

Synergistic inhibition of colon cancer cell growth by a combination of atorvastatin and phloretin

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Abstract. Atorvastatin (ATST), a drug commonly used to reduce the levels of cholesterol and low-density lipoproteins, is a prospective agent for the prevention of colorectal cancer in patients with hyperlipidemia. ATST in combination with functional components is a promising strategy for cancer chemoprevention. In the present study, the growth inhibitory effect of ATST combined with phloretin (PT) on SW620 and HCT116 colon cancer cells was investigated. The results of MTT assays indicated that the combination of PT and ATST markedly reduced cell survival in both cell lines compared with PT or ATST treatment administered individually. The interaction indexes between PT and ATST, which were used to analyze their interaction pattern, were computed by the median-effect equation. The interaction indexes of each PT and ATST concentration pair were <1.0, which indicated a strong synergistic effect between the two compounds. The data obtained by flow cytometry and western blot analysis of cleaved-poly (ADP-ribose) polymerase indicated a synergistic effect resulted in apoptosis and cell cycle arrest at the G₂/M checkpoint. Furthermore, combined treatment with PT and ATST markedly downregulated the expression of cyclin B and upregulated the expression of phospho-cdc2 and Myt1, which suggested that the activation of cdc2 was downregulated. This combined treatment strategy enhanced the anti-cancer activity of ATST at a relatively low dosage and suggested a possible method of preventing colorectal cancer in patients with hyperlipidemia.

Introduction

Colorectal cancer is one of the most common malignancies worldwide; imbalanced dietary patterns, such as the large consumption of red and processed meat, have been identified as a contributor to the increasing incidence of colorectal cancer (1,2). This dietary habit causes increased levels of cholesterol and low-density lipoproteins (LDL) in the blood, which can eventually develop into hyperlipidemia. According to recent data, hyperlipidemia is one factor contributing to the increased incidence of colorectal cancer (3,4) as high levels of cholesterol and LDL can enhance cancer growth and metastasis (5,6). Statin compounds are the most commonly used drugs to reduce the levels of cholesterol and LDL; therefore, using statins to prevent cancer is a recommended treatment for patients with hyperlipidemia. Nielsen, Nordestgaard and Bojesen (7) have reported that the use of statins may decrease cancer mortality in Danish population. Moreover, numerous clinical studies revealed an association between statin consumption and cancer risk; however, the data regarding colorectal cancer were inconsistent (8,9). Consequently, it is worth evaluating the anti-cancer effect of statins for the purpose of applying them in colorectal cancer prevention. Atorvastatin (ATST) is one of the main statins in clinical circulation. Prior studies have indicated the synergistic antitumor effects of ATST when combined with non-steroidal anti-inflammatory drugs (10,11), γ -tocotrienol (12) or green tea polyphenols (13). Therefore, exploring a combinational strategy between statins and other dietary components may be an effective way to prevent colorectal cancer in patients with hyperlipidemia.

Previous studies revealed that apples may reduce the risk of cancer, and that polyphenol and flavonoid compounds can also contribute to this chemopreventive effect (14-17). Phloretin (PT) is one of the most abundant phenolic phytochemicals in apples and apple products. Numerous studies have reported on the antitumor activities of PT, including its ability to suppress cell growth and induce apoptosis in human hepatoma cells, HL-60 human leukemia cells, B16 mouse 4A5 melanoma cells and HT29 human colon cancer cells (18-21). Both *in vitro* and *in vivo* studies have revealed that PT could potentiate the antitumor effects of paclitaxel via the induction of cell apoptosis (22). Another study showed that cytochalasin B could enhance the PT-induced apoptosis of HepG2 cells (21).

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Although, according to these reports, the combination of PT with other compounds may enhance its antitumor effect, little evidence is currently available to support a synergistic effect between PT and statins.

In this study, the potential synergistic inhibitory effect between PT and ATST was evaluated in human colon cancer cells. The synergistic mechanisms involving the cell cycle and apoptosis were also investigated. The results of the present study have provided a potential novel chemoprevention strategy for the hyperlipidemia population, specifically via the combination of dietary functional components and statin compounds.

Materials and methods

Cell lines and reagents. The human colon cancer cells SW620 and HCT116 were purchased from the Institute of Basic Medical Cell Center, Chinese Academy of Medical Sciences. ATST and PT were purchased from the National Institutes for Food and Drug Control (Beijing, China). MTT, propidium iodide (PI) and RNase were purchased from Sigma-Aldrich (Merck KGaA, Darmstadt, Germany). The Annexin V conjugate was purchased from Invitrogen (Thermo Fisher Scientific, Inc., Waltham, MA, USA). Lysis buffer and a BCA assay kit were purchased from Beyotime Co., (Haimen, China). Antibodies for poly-ADP-ribose polymerase (PARP), cleaved-PARP, caspase-3, cyclin B1, phospho-cdc2 (Tyr15) and Myt1 were purchased from Cell Signaling Technology, Inc., (Danvers, MA, USA).

