

Terpinen-4-ol inhibits colorectal cancer growth via reactive oxygen species

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Abstract. Terpinen-4-ol (TP4O) is the main component of the essential oil extracted from *Melaleuca alternifolia*, known as the tea tree, of the botanical family Myrtaceae. The anticancer effects of TP4O have been reported in several cancer cell lines. Previous reports have demonstrated that TP4O exerts anticancer effects by inducing apoptotic cell death in several cell lines; however, the underlying molecular mechanisms of these effects remain unclear. In the present study, the anticancer effects of TP4O against the colorectal cancer (CRC) cell lines HCT116 and RKO were evaluated using WST-8 and bromodeoxyuridine assays. The mechanism of cell death was investigated by the measurement of caspase-3/7, Annexin V and lactate dehydrogenase release. Reactive oxygen species (ROS) levels induced by TP4O were evaluated by electron spin resonance and quantitative measurement of dihydroethidium. Localization of the ROS derived from mitochondria was observed by confocal inverted microscopy. Protein levels of ROS scavengers were assessed by western blotting analysis. To confirm the role of ROS, cell viability was measured in the presence of antioxidant reagents. In an *in vivo* xenograft model of ICR-SCID mice implanted with HCT116 cells, 200 mg/kg TP4O was injected locally, and tumor growth was compared with that of the control. TP4O induced apoptotic cell death in HCT116 and RKO cells in a dose-dependent manner, and TP4O also increased the levels of ROS generated

by mitochondria. TP4O-induced cell death was rescued by administration of antioxidant reagents. *In vivo*, TP4O inhibited the proliferation of HCT116 xenografts compared with that of the control group. The results of the present study suggest that TP4O induces apoptosis in CRC cells through ROS generation. Furthermore, TP4O is potentially useful for the development of novel therapies against CRC.

Introduction

Colorectal cancer (CRC) is the third most common cancer in males and the second in females worldwide (1). For patients in the early stages of CRC, i.e., stages I and II, surgery is the most common treatment, and the 5-year relative survival for patients with early-stage CRC is 90.3%. By contrast, for patients with advanced CRC, chemotherapy and/or radiation therapy is required, and the 5-year survival rate of patients with distant metastasis is 12.5% (2). The ideal anticancer drug would be one that is affordable, has few side effects and is sufficiently potent to achieve complete remission. Inhibitors of vascular endothelial growth factor and monoclonal antibodies that inhibit epidermal growth factor receptor have prolonged the survival of advanced CRC patients (3,4). However, these drugs have severe side effects, and thus, researchers have been searching for novel biochemical compounds (5). Natural products have become an important resource for novel drug discovery (5), and >50% of anticancer drugs are derived from natural products (6).

Essential oils are natural products extracted from seeds, leaves and tree resin. These natural oils are used primarily in perfumes, cosmetics and the food industry (7). Various of these oils have been demonstrated to possess antibiotic, anti-inflammatory and anticancer properties (7,8). The molecular mechanisms underlying their anticancer effects have been revealed to occur via reactive oxygen species (ROS) signaling pathways and cell cycle arrest (9,10). Terpinen-4-ol (TP4O) is the main component of the essential oil extracted from *Melaleuca alternifolia* (of the botanical family Myrtaceae), a

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plant native to Australia that is also known as the tea tree (11). Tea tree oil (TTO) is famous for its scent, and has been demonstrated to have antibacterial (11,12) and anti-inflammatory properties (13). Previous studies have investigated the anticancer effect of TP4O against human melanoma cells (14), human non-small cell lung cancer (15), human leukaemia cells (16) and CRC cells (17). However, the anticancer effects of TPO4 remain unclear. In the present study, the anticancer effects of TP4O against CRC cells, in particular the role of ROS, were evaluated using the HCT116 and RKO cell lines.

Materials and methods

Chemicals and reagents. TP4O, Triton X-100, staurosporine and ebselen (EB) were purchased from Sigma-Aldrich (Merck KGaA, Darmstadt, Germany). TP4O was dissolved in ethanol to prepare stock solutions. The compound was administered at a final concentration of 0.0165% ethanol. High-glucose (4.5 g/l) Dulbecco's modified Eagle's medium (DMEM), phenol red-free DMEM and PBS were purchased from Wako Pure Chemical Industries, Ltd. (Osaka, Japan). Eagle's minimum essential medium (EMEM) was purchased from the American Type Culture Collection (ATCC; Manassas, VA, USA). 5,5-dimethyl-1-pyrroline-N-oxide (DMPO) was purchased from Dojindo Molecular Technologies, Inc. (Kumamoto, Japan). N-acetyl-L-cysteine (NAC) and manganese (III) tetrakis (4-benzoic acid) porphyrin chloride (MnTBAP) were purchased from EMD Millipore (Billerica, MA USA) and Funakoshi Co., Ltd. (Tokyo, Japan), respectively. Phenylmethylsulfonyl fluoride was purchased from Roche Diagnostics GmbH (Mannheim, Germany).

Cell lines and cell culture. Human CRC cell lines (HCT116 and RKO) and a human normal colon epithelial cell line (CCD 841 CoN) were purchased from the ATCC. The cancer cell lines were cultured in DMEM, while CCD 841 CoN cells were cultured in EMEM. The media were supplemented with 10% heat-inactivated foetal bovine serum (FBS) and 1% antibiotics (100 U/ml penicillin). All cell lines were incubated at 37°C and maintained in an incubator with 5% CO₂. All cells were used while within a passage number that ranged between 8 and 21. Cells were seeded at 5,000 cells/well in 96-well plates for the WST-8 and bromodeoxyuridine (BrdU) assays, at 10,000 cells/well in 96-well plates for the lactate dehydrogenase (LDH) release assay, at 2x10⁵ cells/well in 6-well plates for the caspase-3/7 and Annexin V assays, and at 1x10⁶ cells/dish in 6-cm dishes for western blot analysis. Each cell line was seeded in the above culture media and incubated for 24 h prior to each experiment.

