

# Mining the glioma susceptibility genes in children from gene expression profiles and a methylation database

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**Abstract.** Glioma is the most common type of primary brain tumor, which is associated with a poor prognosis due to its aggressive growth behavior and highly invasive nature. Research regarding glioma pathogenesis is expected to provide novel methods of adjuvant therapy for the treatment of glioma. The use of bioinformatics to identify candidate genes is commonly used to understand the genetic basis of disease. The present study used bioinformatics to mine the disease-related genes using gene expression profiles (GSE50021) and dual-channel DNA methylation data (GSE50022). The results identified 17 methylation sites located on 33 transcription factor binding sites, which may be responsible for downregulation of 17 target genes. glutamate metabotropic receptor 2 was one of the 17 downregulated target genes. Furthermore, inositol-trisphosphate 3-kinase A (ITPKA) was revealed to be the gene most associated with the risk of glioma in children. The protein coded by the ITPKA gene appeared in all risk sub-pathways, thus suggesting that ITPKA was the gene most associated with the risk of glioma, and inositol phosphate metabolism may be a key pathway associated with glioma in children. The identification of specific genes helps to determine the pathogenesis and possible therapeutic targets for the treatment of glioma in children.

## Introduction

Glioma commonly occurs in the brain or spine, and is a particularly lethal solid tumor that arises from support cells in the central nervous system. Glioma can be classified into the

following types: Astrocytic tumors, oligodendrogliomas and oligoastrocytomas (1). In addition, patients with glioma often suffer from fatigue, motor difficulties, drowsiness, communication difficulties, headaches and visual problems (2). A previous study reported that the median survival time of patients with the most malignant form of glioma is only 14.6 months, and the 5-year survival rate is only 9.8% (3). The high rates of morbidity and mortality associated with glioma are due to its chemoradiation resistance, and high proliferative and invasive capabilities (4).

In addition to childhood leukemia, glioma is the most common type of tumor in children, and accounts for 80% of malignant brain tumors (5). There is a peak incidence of glioma between the ages of 0 and 8 years (6). Patients with glioma that have survived surgery are more affected by their tumor and its resection than previously appreciated (7). In addition, in patients with childhood brain tumors treated with radiotherapy, memory deficits reflect general cognitive dysfunction (8). At present, the standard approach for treating glioma in children is chemotherapy; however, the safety and effectiveness of chemotherapy in children remains a problem.

Understanding the regulation of the pathological activities of glioma is important for its eradication. Research regarding glioma pathogenesis is expected to provide novel methods of adjuvant therapy for the treatment of glioma (9). With scientific improvements, the concept of epigenetics offers a potential method for cancer therapy. Epigenetics refers to stable alteration in gene expression with no underlying modifications in genetic sequence (10); DNA methylation is central to the aberrant epigenetics associated with cancer (11). DNA methylation is a biochemical process where a methyl group is added to the CpG islands of DNA nucleotides by DNA methyltransferases in mammals. DNA methylation of CpG dinucleotides is considered a distinct inhibitory pathway that may mediate stable silencing (12). CpG islands are located on promoter regions, and the sequences that surround promoter regions. It has previously been reported that methylation at the CpG islands may induce tumorigenesis, since the abnormal expression of genes is involved in tumor development (13). Costello *et al* (14) reported that transcriptional repression of a tumor suppressor gene, cyclin-dependent kinase inhibitor 2A, was mediated by aberrant methylation of the CpG island in glioma. A previous