Cell viability assay. Human SW620 and HCT116 colon cancer cells were seeded into 96-well plates (2,000 cells/well). After 24 h, the cells in each well were treated with a series of concentrations of PT, ATST or a combination (ratio of 10:1, PT and ATST, respectively). After 24 and 48 h, cell viability was determined using an MTT assay.

Analysis of synergy. The synergy analysis was conducted according to the median-effect equation (23). It was assumed that the dose-response model follows the median-effect equation, $f_a/f_u = (D/D_m)^m$; the dose-effect curve, $\log(f_a/f_u) = m \log D - m \log D_m$, is the logarithmic form of the median-effect equation, which is a linear regression model with the independent variable $\log D$ and the dependent variable $\log(f_a/f_u)$. D is the dose; D_m is the dose required for a 50% effect (IC_{50}); f_a is the fraction affected by D , which is the same as the ratio of surviving cells; f_u is the unaffected fraction, which is the same as the ratio of non-surviving cells, m is the slope. This equation is applied for calculating the effective doses of agent 1 and agent 2, and of a fixed ratio of combination agents, using data from an MTT assay. Suppose that the combination (D_1, D_2) exert the same effect x as agent 1 alone at dose level D_{x1} , and agent 2 alone at dose D_{x2} . D_{x1} and D_{x2} were calculated from the dose-effect curve. The interaction index $\alpha = D_1/D_{x1} + D_2/D_{x2} + \alpha + I_1 + I_2 / (D_{x1} \cdot I_{x2})$. If the agents are mutually exclusive, $\alpha = 0$; if the agents are mutually nonexclusive, $\alpha = 1$. The interaction index was used to determine whether the combination dose (D_1, D_2) was additive, synergistic or antagonistic, depending on an interaction index of 1, <1 or >1, respectively.

Measurement of cell apoptosis. Cell apoptosis was assessed using flow cytometry combined with an Annexin V/PI double-staining assay. After being treated with PT (100 μ M), ATST (10 μ M) or a combination of the two (100 μ M + 10 μ M, respectively) for 48 h, cells were harvested, washed with ice-cold PBS and resuspended in 100 μ l Annexin-binding buffer (10 mM HEPES, 140 mM NaCl, and 2.5 mM $CaCl_2$, pH 7.4), which contained 5 μ l Annexin V conjugate and 0.1 μ g PI. After incubation at room temperature for 15 min, the cell suspension was gently mixed with 400 μ l Annexin-binding buffer and analyzed by FACSCalibur flow cytometry (BD Biosciences, Franklin Lakes, NJ, USA) at 488 nm.

Detection of the cell cycle. Flow cytometry was performed to analyze the cell cycle distribution. After being treated with PT (100 μ M), ATST (10 μ M) or a combination of the two (100 μ M + 10 μ M, respectively) for 48 h, cells were harvested and washed with ice-cold PBS, then resuspended in 70% ethanol and stored at 4°C for 24 h. After 24 h, cells were pelleted by centrifugation, incubated with RNase (50 μ M/ml in PBS) and stained with PI (1 mg/ml in PBS) in the dark at 37°C for 30 min. Cells were then evaluated at a wavelength of 550 nm using a FACSCalibur flow cytometer (BD Biosciences), and the data were analyzed with Summit v5.0 software.

Western blot analysis. After being exposed to PT (100 μ M), ATST (10 μ M) or a combination of the two (100 μ M + 10 μ M, respectively) for 48 h, the cells were harvested into tubes. The cell pellet from each tube was incubated on ice for 30 min with 300 μ l lysis buffer containing phenylmethanesulfonyl fluoride (PMSF, 1 mM) and cocktails (1:10). The cell pellets were resuspended and centrifuged at 12,000 rpm for 20 min to collect the supernatants. Proteins were subjected to quantification using a BCA assay kit and then resolved via SDS-PAGE. After electrophoresis, the proteins were transferred to a polyvinylidene fluoride membrane, which was then blocked. The membranes were incubated with different primary antibodies at the concentrations recommended by the manufacturer at 4°C overnight. Subsequently, the membranes were incubated with secondary antibodies at room temperature for 1 h, prior to visualization using an enhanced chemiluminescence reagent. Antibodies for PARP, cleaved-PARP, caspase-3, cyclin B1, Tyr15 and Myt1 were purchased from Cell Signaling Technologies, Inc.

Statistical analysis. All data were obtained from at least three independent experiments and are presented as the mean \pm standard deviation. A Student's t-test was used to assess differences between two groups. One-way analysis of variance and the Dunnett's post hoc test were used to compare differences among multiple groups. $P < 0.05$ was considered to indicate a statistically significant difference.