Evaluation of cell viability and proliferation. To evaluate the effects of TP4O on cell viability and proliferation, WST-8 and BrdU assays were performed. Briefly, cells were pre-incubated for 24 h and treated with various concentrations of TP4O for 24 h (0, 1, 10, 100, 1,000 or 10,000 µM). Cell viability was quantified using a WST-8 assay via the Cell Counting kit-8 (Dojindo Molecular Technologies, Inc.) by measuring the absorbance at 450 nm. Cell proliferation was quantified using a BrdU assay kit purchased from Roche Diagnostics GmbH by measuring the absorbance at 370 nm. The half maximal

inhibitory concentration (IC₅₀) values for each cell line were determined over 24 h following TP4O treatment. Cell viability following treatment with antioxidants was also measured. Following pre-incubation of the cells for 24 h at 37°C, the culture medium was exchanged for medium with antioxidants (20 mM NAC, 20 µM EB and 100 µM MnTBAP) 30 min prior to TP4O administration. The results were obtained from three independent experiments.

Evaluation of apoptosis. To evaluate apoptosis, the following flow cytometry experiment was performed. After 24 h of pre-incubation at 37°C, the cells were treated with various concentrations of TP4O (0, 100 or 1,000 µM). Treatment-induced caspase-3/7 activation was examined in HCT116 and RKO cells using the Muse™ Cell Analyzer (Merck KGaA) and Muse™ Caspase-3/7 Assay kit (Merck KGaA), according to the manufacturer's protocol. Following 6 h of treatment, harvested cells were mixed with the Muse™ Caspase-3/7 reagent, which contains a DNA-binding dye that is linked to a DEVD peptide substrate and a dead cell marker [7-aminoactinomycin D (7-AAD)]. Caspase-3/7 activity was detected with the fluorescence of a DNA-binding dye from the Muse™ Caspase-3/7 Assay kit and cell viability was detected with 7-AAD fluorescence using the Muse™ Cell Analyzer. The results were obtained from three independent experiments.

For the Annexin V assay, cells were treated with TP4O (0, 100 or 1,000 µM) for 12 h. Treatment-induced apoptosis was examined using the Muse™ Annexin V & Dead Cell kit (Merck KGaA) according to the manufacturer's protocol. Phosphatidylserine (PS) was detected using Annexin V, and cell viability was detected using a dead cell marker (7-AAD). The results were obtained from four independent experiments.

LDH-release assay. An LDH-release assay was performed to assess necrosis resulting from TP4O treatment. The LDH Cytotoxicity Detection kit (Takara Bio, Inc., Otsu, Japan) quantifies the activity of LDH released from damaged cells (18). Following 24 h of pre-incubation, the culture medium was exchanged for FBS-free DMEM containing 1% Triton X-100, various concentrations of TP4O (0, 10, 100 or 1,000 µM) and 200 nM staurosporine for 12 h. Following treatment, the plates were centrifuged at 250 x g for 10 min at 4°C. The supernatant was transferred to a clear 96-well plate, and 100 µl of reaction mixture was then added to each well. The plates were incubated in the dark for 30 min at 25°C, the absorbance was measured at 490 nm and the percentage LDH release was calculated. To compare necrotic cell death, samples with 1% Triton X-100 were used as positive controls. To study apoptotic cell death, LDH release induced by 200 nM staurosporine was examined. The results were obtained from three independent experiments.

Electron spin resonance (ESR) measurement. ROS generation in cells was measured using ESR, as previously described (19). Briefly, 10⁶ cells were seeded on a glass cover slide (49.0x5.0x0.2 mm) and incubated overnight as previously described. Cells were treated with 0 or 1,000 µM TP4O for 15 min. ESR was measured in a respiration buffer containing

10 mM DMPO as the spin-trapping agent. All ESR spectra were obtained using a JES-RE1X X-band spectrometer (JEOL, Ltd., Tokyo, Japan).

Detection of mitochondrial superoxide. The source of superoxide was detected by fluorescence microscopy. A total of 2.5×10^4 cells/well were seeded in an 8-well cover glass chamber. After 24 h of pre-incubation, cells were treated with 0 or 1,000 μM TP4O for 24 h. Culture medium was exchanged for phenol red-free DMEM, which contained 5 μM MitoSOX Red™ Mitochondrial Superoxide Indicator (Molecular Probes; Thermo Fisher Scientific, Inc., Waltham, MA, USA), and the cells were incubated for 10 min at 37°C. Following one wash with PBS, the cells were incubated for a further 30 min at 37°C with phenol-free DMEM, which contained 100 nM MitoTracker Green FM™ (Molecular Probes; Thermo Fisher Scientific, Inc.). Next, the cells were washed with PBS and observed using a confocal inverted fluorescence microscope (Olympus Corporation, Tokyo, Japan). Localization of ROS derived from mitochondria was detected by MitoSOX Red™ Mitochondrial Superoxide Indicator fluorescence, and localization of mitochondria was detected by MitoTracker Green FM™ fluorescence. Images of bifocal field, MitoSOX Red™ Mitochondrial Superoxide Indicator and MitoTracker Green FM™ were merged.

Quantification of oxidative stress. The quantitative measurement of cellular populations undergoing oxidative stress was measured using the Muse™ Cell Analyzer and Muse™ Oxidative Stress kit (Merck KGaA). Cells were seeded at 2×10^5 cells/well in a 6-well plate and incubated overnight. Following exposure to 0 or 1,000 μM TP4O for 24 h, according to the manufacturer's protocol, cells were detached, resuspended at 1×10^6 cells/ml and incubated at 37°C for 30 min with Muse™ Oxidative Stress working solution, which contained dihydroethidium (DHE). DHE is cell permeable, and has been proposed to react with superoxide anions, thus undergoing oxidation upon binding to DNA (20). The number of oxidized cells were counted according to the intensity of red fluorescence using the Muse™ Cell Analyzer. The results were obtained from four independent experiments.