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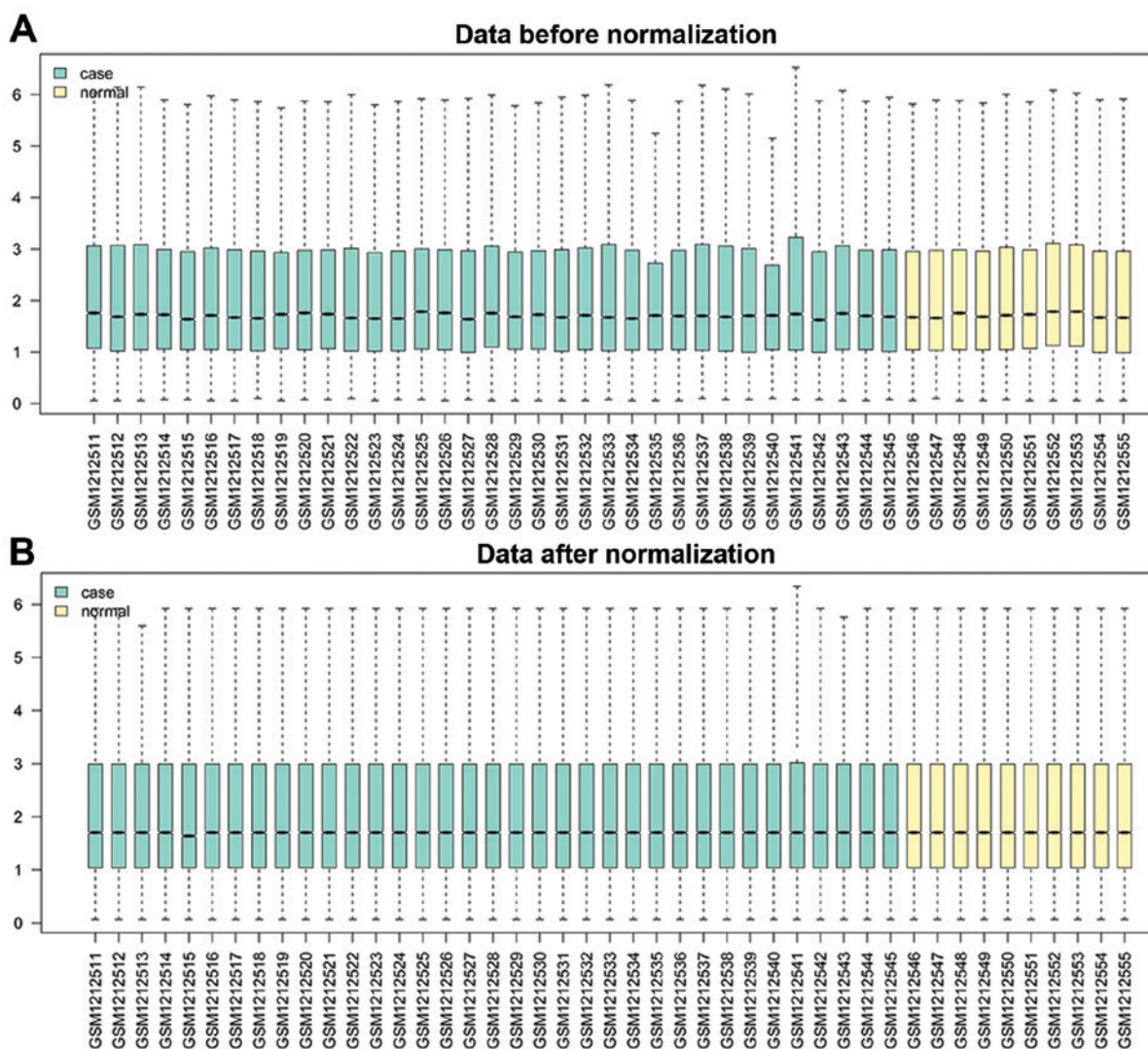


Figure 1. Expression boxplots for glioma-related genes. Box plots represent the mean  $\pm$  interquartile range, with whiskers extending to the minimum and maximum value. (A) Data prior to normalization; (B) data after normalization. GSM1212511-GSM1212545 (green) represent gliomas samples; GSM1212546-GSM1212555 (yellow) represent normal samples.

study also demonstrated that promoter methylation of DNA repair genes is associated with the tumorigenesis and development of glioma (15). However, despite extensive research, the exact mechanisms underlying how DNA methylation induces glioma remain unclear.

