

Candidate miRNAs and pathogenesis investigation for hepatocellular carcinoma based on bioinformatics analysis

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Received November 24, 2015; Accepted January 26, 2017

DOI: 10.3892/ol.2017.5913

Abstract. The present study aimed to explore the mechanisms behind the development and progression of hepatocellular carcinoma (HCC) and identify information regarding HCC-related microRNAs (miRNAs) or marker genes for the gene therapy of HCC. Gene expression profile of GSE67882, generated from 4 hepatitis B virus infected HCC tissue samples (HCC group) and 8 chronic hepatitis B tissue samples with no fibrosis (control group) were downloaded from the Gene Expression Omnibus database. The differentially expressed miRNAs functional enrichment and pathway analyses of HCC were revealed, followed by transcription factor-miRNA interaction network construction and analyses. A total of 14 upregulated miRNAs and 16 downregulated miRNAs between HCC and control samples were obtained. Differentially expressed miRNAs were mainly involved in biological processes like the regulation of histone H3-K9 methylation, and the KEGG pathways in cancer map05200 demonstrates their involvement in cancer. A total of 3 outstanding regulatory networks of miRNAs: *hsa-miR-15a*, *hsa-miR-125b* and *hsa-miR-122* were revealed. A total of 11 differentially expressed miRNAs including *hsa-miR-146b-5p* that regulated the marker genes of HCC were explored. miRNAs such as *hsa-miR-15a*, *hsa-miR-125b*, *hsa-miR-122* and *hsa-miR-146b-5p* may be new biomarkers for the gene therapy of HCC. Furthermore, histone H3-K9 methylation and other pathways in cancer

observed in the KEGG map05200 may be closely related with the development of HCC.

Introduction

Hepatocellular carcinoma (HCC) is a fatal disease (1). Several types of potentially curative treatments have become available due to the advances in technology and surgical techniques (2). Despite these advances in the clinical treatment of HCC, the mortality rate remains high (3). Thus, novel potential therapies for HCC are urgently required.

As potential therapy targets in malignant cells microRNA (miRNA/miR) perform important roles in regulating gene expression (4). Dysregulation of miRNA is widely involved in human cancer, including HCC (5). A previous study indicated that the mutation of the *miR-122* binding site in the interleukin-1 α 3' untranslated region is associated with the incidence rate of HCC (6). As a potential tumor suppressor miRNA, *miR-1285-3p* has been shown to regulate the expression of *JUN* (Jun Proto-Oncogene, AP-1 Transcription Factor Subunit) in the disease progression of HCC (7). The regulation of feedback between miRNAs and target genes performs important roles in the progression of HCC (8). Downregulated *miR-148b* has been shown to be a biomarker for the early detection of HCC (9). Previous miRNA-based HCC gene therapy studies have demonstrated significant inhibition of miRNA in HCC cells, indicating a promising alternative to current therapeutics (10-12). However, the association between miRNA and their target genes, as well as the influence of these associations during the process of HCC is rarely studied.

miRNA expression profiling has proven useful in diagnosing and understanding the development and progression of several diseases including HCC (13-15). The use of miRNA expression profiling as a prognostic biomarker for the detection of HCC is practicable (16). Since chronic hepatitis B virus (HBV) infection is a significant cause of HCC, the present study performed a bioinformatics analysis based on the miRNA expression profile of the HBV infected HCC tissue samples and chronic Hepatitis B (CHB) tissue samples with no fibrosis from 12 patients with HCC. The present study may help to explore the potential disease progression of HCC resulting

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Key words: hepatocellular carcinoma, differentially expressed microRNAs, functional enrichment analysis, pathway analysis, transcription factors

from CHB and provide information regarding HCC-related miRNA/marker genes for the gene therapy of HCC.

Materials and methods

Samples and microarray data. Gene expression profile data GSE67882 was downloaded from the Gene Expression Omnibus (GEO) database (17) (<http://www.ncbi.nlm.nih.gov/geo/>) based on the platform GPL10850; Agilent-021827 Human miRNA Microarray (V3; Agilent Technologies, Inc., Santa Clara, CA, USA). Since the study was performed based on the raw microarray data retrieved from the GEO database, all differentially expressed miRNAs meeting the threshold of adjusted $P < 0.05$ and $\log_2 FC$ (fold change) > 0.52 were analyzed. A total of 14 upregulated miRNAs and 16 downregulated miRNAs were obtained. The present study was performed by using data from 4 HBV infected HCC tissue samples (HCC group) and 8 CHB tissue samples with no fibrosis (control group) of GSE67882 (<https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE67882>). The microarray data was downloaded on July 24, 2015, and the bioinformatic analysis was completed on August 12, 2015.