Western blot analysis. Cells were seeded at 1×10^6 cells/dish in 6-cm dishes. After 24 h, cells were treated with various concentrations of TP4O (0, 100 or 1,000 μM) for 24 h. Cultured cells were washed with PBS and lysed with lysis buffer that contained 1% Protease Inhibitor Cocktail (100X; Cell Signaling Technology, Inc., Danvers, MA, USA) and 1 mM phenylmethylsulfonyl fluoride. Samples were kept on ice for 2 min, followed by sonication for 5 min and centrifugation at $13,000 \times g$ for 2 min at 4°C; the supernatants were then collected. The samples were subjected to 15% SDS-PAGE and transferred to a polyvinylidene fluoride membrane (Merck KGaA). After 30 min blocking with Tris-buffered saline containing 0.1% Tween-20 (T-TBS) and 5% bovine serum albumin (Iwai Chemicals Company, Ltd., Tokyo, Japan), primary antibodies against superoxide dismutase 2 (SOD2; cat. no. 13141; dilution, 1:1,000) glutathione peroxidase 1 (GPX1; cat. no. 3206; dilution 1:500) and GAPDH (endogenous control; cat. no. 2118; dilution, 1:1,000; all from Cell Signalling

Technology, Inc.) were incubated with membranes in Can Get Signal® Immunoreaction Enhancer Solution 1 (Toyobo Co., Ltd., Osaka, Japan) overnight at 25°C. After washing 3 times for 30 min with T-TBS, the membrane was incubated with an anti-rabbit IgG, horseradish peroxidase-linked antibody (cat. no. 7074; dilution, 1:1,000; Cell Signalling Technology, Inc.) in Can Get Signal® Immunoreaction Enhancer Solution 2 (Toyobo Co., Ltd.) for 2 h at 25°C. After washing 3 times for 15 min with T-TBS at 25°C, the bands were incubated with ECL™ Western Blotting Detection reagents as according to the manufacturer's protocol and detected with ImageQuant LAS 4000 mini (both from GE Healthcare Life Sciences, Chalfont, UK). Results were quantified using ImageQuant TL 7.0 software (GE Healthcare Life Sciences).

Animals. A total of 14 male ICR-SCID mice aged 6-7 weeks old and weighing 21-25 g were purchased from Charles River Laboratories Japan, Inc. (Yokohama, Japan). The mice were provided with clean water and food *ad libitum*, and housed in standardized, pathogen-free conditions with a 14 h light/10 h dark cycle, a temperature of $23.5 \pm 2.5^\circ\text{C}$ and humidity of $52.5 \pm 12.5\%$. The mice were used after an acclimation period of ≥ 7 days. All animal experiments were performed with the approval of the Animal Ethics Committee of the University of Tsukuba (Tsukuba, Japan; approval no. 13-385) and according to the guidelines of this committee.

Xenograft model. To evaluate the effect of TP4O *in vivo*, a subcutaneous tumor was induced. HCT116 cells (2×10^6 cells per mouse) were injected subcutaneously into the right flanks of 14 mice. Tumor volume was calculated using the following formula: $0.5 \times \text{length} \times \text{width}^2$. When tumor volume had reached 80-100 mm^3 , the 12 qualifying mice were randomly divided into two groups ($n=6/\text{group}$) and subcutaneously injected with one of the following: 250 μl saline (control group) or 200 mg/kg TP4O dissolved in 250 μl saline (TP4O group). Each mouse was injected once every 3 days (total, five times). The tumor volume was recorded every 3 days. Body weight was recorded the first day of the injection, on day 7 and at the end of experimentation (day 14). On day 14, blood samples were collected under anesthesia, the mice were sacrificed and the tumors were excized. To evaluate TP4O toxicity, serum alanine aminotransferase (ALT) and serum creatinine levels were measured using an autoanalyzer (Dri-chem 7000 V; Fujifilm, Tokyo, Japan).

Immunostaining. The tumor tissue was fixed with 10% formaldehyde. Each 2- μm paraffin-embedded section was stained with an anti-8-hydroxy-2'-deoxyguanosine (8-OHdG) antibody at 4°C overnight (cat. no. MOG-20P; dilution, 1:100, Japan Institute for the Control of Aging, Shizuoka, Japan) or with an anti-cleaved caspase-3 antibody according to the manufacturer's protocol (cat. no. 9661; dilution, 1:300; Cell Signaling Technology, Inc.). 8-OHdG is a marker for oxidative stress (21). To accurately quantify 8-OHdG- and cleaved caspase-3-positive areas, slides from three randomly selected high-power fields (magnification, $\times 200$) of each tumor were visualized using a BZ-X710 microscope (Keyence Corporation, Osaka, Japan). An automated software analysis program, BZ-X analyser version 1.3.0.3 (Keyence Corporation), was