The present study compared DNA methylation and differentially expressed genes (DEGs) between glioma and normal tissues using Gene Expression Omnibus (GEO) data. Subsequently, the important genes and pathways in glioma were screened by enrichment analysis of transcriptional regulatory networks and sub-pathways. The present study aimed to determine the possible molecular mechanisms and the aberrant promoter methylation of tumor-associated genes in glioma, via identification of key genes and pathways. The results may provide a theoretical basis and identify potential molecular biological therapies for the treatment of glioma in children.

## Materials and methods

**Data sources.** GSE50021 DNA microarray data (16) for gene expression profiling and GSE50022 dual-channel

DNA methylation data (16) were downloaded from GEO (<http://www.ncbi.nlm.nih.gov/geo/>). GSE50021 contains 45 samples, including 35 samples from patients with glioma (average age  $1.008 \pm 1.190491$  years) and 10 autopsy samples of normal human brain tissue. Data from the chip (Illumina HumanHT-12 WG-DASL v4.0 Expression Beadchip; Illumina, San Diego, CA, USA) were annotated based on the GPL13938 platform. GSE50022 contains 28 samples from patients with glioma (average age,  $0.9433333 \pm 0.781596$  years). Data from the chip (Illumina HumanMethylation450 BeadChip [UBC enhanced annotation v1.0]; Illumina) were annotated based on the GPL16304 platform.

**Preprocessing of gene expression profiles.** The expression values for genes with various capture probes were used to determine a mean value for each gene. The K-nearest neighbour (KNN) impute missing values module was used to impute missing values by assigning gene expression values based on the nearest neighbors of the gene ( $k=10$ ). All expression values were normalized using quantile normalization, as implemented

Table I. Key methylation sites.

ID	Chromosome	Mapinfo	tfbs_start	tfbs_end	Transcription factor	Distance_closest_TSS	Closest_TSS_gene_name
cg06191091	chr17	30583855	30583848	30583862	USF	-9339	RHBDL3
cg02629157	chr9	138670609	138670546	138670568	TCF11	25013	KCNT1
cg11709150	chr1	2440438	2440431	2440444	TCF11	10256	PLCH2
cg04585209	chr11	6292311	6292257	6292272	TAXCREB	306	CCKBR
cg10707626	chr3	51747098	51747027	51747051	STAT5A	6018	GRM2
cg06191091	chr17	30583855	30583849	30583860	SREBP1	-9339	RHBDL3
cg12603173	chr11	64508421	64508409	64508423	RREB1	-66	RASGRP2
cg11025960	chr3	51749188	51749177	51749195	RFX1	8108	GRM2
cg10692302	chr3	51747227	51747224	51747245	PPARG	6147	GRM2
cg02629157	chr9	138670609	138670558	138670569	POU6F1	25013	KCNT1
cg11014582	chr6	76333727	76333675	76333696	PAX6	-852	LMO7
cg04341461	chr1	2410006	2409978	2410006	PAX5	-1616	PLCH2
cg10692302	chr3	51747227	51747222	51747252	PAX4	6147	GRM2
cg04625615	chr15	41788368	41788310	41788330	P53	2313	ITPKA
cg07200386	chr8	22079169	22079113	22079135	OLF1	10682	PHYHIP
cg11014582	chr6	76333727	76333676	76333683	NKX25	-852	LMO7
cg09864712	chr16	726786	726720	726749	MYOGNF1	712	RHBDL1
cg06191091	chr17	30583855	30583848	30583862	MYCMAX	-9339	RHBDL3
cg00810908	chr3	13612319	13612306	13612320	MEIS1AHOXA9	2080	FBLN2
cg11025960	chr3	51749188	51749181	51749190	LMO2COM	8108	GRM2
cg03358506	chr8	22058702	22058688	22058703	ISRE	31149	PHYHIP
cg07776629	chr16	57989122	57989116	57989129	IRF2	15898	CNGB1
cg07776629	chr16	57989122	57989116	57989129	IRF1	15898	CNGB1
cg10692302	chr3	51747227	51747225	51747244	HNF4	6147	GRM2
cg06632557	chr11	61313548	61313495	61313505	HMX1	-3678	SYT7
cg00155846	chr9	138011566	138011506	138011522	HAND1E47	14081	OLFM1
cg11025960	chr3	51749188	51749181	51749190	GATA3	8108	GRM2
cg11025960	chr3	51749188	51749179	51749193	GATA1	8108	GRM2
cg04625615	chr15	41788368	41788310	41788321	GATA3	2313	ITPKA
cg05392169	chr9	138011814	138011802	138011816	FOXO3	14329	OLFM1

