produce digital photographs for measuring the staining intensity of KIF18A. KIF18A expression was quantified using ImageJ software (Java 1.6.0_30 (32-bit); http://rsb.info.nih. gov/ij/index.html), which calculates the staining intensity of an area by converting RGB pixels to brightness values (11). The 5 most typically stained areas within the tumor were selected for calculating the average value.

**Statistical analysis.** Data from the IHC analysis was analyzed
by JMP 10 software for Windows (version 10.0.1; SAS Institute, Cary, NC, USA). Differences between groups were deter-
mined using the χ2 test and analysis of variance. Disease-free
survival (DFS) rates were calculated actuarially according to
the Kaplan-Meier analysis and compared with the generalized
log-rank test. Variables with a value of P<0.05 in univariate
analysis were used in a subsequent multivariate analysis using
nominal logistic regression. P<0.05 was considered as to indic-
estatistical significance.

**Results**

**KIF18A expression in breast cancer tissues.** KIF18A expression
was assessed by IHC analysis in the primary breast cancer tissue
and normal breast tissue samples. IHC analysis with anti-KIF18A
antibody verified that KIF18A was highly expressed in cancer
cells compared to normal cells (Fig. 1). The cancer and normal
cell types exhibited differential staining patterns: Positive IHC
staining was observed in the nucleus of normal and cancer cells;
positive IHC staining of the cytoplasm was observed predomin-
antly in cancer cells and slightly in normal cells. ImageJ
software was used to quantitatively evaluate the staining inten-
sity of KIF18A in 144 breast cancer samples. Following assay
optimization, a cutoff level of expression was determined as
30.02 (the average expression level of KIF18A in tumor): Breast
cancer specimens <30.02 were assigned to the low expression
group (n=83; 57.6%), whereas those with values ≥30.02 were
assigned to the high expression group (n=61; 42.4%).

Clinicopathological significance of KIF18A expression in
breast cancer tissue. The clinicopathological factors analyzed
in relation to KIF18A expression in breast cancer tissue are shown
in Table I. The incidence of lymph node metastasis was
significantly higher (P=0.047) in the high-expression group
than in the low-expression group. Conversely, no significant
differences were observed with regard to age, menopause
status, tumor stage, lymphovascular invasion, nuclear grade,
hormone status, HER2 status or recurrence.

**DFS analysis.** The 5-year DFS rates in patients with high
KIF18A expression and patients with low KIF18A expression
are presented in Fig. 2. The difference in DFS time between
these two groups was statistically significant (P=0.030; log-rank test). However, the overall survival difference
between these two groups was not statistically significant
datax not shown). Patients received ≥1 postoperative therapy
(anti-hormonal treatment, chemotherapy or radiotherapy).

**Univariate and multivariate analysis.** Univariate and
multivariate logistic regression analyses were performed for
factors affecting lymph node metastasis (Table II). Univariate
analysis revealed a significant association between lymph node
metastasis and the following factors: Tumor size (P=0.009),
lymphatic invasion (P<0.001), venous invasion (P<0.001),
recurrence (P=0.004) and KIF18A expression (P=0.047).
Multivariate analysis of these parameters revealed that
venous invasion (hazard ratio, 9.22; 95% confidence interval,
3.90-23.66; P<0.001) and KIF18A expression (hazard ratio,
3.20; 95% confidence interval, 1.34-6.09; P=0.010) were inde-
dependent predictive factors for lymph node metastasis.

**Discussion**

In recent years, cancer therapy research has focused on
proteins involved in the regulatory events of mitosis (8-11).
Infiltrating growth of cancer cells, which is associated with abnormal, uncontrolled proliferation, requires the biological activation of numerous proteins that serve central roles. Mitotic inhibitor drugs, which include taxanes and vinca alkaloids, act to target microtubules and have yielded various levels of success in the treatment of various types of cancers.

Table I. Clinicopathological significance of the KIF18A expression ratio in breast cancer.

<table>
<thead>
<tr>
<th>Clinicopathological factor</th>
<th>KIF18A expression ratio</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Low</td>
<td>High</td>
</tr>
<tr>
<td>Age, years (mean ± SD)</td>
<td>54.6±12.9</td>
<td>54.0±13.5</td>
</tr>
<tr>
<td>Menopause status</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pre</td>
<td>37</td>
<td>28</td>
</tr>
<tr>
<td>Post</td>
<td>46</td>
<td>33</td>
</tr>
<tr>
<td>Tumor stage</td>
<td></td>
<td></td>
</tr>
<tr>
<td>T1</td>
<td>34</td>
<td>33</td>
</tr>
<tr>
<td>T2-3</td>
<td>49</td>
<td>28</td>
</tr>
<tr>
<td>Lymph node metastasis</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Absent</td>
<td>62</td>
<td>36</td>
</tr>
<tr>
<td>Present</td>
<td>21</td>
<td>25</td>
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<tr>
<td>Lymphatic invasion</td>
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<td></td>
</tr>
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<td>44</td>
<td>30</td>
</tr>
<tr>
<td>Present</td>
<td>39</td>
<td>31</td>
</tr>
<tr>
<td>Venous invasion</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Absent</td>
<td>47</td>
<td>35</td>
</tr>
<tr>
<td>Present</td>
<td>36</td>
<td>26</td>
</tr>
<tr>
<td>Nuclear grade</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Grade 1</td>
<td>47</td>
<td>40</td>
</tr>
<tr>
<td>Grade 2</td>
<td>15</td>
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</tr>
<tr>
<td>Grade 3</td>
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<tr>
<td>Nuclear atypia</td>
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</tr>
<tr>
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</tr>
<tr>
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<td>Score 3</td>
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</tr>
<tr>
<td>Mitotic counts</td>
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<td></td>
</tr>
<tr>
<td>Score 1</td>
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<td>41</td>
</tr>
<tr>
<td>Score 2</td>
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</tr>
<tr>
<td>Score 3</td>
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</tr>
<tr>
<td>Estrogen receptor</td>
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<td></td>
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<tr>
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<td>9</td>
</tr>
<tr>
<td>Present</td>
<td>71</td>
<td>52</td>
</tr>
<tr>
<td>Progesterone receptor</td>
<td></td>
<td></td>
</tr>
<tr>
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<td>22</td>
<td>14</td>
</tr>
<tr>
<td>Present</td>
<td>61</td>
<td>47</td>
</tr>
<tr>
<td>HER2 score</td>
<td></td>
<td></td>
</tr>
<tr>
<td>0-1</td>
<td>69</td>
<td>55</td>
</tr>
<tr>
<td>2-3</td>
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<td>6</td>
</tr>
<tr>
<td>Recurrence</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Absent</td>
<td>69</td>
<td>53</td>
</tr>
<tr>
<td>Present</td>
<td>14</td>
<td>8</td>
</tr>
</tbody>
</table>