Data preprocessing and differential expression analysis. The normalization (robust estimation of variance-stabilizing and calibrating transformations) of gene expression profile data was performed using Vsn (18) (<http://bioconductor.org/help/search/index.html?q=vsu/>) of Bioconductor in R (v.3.0.0). The Linear Models for Microarray Data (limma, <http://www.bioconductor.org/packages/release/bioc/html/limma.html>) package (19) in Bioconductor was applied to identify differentially expressed miRNAs by comparing the expression levels between samples in the HCC group and the control group. The corresponding P-value of miRNAs subsequent to an unpaired *t*-test was defined as the adjusted P-value. Subsequently, the adjusted $P < 0.05$ and $\log_2 FC > 0.52$ were selected as the threshold values for screening.

Analysis for the target genes regulated by differentially expressed miRNAs. Since the inhibition of gene expression following transcription is realized by the combination of miRNA and mRNA, differentially expressed miRNAs in HCC samples were further analyzed to explore the significance of miRNA in liver cancer. In the present study, two experimental certificate databases including miRecords (20) and miWalk (21) were used to explore suitable miRNA-target gene relations. The relations that existed in at least one kind of database aforementioned were selected, then the target genes of differentially expressed miRNAs identified by miRecords or miWalk were obtained.

Gene Ontology (GO) annotation and pathway analysis. The Database for Annotation, Visualization and Integrated Discovery (DAVID; David Bioinformatics Resource) (22) is a gene functional classification tool that provides a comprehensive set of functional annotation tools for investigators to understand the biological meaning behind large lists of genes. GO functional enrichment analysis and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathways (<http://www.genome.jp/kegg/pathway.html>) enrichment analysis (23) of DEGs

were performed using DAVID. GO has 3 functional categories: Molecular function (MF); biological process (BP); and cellular component (CC). $P < 0.05$ was considered to indicate a statistically significant difference.

Transcription factor-miRNA association investigation. Each miRNA may have several target genes, but outstanding target genes are more likely to assemble transcription factors (24). Thus, the analysis between transcription factors and miRNA is beneficial to understand this pathomechanism. iRegulon can be used to detect transcription factors, motifs and their optimal sets of direct targets from a set of genes (25). In the present study, the iRegulon in Cytoscape software was used to explore the potential associations between miRNA and transcription factors. The minimum identity between orthologous genes was 0.05, while the maximum false discovery rate on motif similarity was 0.001. The normalized enrichment score (NES) of > 4 was considered as the threshold value for the selection of potential associations.

HCC-associated miRNA investigation. The Comparative Toxicogenomics Database (CTD) is a robust, publicly available database that is used to advance understanding regarding how environmental exposures affect human health (26). In the present study, the investigation of HCC-associated miRNA was performed using the CTD database. The biomarkers of a disease are generally identified by gene expression traits, abnormalities in gene structure or mutation which could perform roles in the etiology of a disease. Thus miRNAs which regulate these genes may be important for HCC.

Results

Investigation into differentially expressed miRNAs. The original data were analyzed and filtered. A total of 14 upregulated miRNAs and 16 downregulated miRNAs were obtained with thresholds of $P < 0.05$ and $\log_2 FC > 0.52$. The heat map of differentially expressed miRNAs is displayed in Fig. 1.

Analysis of differentially expressed miRNAs and their associated target genes. Based on two experimental certificate databases (miRecords and miWalk), a total of 380 suitable miRNA-target gene relations were obtained. A total of 22 miRNAs including *hsa-miR-15a*, *hsa-miR-125b*, *hsa-miR-122*, *hsa-miR-146b-5p* and *hsa-miR-142-3p* were revealed to have experimental certificate target genes.

GO and KEGG pathway analysis were used to gain further insights into the pathways significantly involved by miRNAs. GO functional enrichment analysis demonstrated that differentially expressed miRNAs, including *hsa-miR-15a* and *hsa-miR-125b*, were mainly involved in the positive regulation of histone H3-K9 methylation (GO, 0051574; BP), fibrillar center (GO, 0001650; CC) and mismatched DNA binding (GO, 0030983; MF; Table I). KEGG pathway analysis in the present study demonstrated that differentially expressed miRNAs were mainly enriched in the pathways in cancer map05200 (http://www.genome.jp/dbget-bin/www_bget?pathway:map05200; Fig. 2). Based on the experimental certificate miRNA-target gene databases aforementioned, the miRNA-target gene regulatory network was constructed

Table I. Results of GO functional enrichment analysis of differentially expressed miRNAs.