tfbs, transcription factor binding sites; TSS, transcription start site.

in the normalize.quantiles function of the preprocessCore R package (17). Expression values were displayed using a box plot.

*Preprocessing of DNA methylation data.* For probes with missing values in the original methylation index matrix the KNN impute missing values module was used to impute the missing values by assigning methylation gene expression values based on the nearest neighbors of the gene ( $k=10$ ).

*Screening of differentially expressed genes (DEGs).* The probe identifiers were converted into gene names. The DEG analysis was conducted using limma package (<http://www.bioconductor.org>).  $|\log_2FC| > 1$  and  $P < 0.05$  were set as the cut-offs to screen for DEGs. DEGs were initially identified following an analysis using Student's t-test. The identified DEGs were considered glioma-related genes in children.

*Screening of key methylated sites.* According to the annotation information, probes were screened to determine which methylated sites were located near DEGs. Subsequently, the preliminarily reserved methylated sites were screened, and  $\geq 80\%$  of samples with a methylation index of  $\geq 0.8$  were reserved. Due to the lack of clear definition of the promoter regions of genes, 50 kb on either side of the transcription start site was considered the promoter region. Finally, the methylated sites were re-screened, and methylated sites were reserved that were located 50 kb on either side of the transcription start site. After screening three times, the remaining methylated sites were considered key methylated sites, which were located in the promoter region of genes and have a high methylation index, thus potentially influencing the expression of downstream genes.

*Screening of significant transcription factors.* Using the transcription factor binding sites (TFBS) prediction tool from