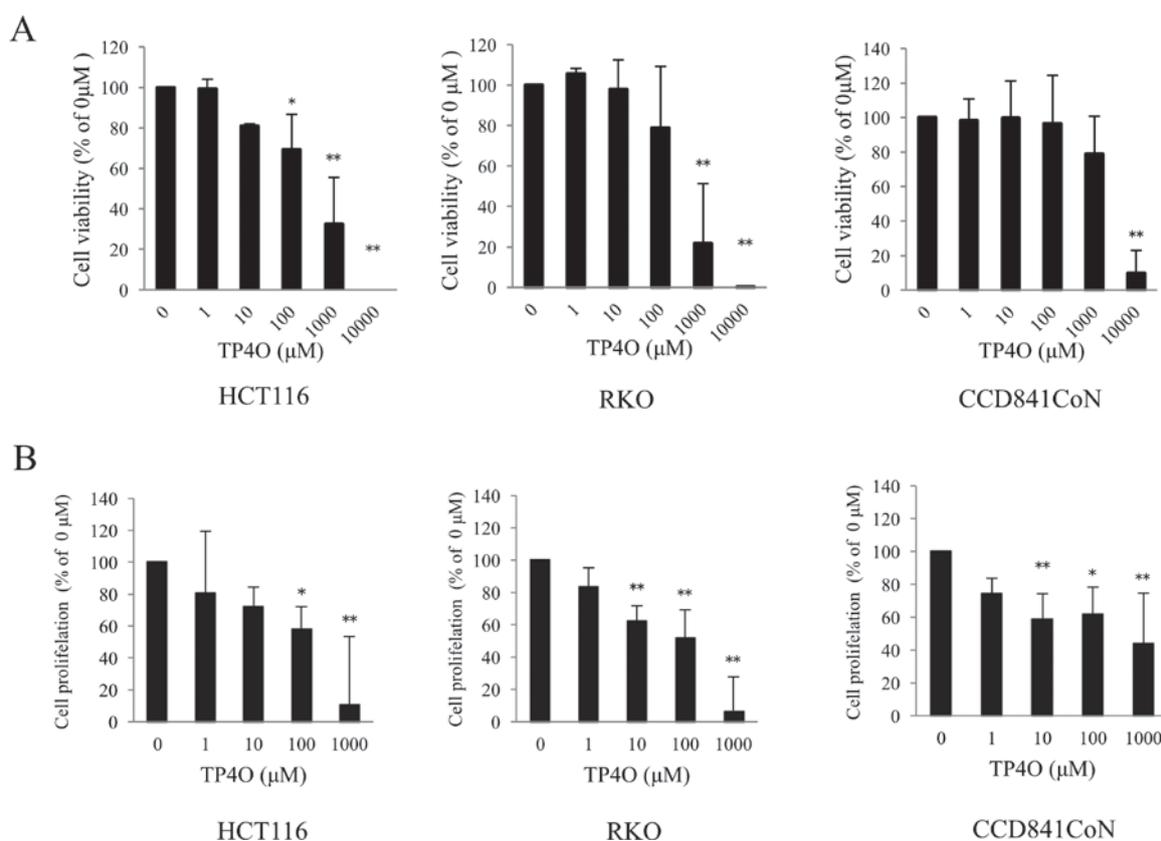


Figure 1. TP4O causes cell death and inhibits cell proliferation in human CRC cell lines. (A) Cell viability was measured using the WST-8 assay. HCT116, RKO and CCD 841 CoN cells were treated with the indicated concentrations of TP4O for 24 h. Values were compared with 0 μ M TP4O as the control, which was considered 100%. Cell viability was decreased in CRC cells in a dose-dependent manner. The IC_{50} values of TP4O in HCT116 and RKO cells were 661 and 381 μ M, respectively. The IC_{50} value of TP4O in the human normal colon epithelial cell line CCD 841 CoN was 5,347 μ M. (B) Cell proliferation was measured using a bromodeoxyuridine assay. HCT116, RKO and CCD 841 CoN cells were treated with the indicated concentrations of TP4O for 24 h. The values were compared with those obtained with 0 μ M TP4O, which served as the control (100%). The percentage of cell proliferation decreased in a dose-dependent manner. The data are represented as the mean \pm standard deviation. * $P < 0.05$. ** $P < 0.01$. The results were obtained from three independent experiments. TP4O, terpinen-4-ol; CRC, colorectal cancer; IC_{50} , half maximal inhibitory concentration.

used to determine the percentage of stained areas in the digital photomicrographs.

Statistical analysis. The data are expressed as the mean \pm standard deviation. For comparison of >2 groups, analysis of variance (ANOVA) was performed. If the ANOVA results were significant, a Dunnett's test for comparison with the control was used. For comparison of two groups, an unpaired Student's t-test was performed. All indicated P-values were two-sided, and $P < 0.05$ was considered to indicate a statistically significant difference. All statistical analyses were performed using GraphPad Prism software version 5.01 (GraphPad Software, Inc., La Jolla, CA, USA).

Results

TP4O inhibits CRC cell viability and proliferation. To evaluate cytotoxicity following TP4O treatment, cell viability was measured using a WST-8 assay. TP4O significantly decreased the viability of the human CRC cell lines HCT116 at 100, 1,000 and 10,000 μ M, and RKO at 1,000 and 10,000 μ M (Fig. 1A). The effect of TP4O was dose dependent. The IC_{50} values of TP4O in HCT116 and RKO cells were 661 and 381 μ M, respectively. By contrast, the IC_{50} value of TP4O in the normal human colon epithelial cell line CCD 841 CoN was 5,347 μ M.

In addition, TP4O significantly inhibited cell proliferation in a dose-dependent manner in CRC cell lines and CCD 841 CoN cell line, particularly in the CRC cell lines at 1,000 μ M (Fig. 1B).

TP4O induces apoptotic cell death in CRC cells.

TP4O-induced apoptotic cell death was determined using a caspase-3/7 activity assay and Annexin V assay. DNA fragmentation and exposure of PS are typical phenomena observed in apoptotic cell death (22). Caspase-3/7 activity was enhanced in TP4O-treated cells in a dose-dependent manner (Fig. 2A). The distribution of apoptotic cells, which were located in the top- and bottom-right quadrants, is presented in a bar graph; the percentage of apoptotic cells was significantly increased in cells treated with 1,000 μ M TP4O compared with 0 μ M TP4O ($P < 0.05$; Fig. 2B). Although there was no statistical difference, the percentage of Annexin V-positive cells tended to increase in HCT116 and RKO cell cultures treated with 1,000 μ M TP4O (Fig. 2C and D). The lower level of LDH release following TP4O treatment indicates that TP4O did not induce necrotic LDH release in HCT116, RKO (Fig. 2E) or CCD 841 CoN (data not shown) cell lines.

TP4O generates ROS in CRC cells, and TP4O-induced cell death is rescued by antioxidant reagents. In the presence of