Results

A synergistic anti-proliferative effect was observed in SW620 and HCT116 cells treated with a combination of PT and ATST. The growth inhibitory effect of PT, ATST and their combination was evaluated in SW620 and HCT116 cells at 24 and 48 h. The tested concentrations of PT were 50, 75, 100, 125, 150 and 200 μ M; the tested concentrations of ATST

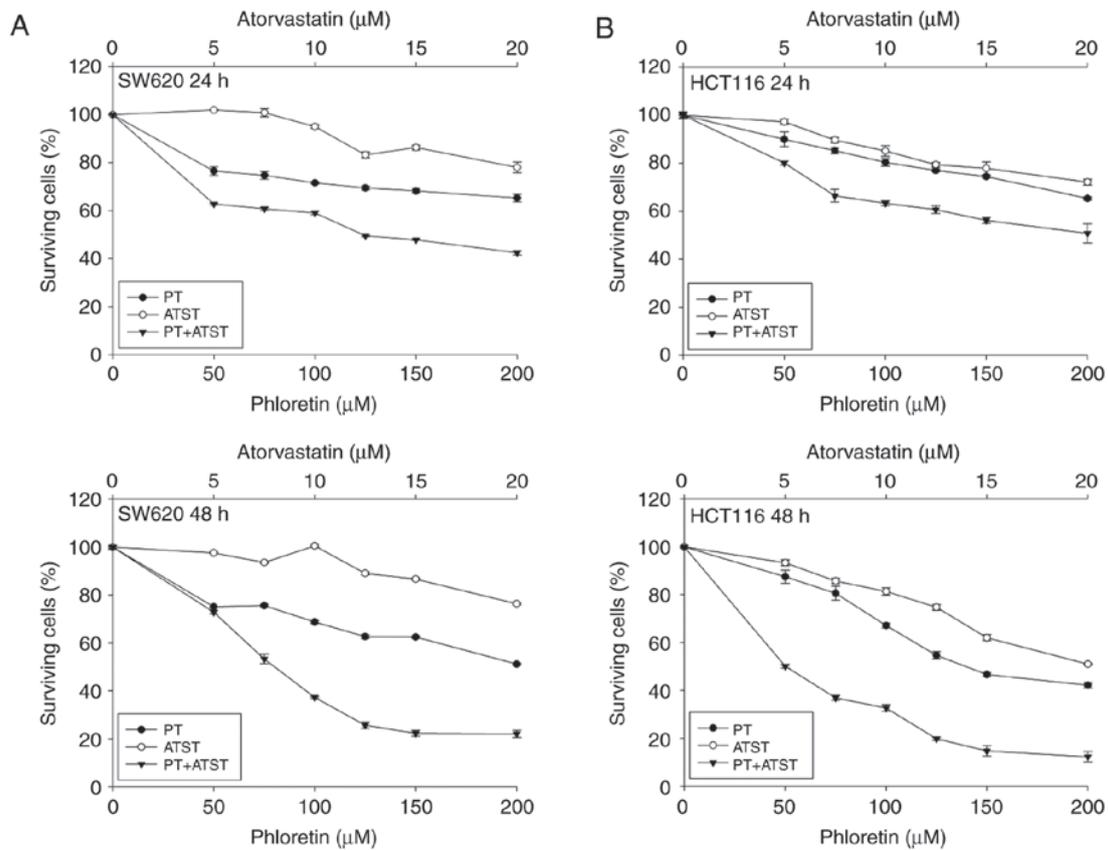


Figure 1. The viability of SW620 (A) and HCT116 cells (B) after treatment with PT, ATST and their combinations. Cells were treated with a series of dosages of PT (0, 50, 100, 150 and 200 μM), ATST (0, 5, 10, 15 and 20 μM), or a combination at a fixed ratio of 10:1 for 24 and 48 h, and then cell viability was measured using an MTT assay. Data are shown as the mean \pm SD ($n=5$), and the surviving cell percentage of the respective controls were set as 100%.

were 5, 7.5, 10, 12.5, 15 and 20 μM ; the combination ratio of PT to ATST was 10:1. After exposure to these treatments for 24 and 48 h, cell survival was measured using an MTT assay. The results (Fig. 1) showed that PT and ATST exhibited time- and dose-dependent growth inhibitory effects on the two cell lines. The IC_{50} values for single PT and ATST treatments in HCT116 cells were 137.48 ± 2.14 and 19.52 ± 1.29 μM , respectively. However, for SW620 cells, the IC_{50} value of PT was 191.70 ± 2.28 μM , and ATST did not show a significant inhibitory effect on cell growth.

Compared with individual PT and ATST treatments, the combination markedly decreased cell viability. The cell viability rates at 24 h and 48 h following treatment with PT (100 μM) combined with ATST (10 μM) were $59.08 \pm 0.73\%$ and $37.27 \pm 0.39\%$, respectively, for SW620 cells, and $63.35 \pm 1.08\%$ and $32.65 \pm 1.34\%$ for HCT116 cells, respectively. Furthermore, to determine whether the enhanced inhibitory effect observed with combined PT/ATST was additive or synergistic, the combination indexes were computed using the aforementioned Chou and Talalay method.

The median-effect plots of these two cells are shown in Fig. 2A and B, and indicate that the linear regression model fit better with the dose-dependent manner of PT, ATST and their combinations. According to the median-effect equation, the effect doses of PT, ATST and their combination at a dosage ratio of 10:1 were computed, and then used to compute the interaction indexes. As presented in Fig. 2C and D, the interaction indexes of each PT and ATST concentration pair

were <1.0 ; notably, a proportion was <0.5 . Consequently, it was determined the combination of PT and ATST generated a strong synergistic inhibitory effect on SW620 and HCT116 cell growth. As the results showed that the inhibitory effect of combination treatment at 48 h were significantly stronger than the ones at 24 h, we therefore selected the time point of 48 h to carry out the following experiments.