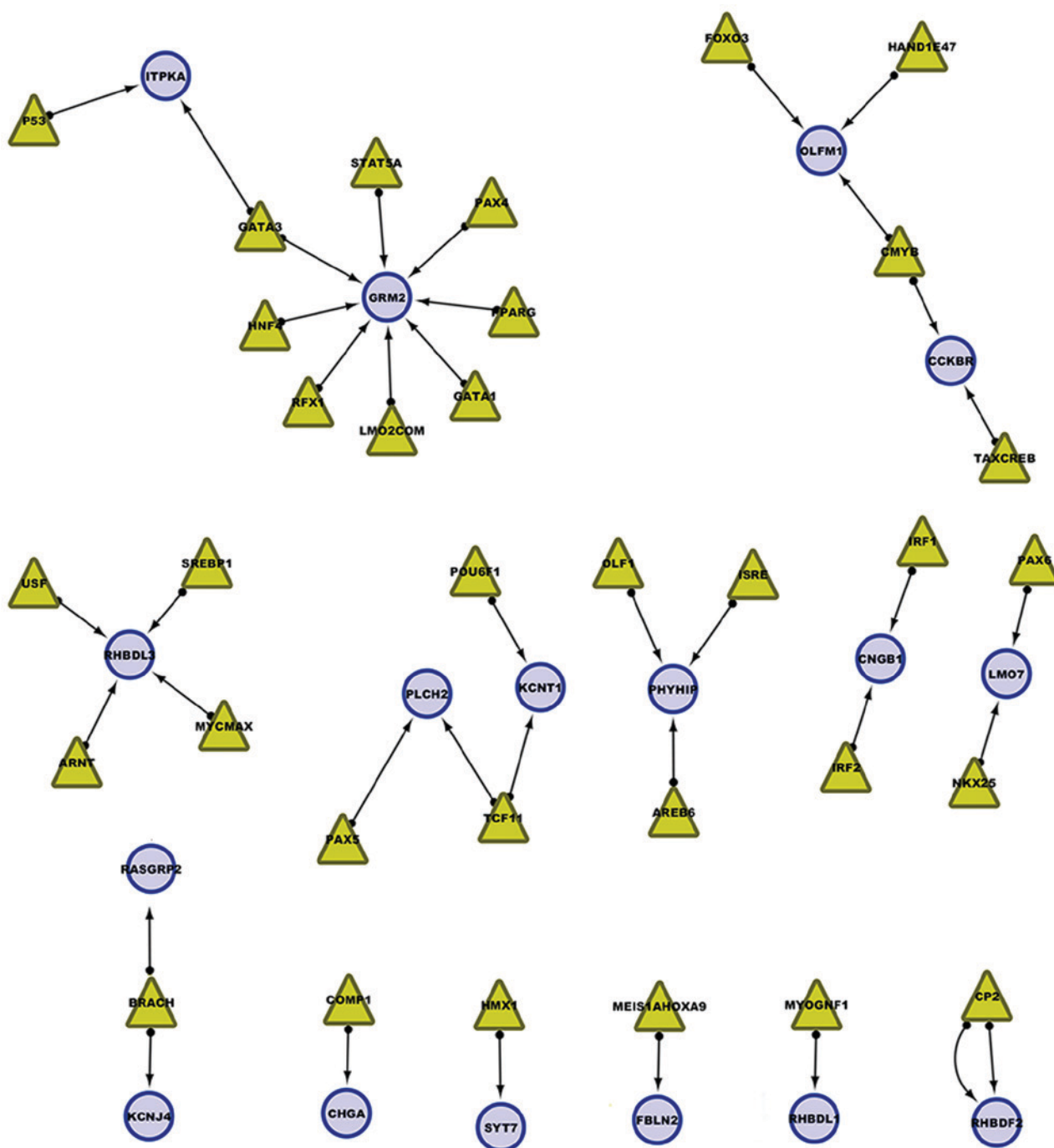


Figure 2. Transcriptional regulatory networks. Yellow triangles represent transcription factors; purple circles represent target genes; arrows represent transcriptional regulation.

University of California, Santa Cruz (UCSC; <http://genome.ucsc.edu>) the methylated sites were screened in TFBS. A transcription factor regulatory network was generated using Cytoscape (<http://cytoscape.org/>), which is an open source desktop tool for integrating, visualizing and analyzing data in the context of biological networks. Enrichment analysis of potential downregulated genes influenced by methylation was performed using the Database for Annotation, Visualization and Integrated Discovery (<http://david.abcc.ncifcrf.gov/>).

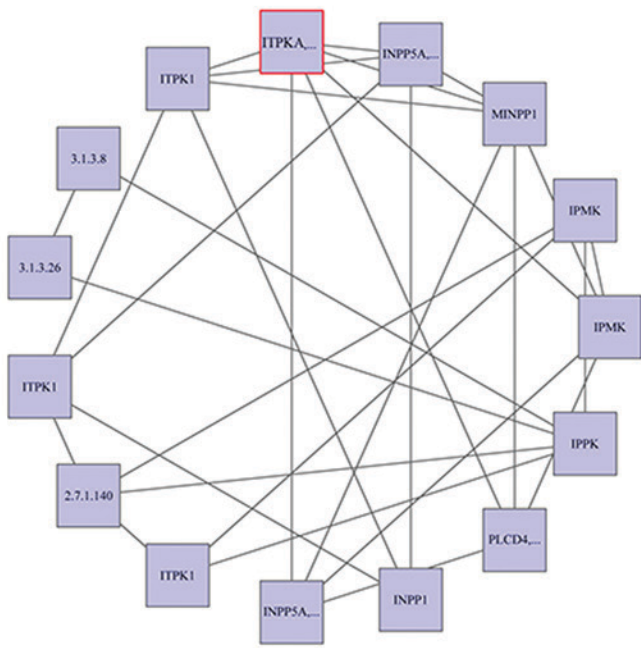
*Analysis of risk sub-pathways.* A hypergeometric test for risk sub-pathways was performed by