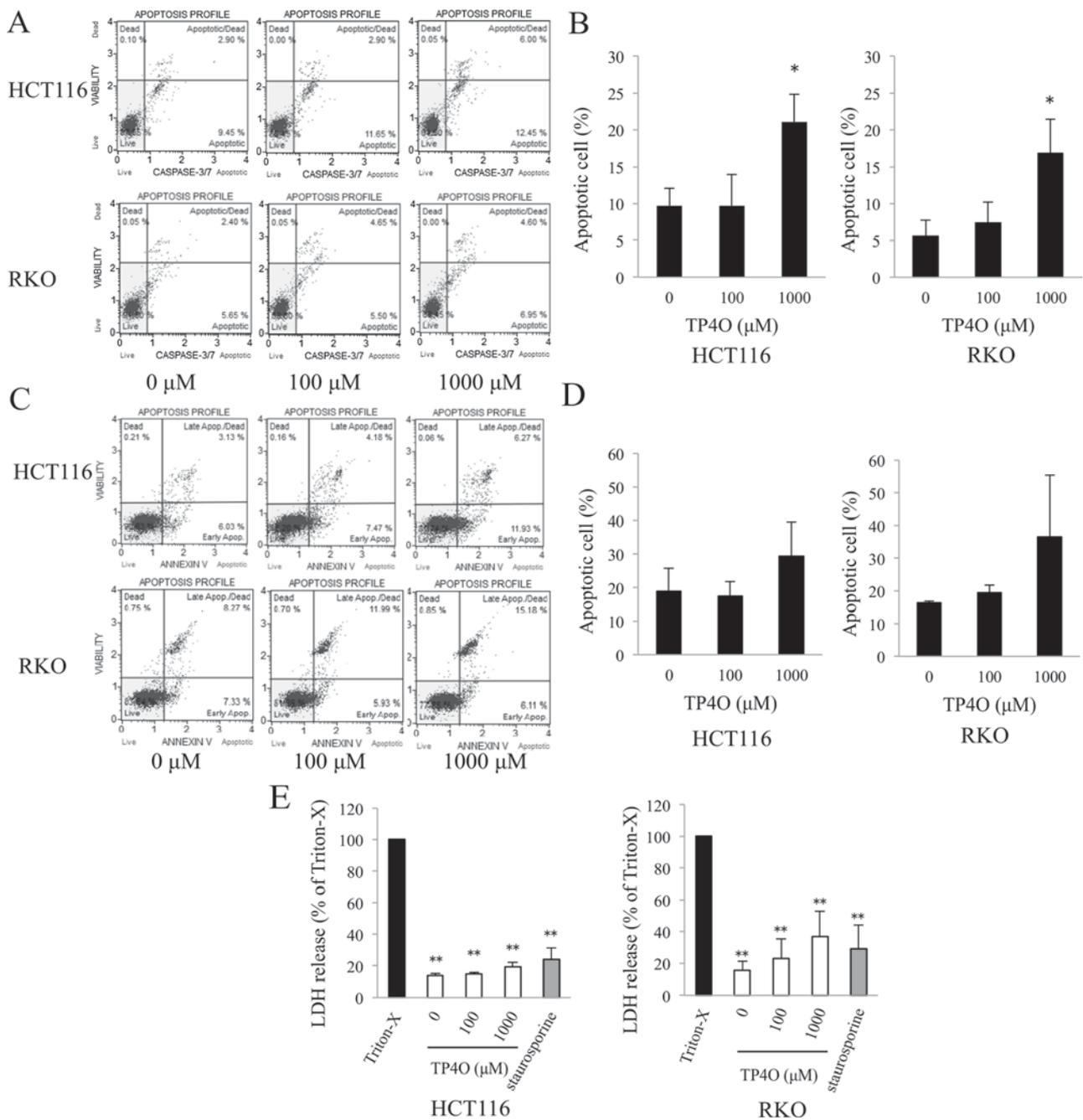


Figure 2. TP40 induces the apoptosis of human CRC cell lines. (A) HCT116 and RKO cells were treated with the indicated concentrations of TP40 for 6 h. Caspase-3/7 activity and cell viability were measured by flow cytometry. Caspase-3/7 activity was detected by fluorescence of a DNA-binding dye and cell viability were detected with the detection of 7-aminoactinomycin D fluorescence using the Muse™ Cell Analyzer. (B) The percentage of apoptotic cells, which are located in the top- and bottom-right quadrants, is presented. The values were compared with those obtained with 0 μM TP40. The data are presented as the mean ± SD. *P<0.05. The results were obtained from three independent experiments. (C) HCT116 and RKO cells were treated with the indicated concentrations of TP40 for 12 h. The extent of apoptosis was measured with Annexin V staining. (D) The percentage of apoptotic cells, which are located in the top- and bottom-right quadrants, is presented. The values were compared with those obtained with 0 μM TP40. The data are presented as the mean ± SD. The results were obtained from four independent experiments. (E) HCT116 and RKO cells were treated with the indicated concentrations of TP40 for 12 h, and the LDH level in the supernatant was measured. The values were compared with those obtained with 1% Triton X-100 as the control (100%). To determine whether LDH release was associated with necrosis or apoptosis, cells were treated with Triton X-100 or staurosporine, respectively. The data are represented as the mean ± SD. **P<0.01. The results were obtained from three independent experiments. TP40, terpinen-4-ol; CRC, colorectal cancer; IC₅₀, half maximal inhibitory concentration; SD, standard deviation; LDH, lactate dehydrogenase; Apop., apoptosis.

DMPO, the hydroxyl radical can be detected as as four spikes in ESR (23). ESR measurements revealed that treatment with 1,000 μM TP40 for 15 min induced the generation of the hydroxyl radical in CRC cells (Fig. 3A), whereas the hydroxyl radical was barely detectable in CCD841 CoN cells (data not shown). The increase in oxidative stress was quantified by DHE

reaction. TP40 administration led to an increase in the number of cells with high ROS (Fig. 3B). There was a significant increase in oxidative stress in HCT116 cells (P<0.05; Fig. 3C). This result implies that TP40 enhances the accumulation of intracellular ROS in HCT116 cells. There was a non-significant increase observed in RKO cells following the administration of TP40.