GO ID	Term	Count	P-value
BP			
0051574	Positive regulation of histone H3-K9 methylation	13	6.04x10 ⁻⁷
0051570	Regulation of histone H3-K9 methylation	13	6.48x10 ⁻⁶
0031061	Negative regulation of histone methylation	12	3.76x10 ⁻⁵
0031062	Positive regulation of histone methylation	13	5.34x10 ⁻⁵
0010001	Glial cell differentiation	21	7.58x10 ⁻⁵
CC			
0001650	Fibrillar center	6	1.32x10 ⁻⁴
0035098	ESC/E(Z) complex	12	8.27x10 ⁻⁴
0005869	Dynactin complex	5	1.09x10 ⁻³
0044452	Nucleolar part	7	4.18x10 ⁻³
0000791	Euchromatin	8	8.40x10 ⁻³
MF			
0030983	Mismatched DNA binding	7	1.84x10 ⁻⁴
0008263	Pyrimidine-specific mismatch base pair DNA N-glycosylase activity	6	2.74x10 ⁻⁴
0043398	HLH domain binding	9	3.92x10 ⁻⁴
0035198	miRNA binding	13	4.56x10 ⁻⁴
0000700	Mismatch base pair DNA N-glycosylase activity	6	6.17x10 ⁻⁴

The top 5 in BP, CC and MF are listed. P<0.05 was considered to indicate a statistically significant difference. GO, Gene Ontology; BP, biological process; CC, cellular component; MF, molecular function.

using Cytoscape software. The results demonstrate that there were 13 significantly upregulated miRNAs and 9 significantly downregulated miRNAs (Fig. 3).

Transcription factor-miRNA relations analysis. The iRegulon in Cytoscape software was used to explore the potential association between miRNA and transcription factors. With NES >4, the 3 miRNAs with the highest number of target genes were *hsa-miR-15a* (Fig. 4), *hsa-miR-125b* (Fig. 5) and *hsa-miR-122* (Fig. 6). There were 61 target genes that were upregulated by *hsa-miR-15a*, including transcription factors such as zinc finger protein 143, poly (ADP-ribose) polymerase 1, V-maf musculoaponeurotic fibrosarcoma oncogene homolog K, DNA polymerase epsilon subunit 3, heat shock factor protein 2 and interferon regulatory factor 8. A total of 54 target genes were downregulated by *hsa-miR-125b*, including transcription factors such as general transcription factor II-I repeat domain-containing protein 1, serum response factor, polymerase (RNA) II (DNA directed) polypeptide A, B-cell lymphoma 6 protein and transcription factor E2F1. A total of 49 target genes were downregulated by *hsa-miR-122*, including transcription factors such as forkhead box protein A2, CCAAT/enhancer-binding protein α and estrogen-associated receptor γ .

HCC-associated miRNA analysis. A total of 11 differentially expressed miRNA that regulated the marker genes of HCC were identified (Table II). Among them, *hsa-miR-146-5p* (n=13), *hsa-miR-15a* (n=5) and *hsa-miR-125b* (n=4) were the miRNAs that regulated the highest number of marker genes of HCC. A

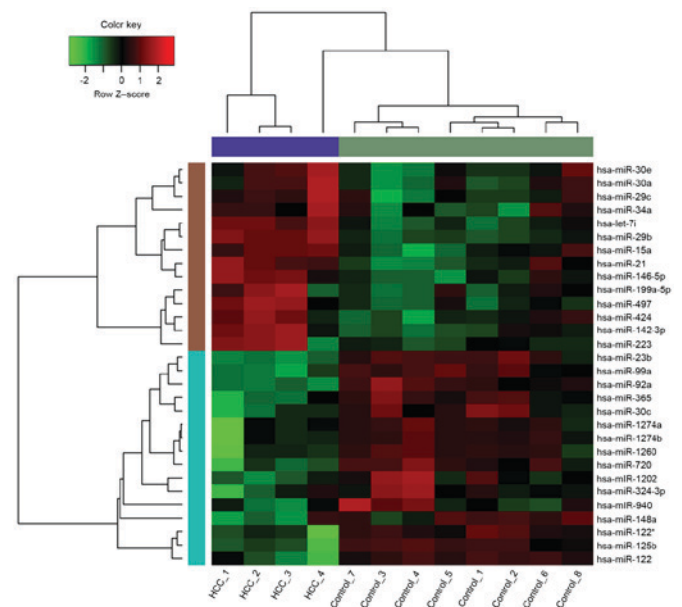


Figure 1. Heat map for the differentially expressed miRNAs. Green represents the low expression level of miRNA, red represents the high expression level of miRNA and blank represents that the expression levels of miRNAs do not change. HCC, hepatocellular carcinoma; hsa, human; miRNA/miR, microRNA.

further investigation was made on other HCC-associated genes (not the marker genes but recorded in databases) in the CTD database. The results indicated that a total of 49 target genes of *miR-122* were HCC-associated genes.