Combined PT and ATST treatment induces apoptosis. The results revealed a strong synergistic anti-proliferative effect exerted by the combined PT and ATST treatment of SW620 and HCT116 cells. This observation prompted us to determine whether the decreased cell survival was related to apoptosis. An Annexin V/PI double staining assay was used to determine the proportion of apoptotic cells. The results are presented in Fig. 3A and B. In both SW620 and HCT116 cells, the percentage of apoptotic cells in the group treated with combined PT and ATST was significantly higher than for those cells treated with each agent alone. Furthermore, the expression levels of PARP and cleaved-PARP were determined (Fig. 3C). The PARP fragments were significant in SW620 and HCT116 cells following the combination treatments. By contrast, there were no significant levels of cleaved-PARP detected in either cell line following treatment with the single agents. These observations, together with the results of the MTT assay, indicated that combined treatment with PT and ATST could induce colon cancer cell apoptosis.

Combined PT and ATST treatment causes cell cycle arrest at the S phase. The results warranted us to hypothesize that the

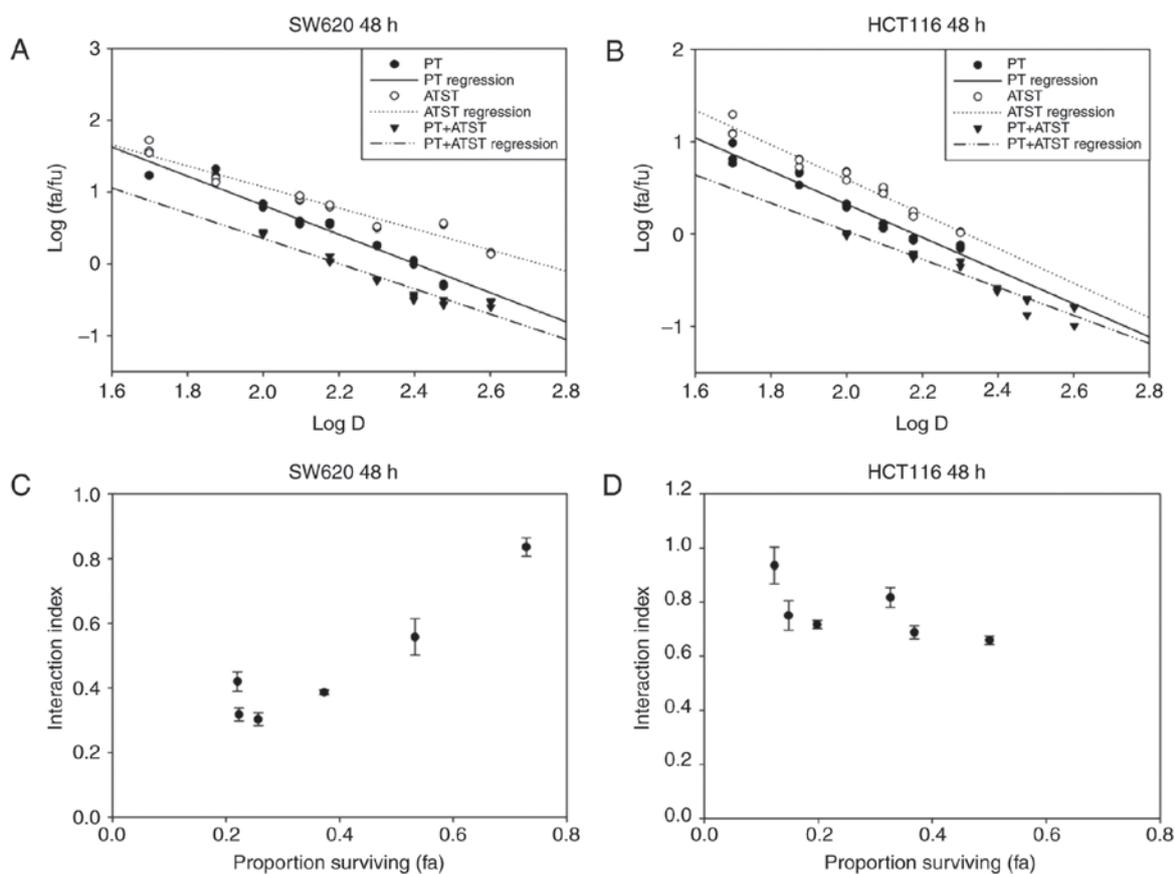


Figure 2. Median-effect and interaction index plots of PT, ATST or a combination in SW620 (A and C) and HCT116 (B and D) cells. Cells were treated with a series of dosages of PT (0, 50, 100, 150 and 200 μ M), ATST (0, 5, 10, 15 and 20 μ M) or a combination at a fixed ratio of 10:1 for 48 h, and then cell viability was measured using an MTT assay. Median-effect plots (A, B) and interaction index plots (C and D) were computed with the median-effect equation. Synergy was defined as an interaction index <1.0 . The data of interaction plots are shown as the mean \pm SD ($n=5$).