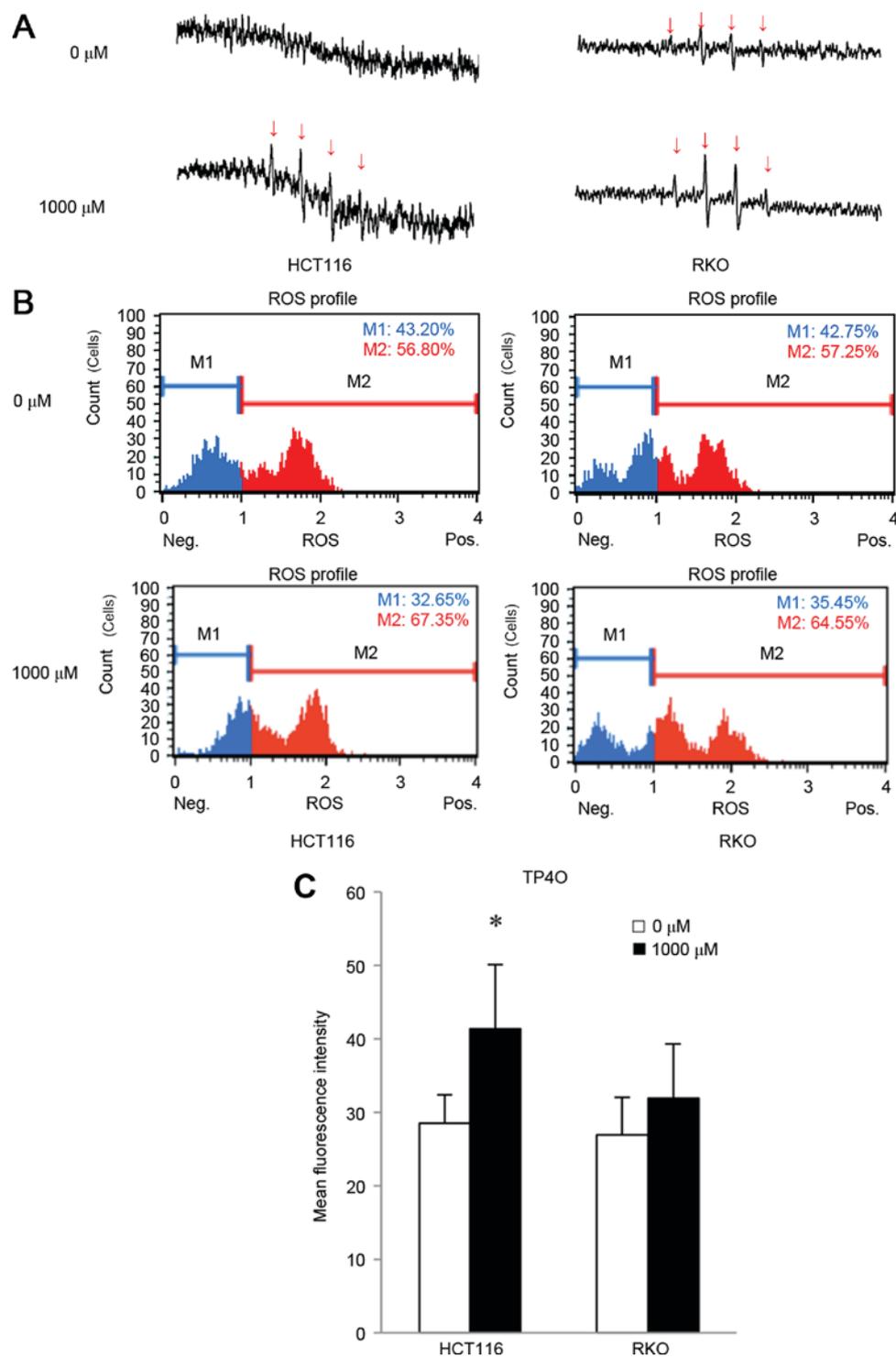


Figure 3. TP4O induces ROS in human CRC cell lines. (A) ESR measurement. HCT116 and RKO cells were treated with 0 or 1,000 μM of TP4O for 15 min. ESR was measured in a respiration buffer containing 5,5-dimethyl-1-pyrroline-N-oxide as the spin-trapping agent. The production of the hydroxyl radical was detected with four spikes in ESR. Spikes derived from the hydroxyl radical (indicated with arrows) became apparent in cells exposed to 1,000 μM TP4O. (B) Quantitative measurements of cellular populations undergoing oxidative stress, based on the detection of ROS. HCT116 and RKO cells were untreated or treated with 1,000 μM TP4O for 24 h. The distribution of cells according to levels of intracellular ROS was represented as a histogram. Cells were divided into two groups: The blue area (M1) represented the cells with low ROS, and the red area (M2) represented the cells with high ROS. The administration of TP4O led to an increase in the number of cells in the red area. (C) The mean fluorescence intensity of each group is presented. The values were compared with those obtained with 0 μM TP4O. The data are presented as the mean \pm SD. * $P < 0.05$. The results were obtained from four independent experiments. TP4O, terpinen-4-ol; CRC, colorectal cancer; IC₅₀, half maximal inhibitory concentration; SD, standard deviation; ROS, reactive oxygen species; DHE, dihydroethidium; ESR, electron spin resonance.

To determine the source of TP4O-induced ROS, CRC cells were observed by confocal inverted fluorescence microscopy. The location of the mitochondria was represented by

MitoTracker Green FM™ fluorescence, while accumulated ROS were represented by MitoSOX Red™ Mitochondrial Superoxide Indicator fluorescence. The merged image

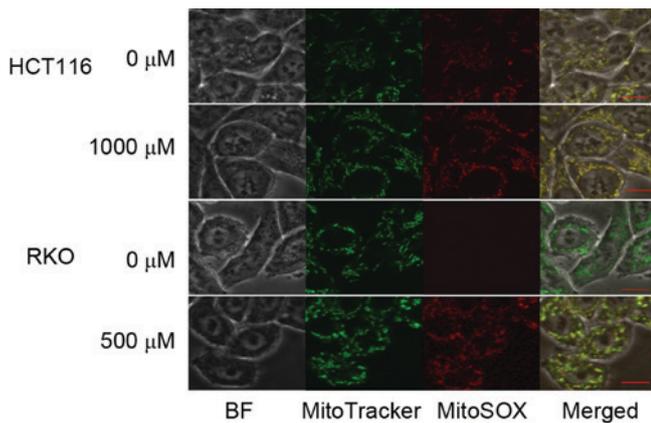


Figure 4. Localization of ROS using MitoSOX Red™ Mitochondrial Superoxide Indicator fluorescence in HCT116 and RKO cells. HCT116 and RKO cells were treated with the indicated concentration of TP4O for 24 h. To locate the mitochondria generating ROS, the same cells were incubated with MitoTracker Green FM™ and MitoSOX Red™ Mitochondrial Superoxide Indicator. Images of merged MitoTracker Green FM™ and MitoSOX Red™ Mitochondrial Superoxide Indicator fluorescence were constructed to identify ROS generated by mitochondria. Scale bars represent 10 μm . ROS, reactive oxygen species; TP4O, terpinen-4-ol.

demonstrated that the ROS induced by TP4O were generated by mitochondria (Fig. 4).