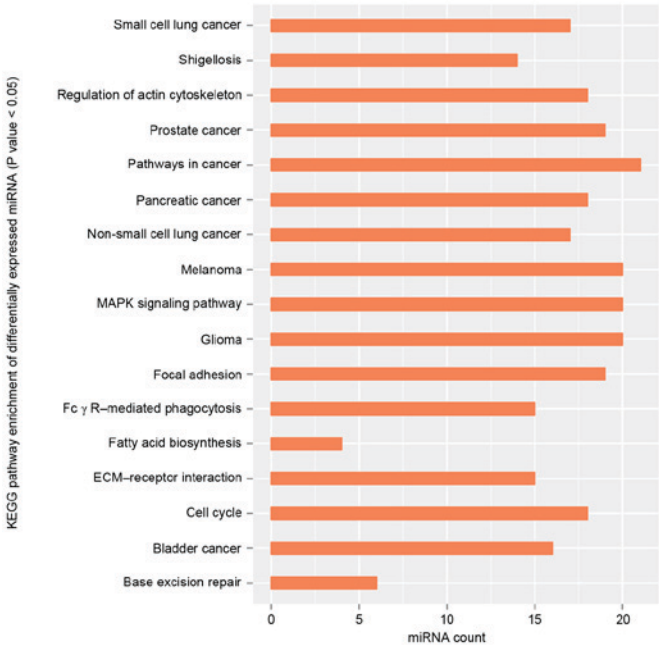


Figure 2. Results of KEGG pathway enrichment analysis. X-axis represents the number of miRNAs, Y-axis represents the KEGG pathways significantly enriched by differentially expressed miRNAs, $P < 0.05$. KEGG, Kyoto Encyclopedia of Genes and Genomes; miRNA/miR, microRNA; MAPK, mitogen activated protein kinase; ECM, extra cellular matrix.

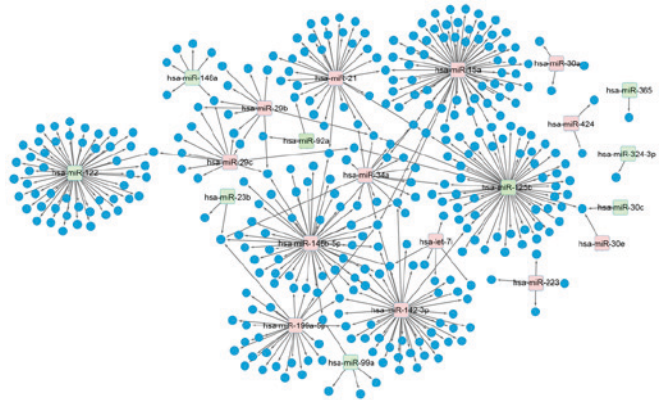


Figure 3. Regulatory network for miRNA and target genes. Green squares represent the upregulated miRNAs, red squares represent the down-regulated miRNAs and blue circles represent the target genes. hsa, human; miRNA/miR, microRNA.

Discussion

HCC is one of the most common types of human cancer worldwide (1). However, the molecular mechanisms behind the pathogenesis and progression of HCC remain unclear. The present study performed a bioinformatics analysis on the miRNA expression profile of HCC. By screening the differentially expressed miRNAs, a total of 14 upregulated miRNAs and 16 downregulated miRNAs between HCC samples and control samples were obtained. These differentially expressed miRNAs were mainly involved in BP like positive regulation of histone H3-K9 methylation, and were observed in KEGG pathways in cancer map05200. A total of 3 outstanding regulatory networks of miRNA (*hsa-miR-15a*, *hsa-miR-125b* and

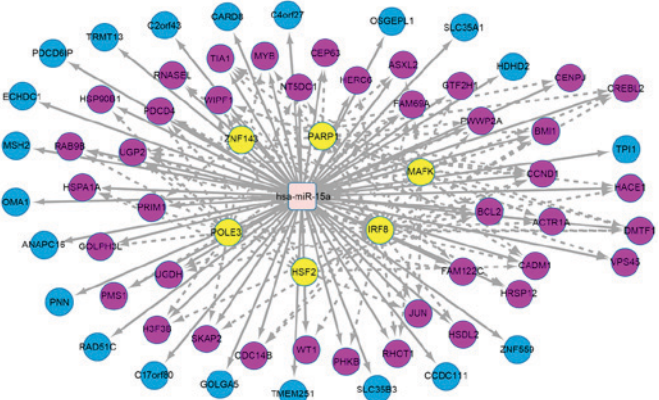


Figure 4. Sub-modules of regulatory networks for *hsa-miR-15a* and associated transcription factors. Yellow circles represent the transcription factors, purple circles represent the co-regulated target genes for transcription factors and blue circles represent the target genes of *miR-15a*. hsa, human; miRNA/miR, microRNA.