synergistic effect of PT and ATST on apoptosis may involve cell cycle arrest. Flow cytometry was performed to analyze the cell cycle distribution for both SW620 and HCT116 cells following treatment with PT, ATST or a combination for 48 h. The results (Fig. 4) indicated that individual PT or ATST treatment did not significantly change the cell cycle distribution for the two cell lines. By contrast, combined PT and ATST could alter the distribution of cell cycle. For SW620 cells, a significant increase in the S phase cell proportion was observed following combined treatment with PT and ATST, which was correspondingly accompanied by a decrease in the G₂/M phase cell proportion. For HCT116 cells, an increase in the S phase proportion was also observed with combined treatment; however, this increase was slighter when compared with the SW620 cells, and no decrease in the G₂/M phase proportion was noted. Therefore, the results indicated that combined PT and ATST treatment could arrest SW620 and HCT116 cells in the S phase.

Combined PT and ATST treatment inhibits cdc2 activation. To further explore the mechanisms underlying the cell cycle arrest, the activity of proteins that perform an important role during progression at the S phase and the G₂/M checkpoint were examined (Fig. 5). In both SW620 and HCT116 cells, the expression of cyclin B1 was not significantly different between the control and PT-treated groups; for ATST-treated cells, the expression of cyclin B1 was decreased by $\sim 50\%$. By contrast,

combined treatments of PT and ATST markedly suppressed the cyclin B expression in all cells (Fig. 5A). The phosphorylation of cdc2 at Tyr15 was also analyzed, as dephosphorylation at this locus results in the activation of cdc2. The results showed that the activity of p-cdc2 were markedly upregulated following combined treatments (Fig. 5B). Consistently, the activity of Myt1, which is responsible for the phosphorylation of cdc2 at Tyr15, showed a similar pattern (Fig. 5C). These results indicated that PT and ATST synergistically inhibit the activation of cdc2 in both SW620 and HCT116 colon cancer cells.

Discussion

Since increased 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) levels have been observed in colon cell lines, previous studies have evaluated the anti-cancer activity of statins. Statins are competitive small-molecule inhibitors of HMG-CoA reductase, and could prevent the transformation of HMG-CoA to mevalonate (24). Although *in vitro* data have suggested that ATST could suppress HCT116 cell growth and induce apoptosis, the effective doses of ATST in these experiments were relatively higher (50 and 100 μ M, respectively) (12,25). In our experimental design, the maximum dose of ATST was only 25 μ M. As expected, the results showed that ATST exhibited little effect on HCT116 cell growth and