Protein levels of intrinsic oxidant scavengers were analysed by western blotting. Protein levels of SOD2 and GPX1 tended to increase following exposure to TP4O (Fig. 5A). However, there was no significant difference in comparison with that of 0 μM TP4O (Fig. 5B). The results suggested that ROS accumulation was not due to suppression of ROS scavengers, but due to increased ROS production. CRC cells were exposed to specific antioxidants (NAC, EB and MnTBAP), which prevented significant TP4O-derived cell death (Fig. 5C), demonstrating that ROS accumulation was the cause for cell death.

TP4O treatment inhibits tumor growth in a xenograft mouse model. A schematic representation of the experimental design is provided in Fig. 6A. Mice treated with TP4O developed significantly smaller tumors compared with the group treated with vehicle ($P < 0.05$; Fig. 6B). TP4O administration did not cause marked complications or body weight loss, and the serum levels of ALT and creatinine were not different in the TP4O group compared with those in the control group (Fig. 6C). For the histological analysis, tumors were stained using antibodies against the oxidative stress marker 8-OHdG. The areas of anti-8-OH-dG staining were larger in tissues from the TP4O group compared with those from the control group (Fig. 6D). In addition, areas stained by anti-cleaved caspase-3 antibody were larger in the TP4O group than in the control group (Fig. 6D).

Discussion

TP4O is an essential oil extracted from aromatic plants and is a type of monoterpene (10). Monoterpenes, the C10 class of isoprenoids, are a diverse family of natural products, ranging in structure from linear to polycyclic (24). Menthol, limonene, eucalyptol and citral are familiar monoterpenes that are used as food and cosmetic additives (8,24).

The anticancer effects of essential oils have previously been investigated by biological approaches focusing on pharmaceutical and therapeutic potentials of essential oils (7,9,10,25). Previous studies have indicated that the anticancer effects of TP4O are due to the induction of apoptosis in human melanoma cells (14), human non-small cell lung cancer (15), human leukaemia cells (16) and CRC cells (17). However, Greay *et al* (26) reported that TP4O induces necrosis and cell cycle arrest in murine mesothelioma and melanoma cells. In the present study, TP4O elevated caspase-3/7 activity and increased the number of Annexin V-positive cells without inducing necrotic LDH release in CRC cells. In the *in vivo* experiments, the number of cleaved caspase-3-positive cells increased significantly in the TP4O group compared with that in the control group. These results indicated that TP4O induced cell death in human CRC cell lines by apoptosis *in vitro* and *in vivo*.

In addition, the data from the present study revealed the source of ROS generation in the *in vitro* and *in vivo* TP4O treatments. Previous studies have demonstrated the antioxidant activity of essential oils [reviewed in (7)], including TTO, which contains TP4O (27). Kim *et al* (28) investigated the antioxidant activity of TTO, and determined that three compounds of TTO serve important roles in the antioxidant mechanism, whereas TP4O did not possess antioxidant activity. In the current study, the hydroxyl radical was directly detected by ESR. Moreover, the protein levels of the major ROS scavengers and GPX1 were not decreased, which indicates that the increase in ROS following TP4O treatment is not due to inhibition of the redox system but is the result of an increase in ROS production. The addition of antioxidant reagents inhibited TP4O-induced cell death. Taken together, these results suggested that the ROS induced following TP4O exposure serve a role in the anticancer effect of TP4O.

To maintain active proliferation, cancer cells produce more oxidative metabolites than normal cells (29). The redox system is activated in cancer, but cancer cells are not able to adapt to excess oxidative stress due to a decreased capacity for ROS metabolism compared with that of normal cells (29). However, normal cells produce lower levels of ROS than cancer cells; thus, they can adapt to increases in ROS (30,31). Previous reports have demonstrated that the differences in cellular metabolism of ROS could be a target for cancer treatment (31-37). There are three strategies to amplify intracellular oxidative stress. The first one is the inhibition of redox enzymes, i.e., GPX (35) or SOD (31). The second one is facilitating ROS production (36); the use of TP4O is included in this category. The third strategy is a combination of the two strategies mentioned above (37). These previous studies have revealed IC_{50} discrepancies between cancer cells and normal cells. The results of the present study support those from previous reports suggesting that ROS-mediated compounds have therapeutic potential against malignant neoplasms.

A number of limitations of the present study should be acknowledged. First, the underlying molecular mechanisms of ROS production by TP4O or the apoptotic pathway involved have not been detailed. As indicated by previous reports, excessive production and accumulation of ROS affects several mitogen-activated protein kinase signalling pathways, e.g., p38 mitogen-activated protein kinase, extracellular signal-regulated