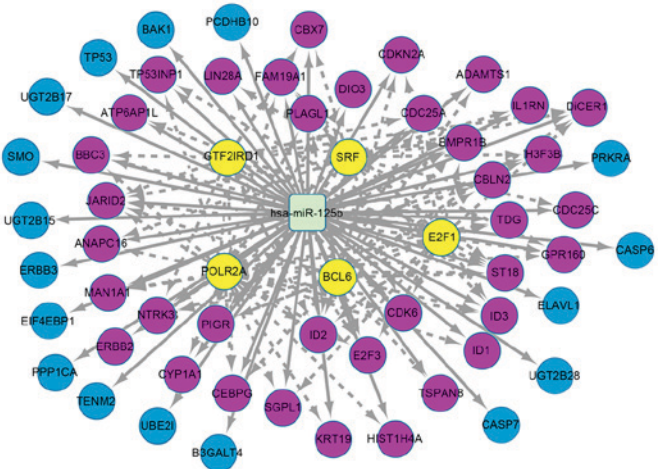


Figure 5. Sub-modules of regulatory network for *hsa-miR-125b* and associated transcription factors. Yellow circles represent the transcription factors, purple circles represent the co-regulated target genes for transcription factors and blue circles represent the target genes of *miR-125b*. hsa, human; miRNA/miR, microRNA.

hsa-miR-122) were further investigated. A total of 11 differentially expressed miRNAs that regulated the candidate marker genes of HCC were explored.

In the present study, GO enrichment analysis demonstrated that the target genes that were regulated by the differentially expressed miRNA, such as *hsa-miR-15a* and *hsa-miR-125b*, were mainly enriched in functions such as histone H3-K9 methylation. DNA methylation and histone H3-K9 modifications perform important roles in the process of tumor formation by the dysregulation of target genes (27). A previous study demonstrated that H3-K9 methylation contributed to chromosomal targeting of polycomb group proteins (28). DNA methylation profiling for serum DNA of patients with HCC revealed a potential association between HCC and DNA methylation (29). Peng *et al* (30) indicated that heterochromatin (essential for chromosome organization and inheritance) stability requires regulators of histone H3-K9 methylation, which indicates that regulation of histone

Table II. Differentially expressed miRNAs that regulated the marker genes of hepatic cell carcinoma (HCC).

miRNA	Marker gene
<i>hsa-miR-99a</i>	<i>MTOR, IGF1R</i>
<i>hsa-miR-424</i>	<i>KDR</i>
<i>hsa-miR-34a</i>	<i>CCND1, MYC, MET</i>
<i>hsa-miR-23b</i>	<i>MET</i>
<i>hsa-miR-21</i>	<i>PPARA, E2F1, BTG2</i>
<i>hsa-miR-199a-5p</i>	<i>CCND1, MET, FASN</i>
<i>hsa-miR-15a</i>	<i>UGDH, BCL2, CCND1, HSDL2, JUN</i>
<i>hsa-miR-148a</i>	<i>CDKN1B</i>
<i>hsa-miR-146b-5p</i>	<i>JUN, IL6, FOS, MET, TNF, SOCS1, CCND1, MYC, BRAF, MMP9, IGF1R, BCL2, FASN</i>
<i>hsa-miR-142-3p</i>	<i>APCS, KRAS, JUN, CCND1</i>
<i>hsa-miR-125b</i>	<i>ADAMTS1, TP53, CDKN2A, CYP1A1</i>

hsa, human; miR/miRNA, microRNA.

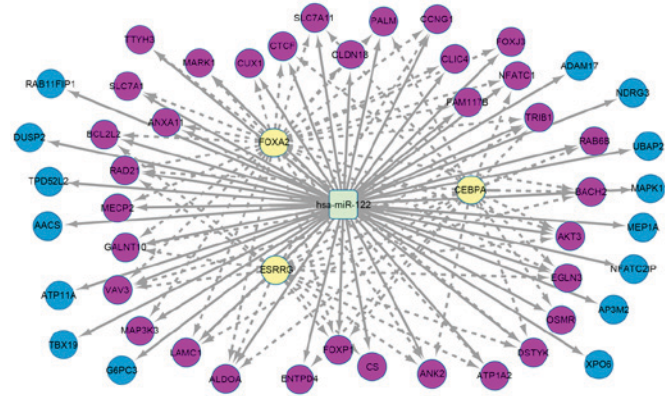


Figure 6. Sub-modules of regulatory network for *hsa-miR-122* and associated transcription factors. Yellow circles represent the transcription factors, purple circles represent the co-regulated target genes for transcription factors and blue circles represent the target genes of *miR-122*. hsa, human; miRNA/miR, microRNA.