application of the R package *iSubpathwayMiner* (<http://cran.r-project.org/web/packages/iSubpathwayMiner/>). All genes that were mapped to sub-pathways with a significant enrichment were reported. These pathways were used to identify glioma-related pathways.

## Results

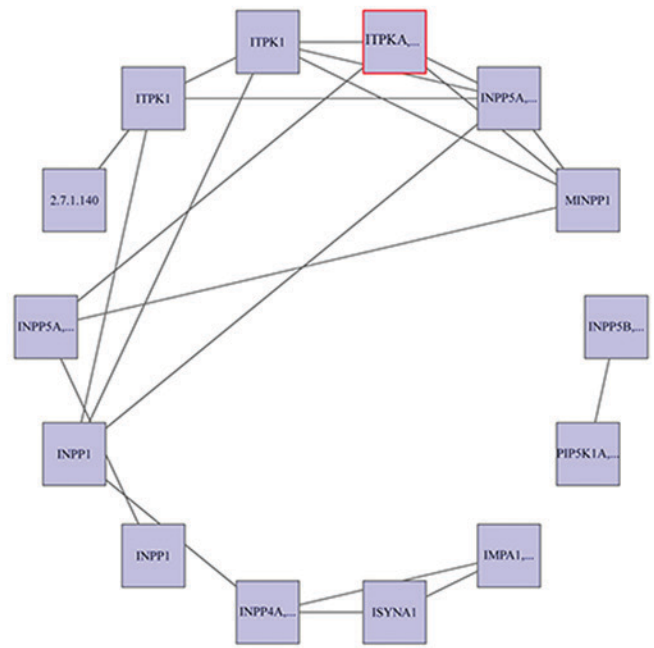
*Preprocessing of gene expression profiles and DNA methylation data.* To explore glioma-related genes in children, 19,252 genes were identified from 45 samples following preprocessing of gene expression profiles. As shown in Fig. 1 expression boxplots were