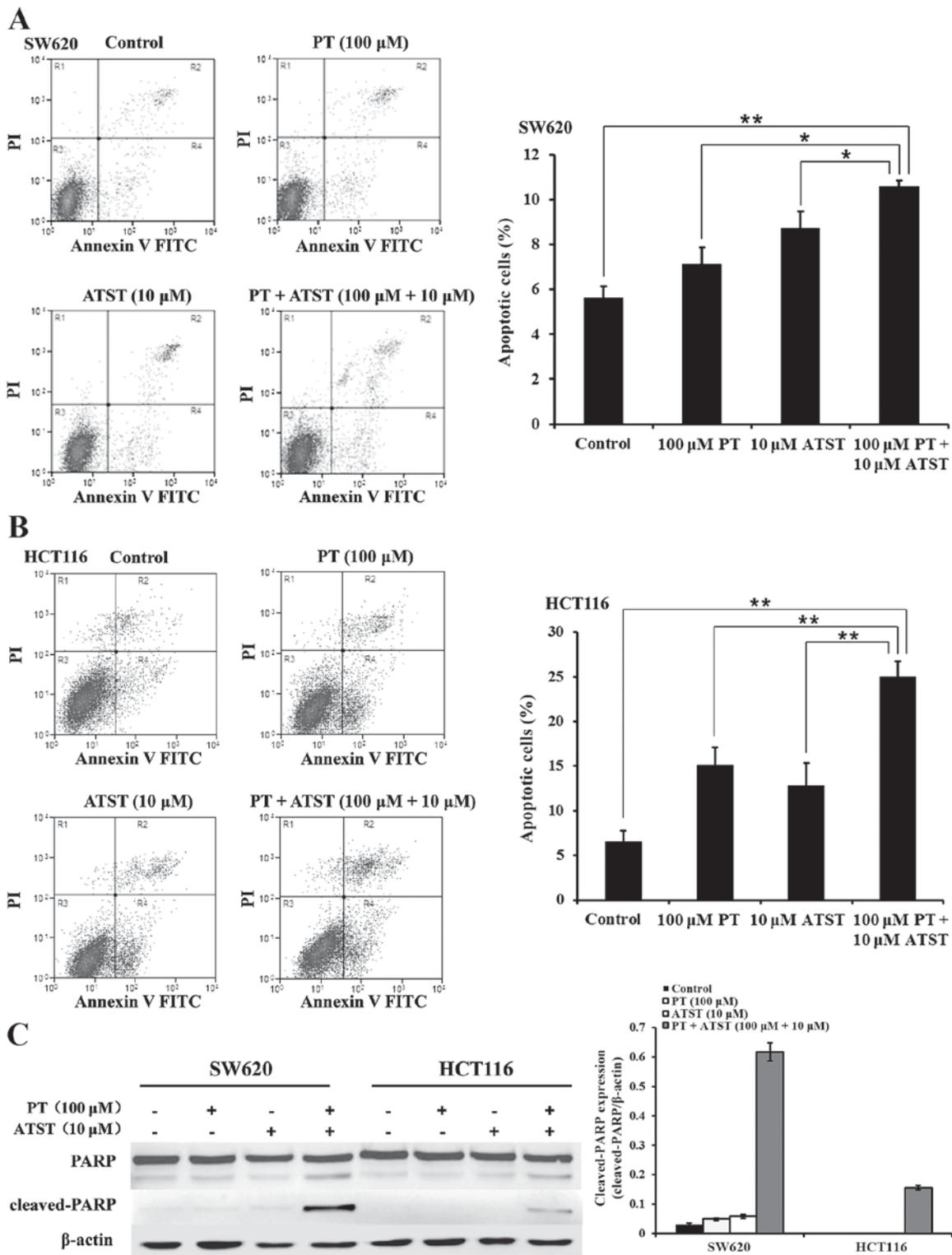


Figure 3. Effects of PT, ATST or a combination on apoptosis (A, SW620; B, HCT116) and cleaved-PARP levels (C). Cells were treated with PT (100 μM) ATST (10 μM) or a combination of the two for 48 h. Apoptosis was detected by flow cytometry after Annexin V/PI co-staining, and protein levels were analyzed by western blotting. Data are shown as the mean ± SD (n=3). *P<0.05 and **P<0.01 indicated statistical significance, compared with controls, as analyzed by the Student's t-test.

apoptosis at this relatively low dosage. Previous evidence has suggested that the safe and tolerated therapeutic dosage range of ATST is 10-80 mg/day (26). This dosage range is lower for ATST when administered to exhibit a protective effect against colorectal cancer. Our results suggested a combination strategy for the prevention of colorectal cancer based on the synergy

between ATST and PT. Through this combination, the growth inhibition effect of ATST would be substantially increased at a relatively low dosage.

Similar to ATST, this enhancement effect is also applicable to PT. The *in vivo* activities of phytochemicals are usually restricted due to poor bioavailability. Although previous studies