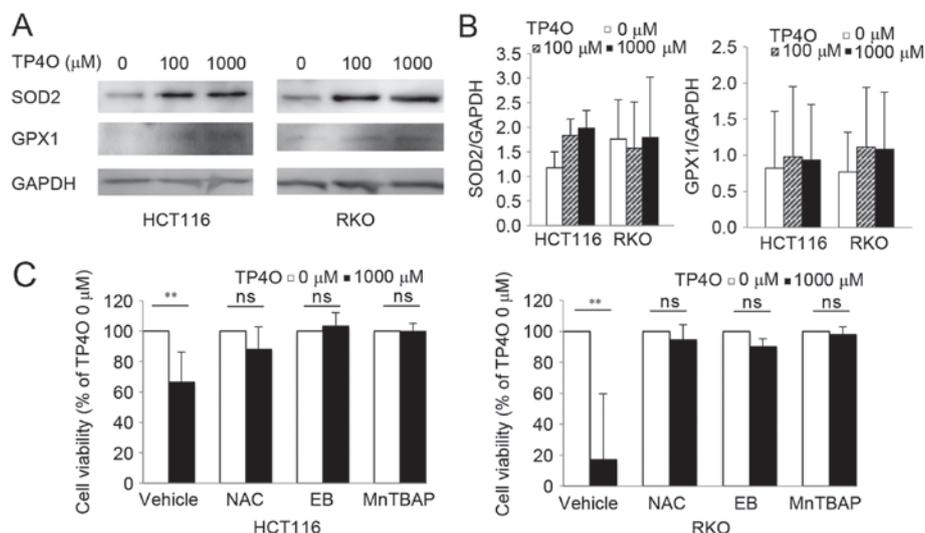


Figure 5. TP4O induces the activation of antioxidants, and antioxidant reagents rescue TP4O cytotoxicity. (A) Western blot analysis of antioxidant proteins. Lysates of HCT116 and RKO cells were examined following treatment with the indicated concentrations of TP4O for 24 h. GAPDH was used as a loading control. (B) HCT116 and RKO cells were treated with 0 or 1,000 μM TP4O in the presence of vehicle, 20 mM NAC, 20 μM EB or 100 μM MnTBAP for 24 h, and cell viability was measured by WST-8 assay. The values for each group were compared with those obtained with 0 μM TP4O. The data are represented as the mean \pm standard deviation. ** $P < 0.01$. The results were obtained from three independent experiments. TP4O, terpinen-4-ol; MnTBAP, manganese (III) tetrakis (4-benzoic acid) porphyrin chloride; SOD2, superoxide dismutase 2; GPX1, glutathione peroxidase 1; NAC, N-acetyl-L-cysteine; EB, ebselen; ns, not significant.

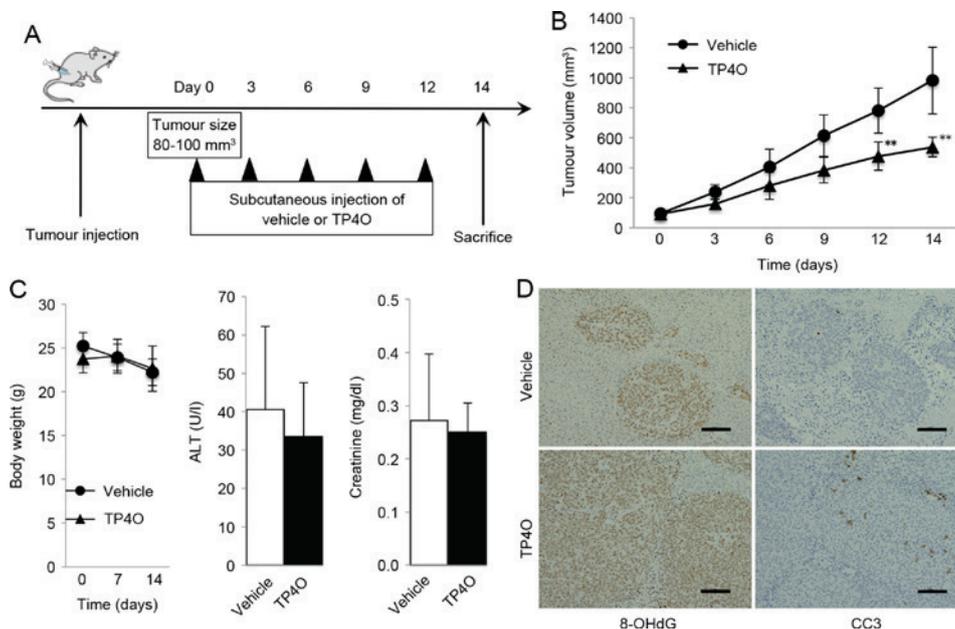


Figure 6. *In vivo* antitumor activity of TP4O in a xenograft model. (A) Schematic experimental design. A total of 2×10^6 HCT116 cells were subcutaneously injected into the right flank of ICR-SCID mice receiving 250 μl of vehicle or TP4O (200 mg/kg) every 3 days by subcutaneous injection. Mice were sacrificed 14 days after the first day of administration. (B) Tumor volume was measured over time. Increase in tumor volume was suppressed in the TP4O group. The data are presented as the mean \pm standard deviation. ** $P < 0.01$. (C) Mice were sacrificed after 14 days, and body weight as well as serum levels of ALT and creatinine were measured. (D) Tumors were stained with antibodies against 8-OHdG and CC3. 8-OHdG- and CC3-stained positive areas were assessed by the ratio of the stained area to the entire tumor area in high-magnification ($\times 200$) digital images in three randomly selected fields for each sample. Scale bars represent 100 μm . TP4O, terpinen-4-ol; CC3, cleaved caspase-3; 8-OHdG, 8-hydroxy-2'-deoxyguanosine; ALT, alanine aminotransferase.

kinase and c-Jun N-terminal kinase signalling pathways, which may be associated with ROS production (31,34,38,39). Thus, further studies are required to elucidate the association between TP4O and ROS generation or signalling pathways. Second, the activities of ROS scavengers were estimated using western blot analysis of SOD2 and GPX1; however, the actual activities of these enzymes or other redox systems have not been measured.

The possibility that TP4O suppresses other redox systems cannot be ruled out. Third, ROS cause DNA damage and can promote tumorigenic signalling pathways (40). ROS are also known to lead to Alzheimer's disease (41), ischaemic vascular disease (42), non-alcoholic steatohepatitis (43) and other diseases (44). Although the data from the current study demonstrated that TP4O inhibited CRC xenograft growth without marked liver

or renal injury *in vivo*, more precise assessment of the risk of the use of ROS for cancer therapy is required. The therapeutic concentration in the present study of TP4O was similar to that of a previous report (17), which is considered high in clinical use. Therefore, consideration of a practical dose for future clinical use is crucial. As Shapira *et al* (17) reported, combination therapy may reduce the dose of TP4O and existing cancer drugs. Additionally, Berndtsson *et al* (45) reported the ROS accumulation effect of oxaliplatin, which is typically used for the treatment of CRC (4). Thus, it is expected that combination therapy with TP4O and oxaliplatin would result in a synergistic effect. Upon sufficiently addressing the aforementioned issues, this new therapeutic strategy could be applicable in a clinical setting.

In conclusion, the results from the present study suggest that TP4O induces apoptosis in CRC cells through the generation of ROS *in vitro* and *in vivo* without damaging normal cells. Furthermore, TP4O is potentially useful for the future development of therapies against CRC.

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