path:00562\_1 Inositol phosphate metabolism



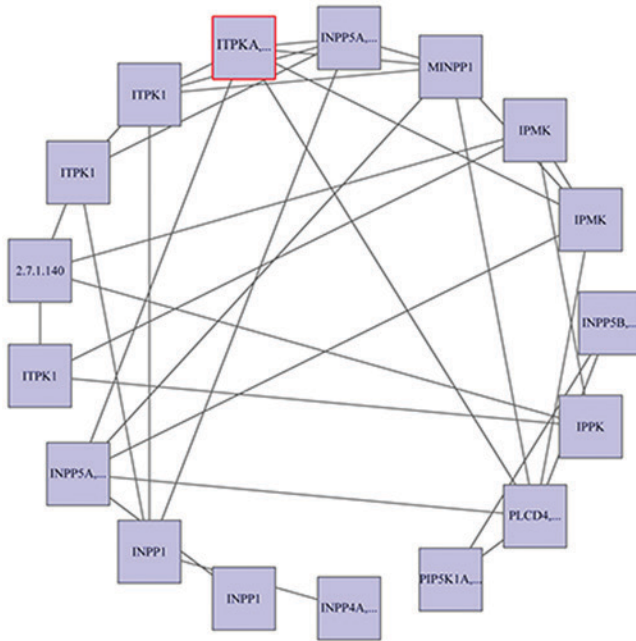
p-value= 0.0182007558864037

path:00562\_2 Inositol phosphate metabolism



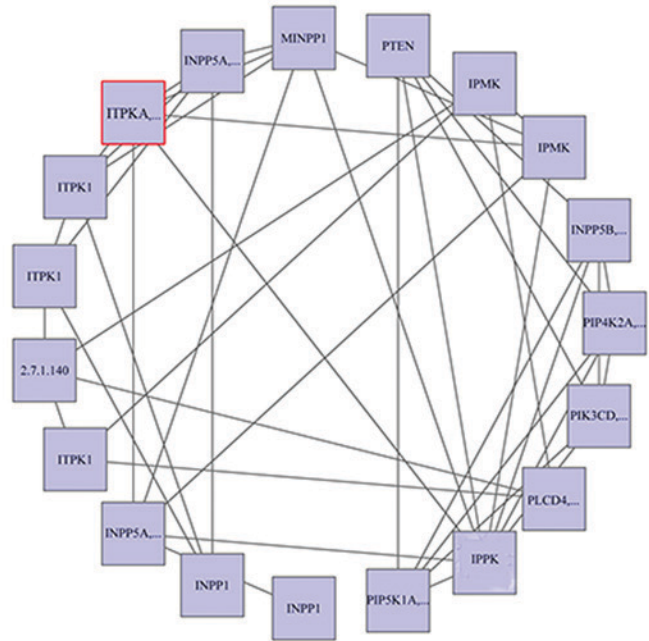
p-value= 0.0209035740380185

path:00562\_3 Inositol phosphate metabolism



p-value= 0.0280765742267587

path:00562\_4 Inositol phosphate metabolism



p-value= 0.0334235323944514

Figure 3. Risk sub-pathways in inositol phosphate metabolism. ITPKA gene is marked with a red rim. ITPKA, inositol-trisphosphate 3-kinase.

generated before and after normalization. The original expression datasets from all conditions were processed into expression estimates using the median polish algorithm. As shown in a box plot (Fig. 1B), the median of different samples was almost on the

same line after normalization, which shows an excellent degree of standardization.

Data located in the X/Y and mitochondrial chromosomes were removed. Subsequently, a methylation index array

identified 371,920 sites from the 28 samples, following imputation of missing values.

**Screening of DEGs.** DEGs were analyzed using limma package. Using the selective conditions ( $\log_2FC > 1$  and  $P < 0.05$ ), 58 DEGs were identified, all of which were downregulated.

**Identification of DNA methylation in the promoter regions of downregulated genes.** TFBS methylation has a strong impact on the regulatory role of transcription factors and the normal expression of downstream genes, thus affecting body functioning. DNA methylation in the promoter region is an important aspect of DNA methylation in cancer (18,19).

The 58 downregulated DEGs identified in the present study were overlapped with DNA methylation data; 1,123 methylation sites were revealed to be associated with the 58 downregulated genes. The sites were distributed evenly over the 22 autosomes. The methylation index of the 1,123 methylation sites was analyzed, and  $\geq 80\%$  of samples with a methylation index of  $\geq 0.8$  were reserved. This resulted in the reservation of 168 methylation sites. To further analyze the 168 methylation sites, methylated sites located 50 kb on either side of the transcription start sites were reserved. This resulted in the reservation of 119 methylation sites located on the transcription start sites of 43 genes. These 43 genes are considered potential glioma susceptibility genes in children. Finally, 17 methylation sites located on 33 TFBS were identified, which may regulate 17 of the downregulated target genes. These results were determined using the TFBS prediction tool from UCSC (Table I).