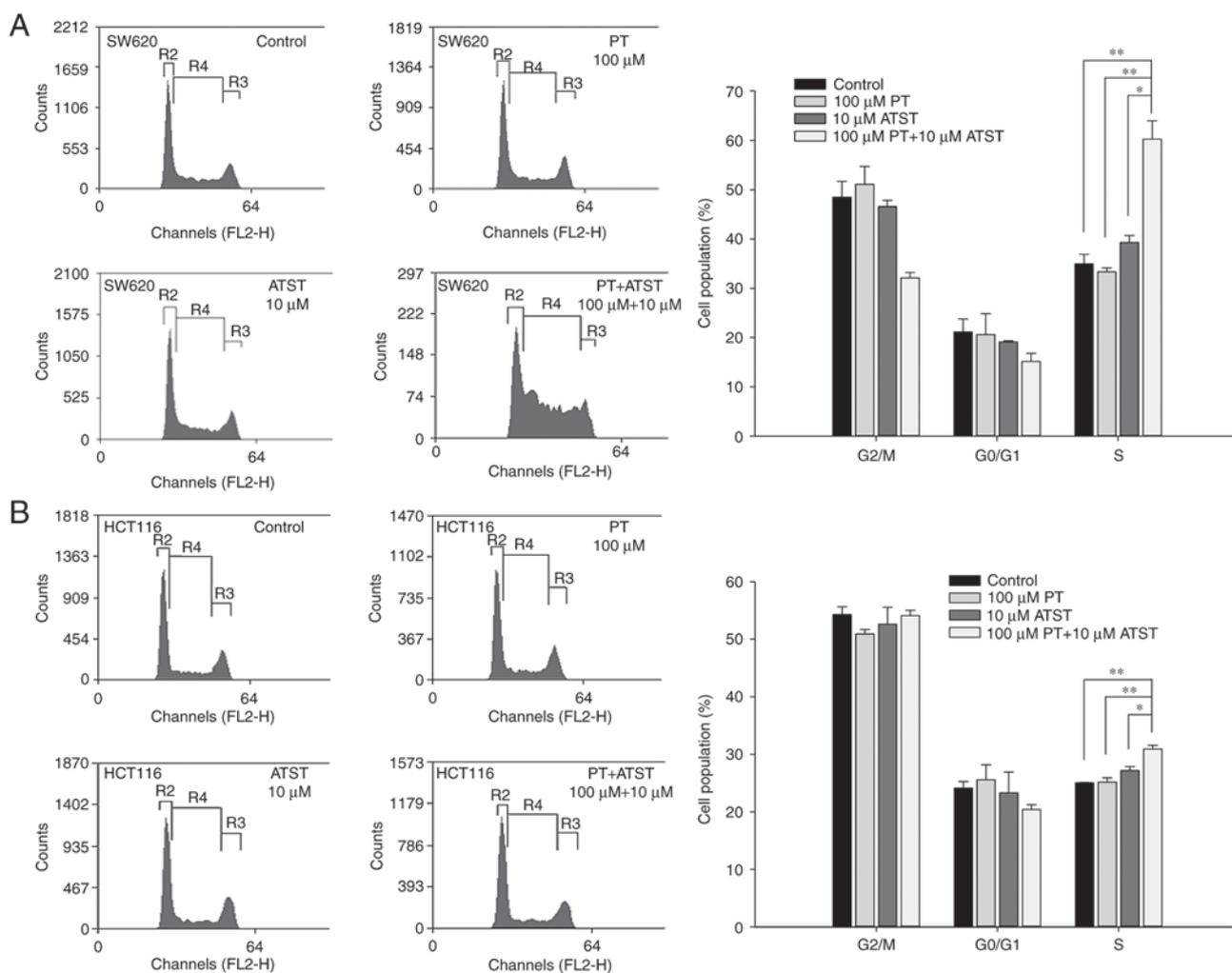


Figure 4. Effects of PT, ATST or a combination on the cell cycle in SW620 (A) and HCT116 (B) cells. Cells were treated with PT (100 μ M), ATST (10 μ M) or a combination for 48 h, and the cell distribution was detected by flow cytometry. The quantified data of the cell population in the S phase are shown as the percentage compared with the total cell counts. Data are shown as the mean \pm SD ($n=3$). * $P<0.05$ and ** $P<0.01$ indicated statistical significance, compared with controls, as analyzed by the Student's *t*-test.

have reported the anti-proliferative effects of PT on HL-60, HT29 and HepG2 cells, the effective dosages of PT were all $>100 \mu\text{M}$ (18,20,22). Concordantly, our results showed that, in both SW620 and HCT116 cells, PT could not efficiently inhibit cell growth unless the dose was higher than $100 \mu\text{M}$. Although previous studies have reported that cytochalasin B could enhance the PT-induced apoptosis of HepG2 cells (21), and that PT can potentiate the anticancer activity of paclitaxel (22), no prior research has focused on synergy involving PT. The results of the present study demonstrated that the antitumor efficacy of ATST could be enhanced at a relatively low dosage through the synergistic action with PT, which suggested the potential interaction of statins with other compounds in the food matrix. This interaction affects the efficacy of statins, and may explain the controversial results obtained in prior studies regarding the associations between statin use and the risk of colon cancer-associated mortality (27,28). As the dietary composition is different for each individual, this can result in varying statin efficacy. Conversely, different statins have different antitumor effects. In six colorectal cancer cell lines, including DLD1, HT29, SW620, HCT116, LoVo and colo320, simvastatin and fluvastatin showed strong growth suppressive

effects. Atorvastatin demonstrated a relatively weak growth suppressive effect, whereas no growth suppressive effect was observed with pravastatin (29). This may be another reason for the paradoxical results regarding the antitumor effects of statins.

A close relationship is believed to exist between the cell cycle and apoptosis in cancer cells (30). Numerous studies have reported PT-induced apoptosis; however, few of these results involved the cell cycle, especially in colon cancer cells. Some data supported that the presence of HMG-CoA reductase inhibitors may influence the cell cycle distribution of cancer cells. Known as the typical statin family compound, ATST has been shown to induce colon cancer cell cycle arrest in the G_0/G_1 phase when combined with other compounds (11,12). Therefore, we decided to determine whether treatments with PT and ATST could regulate the cell cycle distribution of SW620 and HCT116 cells. The results partly confirmed our hypothesis, in that the cell cycle was arrested, but also demonstrated that the cells were arrested at the G_2/M checkpoint and accompanied by an increased cell population in the S phase, rather than arrested in the G_0/G_1 phase. Specifically, *cdc2* kinase activation is the pivotal regulator mechanism