H3-K9 methylation is vital for cancer-associated gene expression. Thus, the present study speculated that histone H3-K9 modifications may relate to the progression of HCC. KEGG pathway analysis in the present study revealed that target genes of differentially expressed miRNAs were mainly enriched in multiple cellular processes related to cancer and MAPK signaling. Daroqui *et al* (31) proved that MAPK signaling promotes cell invasiveness and *in vivo* tumor progression via actin remodeling in response to transforming growth factor- β . Tumor necrosis factor has been revealed to activate the cyclic adenosine monophosphate response element-binding protein through the p38 MAPK/mitogen and stress activated protein kinase 1 signaling pathway (32). MAPK signaling and pathways in cancer had a close association with HCC processes, and so the differentially expressed miRNA that relate with the MAPK signaling pathway and cancer-related pathways (as determined by KEGG analysis) may take part in the development of HCC.

It is considered that the regulation of transcription factors and miRNAs is essential for the appropriate execution of biological events and developmental processes (33). By incorporating prior information from predicted regulatory interactions with parallel miRNA/mRNA expression datasets, Peng *et al* (34) indicated that the transcription factor-miRNA co-regulatory based on feed-forward loops served an important role in the development of prostate cancer. In the present study, the regulatory networks of 3 outstanding miRNAs (*hsa-miR-15a*, *hsa-miR-125b* and *hsa-miR-122*) were obtained. This result demonstrates that these transcription factors had close associations with the 3 miRNAs in the co-regulation of target genes, which further indicated the complex regulatory roles of them during the development of HCC. Thus, it is speculated that *hsa-miR-15a*, *hsa-miR-125b* and *hsa-miR-122* may be the miRNAs to be investigated for their role in HCC gene therapy.

Human *miR-146b-5p* is an miRNA which is located on chromosome 10q24.3 (35). *miR-146b-5p* is frequently downregulated in solid tumors, including prostate cancer, pancreatic cancer and glioblastoma (36). In the present study, HCC-associated miRNA analysis revealed that there were 11 differentially expressed miRNAs that regulate the marker genes of HCC including *miR-146b-5p*, which had the highest number of marker genes. Furthermore, the HCC-associated gene (not recorded marker genes) investigation based on the CTD database revealed that *miR-122* had the highest number of HCC-associated genes. A previous study indicated that *miR-122* negatively correlates with liver fibrosis as detected by histology and FibroScan software (37). In animal models, *miR-122* has been proven to be a marker of liver cell damage, inflammation and regeneration in acetaminophen-induced liver injury (38). In the present study, *miR-122* was revealed to be closely associated with HCC, thus the target gene of *miR-122* may be the potential novel marker gene for HCC. Therefore, it was speculated that *hsa-miR-15a*, *hsa-miR-125b*, *hsa-miR-122* and *hsa-miR-146b-5p* may perform important roles in the progression of HCC, and may be new biomarkers for gene therapy of HCC. However, there were limitations to the present this study, such as lack of experimental verification and joint analysis of expression profile data. An investigation based on experimental verification is required in future studies.

In conclusion, miRNAs such as *hsa-miR-15a*, *hsa-miR-125b*, *hsa-miR-122* and *hsa-miR-146b-5p* may be new biomarkers for gene therapy of HCC. Histone H3-K9 methylation and miRNAs identified through KEGG analysis to be involved in cancer formation may be closely associated with the development of HCC.

Acknowledgements

The present project was supported by the social development fund of Nantong (grant no. HS2014063) and The 12th Talent Summit of Top Six Industries in Jiangsu Province (grant no. WSW-080).

References

- Chen KW, Ou TM, Hsu CW, Horng CT, Lee CC, Tsai YY, Tsai CC, Liou YS, Yang CC, Hsueh CW, *et al*: Current systemic treatment of hepatocellular carcinoma: A review of the literature. *World J Hepatol* 7: 1412-1420, 2015.