<sup>a</sup>P<0.05, statistical significance. KIF18a, kinesin family member 18A; SD, standard deviation; HER2, human epidermal growth factor receptor 2.
Several next-generation mitotic drug targets have been developed, and small molecule inhibitors that have been identified are already under investigation in clinical trials (11).

KIF18A is a member of the kinesin 8 family and has been demonstrated to play important roles in chromosome alignment during mitosis (18-20). Through several in vitro assays, it has been revealed that upregulation of KIF18A may affect the biological characteristics of cancer cells (21-24).

In the current study, the expression of KIF18A in breast cancer tissues and the association between KIF18A expression and clinicopathological factors in breast cancer were explored using IHC analysis. The results revealed that KIF18A protein expression was significantly higher in breast cancer tissues than in normal breast tissues (21). This suggests that breast cancer cells may take advantage of KIF18A overexpression to control mitotic chromosome alignment and increase their rate of repetitive cell division.

Table II. Univariate and multivariate analyses of clinicopathological factors affecting lymph node metastasis.

<table>
<thead>
<tr>
<th>Factors</th>
<th>Univariate analysis</th>
<th>Multivariate analysis</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>HR 95% CI</td>
<td>P-value</td>
</tr>
<tr>
<td>Age (&lt;50/≥51)</td>
<td>0.97 0.48-1.97</td>
<td>0.93</td>
</tr>
<tr>
<td>T stage (T1/T2-3)</td>
<td>2.69 1.30-5.79</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>LI (absent/present)</td>
<td>19.51 7.57-61.01</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>VI (absent/present)</td>
<td>9.97 4.48-23.92</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>ER (absent/present)</td>
<td>1.60 0.58-5.16</td>
<td>0.39</td>
</tr>
<tr>
<td>PgR (absent/present)</td>
<td>0.92 0.41-2.10</td>
<td>0.84</td>
</tr>
<tr>
<td>HER2 (absent/present)</td>
<td>1.51 0.55-3.95</td>
<td>0.41</td>
</tr>
<tr>
<td>Recurrence</td>
<td>3.90 1.54-10.27</td>
<td>0.047</td>
</tr>
<tr>
<td>KIF18A (low/high)</td>
<td>2.05 1.01-4.21</td>
<td>0.047</td>
</tr>
</tbody>
</table>

LI, lymphatic invasion; VI, venous invasion; ER, estrogen receptor; PgR, progesterone receptor; HER2, human epidermal growth factor receptor 2; KIF18A, kinesin family member 18A; HR, hazard ratio; CI, confidence interval.

Figure 1. Representative KIF18A immunohistochemistry images of breast cancer tissues. Positive staining was observed in cancer cells, but very limited in normal cells. (A) Normal breast, magnification x400. (B) Breast cancer, magnification x400. Enlarged views of (C) normal breast and (D) breast cancer tissues, magnification x1,000.
The present study also revealed that KIF18A overexpression in breast cancer was associated with lymph node metastasis and poor prognosis. The group with high KIF18A expression had a poorer prognosis compared with that of the low-expression group in terms of DFS. KIF18A overexpression was prevalent in breast cancer cells and was also associated with prognostic factors and shorter survival time; these results may suggest that the overexpression of this mitotic protein is associated with aggressive primary tumors. To the best of our knowledge, this is the first study to demonstrate the clinical relevance of KIF18A in invasive breast cancer and its relation to disease outcome.

The axillary lymph node status is the most consistent prognostic factor used in adjuvant therapy decision-making (29). Currently, the sentinel node biopsy is a common surgical procedure to determine the stage of the cancer and select an appropriate treatment plan (30,31). In multivariate analysis, KIF18A overexpression in breast cancer was determined to be an independent and significant predictive factor for lymph node metastasis. Based on these findings, in cases where low KIF18A expression is identified prior to breast surgery, it may be possible to avoid performing the sentinel node biopsy in selected patients with clinically and radiologically normal axilla.

In summary, this is the first report of clinicopathological analysis of KIF18A in breast cancer patients. KIF18A expression was correlated with lymph node metastasis and was an independent predictive factor for the lymph node metastasis and DFS. Kaplan-Meier analyses revealed that the DFS rate was significantly lower in the high KIF18A expression group. These findings suggest that KIF18A may be a useful predictive biomarker of lymph node metastasis, which could aid in the development of optimal adjuvant treatments.

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References