**Analysis of transcriptional regulatory networks.** To construct a transcription factor regulatory network, predictive models of transcriptional regulatory networks were generated using Cytoscape. A transcriptional regulatory network including 33 transcription factors and target genes is presented in Fig. 2. The eight methylation sites in the TFBS of glutamate metabotropic receptor 2 (GRM2) potentially influenced the binding of eight transcription factors to TFBS. It may therefore be hypothesized that the downregulation of GRM2 is caused by methylation of these TFBS. In addition, methylation sites in the TFBS of rhomboid, veinlet-like 3 (*Drosophila*) (RHBDL3) resulted in the failure of four transcription factors binding with TFBS, thus affecting transcriptional regulation. Furthermore, the transcription factor CP2 regulates rhomboid 5 homolog 2 (*Drosophila*) (RHBDF2) expression; in the present study, two methylation sites were detected in TFBS of CP2, which may influence transcriptional regulation of RHBDF2.

**Functional analysis of key genes.** To generate the downregulated target gene-related risk sub-pathways, a hypergeometric test was performed using R package iSubpathwayMiner. Four sub-pathways were generated, which were associated with inositol phosphate metabolism (hsa00562). As shown in Fig. 3 the inositol-trisphosphate 3-kinase protein (EC 2.7.1.127), which is coded by the ITPKA gene (marked with a red rim), appeared in all of the risk sub-pathways. In conclusion, ITPKA was considered to be the gene most associated with the risk of glioma in children.

## Discussion

Bioinformatics is an interdisciplinary field that develops methods and software tools for understanding biological data. The identification of candidate genes and single nucleotide polymorphisms are common uses of bioinformatics to understand the genetic basis of disease (20). The present study identified 17 methylation sites located on 33 TFBS, which may regulate the 17 downregulated target genes. GRM2 was identified as one of the 17 downregulated target genes. Furthermore, ITPKA was revealed to be the gene most associated with glioma risk in children. The protein coded by the ITPKA gene appeared in all risk sub-pathways, thus suggesting that ITPKA is the gene most associated with glioma risk, and inositol phosphate metabolism is the key pathway associated with glioma in children.

Glioma is the most common type of primary brain tumor, which is associated with a poor prognosis for patients due to its aggressive growth behavior and highly invasive nature (21). Abnormal metabolic networks are closely associated with the pathogenesis of glioma. A metabolic pathway is a series of chemical reactions, which are catalyzed by enzymes, where the product of one enzyme acts as a substrate for the next. Abnormal gene expression of specific enzymes may affect other enzymatic genes in the network. Therefore, identification of disease-related genes in the metabolic pathway is of great importance.

The correct targeting, localization, regulation and signaling of metabotropic glutamate receptor 2 (mGluR2), which is coded by the GRM2 gene, represents a major mechanism underlying the complex function of neuronal networks (22). Dysfunction of mGluR and associated proteins is associated with neurodegenerative and neuropsychiatric disorders. Several types of glioma have been reported to release high levels of glutamate, which promotes malignancy. A previous study reported that activation of mGluR2 supports the growth of human glioma cells in culture, and that antagonists of mGluR2 should be assessed for their ability to reduce tumour growth *in vivo* (23). Arcella *et al* (24) demonstrated that mGluR2/3 receptor antagonists are of potential use in the experimental treatment of malignant gliomas. Furthermore, ectopic expression of ITPKA in lung cancer cells has been shown to increase their metastatic potential, since the protein exhibits two actin-regulating activities (25). The ITPKA gene has also been reported to be hypermethylated in human patients with normal karyotype acute myeloid leukemia (26). Furthermore, the remaining 15 downregulated genes identified in the present study may be beneficial for the generation of a target therapy for the treatment of patients with glioma.

In conclusion, the present study proposed a novel method for extracting DEGs from gene expression datasets combined with DNA methylation data, and applied it to the occurrence of glioma in children. Results of the functional analysis revealed that ITPKA was considered the gene most associated with the risk of glioma, and inositol phosphate metabolism may serve an important role in glioma in children. The identification of specific genes may improve understanding regarding the pathogenesis and possible therapeutic targets of glioma in children.

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