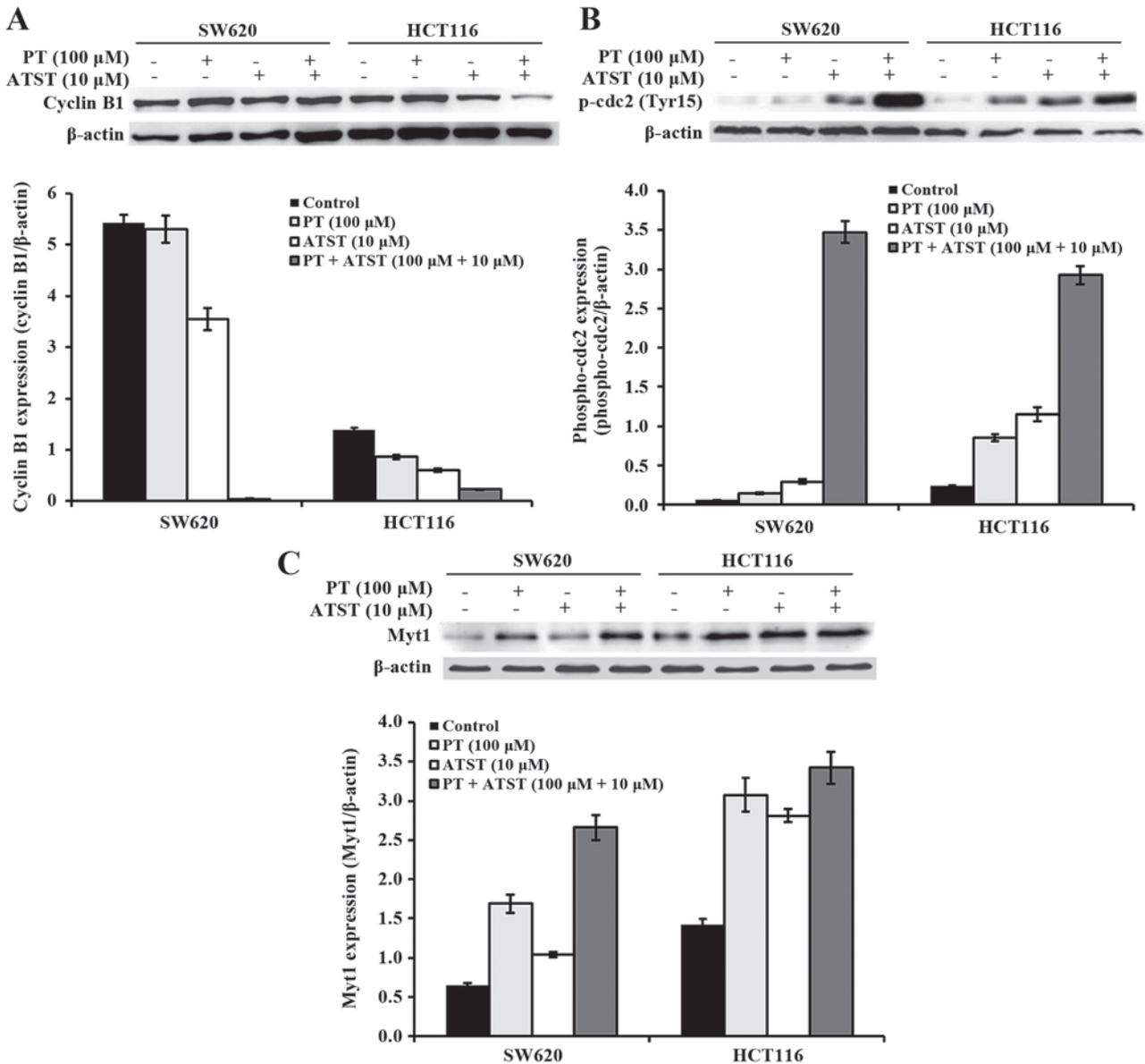


Figure 5. Effects of PT, ATST or a combination on the levels of cyclin B1 (A), p-cdc2 (B) and Myt1 (C) in SW620 and HCT116 cells. Cells were treated with PT (100 μ M), ATST (10 μ M) or a combination of the two for 48 h, and the protein levels were analyzed by western blotting. Combined treatments suppressed the levels of cyclin B1, and increased the levels of p-cdc2 and Myt1. The quantified levels of cyclin B1, p-cdc2 and Myt1 are shown as the mean \pm SD ($n=3$).

responsible for the G₂/M checkpoint. Activation of cdc2 is controlled via two steps: One is cyclin binding; the other is the dephosphorylation of cdc2 at Tyr15, which is the core regulatory step (31).

In the present study, we observed that PT and ATST synergistically downregulated the expression of cyclin B1, which indicated that formation of the cyclin B-cdc2 complex might be inhibited. In addition, our data showed that the level of cdc2 phosphorylation at Tyr15 was markedly increased by combined treatment with PT and ATST. These results demonstrated that the cdc2 kinase was inactivated, possibly due to a failure to bind to cyclin B and the increased levels of p-cdc2 at Tyr15. Myt1 protein kinase is regarded as a negative modulator of cdc2, and carries out the phosphorylation of cdc2 at Tyr15 (32). The p21 gene is an inhibitor of cyclin; hyper-phosphorylation of p21 activates cdc2 kinase in the G₂/M transition (33,34). Previous studies have shown

that ATST can increase p21 levels in A549 cells (35) and the pancreatic cancer (36). However, Buranrat *et al* reported opposing results, stating that ATST reduced p21 expression in KKU-100 cells and did not alter p21 expression in KKU-M214 cells (37). In terms of the synergistic effect, p21 levels were increased in HT29 and HCT116 cells following treatment with combined ATST and celecoxib (11), and with combined ATST and γ -tocotrienol (12). Therefore, the p21 gene may be the potential regulatory target underlying the G₂/M phase arrest following the synergistic action of ATST and PT; more in depth future investigations are warranted.

In summary, the present study demonstrated that PT and ATST produce a powerful synergistic interaction in suppressing colon cancer cell growth. This process was accomplished via the synergistic induction of apoptosis and the arrest of the cell cycle at the G₂/M checkpoint, which resulted from downregulated cdc2 activation following combined treatment.

Acknowledgements

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