2. Lin S, Hoffmann K and Schemmer P: Treatment of hepatocellular carcinoma: A systematic review. *Liver Cancer* 1: 144-158, 2012.
3. Cabrera R and Nelson DR: Review article: The management of hepatocellular carcinoma. *Aliment Pharmacol Ther* 31: 461-476, 2010.
4. Liang HH, Wei PL, Hung CS, Wu CT, Wang W, Huang MT and Chang YJ: MicroRNA-200a/b influenced the therapeutic effects of curcumin in hepatocellular carcinoma (HCC) cells. *Tumour Biol* 34: 3209-3218, 2013.
5. Cao W, Guo Q, Zhang T, Zhong D and Yu Q: Prognostic value of microRNAs in acute myocardial infarction: A systematic review and meta-analysis. *Int J Cardiol* 189: 79-84, 2015.
6. Gao Y, He Y, Ding J, Wu K, Hu B, Liu Y, Wu Y, Guo B, Shen Y, Landi D, *et al*: An insertion/deletion polymorphism at miRNA-122-binding site in the interleukin-1 α 3' untranslated region confers risk for hepatocellular carcinoma. *Carcinogenesis* 30: 2064-2069, 2009.
7. Liu J, Yan J, Zhou C, Ma Q, Jin Q and Yang Z: miR-1285-3p acts as a potential tumor suppressor miRNA via downregulating JUN expression in hepatocellular carcinoma. *Tumour Biol* 36: 219-225, 2015.
8. Yang Z, Zhang Y and Wang L: A feedback inhibition between miRNA-127 and TGF β /c-Jun cascade in HCC cell migration via MMP13. *PLoS One* 8: e65256, 2013.
9. Ziari K, Zarea M, Gity M, Fayyaz AF, Yahaghi E, Darian EK and Hashemian AM: Downregulation of miR-148b as biomarker for early detection of hepatocellular carcinoma and may serve as a prognostic marker. *Tumour Biol* 37: 5765-5768, 2016.
10. Huang X and Jia Z: Construction of HCC-targeting artificial miRNAs using natural miRNA precursors. *Exp Ther Med* 6: 209-215, 2013.
11. Borel F, Konstantinova P and Jansen PL: Diagnostic and therapeutic potential of miRNA signatures in patients with hepatocellular carcinoma. *J Hepatol* 56: 1371-1383, 2012.
12. Yang N, Ekanem NR, Sakyi CA and Ray SD: Hepatocellular carcinoma and microRNA: New perspectives on therapeutics and diagnostics. *Adv Drug Deliv Rev* 81: 62-74, 2015.
13. Murakami Y, Tanahashi T, Okada R, Toyoda H, Kumada T, Enomoto M, Tamori A, Kawada N, Taguchi YH and Azuma T: Comparison of hepatocellular carcinoma miRNA expression profiling as evaluated by next generation sequencing and microarray. *PLoS One* 9: e106314, 2014.
14. Xie Y, Yao Q, Butt AM, Guo J, Tian Z, Bao X, Li H, Meng Q and Lu J: Expression profiling of serum microRNA-101 in HBV-associated chronic hepatitis, liver cirrhosis, and hepatocellular carcinoma. *Cancer Biol Ther* 15: 1248-1255, 2014.
15. Wang Y, Lee AT, Ma JZ, Wang J, Ren J, Yang Y, Tantoso E, Li KB, Ooi LL, Tan P and Lee CG: Profiling microRNA expression in hepatocellular carcinoma reveals microRNA-224 up-regulation and apoptosis inhibitor-5 as a microRNA-224-specific target. *J Biol Chem* 283: 13205-13215, 2008.
16. Wei L, Lian B, Zhang Y, Li W, Gu J, He X and Xie L: Application of microRNA and mRNA expression profiling on prognostic biomarker discovery for hepatocellular carcinoma. *BMC Genomics* 15 (Suppl 1): S13, 2014.
17. Edgar R, Domrachev M and Lash AE: Gene expression omnibus: NCBI gene expression and hybridization array data repository. *Nucleic Acids Res* 30: 207-210, 2002.
18. Huber W, Von Heydebreck A, Sültmann H, Poustka A and Vingron M: Variance stabilization applied to microarray data calibration and to the quantification of differential expression. *Bioinformatics* 18 (Suppl 1): S96-S104, 2002.
19. Ritchie ME, Phipson B, Wu D, Hu Y, Law CW, Shi W and Smyth GK: limma powers differential expression analyses for RNA-sequencing and microarray studies. *Nucleic Acids Res* 43: e47, 2015.
20. Xiao F, Zuo Z, Cai G, Kang S, Gao X and Li T: miRecords: An integrated resource for microRNA-target interactions. *Nucleic Acids Res* 37 (Database Issue): D105-D110, 2009.
21. Dweep H, Sticht C, Pandey P and Gretz N: miRWalk-database: Prediction of possible miRNA binding sites by 'walking' the genes of three genomes. *J Biomed Inform* 44: 839-847, 2011.
22. Dennis G Jr, Sherman BT, Hosack DA, Yang J, Gao W, Lane HC and Lempicki RA: DAVID: Database for annotation, visualization, and integrated discovery. *Genome Biol* 4: P3, 2003.
23. Kanehisa M and Goto S: KEGG: Kyoto encyclopaedia of genes and genomes. *Nucleic Acids Res* 28: 27-30, 2000.
24. Enright AJ, John B, Gaul U, Tuschl T, Sander C and Marks DS: MicroRNA targets in *Drosophila*. *Genome Biol* 5: R1, 2003.
25. Janky R, Verfaillie A, Imrichová H, Van de Sande B, Standaert L, Christiaens V, Hulselmans G, Hertzen K, Naval Sanchez M, Potier D, *et al*: iRegulon: From a gene list to a gene regulatory network using large motif and track collections. *PLoS Comput Biol* 10: e1003731, 2014.
26. Davis AP, Grondin CJ, Lennon-Hopkins K, Saraceni-Richards C, Sciaky D, King BL, Wieggers TC and Mattingly CJ: The comparative toxicogenomics database's 10th year anniversary: Update 2015. *Nucleic Acids Res* 43 (Database Issue): D914-D920, 2015.
27. Kitamoto S, Yamada N, Yokoyama S, Houjou I, Higashi M, Goto M, Batra SK and Yonezawa S: DNA methylation and histone H3-K9 modifications contribute to MUC17 expression. *Glycobiology* 21: 247-256, 2011.
28. Sewalt RG, Lachner M, Vargas M, Hamer KM, den Blaauwen JL, Hendrix T, Melcher M, Schweizer D, Jenuwein T and Otte AP: Selective interactions between vertebrate polycomb homologs and the SUV39H1 histone lysine methyltransferase suggest that histone H3-K9 methylation contributes to chromosomal targeting of Polycomb group proteins. *Mol Cell Biol* 22: 5539-5553, 2002.
29. Tu T, Shackel NA and McCaughan G: 'Testing your methyl': DNA methylation profiling of serum DNA of HCC patients. *Hepatol Int* 7: 785-787, 2013.
30. Peng JC and Karpen GH: Heterochromatic genome stability requires regulators of histone H3 K9 methylation. *PLoS Genet* 5: e1000435, 2009.
31. Daroqui MC, Vazquez P, Bal de Kier Joffé E, Bakin AV and Puricelli LI: TGF- β autocrine pathway and MAPK signaling promote cell invasiveness and in vivo mammary adenocarcinoma tumor progression. *Oncol Rep* 28: 567-575, 2012.
32. Gustin JA, Pincheira R, Mayo LD, Ozes ON, Kessler KM, Baerwald MR, Korgaonkar CK and Donner DB: Tumor necrosis factor activates CRE-binding protein through a p38 MAPK/MSK1 signaling pathway in endothelial cells. *Am J Physiol Cell Physiol* 286: C547-C555, 2004.
33. Wang S, Li W, Lian B, Liu X, Zhang Y, Dai E, Yu X, Meng F, Jiang W and Li X: TMREC: A database of transcription factor and miRNA regulatory cascades in human diseases. *PLoS One* 10: e0125222, 2015.
34. Peng C, Wang M, Shen Y, Feng H and Li A: Reconstruction and analysis of transcription factor-miRNA co-regulatory feed-forward loops in human cancers using filter-wrapper feature selection. *PLoS One* 8: e78197, 2013.
35. Katakowski M, Zheng X, Jiang F, Rogers T, Szalad A and Chopp M: MiR-146b-5p suppresses EGFR expression and reduces in vitro migration and invasion of glioma. *Cancer Invest* 28: 1024-1030, 2010.
36. Li Y, Wang Y, Yu L, Sun C, Cheng D, Yu S, Wang Q, Yan Y, Kang C, Jin S, *et al*: miR-146b-5p inhibits glioma migration and invasion by targeting MMP16. *Cancer Lett* 339: 260-269, 2013.
37. Halász T, Horváth G, Pár G, Werling K, Kiss A, Schaff Z and Lendvai G: miR-122 negatively correlates with liver fibrosis as detected by histology and FibroScan. *World J Gastroenterol* 21: 7814-7823, 2015.
38. Park HK, Jo W, Choi HJ, Jang S, Ryu JE, Lee HJ, Lee H, Kim H, Yu ES and Son WC: Time-course changes in the expression levels of miR-122, -155 and -21 as markers of liver cell damage, inflammation, and regeneration in acetaminophen-induced liver injury in rats. *J Vet Sci* 17: 45-51, 2016.